The biomimetic synthesis of balsaminone A and ellagic acid via oxidative dimerization

Sharna-kay Daley and Nadale Downer-Riley*

Full Research Paper

Address:

Department of Chemistry, The University of the West Indies, Mona, Jamaica

Email:

Nadale Downer-Riley* - nadale.downer02@uwimona.edu.jm

* Corresponding author

Keywords:

balsaminone A; biomimetic synthesis; ellagic acid; oxidative dimerization

Beilstein J. Org. Chem. 2020, 16, 2026-2031.

doi:10.3762/bjoc.16.169

Received: 27 April 2020 Accepted: 06 August 2020 Published: 18 August 2020

Associate Editor: J. Aubé

© 2020 Daley and Downer-Riley; licensee Beilstein-Institut.

License and terms: see end of document.

Abstract

The application of oxidative dimerization for the biomimetic synthesis of balsaminone A and ellagic acid is described. Balsaminone A is synthesized via the oxidative dimerization of 1,2,4-trimethoxynaphthalene under anhydrous conditions using CAN, PIDA in BF₃·OEt₂ or PIFA in BF₃·OEt₂ in 7–8% yields over 3 steps. Ellagic acid is synthesized from its biosynthetic precursor gallic acid, in 83% yield over 2 steps.

Introduction

Over the last century, the formation of an aryl to aryl bond has garnered considerable synthetic attention due to the applications of biaryls as pharmaceutical agents, as well as chiral auxiliaries in asymmetric synthesis [1-3]. Methods such as the Ullman coupling, Suzuki–Miyaura coupling and Stille coupling have dominated the field for the synthesis of biaryls [1,4]. Throughout the years, the exploration of oxidative dimerization reactions of electron-rich aromatic compounds, such as thiophenes, anilines and alkoxyarenes, in an attempt to establish high-yielding and selective oxidative coupling reactions, has afforded new and greener synthetic protocols for biaryls [5-7]. Several oxidants, such as the salts of Ag(I&II) [8], Ti(III&IV) [9], Mn(III) [10], Ce(IV) [11], Sn(IV) [12] and Fe(III) [13], as well as the hypervalent iodine reagents phenyliodine diacetate

(PIDA) and phenyliodine bis(trifluoroacetate) (PIFA), have been utilized for oxidative dimerization reactions [14,15]. The use of these one-electron oxidants, as well as non-metallic reagents, plays an important role in accessing symmetrical and asymmetrical biaryls and polyaryls [1]. There are many important applications of oxidative dimerization reactions, one of which is the direct synthesis of natural product scaffolds [1,7,16-18]. Examples of this application include the biomimetic synthesis of the bioactive natural products bismurrayaquinone A (1) [16], parvistemin A (2) [17] and violetquinone (3) [18] (Figure 1).

Similar to those natural products, the biomimetic synthesis of balsaminone A (4) and ellagic acid (5) can be attained using ox-

Open Access

idative dimerization reactions, based on their proposed biosynthesis (Scheme 1 and Scheme 2) [19,20]. These pathways, which start with shikimic acid, feature the dimerization of monomeric phenolic precursors by a laccase enzyme, a single-electron enzyme complex within macro-organisms which facilitates oxidative dimerization through phenolic coupling [19]. In

the case of balsaminone A (4), lawsone (6) is methylated to

ether 7 which undergoes reduction to the dihydroxynaphthalene

8. This is then dimerized by the copper-rich enzyme to quinone

arenol 9, which, following intramolecular cyclization, affords balsaminone A (4, Scheme 1). Similarly, the biosynthetic precursor of ellagic acid (5), gallic acid (10), undergoes dimerization to the dimeric polyphenolic acid 11, followed by intramolecular cyclization to yield ellagic acid (5, Scheme 2) [20].

