## Accepted Manuscript

Toward the discovery of dual HCMV-VZV inhibitors: synthesis, structure activity relationship analysis, and cytotoxicity studies of long chained 2-uracil-3yl-N-(4-phenoxyphenyl)acetamides

Denis A. Babkov, Anastasia L. Khandazhinskaya, Alexander O. Chizhov, Graciela Andrei, Robert Snoeck, Katherine L. Seley-Radtke, Mikhail S. Novikov

PII:	S0968-0896(15)30058-4
DOI:	http://dx.doi.org/10.1016/j.bmc.2015.09.033
Reference:	BMC 12580
To appear in:	Bioorganic & Medicinal Chemistry
Received Date:	8 August 2015
Revised Date:	15 September 2015
Accepted Date:	19 September 2015



Please cite this article as: Babkov, D.A., Khandazhinskaya, A.L., Chizhov, A.O., Andrei, G., Snoeck, R., Seley-Radtke, K.L., Novikov, M.S., Toward the discovery of dual HCMV-VZV inhibitors: synthesis, structure activity relationship analysis, and cytotoxicity studies of long chained 2-uracil-3-yl-N-(4-phenoxyphenyl)acetamides, *Bioorganic & Medicinal Chemistry* (2015), doi: http://dx.doi.org/10.1016/j.bmc.2015.09.033

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

#### **Graphical Abstract**

CC

To create your abstract, type over the instructions in the template box below. Fonts or abstract dimensions should not be changed or altered.

Toward the discovery of dual HCMV-VZV inhibitors: synthesis, structure-activity relationship analysis, and cytotoxicity studies of long chained 2-uracil-3-yl-N-(4phenoxyphenyl)acetamides Leave this area blank for abstract info.

Denis A. Babkov<sup>a</sup>, Anastasia L. Khandazhinskaya<sup>b</sup>, Alexander O. Chizhov<sup>c</sup>, Graciela Andrei<sup>d</sup>, Robert Snoeck<sup>d</sup>, Katherine L. Seley-Radtke<sup>e</sup>\*, and Mikhail S. Novikov<sup>a</sup>

<sup>a</sup>Department of Pharmaceutical & Toxicological Chemistry, Volgograd State Medical University, Pavshikh Bortsov Sq., 1, Volgograd 400131, Russia

<sup>b</sup>Engelhardt Institute of Molecular Biology, Russian Academy of Science, Vavilova 32, Moscow 119991, Russia <sup>c</sup>Zelinsky Institute of Organic Chemistry, Russian Academy of Science, Leninsky pr., 47, Moscow 119991, Russia

<sup>d</sup>Rega Institute for Medical Research, KU Leuven, Minderbroedersstraat 10, Leuven B-3000, Belgium <sup>e</sup>Department of Chemistry & Biochemistry, University of Maryland, Baltimore County, 1000 Hilltop Circle, Baltimore, MD 21250, USA

#### **Dual inhibitors**





Bioorganic & Medicinal Chemistry journal homepage: www.elsevier.com

# Toward the discovery of dual HCMV-VZV inhibitors: synthesis, structure activity relationship analysis, and cytotoxicity studies of long chained 2-uracil-3-yl-N-(4-phenoxyphenyl)acetamides

Denis A. Babkov<sup>a</sup>, Anastasia L. Khandazhinskaya<sup>b</sup>, Alexander O. Chizhov<sup>c</sup>, Graciela Andrei<sup>d</sup>, Robert Snoeck<sup>d</sup>, Katherine L. Seley-Radtke<sup>e\*</sup>, and Mikhail S. Novikov<sup>a</sup>

<sup>a</sup>Department of Pharmaceutical & Toxicological Chemistry, Volgograd State Medical University, Pavshikh Bortsov Sq., 1, Volgograd 400131, Russia <sup>b</sup>Engelhardt Institute of Molecular Biology, Russian Academy of Science, Vavilova 32, Moscow 119991, Russia <sup>c</sup>Zelinsky Institute of Organic Chemistry, Russian Academy of Science, Leninsky pr., 47, Moscow 119991, Russia <sup>d</sup>Rega Institute for Medical Research, KU Leuven, Minderbroedersstraat 10, Leuven B-3000, Belgium <sup>e</sup>Department of Chemistry & Biochemistry, University of Maryland, Baltimore County, 1000 Hilltop Circle, Baltimore, MD 21250, USA

#### ARTICLE INFO

Article history: Received Revised Accepted Available online Keywords: human cytomegalovirus varicella zoster virus antiviral non-nucleoside inhibitor uracil \* corresponding author: <u>kseley@umbc.edu</u> 1-410-455-8684 (O); 1-410-455-2608 (Fax)

#### ABSTRACT

The need for novel therapeutic options to fight herpesvirus infections still persists. Herein we report the design, synthesis and antiviral evaluation of a new family of non-nucleoside antivirals, derived from  $1-[\omega-(4-bromophenoxy)alkyl]uracil derivatives – previously reported inhibitors of human cytomegalovirus (HCMV). Introduction of the N-(4-phenoxyphenyl)acetamide side chain at N3 increased their potency and widened activity spectrum. The most active compounds in the series exhibit submicromolar activity against different viral strains of HCMV and varicella zoster virus (VZV) replication in HEL cell cultures. Inactivity against other DNA and RNA viruses, including herpes simplex virus 1/2, points to a novel mechanism of antiviral action.2009 Elsevier Ltd. All rights reserved.$ 

#### 1. Introduction

cytomegalovirus Human (HCMV, or human herpesvirus-5) and varicella zoster virus (VZV, also known as human herpesvirus-3) belong to the viral family known as Herpesviridae. Despite modern prevention and treatment strategies, they remain a common opportunistic pathogen associated with serious morbidity and mortality, particularly in immunocompromised individuals such as transplant recipients<sup>1</sup> and AIDS patients.<sup>2,3</sup> All drugs currently licensed for the treatment of HCMV and VZV infections (Figure 1) target the viral DNA polymerase. Unfortunately they are associated with severe toxicity issues, including marrow toxicity for ganciclovir, valganciclovir, and cidofovir, and renal toxicity for foscarnet and cidofovir.<sup>4,5</sup> The emergence of drug resistance is also a significant problem.<sup>6</sup> Moreover, our understanding of the full spectrum of risks of HCMV infection and its interactions with the host immune system remains far from complete.7 While the past two decades have seen progress toward novel treatments for herpesviruses, the need for better drugs that exhibit an improved toxicity profile persists.8





Previously, we have described a series of  $1-[\omega-(phenoxy)alkyl]$ uracil derivatives that were found to exhibit high specificity and promising inhibitory activity against HCMV replication in HEL cell cultures with EC<sub>50</sub> values within 5.5–12  $\mu$ M range.<sup>9</sup> These results provided strong impetus to further explore structure-activity relationships and the antiviral activity spectrum of similar scaffolds. The present paper describes lead development of N-(4-phenoxyphenyl)acetamide derivatives (Figure 2).



Previously reported anti-HCMV compounds



Target compounds

Figure 2. Target compounds.

#### 2. Results and Discussion

#### 2.1. Chemistry

The synthesis of the compound library relied on three key steps: synthesis of  $1-[\omega-(phenoxy)alkyl]uracil$ derivatives and related compounds, synthesis of 2-chloro-N-(4-phenoxyphenyl)acetamides, and their subsequent conjugation to realize the target compounds. Based on the successful synthesis of several previously reported compounds,9 a series of analogues were synthesized following the classical synthetic approach reported in Scheme 1. This first set of derivatives is characterized by focused modifications on the aromatic ring. The synthesis of compounds **4a-j** started from commercially available phenols **1** ( $R_1 = H$ , 3-Br, 4-Br, 4-Ph or 4-CN), which were treated with 4-fold excess of  $\alpha, \omega$ -dibromoalkanes 2 (n = 1-4, 6 or 8) to produce bromides 3a-i according to known procedures.<sup>10-12</sup> A modified silvl Hilbert-Johnson reaction, i.e. condensation of equimolar amounts of 2,4bis(trimethylsilyloxy)pyrimidines<sup>13</sup> with bromides 3a-e, was performed at 160-170 °C in the absence of solvent9 to afford target compounds 4a-e in 76-88% yield. Compounds 4f-j comprising 3, 4, 6, and 8 methylene units, respectively, were obtained in an analogous manner as was used for compounds 4a-e (Scheme 1).

Intermediate 2-chloroacetamides **6a-c** were readily obtained via acylation of the commercially available 4-(phenoxy)- **(5a)**, 4-(benzyloxy)- **(5b)** and 4-benzyl- **(5c)** anilines with chloroacetyl chloride promoted by anhydrous K<sub>2</sub>CO<sub>3</sub> in aprotic media.<sup>14</sup> 2-Chloroacetamide **6d** was obtained as we described previously.<sup>15</sup> Treatment of potassium salts of uracil derivatives **4a-j** with 2-chloroacetamides **6a-d** in anhydrous DMF afforded target compounds **7a-i** and **7k-m** in good yields (Scheme 2). To avoid possible complications due to iodine elimination, compound **7j** was obtained from **4j** and 2-chloroacetamide **6a** employing NaH as the base.<sup>16</sup> Target compound **7j** was subsequently isolated in 86% yield.<sup>15</sup>



 $\label{eq:scheme1} \begin{array}{l} \mbox{Scheme1. Reagents and conditions: (a) $K_2CO_3$, acetone, reflux, 12 $h; (b) $2,4-bis(trimethylsilyloxy)-5-$R_2-pyrimidine, neat, 160–170 °C, 1 $h$.} \end{array}$ 



Scheme 2. Reagents and conditions: (a) chloroacetyl chloride,  $K_2CO_3$ , DCE, 0 °C, 2 h; (b) 4a-i,  $K_2CO_3$ , DMF, 80 °C  $\rightarrow$  RT, 24 h (for 7a-i and 7k-m) or 4j, NaH, DMF, RT, 24 h (for 7j).

In order to obtain benzophenone derivative **9**, the appropriate building blocks were obtained in a somewhat different order. First, acid **8** was synthesized quantitatively via treatment of **4b** with ethyl bromoacetate in the presence of  $K_2CO_3$ , followed by hydrolysis using LiOH according to published procedures.<sup>17</sup> Subsequent conversion of acid **8** into the corresponding acyl chloride, followed by condensation with the *N*-trimethylsilyl derivative of 4-(benzoyl)aniline led to **9** in 57% yield (Scheme 3).<sup>15</sup> It should be noted that utilization of base-free conditions for the synthesis of **9** avoids the by-products that typically arise from Knoevenagel condensation between the benzophenone carbonyl and the active methylene of the acetic acid residue.<sup>18</sup>



**Scheme 3.** Reagents and conditions: (a) (i) ethyl bromoacetate,  $K_2CO_3$ , DMF, 80 °C  $\rightarrow$  RT, 24 h; (ii) LiOH, EtOH/H<sub>2</sub>O, RT, 2 h; (b) (i) SOCl<sub>2</sub>, DCE, reflux, 1 h; (ii) DCE, -15 °C, overnight.

To further explore the SAR of these compounds we then designed a second set of analogues featuring a modified spacer linking the N<sup>1</sup> of the uracil with the aromatic moiety. Synthesis of the various target structures demanded different, but related approaches. Copper–catalyzed coupling of phenylmagnesium bromide (**10**) with excess of 1,6dibromohexane<sup>19</sup> in THF media produced bromide **11**, which was condensated with 2,4-bis(trimethylsilyloxy)pyrimidine to afford 1-(6-phenylhexyl)uracil (**12**), however in an unexpectedly mediocre 30% yield (Scheme 4). This was somewhat surprising since our previous observations for this reaction involving similar  $\omega$ -(phenoxy)alkyl bromides **3a-i** as alkylating agents were, in contrast, quite efficient.<sup>9</sup>



Scheme 4. Reagents and conditions: (a) 1,6-dibromohexane, 5 mol% Li<sub>2</sub>[CuICl<sub>2</sub>], THF, RT, 2 h; (b) 2,4-bis(trimethylsilyloxy)pyrimidine, neat, 160–170 °C, 1 h; (c) 6a, K<sub>2</sub>CO<sub>3</sub>, DMF, 80 °C  $\rightarrow$  RT, 24 h.

Silyl Hilbert-Johnson reactions between 2,4bis(trimethylsilyloxy)pyrimidine and alkyl bromides are known to be associated with the release of trimethylsilyl bromide. This subsequently, and efficiently, cleaves dialkyl ethers at elevated temperatures.<sup>20</sup> As a result, in order to obtain 1-[4-(4-bromobenzyloxy)buty]]uracil (16), it was necessary to alkylate uracil with tosylate 15, which was obtained via a Williamson ether synthesis between 1,4butanediol and 4'-bromobenzyl bromide<sup>18</sup> followed by condensation with 4-toluenesulfonyl chloride in the presence of pyridine (Scheme 5).



Scheme 5. Reagents and conditions: (a) HO(CH<sub>2</sub>)<sub>4</sub>OH, KOH, 24 h; (b) TsCl, Py, DCE, 0–5 °C; (c) uracil, K<sub>2</sub>CO<sub>3</sub>, DMF, 80 °C, 24 h; (d) **6a**, K<sub>2</sub>CO<sub>3</sub>, DMF, 80 °C  $\rightarrow$  RT, 24 h.

In addition, an analogue featuring two oxygen atoms in the spacer chain was pursued as shown in Scheme 6. Chloromethyl ether **18** was obtained via the classical Henry method.<sup>21</sup> Condensation with equimolar amount of 2,4bis(trimethylsilyloxy)pyrimidine led to 1-([2-(4-bromobenzyl)ethoxy]methyl)uracil (**19**) in 67% yield. Being more reactive than bromides **3a-i** and **11**, the reaction proceeds under rather mild conditions in 1,2-dichloroethane at ambient temperature.<sup>22</sup>



Scheme 6. Reagents and conditions: (a)  $HO(CH_2)_2OH$ , KOH, 12 h; (b)  $(CH_2O)_n$ , HCl, DCE, 0 °C, 2 h; (c) 2,4-bis(trimethylsilyloxy)pyrimidine, DCE, RT, 16 h; (d) 6a, K<sub>2</sub>CO<sub>3</sub>, DMF, 80 °C  $\rightarrow$  RT, 24 h.

Next, the flexible spacer was replaced with a rigid 4-(4-bromophenoxy)benzyl moiety, which was introduced at the N<sup>1</sup> of uracil employing previously reported procedures.<sup>23</sup> As outlined in Scheme 7, 2,4-bis(trimethylsilyloxy)pyrimidine and 4-(4-bromophenoxy)benzyl bromide (**21**) were reacted in DCE at reflux for 20 h to give **23**. Finally, to investigate the potential role of an amide nitrogen, methylation of **7b** with methyl iodide/NaH in DMF successfully produced compound **24** (92% yield, Scheme 8).



**Scheme 7.** Reagents and conditions: (a) 2,4-bis(trimethylsilyloxy)pyrimidine, DCE, reflux, 24 h; (b) **6a**, K<sub>2</sub>CO<sub>3</sub>, DMF, 80 °C  $\rightarrow$  RT, 24 h.



Scheme 8. Reagents and conditions: (a) MeI, NaH, DMF, 0  $^\circ\text{C}$ , 4 h.

#### 2.2. Biological activities

#### 2.2.1. Anti-HCMV activity

The antiviral activity of the target compounds was evaluated *in vitro* against different human herpesviruses [i.e. HCMV (AD-169 and Davis strains), VZV (OKA and 07-1

Table 1. Anti-HCMV activity in human embryonic lung (HEL) cells

strains), and herpes simplex virus 1 and 2 (HSV-1 and HSV-2)] in HEL cell cultures. The results for HCMV and VZV are summarized, respectively, in Tables 1 and 2. The results reveal that the majority of the target compounds share marked inhibitory properties. In examining the results for HCMV, the SAR studies revealed that the nature of the R<sub>1</sub> substituent significantly influences the antiviral activity: **7a** (R<sub>1</sub> = H)  $\approx$  **7c** (R<sub>1</sub> = 3-Br) < **7b** (R<sub>1</sub> = 4-Br) < **7e** (R<sub>1</sub> = 4-Ph)  $\approx$  **7d** (R<sub>1</sub> = 4-CN). Since compound **7b** showed a selectivity (ratio CC<sub>50</sub>/EC<sub>50</sub>) of about 9 for both HCMV strains and lower toxicity (alteration of cell morphology) than compound **7d**, it was considered as the basis for further modifications.