Because of the bioactivity of both natural products, there have been previous investigations of the synthesis of balsaminone A (4) [21,22] and ellagic acid (5) [23]. In our previous synthesis

of balsaminone A (4) [22] (Scheme 3), silver oxide was used as the oxidative dimerizing agent for the formation of the binaphthoquinone intermediate 13. This was followed by photolytic cyclisation and *ortho*-formylation to give carbaldehyde 14. Conversion to balsaminone A (4) could then be achieved in 57% yield over 5 steps. The synthesis of ellagic acid (5) on the other hand featured an *o*-chloranil-mediated dimerization of methyl gallate (15), which yielded the natural product in 48% yield over 5 steps, as reported by Tsuboi et al. (Scheme 3) [23].

Results and Discussion Biomimetic synthesis of balsaminone A

With the initial focus being the biomimetic synthesis of balsaminone A (4), well-established methods for the dimerization of phenolic compounds were explored [8,9]. As such, a

methanolic solution of 2-methoxy-1,4-dihydroxynaphthalene (8), obtained from the reduction of lawsone (6), was subjected to the oxidants activated carbon (Act-C), potassium ferricyanide (K₃[Fe(CN)₆]), *p*-benzoquinone and stannic chloride (SnCl₄). As shown in Scheme 4, with the exception of SnCl₄, all oxidants resulted in the re-oxidation of the hydroxylated substrate to naphthoquinone 7. SnCl₄, however, while not successful for the synthesis of the desired biomimetic precursor 9, afforded the binaphthylquinone 16 in 22% yield.

Given the initial unsuccessful attempt at the synthesis of the biomimetic precursor of balsaminone A (4), it was evident that alternative oxidative dimerization conditions needed to be explored. The oxidants cerium(IV) ammonium nitrate (CAN), ferric chloride hexahydrate (FeCl₃·6H₂O), vanadium pentoxide

(V₂O₅), PIFA, and PIDA, in addition to SnCl₄, were considered. Also investigated were 2-iodoxybenzoic acid (IBX) because of its implication in single-electron oxidation [24], and chromium trioxide (CrO₃), which, based on its Cr(VI) oxidation state, should be able to facilitate single-electron transfer in the presence of electron-rich arenes.

The dimerization of 1,2,4-trimethoxynaphthalene (17) in the presence of the metal oxidants CAN, V2O5, and CrO3, afforded binaphthyl 16 in 29, 34, and 19% yields, respectively (Table 1). As anticipated, the reaction of naphthalene 17 with CAN under aqueous conditions resulted in preferential oxidative demethylation to give quinone 7, as opposed to oxidative dimerization. The elimination of water from the reaction posed a few challenges since the solubility of CAN in organic solvents determines the likelihood of nitration or oxidative dimerization [1]. It was found that dry acetonitrile was more conducive to nitration, whereas methanol afforded the desired biaryl in 55% yield. In contrast to CAN, both V2O5 and CrO3 required aqueous conditions to prevent complexation of the reagent and the starting material. Of the Lewis acids used, stannic chloride proved to be the most effective oxidant for dimerization (Table 1). However, the hypervalent iodine reagents PIFA and PIDA gave better results overall, affording dimer 18 in 63% and 59% yields, respectively (Table 1).

Following the successful oxidative dimerization of 1,2,4-trimethoxynaphthalene 17 to biaryl 18, the biaryl was con-

verted quantitatively to binaphthyl 16 in the presence of an aqueous solution of CAN at 0 °C. Alternatively, the binaphthylquinone 16 was synthesized through the oxidation of lawsone (6), in the presence of sodium persulfate, followed by methylation [25]. The cyclization of binaphthyl 16 was then attempted taking into consideration the photolytic cyclization of binaphthyls to form pentacyclic furan derivatives [8,18,22]. However, the photolytic cyclization of binaphthyl 16 using a 100 W bulb was attempted without success, due to degradation of the starting material. This led to a different approach employing a reductive, base-mediated cyclization using alkaline aqueous sodium dithionite. Interestingly, the typical conditions for reduction using dithionite proved too harsh for the substrate, and like the photolysis of the binaphthyl 16, degradation occurred. However, the use of triethylamine instead of aqueous sodium hydroxide resulted in the isolation of balsaminone A (4) in 13% yield, an overall 7-8% yield over 3 steps, completing the biomimetic synthesis of the natural product as shown in Scheme 5.