Comp	$R_1$	Spacer	D	v	Antiviral activity, $EC_{50}$ (µM) <sup>a</sup>		Cytotoxicity (µM)	
			112	Λ.	AD-169 strain	Davis strain	Cell morphology (MCC) <sup>b</sup>	Cell growth (CC <sub>50</sub> ) <sup>c</sup>
7a	Н	O(CH <sub>2</sub> ) <sub>5</sub>	Н	0	3.6 ± 0.6	4.0 ± 2.8	≥20	$5.4 \pm 2.0$
7b	4-Br	O(CH <sub>2</sub> ) <sub>5</sub>	Н	0	0.93 ± 0.75	0.97 ± 0.95	20	$8.7 \pm 5.4$
7c	3-Br	O(CH <sub>2</sub> ) <sub>5</sub>	Н	0	2.0 ± 0.3	3.1 ± 1.3	≥100	$6.3 \pm 1.3$
7d	4-CN	O(CH <sub>2</sub> ) <sub>5</sub>	Н	0	0.53 ± 0.12	$0.44 \pm 0.11$	≥4	$4.9 \pm 1.3$
7e	4-Ph	O(CH <sub>2</sub> ) <sub>5</sub>	Н	0	0.51 ± 0	$1.04 \pm 0.33$	20	$2.8 \pm 0.5$
7f	4-Br	O(CH <sub>2</sub> ) <sub>3</sub>	Н	0	$3.8 \pm 2.0$	$3.9 \pm 3.4$	≥100	15.2 ± 6.8
7g	4-Br	O(CH <sub>2</sub> ) <sub>4</sub>	Н	0	$1.79 \pm 0$	$1.64 \pm 0$	≥20	7.7 ± 7.2
7h	4-Br	O(CH <sub>2</sub> ) <sub>6</sub>	Н	0	0.29 ± 0.06	$0.20 \pm 0.18$	4	11.7 ± 11.7
7i	4-Br	O(CH <sub>2</sub> ) <sub>8</sub>	Η	0	< 0.032	< 0.032	≥0.16	-
7j	4-Br	O(CH <sub>2</sub> )5	Ι	0	>20	>20	100	-
7k	4-Br	O(CH <sub>2</sub> ) <sub>5</sub>	Н	$CH_2$	$0.44 \pm 0.11$	$0.57 \pm 0.33$	4	8.7 ± 1.7
71	4-Br	O(CH <sub>2</sub> )5	Н	OCH <sub>2</sub>	>20	4	≥20	-
7m	-	-	-	-	>4	>4	20	-
9	4-Br	O(CH <sub>2</sub> ) <sub>5</sub>	Н	CO	$1.79 \pm 0$	$1.64 \pm 0$	≥20	$5.8 \pm 0.6$
13	Н	(CH <sub>2</sub> ) <sub>6</sub>	Н	0	>20	>20	100	-
17	4-Br	$CH_2O(CH_2)_4$	Н	0	$1.8 \pm 0.3$	≥1.2 ± 0.6	≥4	$4.8 \pm 1.1$
20	4-Br	CH <sub>2</sub> O(CH <sub>2</sub> ) <sub>2</sub> OCH <sub>2</sub>	Н	0	>20	>20	100	-
23	4-Br	p-OC <sub>6</sub> H <sub>4</sub> CH <sub>2</sub>	Н	0	>0.8	>0.8	4	-
24	-	-	-	-	0.98 ± 0.93	1.29 ± 1.27	≥4	$6.1 \pm 2.3$
GCV					13.9 ± 10.6	7.8 ± 3.4	>350	≥319 ± 102
CDV					$0.83 \pm 0.40$	$0.84 \pm 0.29$	>300	≥208 ± 116

<sup>a</sup> Effective concentration required to reduce virus plaque formation by 50%. Virus input was 100 plaque forming units (PFU).

<sup>b</sup> Minimum cytotoxic concentration that causes a microscopically detectable alteration of cell morphology.

 $^{\rm c}$  Cytotoxic concentration required to reduce cell growth by 50%.

Notably, the nature of the spacer between the uracil and the left aromatic "wing" significantly influences the anti-HCMV activity of the compounds. For example, elongation of the spacer from 3 to 8 methylene groups progressively lowers EC<sub>50</sub> values, as demonstrated by **7f** (n = 3) < **7g** (n = 4) < **7b** (n = 5) < **7h** (n = 6) < **7i** (n = 8). Specifically, introduction of five methylene groups to **7f** to give **7i** increases the activity 43-73 fold, however a concomitant increase in cytotoxicity was also observed.

Replacement of the oxygen atom for a methylene (compound **13**) resulted in a loss of activity, which points to the potential role of an ether functionality in the spacer region. A shift in position of the oxygen on the aromatic moiety did not markedly affect activity (1.2 to 2-fold change of **17** as compared to **7b**), while introduction of an additional oxygen (compound **20**) renders the compound completely inactive. Use of a rigid spacer (compound **23**) also proved unfavorable. Overall, incorporation of an alkoxyphenyl side chain proved to be optimal.

Interestingly, the one modification made to the uracil moiety proved deleterious. Substitution of the H5 of the uracil with iodine rendered **7j** completely inactive.

Next, the SAR studies involving modifications to the right "wing" revealed that the presence of a second benzene core in the acetamide side chain is mandatory for inhibitory properties (compound 7k). Investigation of the role of the linker between the aromatic residues shows that oxygen provides the optimal activity profile. The corresponding methylene analogue retains a similar level of activity, but is more cytotoxic (7k), while use of a carbonyl (9) or OCH<sub>2</sub> (7I) group renders the compound significantly less active. *N*-Methylation of the parent 7b has no significant influence on the potency as compound 24 shows.

#### 2.2.2. Anti-VZV activity

Similar activity trends were observed for the anti-VZV properties of the target compounds. The 4-bromosubstituted compound **7b** blocks VZV replication at 1.14 µM (OKA strain), which is comparable with acyclovir. At the same time, thymidine kinase deficient strain 07-1 was also susceptible to **7b**. Since thymidine kinase is required for the activation of nucleoside analogues (e.g. acyclovir and brivudin), it is likely these compounds are acting as nonnucleoside inhibitors.<sup>24</sup> Other substituents at R1 were found inactive (7a, 7c-e). As was noted for the HCMV inhibition, elongation of the spacer has a pronounced positive impact on the anti-VZV properties. Compound 7i featuring 8 methylene units proved the most active, with  $EC_{50}$ 's of  $\ge 0.12$  $\mu$ M and  $\geq$ 0.16  $\mu$ M for the OKA and 07-1 strains, respectively. Again however, an increase in activity was accompanied by higher cytotoxicity for both 7h and 7i. Other spacer modifications, including oxygen (13), a position shift (17), and introduction of additional oxygen (20), or a benzene moiety (23) all resulted in a loss of activity. Thus, the nature of the spacer once again plays a crucial role in the antiviral properties.

In terms of substituents, introduction of a substituent at C5 of uracil renders compound **7**j completely inactive. Substitution of the oxygen linker between aromatic residues on the acetamide side chain (right wing) led to more

the cytotoxic methylene derivative **7k** and the less potent **7l** (X = OCH<sub>2</sub>) and **9** (X = CO). In addition, *N*-methyl derivative **24** and compound **7m**, both lacking the phenoxy core, indicated that the *N*-(4-phenoxyphenyl)acetamide side chain is likely a key element required for antiviral properties of the scaffold.

 Table 2. Anti-VZV activity in human embryonic lung (HEL)

 cells

cens					
	Antiviral activi	ity, EC <sub>50</sub> (μM) <sup>a</sup>	Cytotoxicity (µM)		
Comp	TK⁺ VZV (OKA strain)	TK <sup>-</sup> VZV (07- 1 strain)	Cell morphology (MCC) <sup>b</sup>	Cell growth (CC <sub>50</sub> )¢	
7a	>100	>100	>100	-	
7b	$1.6 \pm 0.7$	2.34 ± 0.18	>100	8.7 ± 5.4	
7c	>100	>100	>100	-	
7d	>100	>100	>100	-	
7e	>20	>20	100	-	
7f	5.4 ± 0.5	$4.6 \pm 0.9$	≥100	$15.2 \pm 6.8$	
7g	3.1 ± 0.1	$2.8 \pm 0.6$	>100	7.7 ± 7.2	
7h	0.8 ± 0	$0.51 \pm 0.01$	4	11.7 ± 11.7	
7i	≥0.12 ± 0.05	≥0.16 ± 0	0.8	11.7 ± 11.7	
7j	>100	>100	>100	-	
7k	0.8	4	≥4	-	
71	>4	11.7	≥20	-	
7m	>4	15	≥20	-	
9	>100	>100	>100	-	
13	>100	>100	>100	-	
17	>100	>100	>100	-	
20	>100	>100	>100	-	
23	>4	>4	20	-	
24	8.0 ± 0.3	7.7 ± 1.5	≥100	$6.0 \pm 2.4$	
Acyclovir	$1.82 \pm 0.67$	54.5 ± 50.6	>440	>440	
Brivudin	0.029 ± 0.017	38.6 ± 45.5	>300	≥79.3	

<sup>a</sup> Effective concentration required to reduce virus plaque formation by 50%. Virus input was 100 plaque forming units (PFU).

<sup>b</sup> Minimum cytotoxic concentration that causes a microscopically detectable alteration of cell morphology.

 $^{\rm c}$  Cytotoxic concentration required to reduce cell growth by 50%.

Interestingly, the compounds were inactive against HSV-1 and HSV-2. This is somewhat surprising since most antiherpes drugs targeting viral DNA polymerase share a broad spectrum activity against the various herpesviruses.<sup>25-29</sup> The mechanism of action for compounds **7a-m**, **9**, **13**, **17**, **20**, **23**, and **24** however, remains unclear, thus further investigation is warranted.

In addition, the target compounds were screened against a large panel of additional DNA and RNA viruses. No activity was observed for Vaccinia virus, Vesicular stomatitis virus, Coxsackie virus B4, Influenza A virus H1N1 subtype, Influenza A virus H3N2 subtype, Influenza B virus, Parainfluenza-3 virus, Reovirus-1, Sindbis virus, Punta Toro virus, Feline Corona Virus, HIV-1 and HIV-2.

#### 3. Conclusions

Herein we have described the synthesis, preliminary biological evaluation and SAR studies for a series of novel uracil derivatives as potential dual HCMV-VZV agents. The majority of the synthesized compounds exhibited potent antiviral activity in cell cultures. Experimental data undoubtedly shows that  $\omega$ -(4-bromophenoxy)alkyl substituent at  $N^1$  and N-(4-phenoxyphenyl)acetamide side chain at N<sup>3</sup> of uracil are the key elements responsible for both anti-HCMV and anti-VZV activity. The mode of action for the reported series is not yet fully elucidated and will be published elsewhere once it has been studied further. In the meantime, the most active compounds in the series 7b, 7h and 7i represent an excellent starting point for further optimization.

#### 4. Experimental

#### 4.1. General

All reagents were obtained at the highest grade available from Sigma and Acros Organics, and were used without further purification unless otherwise noted. Anhydrous DMF and isopropyl alcohol were purchased from Sigma-Aldrich Co. Anhydrous acetone, DCE, and EtOAc were obtained by distillation over P2O5. TLC was performed on Merck TLC Silica gel 60 F<sub>254</sub> plates eluting with the specified solvents and samples were made visual with a UV lamp, VL-6.LC (France). Acros Organics (Belgium) silica gel (Kieselgur 60-200 µm, 60A) was used for column chromatography. Yields refer to spectroscopically (<sup>1</sup>H and <sup>13</sup>C NMR) homogeneous materials. Melting points were determined in glass capillaries on a Mel-Temp 3.0 (Laboratory Devices Inc., USA). NMR spectra were obtained using Bruker Avance 400 (400 MHz for <sup>1</sup>H and 100 MHz for <sup>13</sup>C) and Bruker Avance 600 (600 MHz for <sup>1</sup>H and 150 MHz for <sup>13</sup>C) spectrometers in DMSO-*d*<sub>6</sub> or CDCl<sub>3</sub> with tetramethylsilane as an internal standard. High-resolution mass spectra were measured on Bruker micrOTOF II instruments using electrospray ionization (HRESIMS). The measurements were run in positive ion mode (interface capillary voltage -4500 V) in a mass range from m/z 50 to m/z 3000 Da; external or internal calibration was performed with ESI Tuning MixTM (Agilent Technologies). A syringe injection was used for solutions in MeCN (flow rate = 3  $\mu$ L/min). N<sub>2</sub> was applied as a dry gas; the interface temperature was set at 180 °C.

4.2. Synthesis

#### 4.2.1. Compounds 4a, 4b, 4d-k

The synthesis and characterization data for were reported previously.<sup>9</sup>

#### 4.2.1.1. 1-[5-(3-Bromophenoxy)pentyl]uracil (4c)

equimolar mixture of the 1-bromo-5-(3-An bromophenoxy)pentane (3c) (4.35 g, 13.51 mmol) and 2,4bis(trimethylsilyloxy)pyrimidine (3.43 g, 13.38 mmol) was heated at 160-170 °C for 1 h. The resulting melt was dissolved in EtOAc (50 mL) and treated with *i*-PrOH (10 mL). The precipitated product was collected and purified by shortcolumn flash chromatography using EtOAc/DCE (1:5). Subsequent recrystallization from a mixture of EtOAc/hexane (2:1) provided the desired product as white crystals (3.87 g, 82%); mp 124.5-126 °C; Rf 0.42 (ethyl acetate); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 1.38 (2H, quin, *J* = 7.6 Hz, CH<sub>2</sub>), 1.62 (2H, quin, J = 7.5 Hz, CH<sub>2</sub>), 1.71 (2H, quin, J = 7.5 Hz, CH<sub>2</sub>), 3.66 (2H, t, J = 7.2 Hz, NCH<sub>2</sub>), 3.97 (2H, t, J = 6.5 Hz, OCH<sub>2</sub>), 5.53

(1H, dd, J = 7.8 and 2.2 Hz, Ura-H-5), 6.93 (1H, dd, J = 8.3 and 2.2 Hz, H-6'), 7.06-7.14 (2H, m, H-2', H-4'), 7.22 (1H, t, J = 8.2 Hz, H-5'), 7.65 (1H, d, J = 7.8 Hz, Ura-H-6), 11.22 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  22.6, 28.4, 47.6, 67.9, 101.1, 114.3, 117.5, 122.4, 123.6, 131.5, 146.0, 151.3, 160.0, 164.1.

#### 4.2.1.2. 1-(6-Phenylhexyl)uracil (12)

An equimolar mixture of the 1-bromo-6-phenylhexane (2.15 8.91 mmol) and (11)g, 2,4bis(trimethylsilyloxy)pyrimidine (2.26 g, 8.81 mmol) was heated at 160-170 °C for 1 h. The resulting melt was dissolved in EtOAc (50 mL) and treated with *i*-PrOH (10 mL). Precipitate was filtered off, and the filtrate evaporated, taken up into acetone (30 mL) and EtOAc (5 mL) and purified by short-column flash chromatography using EtOAc to give 12 as white crystals (0.72 g, 30%); mp 78-79 °C, R<sub>f</sub> 0.61 (ethyl acetate); <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ 1.26-1.34 (4H, m, CH<sub>2</sub>), 1.55-1.59 (4H, m, CH<sub>2</sub>), 2.55 (2H, t, *J* = 7.4 Hz, PhCH<sub>2</sub>), 3.64 (2H, t, J = 7.2 Hz, NCH<sub>2</sub>), 5.53 (1H, dd, J = 7.8 and 1.6 Hz, Ura-H-5), 7.13-7.17 (3H, m, H-2', H-4', H-6'), 7.26 (2H, t, J = 7.4 Hz, H-3', H-5'), 7.59 (2H, d, J = 7.8 Hz, Ura-H-6), 11.11 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>): δ 25.6, 28.2, 28.4, 30.7, 35.1, 47.5, 100.8, 125.6, 128.2, 142.2, 145.6, 151.0, 163.8.

# 4.2.1.3. 1-[4-[(4-Bromobenzyl)oxy]butyl]uracil (16)

A mixture of uracil (3.1 g, 27.66 mmol) and K<sub>2</sub>CO<sub>3</sub> (1.40 g, 10.13 mmol) in anhydrous DMF (10 mL) was stirred at 80°C for 40 min. After cooling to room temperature, 4-(4bromobenzyloxy)butyl p-toluenesulfonate (15) (3.80 g, 9.19 mmol) was added and stirring continued for 24 h. The reaction mixture was filtered, concentrated under reduced pressure and purified by short-column flash chromatography using a mixture of CHCl<sub>3</sub>/EtOH (10:1). Analytical sample was recrystallized from EtOAc to give compound 16 as white crystals (1.20 g, 37%), mp 95.5-97.5 °C, Rf 0.44 (ethyl acetate); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  1.51 (2H, quin, J = 7.4 Hz, CH<sub>2</sub>), 1.63 (2H, quin, J = 7.4 Hz, CH<sub>2</sub>), 3.42 (2H, t, J = 6.1 Hz, NCH<sub>2</sub>), 3.66 (2H, t, J = 7.1 Hz, OCH<sub>2</sub>), 4.41 (2H, s, CH<sub>2</sub>O), 5.53 (1H, d, J = 7.8 Hz, Ura-H-5), 7.27 (2H, d, J = 8.1 Hz, H-2, H-6), 7.53 (2H, d, J = 8.1 Hz, H-3, H-5), 7.63 (1H, d, J = 7.8 Hz, Ura-H-6), 11.21 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>): δ 25.8, 26.4, 47.6, 69.6, 71.3, 101.1, 120.7, 129.8, 131.5, 138.4, 146.0, 151.3, 164.1.