Synthesis of ellagic acid

With the synthesis of balsaminone A (4) accomplished, the direct biomimetic synthesis of ellagic acid (5) was targeted. While the synthesis of ellagic acid (5) has been achieved in 48% yield over four steps by Alam et al. [23], starting from methyl gallate (15), it was anticipated that a more concise and efficient synthesis could be attained. Methyl gallate (15), which may be obtained commercially or from the

Table 1: Reagents and products in the oxidative dimerization of 1,2,4-trimethoxynaphthalene (17).			
OMe OMe OMe	O OMe	OMe + OMe - +	OMe
17	16	18	7
	16 (%)	18 (%)	7 (%)
CAN ^a	29 ^b	55 ^c	67 ^b
V ₂ O ₅ ^{a,b} CrO ₃ ^{a,b}	34	=	-
CrO ₃ ^{a,b}	19	=	62
FeCl ₃ ·SiO ₂ ^{a,c}	15	43	-
FeCl ₃ ·6H ₂ O ^{a,c}	-	8	-
SnCl ₄ ^d	22	48 ^d	_
PIDA, BF ₃ ·OEt ₂ ^{e,f}	21	59	_
PIFA, BF ₃ ·OEt ₂ e,f	12	63	-
IBX, BF ₃ ·OEt ₂ ^{e,f}	13	_	45

^aReaction of naphthalene **17** (0.5 mmol) with oxidant (1.65 mmol) at rt. ^bAddition of oxidant in H₂O (1 mL, dropwise) to substrate in CH₃CN or CH₃OH (1 mL). ^cAddition of oxidant (neat) in thirds, to substrate dissolved in CH₃CN. ^dReaction was carried out in CH₂Cl₂ at 100 °C in a sealed tube. ^eReaction of naphthalene **17** (0.5 mmol) with oxidant (0.65 mmol). ^fReaction carried out under N₂.

Scheme 5: Biomimetic synthesis of balsaminone A (4).

methylation of gallic acid (10) [23], was subjected to the oxidants CAN, PIDA, FeCl₃·6H₂O, and FeCl₃·SiO₂. Treatment of the polyphenol with PIDA (1.5 equiv) in boron trifluoride etherate (1.5 mL) at room temperature proved to be the most successful dimerization protocol, as the dimeric precursor 19 was formed in 80% yield after 16 hours. The biaryl 19 was converted quantitatively to the natural product when heated at reflux in aqueous methanol (1:1) for 24 hours. However, allowing the PIFA reaction mixture to stir at room temperature for 24 hours resulted in complete cyclization, affording the natural product 5 in 83% yield directly from methyl gallate (15, Scheme 6), an improvement for the synthesis of the bioactive natural product.

Conclusion

The synthesis of balsaminone A (4) is reported in 7-8% yield over 3 steps from 1,2,4-trimethoxynaphthalene (17). Also, the one-step synthesis of ellagic acid (5) from methyl gallate (15) in 83% yield is described. These biosynthetically-driven syntheses represent the most efficient routes to these bioactive natural products to date.

Supporting Information

Supporting Information File 1

Experimental part.

[https://www.beilstein-journals.org/bjoc/content/ supplementary/1860-5397-16-169-S1.pdf]

Funding

The authors thank the Department of Chemistry, The University of the West Indies, Mona, Jamaica for support of this work.