#### 4.2.1.4. 1-[[2-(4-Bromobenzyloxy)ethoxy]methyl]uracil (19)

To a solution of 2,4-bis(trimethylsilyloxy)pyrimidine (4.58 g, 17.84 mmol) in anhydrous methylene chloride (30 mL) was solution [[2-(4added of а bromobenzyloxy)ethoxy]methoxy]methyl chloride (18) (5.0 g, 17.88 mmol) in methylene chloride (20 mL) and stirred for 16 h at room temperature. Ethanol (95%, 10 ml) was added and the resulting mixture was stirred for 30 min, filtered, evaporated to dryness under reduced pressure and purified by short-column flash chromatography using a mixture of CHCl<sub>3</sub>/EtOH (10:1). Analytical sample was recrystallized from ethyl acetate to give compound **19** as white crystals (4.25 g, 67%), mp 103.5-104.5 °C, R<sub>f</sub> 0.39 (ethyl acetate); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  3.54 (2H, dd, J = 5.9 and 3.2 Hz, CH<sub>2</sub>), 3.66 (2H, dd, J = 5.5 and 3.5 Hz, CH<sub>2</sub>), 4.44 (2H, s, NCH<sub>2</sub>), 5.10 (2H, s, OCH<sub>2</sub>), 5.60 (1H, d, J = 7.8 Hz, Ura-H-5), 7.26 (2H, d, J = 8.3 Hz, H-2', H-6'), 7.52 (2H, d, J = 8.3 Hz, H-3', H-5'), 7.70 (1H, d, J = 8.1 Hz, Ura-H-6), 11.33 (1H, s, NH); <sup>13</sup>C NMR (100 MHz,

# DMSO-*d*<sub>6</sub>): δ 68.4, 69.2, 71.5, 76.9, 101.9, 120.7, 129.8, 131.5, 138.2, 145.3, 151.4, 163.9.

#### 4.2.1.5. 1-[4-(4-Bromophenoxy)benzyl]uracil (22)

A solution of 2,4-bis(trimethylsilyloxy)-pyrimidine (3.44 g, 13.40 mmol) and 4-(bromophenoxy)benzylbromide (21) (4.6 g, 13.45 mmol) in anhydrous DCE (50 mL) was heated at reflux for 24 h, cooled to room temperature and treated with *i*-PrOH (15 mL). The resulting precipitate was collected and purified by short-column flash chromatography using a mixture of CHCl<sub>3</sub>/EtOH (10:1). An analytical sample was recrystallized from a mixture of DMF/i-PrOH/H<sub>2</sub>O (2:2:1) to give compound 22 as white crystals (2.60 g, 52%), mp 164-165.5 °C, R<sub>f</sub> 0.46 (ethyl acetate); <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>)  $\delta$  4.85 (2H, s, CH<sub>2</sub>), 5.60 (Hz, dd, I = 7.8 and 2.2 1H, Ura-H-5), 6.95 (2H, d, J = 9.0 Hz, H-2", H-6"), 7.03 (d, J = 8.6 Hz, 2H, H-3", H-6"), 7.35 (d, J = 8.8 Hz, 2H, H-2', H-6'), 7.54 (d, J = 8.8 Hz, 2H, H-3', H-5'), 7.78 (d, J = 8.1 Hz, 1H, Ura-H-6), 11.34 (d, J = 2.0 Hz, 1H, NH); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>): δ 50.0, 101.7, 115.4, 119.4, 120.9, 129.9, 132.7, 133.1, 145.9, 151.3, 156.0, 156.4, 164.0.

#### 4.2.2. General procedure for the synthesis of 2chloro-N-(4-substituted phenyl)acetamide **6a-c**

Chloroacetyl chloride (0.47 mL, 5.90 mmol) was added dropwise to a stirred mixture of the appropriate aniline **5a-c** (5.56 mmol) and  $K_2CO_3$  (0.90 g, 6.51 mmol) in anhydrous DCE (20 mL) at 0 °C. The reaction mixture was stirred for 2 hours and allowed to warm to room temperature overnight. The inorganic materials were filtered through a pad of silica gel and washed with DCE (25 mL). The filtrate was evaporated under reduced pressure and the residue was purified by recrystallization from a mixture of hexane/ethyl acetate (3 : 2).

# 4.2.2.1. 2-Chloro-N-(4-phenoxyphenyl)acetamide (6a)

Yield 80%, mp 105-106 °C,  $R_f$  0.62 (hexane/ethyl acetate, 1:1); <sup>1</sup>H NMR (600 MHz, DMSO- $d_6$ ):  $\delta$  4.24 (2H, s, COCH<sub>2</sub>), 6.97 (2H, d, J = 8.7 Hz, H-3', H-5'), 7.00 (2H, d, J = 9.0 Hz, H-2", H-6"), 7.10 (1H, t, J = 7.4 Hz, H-4"), 7.36 (2H, t, J = 8.5 Hz, H-3", H-5"), 7.61 (2H, d, J = 9.0 Hz, H-2', H-6'), 10.30 (1H, s, NH); <sup>13</sup>C NMR (150 MHz, DMSO- $d_6$ ):  $\delta$  47.7, 122.2, 123.6, 125.4, 134.2, 138.5, 156.6, 161.4, 168.7.

# 4.2.2.2. N-(4-Benzylphenyl)-2-chloroacetamide (6b)

Yield 78%, mp 142.5-144 °C,  $R_f$  0.52 (hexane/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  3.89 (2H, s, PhCH<sub>2</sub>), 4.23 (2H, s, COCH<sub>2</sub>), 7.15-7.21 (m, 5H, C<sub>6</sub>H<sub>5</sub>), 7.25-7.29 (2H, m, aromatic H), 7.51 (2H, d, J = 8.6 Hz, H-2', H-6'), 10.24 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  40.9, 43.9, 119.9, 126.3, 128.7, 128.9, 129.4, 136.8, 137.1, 141.7, 164.8.

#### 4.2.2.3.N-[4-(Benzyloxy)phenyl]-2chloroacetamide (**6c**)

Yield 88%, mp 142-144 °C,  $R_f$  0.63 (hexane/ethyl acetate, 1:1); <sup>1</sup>H NMR (600 MHz, DMSO- $d_6$ )  $\delta$  4.19 (2H, s, COCH<sub>2</sub>), 5.05 (2H, 2H, s, PhCH<sub>2</sub>), 6.97 (2H, d, J = 9.1 Hz, H-3', H-5'), 7.31 (1H, t, J = 7.4 Hz, H-4"), 7.37 (2H, t, J = 7.6 Hz, H-3", H-5"), 7.42 (2H, d, J = 7.0 Hz, H-2", H-6"), 7.49 (2H, d, J = 9.0 Hz, H-2', H-6'), 10.16 (1H, s, NH); <sup>13</sup>C NMR (150 MHz, DMSO- $d_6$ ):  $\delta$  47.7, 73.6, 119.2, 125.3, 131.9, 132.0, 132.6, 135.9, 141.3, 159.0, 168.5.

4.2.3. General procedure for the synthesis of **7a-l**, **9**, **13**, **17**, **20**, and **23** 

A mixture of the appropriate 1-substituted uracil **4a-i**, **12**, **16**, **19** or **22** (1.42 mmol) and  $K_2CO_3$  (0.29 g, 2.10 mmol) in anhydrous DMF (10 mL) was stirred at 80°C for 40 min. After cooling to room temperature, the corresponding 2-chloroacetamide **6a-c** (1.56 mmol) was added and stirring continued for 24 h. The reaction mixture was filtered, concentrated under reduced pressure and purified by short-column flash chromatography using DCE. Analytical samples were recrystallized from a mixture of hexane/ethyl acetate (1:1).

#### 4.2.3.1. 2-[2,6-Dioxo-3-(5-phenoxypentyl)-3,6dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl) acetamide (**7a**)

Yield 92%, mp 114-115.5 °C,  $R_f$  0.39 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  1.41 (2H, quin, J = 7.1 Hz, CH<sub>2</sub>), 1.67 (2H, quin, J = 7.1 Hz, CH<sub>2</sub>), 1.74 (2H, quin, J = 7.8 Hz, CH<sub>2</sub>), 3.76 (2H, t, J = 7.0 Hz, NCH<sub>2</sub>), 3.94 (2H, t, J = 6.3 Hz, OCH<sub>2</sub>), 4.60 (2H, s, COCH<sub>2</sub>), 5.75 (1H, d, J = 7.9 Hz, H-5), 6.87-6.91 (3H, m, H-4', H-2", H-6"), 6.95-6.99 (4H, m, H-2", H-6"), H-2', H-6'), 7.10 (1H, dt, J = 7.3 and 1.0 Hz, H-4"'), 7.36 (2H, dt, J = 7.3 and 1.8 Hz, H-3"', H-5"'), 7.36 (2H, dt, J = 8.6 and 1.3 Hz, H-3', H-5'), 7.57 (2H, d, J = 9.0 Hz, H-3", H-5"), 7.78 (1H, d, J = 7.8 Hz, H-6), 10.26 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  22.7, 28.5, 28.6, 43.4, 48.8, 67.4, 100.3, 114.7, 118.2, 119.8, 120.7, 121.0, 123.3, 130.0, 130.3, 135.0, 144.9, 151.4, 152.1, 157.6, 158.9, 162.6, 165.4; HRMS: found m/z 500.2181; calcd for C<sub>29</sub>H<sub>29</sub>N<sub>3</sub>O<sub>5</sub> [M+H]+ 500.2180.

#### 4.2.3.2. 2-[3-[5-(4-Bromophenoxy)pentyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**7b**)

Yield 87%, mp 128-130 °C,  $R_f$  0.47 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  1.40 (2H, quin, J = 7.8 Hz, CH<sub>2</sub>), 1.66 (2H,quin, J = 7.2 Hz, CH<sub>2</sub>), 1.72 (2H, quin, J = 7.3 Hz, CH<sub>2</sub>), 3.76 (2H, t, J = 7.0 Hz, NCH<sub>2</sub>), 3.93 (2H, t, J = 6.4 Hz, OCH<sub>2</sub>), 4.61 (2H, s, COCH<sub>2</sub>), 5.75 (1H, d, J = 7.8 Hz, H-5), 6.88 (2H, d, J = 8.9 Hz, H-2", H-6"), 6.96 (2H, d, J = 8.2 Hz, H-2", H-6"), 6.96 (2H, d, J = 8.2 Hz, H-2", H-6"), 6.98 (2H, d, J = 8.4 Hz, H-2', H-6'), 7.09 (1H, dt, J = 7.3 and 0.9 Hz, H-4"), 7.36 (2H, t, J = 7.6 Hz, H-3", H-5"), 7.40 (2H, d, J = 8.8 Hz, H-3', H-5'), 7.57 (2H, d, J = 8.9 Hz, H-3", H-5"), 7.77 (1H, d, J = 8.0 Hz, H-6), 10.27 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  22.3, 25.5, 28.1, 43.2, 48.5, 67.6, 100.0, 111.8, 116.7, 118.0, 119.5, 120.7, 123.0, 130.0, 132.1, 134.7, 144.6, 151.1, 151.8, 157.3, 157.9, 162.3, 165.1; HRMS: found m/z 578.1287; calcd for C<sub>29</sub>H<sub>28</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> 578.1285.

#### 4.2.3.3. 2-[3-[5-(3-Bromophenoxy)pentyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**7c**)

Yield 91%, mp 125-126.5 °C,  $R_f$  0.53 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  ppm 1.40 (quin, *J* = 6.4 Hz, 2H, CH<sub>2</sub>), 1.66 (quin, *J* = 7.3 Hz, 2H, CH<sub>2</sub>), 1.73 (quin, *J* = 7.8 Hz, 2H, CH<sub>2</sub>), 3.76 (t, *J* = 7.3 Hz, 2H, CH<sub>2</sub>N), 3.96 (t, *J* = 6.4 Hz, 2H, CH<sub>2</sub>O), 4.61 (s, 2H, CH<sub>2</sub>CO), 5.76 (d, *J* = 7.8 Hz, 1H, H-5), 6.93 (dd, *J* = 8.3, 2.3 Hz, 1H, H-4'), 6.95-7.00 (m, 4H, H-2", H-6", H-2"', H-6"'), 7.07-7.12 (m, 3H, H-2', H-6', H-4"''), 7.20 (t, *J* = 8.0 Hz, 1H, H-5'), 7.36 (dt, *J* = 7.6, 2.4 Hz, 2H, H-3"'', H-5"'), 7.57 (d, *J* = 9.0 Hz, 2H, H-3", H-5"'), 7.77 (d, *J* = 8.0 Hz, 1H, H-6), 10.27 (s, 1H, NH); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  ppm 22.6, 28.4, 40.7, 43.4, 48.8, 67.9, 100.3, 114.3, 117.5, 118.2, 119.8, 121.0, 122.4, 123.3, 123.6, 130.3, 131.5, 135.0, 144.8, 151.4, 152.1, 157.6, 160.0, 162.6, 165.4; HRMS: found *m*/*z* 578.1280; calcd for C<sub>29</sub>H<sub>28</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> 578.1285.

#### 4.2.3.4. 2-[3-[5-(4-Cyanophenoxy)pentyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**7d**)

Yield 83%, mp 127-128.5 °C,  $R_f$  0.33 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  1.41 (2H, quin, J = 8.5 Hz, CH<sub>2</sub>), 1.67 (2H, quin, J = 7.4 Hz, CH<sub>2</sub>), 1.76 (2H, quin, J = 7.6 Hz, CH<sub>2</sub>), 3.76 (2H, t, J = 7.1 Hz, NCH<sub>2</sub>), 4.05 (2H, t, J = 6.4 Hz, OCH<sub>2</sub>), 4.60 (2H, s, COCH<sub>2</sub>), 5.75 (1H, d, J = 8.0 Hz, H-5), 6.96 (2H, d, J = 8.8 Hz, H-2", H-6"), 6.98 (2H, d, J = 8.8 Hz, H-2", H-6"), 7.09 (1H, t, J = 7.6 Hz, H-4"), 7.36 (2H, dt, J = 7.4 and 1.2 Hz, H-3", H-5"), 7.57 (2H, d, J = 8.8 Hz, H-3', H-5'), 7.72 (2H, d, J = 8.8 Hz, H-3", H-5"), 7.77 (1H, d, J = 7.8 Hz, H-6), 10.27 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  22.5, 28.3, 28.4, 43.4, 48.7, 68.2, 100.3, 103.0, 115.8, 118.2, 119.5, 119.8, 121.0, 123.3, 130.3, 134.5, 135.0, 144.8, 151.4, 152.1, 157.6, 162.4, 162.5, 165.4; HRMS: found m/z 525.2126; calcd for C<sub>30</sub>H<sub>28</sub>N<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup> 525.2132.

#### 4.2.3.5. 2-[3-[5-(4-Phenylphenoxy)pentyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**7e**)

Yield 78%, mp 138.5-139.5 °C,  $R_f$  0.42 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  1.43 (2H, quin, *J* = 8.1 Hz, CH<sub>2</sub>), 1.68 (2H, quin, *J* = 7.1 Hz, CH<sub>2</sub>), 1.76 (2H, quin, *J* = 7.6 Hz, CH<sub>2</sub>), 3.78 (2H, t, *J* = 7.0 Hz, NCH<sub>2</sub>), 3.99 (2H, t, *J* = 6.3 Hz, OCH<sub>2</sub>), 4.61 (2H, s, COCH<sub>2</sub>), 5.76 (1H, d, *J* = 8.0 Hz, H-5), 6.94-7.01 (6H, m, H-2', H-6', H-2", H-6", H-2"', H-6"'), 7.09 (1H, dt, *J* = 7.3 and 1.0 Hz, H-4"'), 7.29 (1H, dt, *J* = 7.3 and 1.0 Hz, Ph-H<sup>4</sup>), 7.35 (dt, *J* = 7.6, 1.0 Hz, 2H, H-3"', H-5"'), 7.41 (t, *J* = 7.3 Hz, 2H, Ph-H<sup>3</sup>, Ph-H<sup>5</sup>), 7.54-7.59 (6H, m, H-3', H-5', H-3", H-5", Ph-H<sup>2</sup>, Ph-H<sup>6</sup>), 7.79 (1H, d, *J* = 7.8 Hz, H-6), 10.27 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  22.7, 28.5, 28.6, 43.4, 45.4, 48.8, 67.6, 100.3, 115.2, 118.2, 119.8, 121.0, 123.3, 126.5, 127.0, 128.0, 129.2, 130.3, 132.7, 135.1, 140.2, 144.9, 151.4, 152.1, 157.7, 158.6, 162.6, 165.4; HRMS: found *m*/*z* 576.2487; calcd for C<sub>35</sub>H<sub>33</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]\* 576.2493.

#### 4.2.3.6. 2-[3-[3-(4-Bromophenoxy)propyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**7f**)

Yield 71%, mp 127.5-129 °C,  $R_f$  0.38 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) d 1.96-2.13 (2H, m, CH<sub>2</sub>), 3.92 (2H, t, *J* = 6.7 Hz, NCH<sub>2</sub>), 3.96-4.06 (2H, m, OCH<sub>2</sub>), 4.60 (2H, s, CH<sub>2</sub>CO), 5.74 (1H, d, *J*=7.9 Hz, H-5), 6.84-6.91 (2H, m, H-2", H-6"), 6.93-7.03 (4 H, m, H-2", 'H-6"), 7.07-7.14 (1H, m, H-4"'), 7.30-7.47 (4H, m, H-3', H-5', 'H-3"', H-5''), 7.53 - 7.64 (2H, m, H-3", 'H-5"), 7.74 (1 H, d, *J*=7.9 Hz, H-6), 10.25 (1 H, s, NH); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  27.8, 43.1, 46.3, 65.2, 100.0, 100.8, 112.0, 116.7, 117.9, 119.4, 120.7, 123.0, 129.9, 132.0, 134.6, 144.6, 145.7, 151.1, 157.6, 162.2, 165.0; HRMS: found *m/z* 550.0968; calcd for C<sub>27</sub>H<sub>24</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]+ 550.0972.