ORCID® iDs

Nadale Downer-Riley - https://orcid.org/0000-0002-2850-1726

References

- Cepanec, I. The Synthesis of Biaryls; Elsevier Science: Oxford, U.K., 2004: pp 2–3.
- Smrcina, M.; Polakova, J.; Vyskocil, S.; Kocovsky, P. J. Org. Chem. 1993, 58, 4534–4538, doi:10.1021/io00069a010
- Chen, Y.; Yekta, S.; Yudin, A. K. Chem. Rev. 2003, 103, 3155–3212. doi:10.1021/cr020025b
- Sperotto, E.; van Klink, G. P. M.; van Koten, G.; de Vries, J. G. Dalton Trans. 2010, 39, 10338–10351. doi:10.1039/c0dt00674b
- Sarhan, A. A. O.; Bolm, C. Chem. Soc. Rev. 2009, 38, 2730–2744. doi:10.1039/b906026j
- Ashenhurst, J. A. Chem. Soc. Rev. 2010, 39, 540–548. doi:10.1039/b907809f
- Yang, Y.; Lan, J.; You, J. Chem. Rev. 2017, 117, 8787–8863. doi:10.1021/acs.chemrev.6b00567
- Takeya, T.; Kondo, H.; Otsuka, T.; Tomita, K.; Okamoto, I.; Tamura, O. Org. Lett. 2007, 9, 2807–2810. doi:10.1021/ol070951i
- Otsuka, T.; Okamoto, I.; Kotani, E.; Takeya, T. Tetrahedron Lett. 2004, 45, 2643–2647. doi:10.1016/j.tetlet.2004.01.133
- 10. Jasch, H.; Scheumann, J.; Heinrich, M. R. J. Org. Chem. **2012**, *77*, 10699–10706. doi:10.1021/jo301980j
- 11. Love, B. E.; Duffy, B. C.; Simmons, A. L. *Tetrahedron Lett.* **2014**, *55*, 1994–1997. doi:10.1016/j.tetlet.2014.02.017
- Takeya, T.; Doi, H.; Ogata, T.; Okamoto, I.; Kotani, E. Tetrahedron 2004, 60, 9049–9060. doi:10.1016/j.tet.2004.07.073
- Li, X.-L.; Huang, J.-H.; Yang, L.-M. Org. Lett. 2011, 13, 4950–4953. doi:10.1021/ol202058r
- Kar, A.; Mangu, N.; Kaiser, H. M.; Beller, M.; Tse, M. K.
 Chem. Commun. 2008, 386–388. doi:10.1039/b714928j
- 15. Guo, W.; Faggi, E.; Sebastián, R. M.; Vallribera, A.; Pleixats, R.; Shafir, A. *J. Org. Chem.* **2013**, *78*, 8169–8175. doi:10.1021/jo401001k
- 16. Bringmann, G.; Tasler, S. *Tetrahedron* **2001**, *57*, 331–343. doi:10.1016/s0040-4020(00)00940-6
- Smith, M. J.; Nawrat, C. C.; Moody, C. J. Org. Lett. 2011, 13, 3396–3398. doi:10.1021/ol201246e
- Ogata, T.; Okamoto, I.; Kotani, E.; Takeya, T. Tetrahedron 2004, 60, 3941–3948. doi:10.1016/j.tet.2004.03.038
- Manitto, P. Biosynthesis of Natural Products; Ellis Horwood Ltd.: Chichester, U.K., 1981; pp 98–100.
- Ishikura, N.; Hayashida, S.; Tazaki, K. Bot. Mag., Tokyo 1984, 97, 355–367. doi:10.1007/bf02488668
- Padwal, J.; Lewis, W.; Moody, C. J. J. Org. Chem. 2011, 76, 8082–8087. doi:10.1021/jo201395n
- Daley, S.-k. A.; Downer-Riley, N. K. Synlett 2019, 30, 325–328. doi:10.1055/s-0037-1611975
- Alam, A.; Takaguchi, Y.; Tsuboi, A.
 J. Fac. Environ. Sci. Technol., Okayama Univ. 2005, 10, 111–117.
- 24. Wirth, T. *Angew. Chem., Int. Ed.* **2001**, *40*, 2812–2814. doi:10.1002/1521-3773(20010803)40:15<2812::aid-anie2812>3.0.co;2-
- Qi, C.; Wang, W.; Reichl, K. D.; McNeely, J.; Porco, J. A., Jr. *Angew. Chem., Int. Ed.* 2018, 57, 2101–2104. doi:10.1002/anie.201711535

License and Terms

This is an Open Access article under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0). Please note that the reuse, redistribution and reproduction in particular requires that the authors and source are credited.

The license is subject to the *Beilstein Journal of Organic Chemistry* terms and conditions: (https://www.beilstein-journals.org/bjoc)

The definitive version of this article is the electronic one which can be found at: doi:10.3762/bjoc.16.169