#### 4.2.3.7. 2-[3-[4-(4-Bromophenoxy)butyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**7g**)

Yield 69%, mp 151-152 °C,  $R_f$  0.39 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  1.72 (4H, m, CH<sub>2</sub>CH<sub>2</sub>), 3.80 (2H, t, J = 6.6 Hz, CH<sub>2</sub>), 3.97 (2H, t, J = 5.9 Hz, CH<sub>2</sub>), 4.60 (2H, s, CH<sub>2</sub>CO), 5.76 (1H, d, J = 7.8 Hz, H-5), 6.90 (2H, d, J = 9.1 Hz, H-2", H-6"), 6.94-7.00 (4H, m, H-2", H-6", H-2', H-6'), 7.10 (1H, dt, J = 7.2 and 1.0 Hz, H-4"), 7.36 (2H, t, 7.6 Hz, H-3", H-5"), 7.42 (2H, d, J = 9.0 Hz, H-3', H-5'), 7.57 (2H, d, J = 9.1 Hz, H-3", H-5"), 7.79 (1H, d, J = 8.1 Hz, H-6), 10.27 (1H, s, NH); <sup>13</sup>C

NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  25.5, 25.8, 43.5, 48.7, 67.7, 100.4, 112.2, 117.1, 118.2, 119.8, 121.0, 123.3, 130.3, 132.4, 135.0, 144.8, 151.4, 152.1, 157.6, 158.2, 162.5, 165.4; HRMS: found m/z 564.1127; calcd for  $C_{28}H_{26}BrN_3O_5$  [M+H]<sup>+</sup> 564.1129.

4.2.3.8.2-[3-[6-(4-Bromophenoxy)hexyl]-2,6-dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**7h**)

Yield 77%, mp 143-144 °C,  $R_f$  0.48 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.42 (2H, quin, J = 7.6 Hz, CH<sub>2</sub>), 1.52 (2H, quin, J = 7.1 Hz, CH<sub>2</sub>), 1.73-1.79 (4H, m, CH<sub>2</sub>CH<sub>2</sub>), 3.76 (2H, t, J = 7.8 Hz, NCH<sub>2</sub>), 3.93 (2H, t, J = 6.3 Hz, OCH<sub>2</sub>), 4.77 (2H, s, COCH<sub>2</sub>), 5.77 (1H, d, J = 7.8 Hz, H-5), 6.76 (2H, d, J = 8.8 Hz, H-2", H-6"), 6.93 (2H, d, J = 8.8 Hz, H-2", H-6"), 6.97 (2H, d, J = 8.3 Hz, H-2', H-6'), 7.07 (1H, dt, J = 7.1 and 0.9 Hz, H-4"'), 7.14 (1H, d, J = 7.9 Hz, H-6), 7.28-7.37 (4H, m, H-3"', H-5"', H-3', H-5'), 7.44 (2H, d, J = 8.8 Hz, H-3", H-5"), 7.95 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  25.4, 25.9, 28.7, 28.8, 49.6, 67.9, 101.2, 116.3, 118.4, 119.3, 122.9, 129.5, 132.1, 142.5, 151.3, 162.6; HRMS: found m/z 592.1437; calcd for C<sub>30</sub>H<sub>30</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> 592.1442.

#### 4.2.3.9. 2-[3-[8-(4-Bromophenoxy)octyl]-2,6-dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**7i**)

Yield 71%, mp 128-129 °C,  $R_f$  0.54 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.34 (6H, m, CH<sub>2</sub> × 6), 1.42 (2H, quin, J = 7.6 Hz, CH<sub>2</sub>), 1.71 (2H, quin, J = 7.1 Hz, CH<sub>2</sub>), 1.75 (2H, quin, J = 8.1 Hz, CH<sub>2</sub>), 3.73 (2H, t, J = 7.4 Hz, NCH<sub>2</sub>), 3.90 (2H, t, J = 6.6 Hz, OCH<sub>2</sub>), 4.81 (2H, s, COCH<sub>2</sub>), 5.80 (1H, d, J = 7.8 Hz, H-5), 6.77 (2H, d, J = 9.0 Hz, H-2", H-6"), 6.88 (2H, d, J = 8.8 Hz, H-2", H-6"), 6.94 (d, J = 7.8 Hz, 2H, H-2', H-6'), 7.06 (t, J = 7.3 Hz, 1H, H-4"), 7.17 (d, J = 7.8 Hz, 1H, H-6), 7.30-7.37 (4H, m, H-3", H-5", H-3', H-5'), 7.43 (2H, d, J = 8.8 Hz, H-3", H-5"), 8.52 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  25.7, 26.2, 28.8, 28.91, 28.94, 29.0, 44.2, 49.9, 68.0, 76.6, 76.9, 77.3, 101.2, 112.4, 116.2, 118.2, 119.3, 121.2, 122.8, 129.6, 132.1, 133.3, 143.0, 151.3, 153.1, 156.9, 158.1, 162.9, 164.8; HRMS: found m/z 620.1749; calcd for C<sub>32</sub>H<sub>34</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> 620.1755.

#### 4.2.3.10. 2-[3-[5-(4-Bromophenoxy)pentyl]-5-iodo-2,6-dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4phenoxyphenyl)acetamide (**7***j*)

A mixture of 1-[5-(4-bromophenoxy)pentyl]-5-iodouracil (4j) (0.48 g, 1.00 mmol) and NaH (60% dispersion in mineral oil, 0.05 g, 1.25 mmol) in anhydrous DMF (15 mL) was stirred at room temperature for 1 h followed by addition of 2-chloro-N-(4-phenoxyphenyl)acetamide (6a, 0.26 g, 0.99 mmol). After 4 h, the reaction mixture was filtered, concentrated under reduced pressure and purified by short-column flash chromatography using DCE. Analytical sample was recrystallized from a mixture of hexane/EtOAc (1:1) to give compound **7j** as a white powder (0.60 g, 86%), mp 151-153 °C, R<sub>f</sub> 0.74 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 1.39 (quin, *J* = 8.1 Hz, 2H, CH<sub>2</sub>), 1.66 (quin, *J* = 7.1 Hz, 2H, CH<sub>2</sub>), 1.72 (quin, J = 7.9 Hz, 2H, CH<sub>2</sub>), 3.78 (t, J = 7.0 Hz, 2H, NCH<sub>2</sub>), 3.93 (t, J = 6.6 Hz, 2H, OCH<sub>2</sub>), 4.65 (s, 2H, COCH<sub>2</sub>), 6.88 (d, J = 9.0 Hz, 2H, H-2", H-6"), 6.96 (d, J = 8.1 Hz, 2H, H-2"", H-6""), 6.98 (d, J = 8.8 Hz, 2H, H-2', H-6'), 7.10 (dt, J = 7.3, 0.9 Hz, 1H, H-4""), 7.36 (dt, J = 7.5, 1.0 Hz, 2H, H-3"', H-5""), 7.40 (d, J = 9.0 Hz, 2H, H-3', H-5'), 7.56 (d, J = 9.0 Hz, 2H, H-3" H-5"), 8.37 (s, 1H, H-6), 10.28 (s, 1H, NH); 13C NMR (100 MHz, DMSO-d<sub>6</sub>): δ 22.5, 28.4, 28.5, 45.0, 49.1, 67.2, 67.9, 112.1, 117.1, 118.3, 119.8, 121.0, 123.4, 130.3, 132.4, 134.9, 149.1,

151.0, 152.2, 157.6, 158.2, 160.2, 165.2; HRMS: found *m*/*z* 704.0248; calcd for C<sub>29</sub>H<sub>27</sub>BrIN<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> 704.0252.

4.2.3.11. N-(4-Benzylphenyl)-2-[3-[5-(4bromophenoxy)pentyl]-2,6-dioxo-3,6dihydropyrimidin-1(2H)-yl]acetamide (**7k**)

Yield 80%, mp 117-118 °C,  $R_f$  0.42 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  ppm 1.39 (2H, quin, J = 6.9 Hz, CH<sub>2</sub>), 1.65 (2H, quin, J = 7.1 Hz, CH<sub>2</sub>), 1.72 (2H, quin, J = 7.1 Hz, CH<sub>2</sub>), 3.75 (2H, t, J = 6.9 Hz, NCH<sub>2</sub>), 3.87 (2H, s, PhCH<sub>2</sub>), 3.92 (2H, t, J = 6.4 Hz, OCH<sub>2</sub>), 4.59 (2H, s, COCH<sub>2</sub>), 5.75 (1H, d, J = 7.8 Hz, H-5), 6.88 (2H, d, J = 9.1 Hz, H-2", H-6"), 7.13-7.20 (5H, m, aromatic H), 7.25-7.28 (2H, m, aromatic H), 7.40 (2H, d, J = 9.0 Hz, H-3", H-5"), 7.46 (2H, d, J = 8.5 Hz, H-3', H-5'), 7.77 (1H, d, J = 7.8 Hz, H-6), 10.20 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  ppm 22.6, 28.4, 43.4, 48.7, 67.8, 100.28, 112.1, 117.0, 119.4, 126.2, 128.7, 128.9, 129.3, 132.4, 136.5, 137.1, 141.7, 144.8, 151.4, 158.2, 162.6, 165.4; HRMS: found m/z 576.1487; calcd for C<sub>30</sub>H<sub>30</sub>BrN<sub>3</sub>O<sub>4</sub> [M+H]+ 576.1492.

#### 4.2.3.12. N-[4-(Benzyloxy)phenyl]-2-[3-[5-(4bromophenoxy)pentyl]-2,6-dioxo-3,6-dihydropyrimidin-1(2H)-yl]acetamide (**7l**)

Yield 80%, mp 172.5-174.5 °C,  $R_f$  0.46 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  1.38 (2H, quin, J = 7.8 Hz, CH<sub>2</sub>), 1.64 (2H, quin, J = 7.4 Hz, CH<sub>2</sub>), 1.71 (2H, quin, J = 7.5 Hz, CH<sub>2</sub>), 3.74 (2H, t, J = 7.1 Hz, NCH<sub>2</sub>), 3.91 (2H, t, J = 6.4 Hz, OCH<sub>2</sub>), 4.58 (2H, s, COCH<sub>2</sub>), 5.05 (2H, s, OCH<sub>2</sub>Ph), 5.74 (1H, d, J = 8.1 Hz, H-5), 6.87 (2H, d, J = 9.1 Hz, H-3', H-5'), 6.94 (2H, d, J = 8.2 Hz, H-2"', H-6"'), 7.31 (1H, t, J = 7.1 Hz, H-4"), 7.35-7.46 (8H, m, aromatic H), 7.73 (1H, d, J = 8.0 Hz, H-6), 10.13 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  22.6, 28.4, 43.4, 48.8, 67.9, 69.7, 100.3, 112.0, 115.2, 117.0, 120.8, 128.0, 128.1, 128.7, 132.4, 137.4, 144.8, 151.4, 154.6, 158.2, 162.6, 165.1; HRMS: found m/z 592.1437; calcd for C<sub>30</sub>H<sub>30</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> 592.1442.

#### 4.2.3.13. 2-[3-[5-(4-Bromophenoxy)pentyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-phenylacetamide (**7m**)

Yield 78%, mp 157-158.5 °C,  $R_f$  0.37 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  1.39 (2H, quin, J = 7.6 Hz, CH<sub>2</sub>), 1.72 (2H, quin, J = 7.2 Hz, CH<sub>2</sub>), 1.66 (2H, quin, J = 7.6 Hz, CH<sub>2</sub>), 3.75 (2H, t, J = 7.2 Hz, NCH<sub>2</sub>), 3.93 (2H, t, J = 6.4 Hz, OCH<sub>2</sub>), 4.61 (2H, s, CH<sub>2</sub>CO), 5.76 (1H, d, J = 7.8 Hz, Ura-H-5), 6.88 (2H, d, J = 8.8 Hz, H-2, H-6), 7.04 (1H, t, J = 7.3 Hz, H-4'), 7.30 (2H, t, J = 8.1 Hz, H-3', H-5'), 7.40 (2H, d, J = 9.0 Hz, H-3, H-5), 7.55 (2H, dd, J = 8.4 and 1.1 Hz, H-2, H-6), 7.77 (1H, d, J= 8.1 Hz, Ura-H-6), 10.25 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  22.6, 28.4, 43.5, 48.8, 67.8, 100.3, 112.1, 117.0, 119.3, 123.6, 129.1, 132.4, 139.1, 144.8, 151.4, 158.2, 162.6, 165.6; HRMS: found m/z 486.1020; calcd for C<sub>23</sub>H<sub>24</sub>BrN<sub>3</sub>O<sub>4</sub> [M+H]<sup>+</sup> 486.1023.

#### 4.2.3.14. 2-[2,6-Dioxo-3-(6-phenylhexyl)-3,6dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**13**)

Yield 90%, mp 135-136 °C,  $R_f$  0.48 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  1.22-1.37 (4H, m, CH<sub>2</sub>), 1.49-1.65 (4H, m, CH<sub>2</sub>), 2.55 (2H, t, *J* = 7.4 Hz, CH<sub>2</sub>), 3.72 (2H, t, *J* = 7.1 Hz, CH<sub>2</sub>), 4.61 (2H, s, CH<sub>2</sub>), 5.74 (1H, d, *J* = 7.8 Hz, Ura-H-5), 6.93-7.00 (4H, m, H-3', H-5', H-2", H-6"), 7.07-7.12 (1H, m, H-4"), 7.13-7.19 (3H, m, H-4, H-2, H-6), 7.21-7.27 (2H, m, H-3, H-5), 7.33-7.39 (2H, m, H-2', H-6'), 7.54-7.60 (2H, m, H- 3", H-5"), 7.75 (1H, d, J = 7.8 Hz, Ura-H-6), 10.27 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  25.9, 28.5, 28.6, 31.1, 35.3, 43.4, 48.9, 100.2, 118.2, 119.8, 121.0, 123.3, 125.9, 128.5, 128.5, 130.3, 135.0, 142.5, 144.8, 151.4, 152.1, 157.6, 162.6, 165.4; HRMS: found m/z 498.2386; calcd for C<sub>30</sub>H<sub>31</sub>N<sub>3</sub>O<sub>4</sub> [M+H]<sup>+</sup> 498.2387.

4.2.3.15. 2-[3-[4-(4-Bromobenzyloxy)butyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**17**)

Yield 63%, mp 114-116 °C,  $R_f$  0.28 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  1.54 (2H, quin, J = 6.3 Hz, CH<sub>2</sub>), 1.67 (2H, quin, J = 7.4 Hz, CH<sub>2</sub>), 3.44 (2H, t, J = 6.1 Hz, CH<sub>2</sub>), 3.75 (2H, t, J = 7.1 Hz, CH<sub>2</sub>), 4.42 (2H, s, ArCH<sub>2</sub>), 4.60 (2H, s, CH<sub>2</sub>CO), 5.75 (1H, d, J = 7.9 Hz, H-5), 6.96 (2H, d, J = 8.1 Hz, H-2", H-6"), 6.98 (2H, d, J = 9.1 Hz, H-2", H-6"), 7.09 (t, J = 7.3 Hz, 1H, H-4"), 7.27 (d, J = 8.3 Hz, 2H, H-2', H-6'), 7.36 (2H, dt, J = 7.6 and 2.0 Hz, H-3", H-5"), 7.52 (2H, d, J = 8.3 Hz, H-3', H-5'), 7.57 (2H, d, J = 7.1 Hz, H-3", H-5"), 7.76 (1H, d, J = 7.8 Hz, H-6), 10.27 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  25.7, 26.4, 43.4, 45.4, 48.8, 69.6, 71.3, 100.3, 118.2, 119.8, 120.7, 121.0, 123.3, 129.9, 130.3, 131.5, 135.0, 138.4, 144.8, 151.4, 152.1, 157.6, 162.6, 165.4; HRMS: found m/z 578.1287; calcd for C<sub>29</sub>H<sub>28</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> 578.1285.

#### 4.2.3.16. 2-[3-[[2-(4-Bromobenzyloxy)ethoxy]methyl]-2,6-dioxo-3,6dihydropyrimidin-1(2H)-yl]-N-(4phenoxyphenyl)acetamide (**20**)

Yield 75%, mp 147.5-149.5 °C,  $R_f$  0.53 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  3.55 (2H, t, J = 5.9 Hz, CH<sub>2</sub>), 3.68 (2H, t, J = 4.9 Hz, CH<sub>2</sub>), 4.45 (2H, s, ArCH<sub>2</sub>), 4.61 (2H, s, CH<sub>2</sub>CO), 5.19 (2H, s, NCH<sub>2</sub>O), 5.82 (1H, d, J = 7.8 Hz, H-5), 6.96 (2H, d, J = 9.0 Hz, H-2"', H-6"'), 6.98 (d, J = 9.0 Hz, 2H, H-2", H-6'), 7.09 (t, J = 7.3 Hz, 1H, H-4"'), 7.27 (d, J = 8.3 Hz, 2H, H-2', H-6'), 7.36 (2H, dt, J = 8.6 and 1.0 Hz, H-3"', H-5"'), 7.52 (2H, d, J = 8.3 Hz, H-3', H-5'), 7.56 (2H, d, J = 7.1 Hz, H-3", H-5"), 7.83 (1H, d, J = 7.9 Hz, H-6), 10.30 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  43.4, 68.6, 69.2, 71.5, 78.0, 101.1, 118.3, 119.8, 120.7, 121.0, 123.3, 129.9, 130.3, 131.5, 135.0, 138.2, 144.2, 151.6, 152.1, 157.6, 162.4, 165.3; HRMS: found m/z 580.1073; calcd for C<sub>28</sub>H<sub>26</sub>BrN<sub>3</sub>O<sub>6</sub> [M+H]<sup>+</sup> 580.1078.

4.2.3.17. 2-[3-[4-(4-Bromophenoxy)benzyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-(4-phenoxyphenyl)acetamide (**23**)

Yield 64%, mp 201.5-202.5 °C,  $R_f$  0.43 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  4.62 (2H, s, COCH<sub>2</sub>), 4.95 (2H, s, ArCH<sub>2</sub>), 5.82 (2H, 1H, d, J = 7.8 Hz, H-5), 6.94-6.98 (6H, m, H-2", H-6", H-2", H-6", H-3", H-5"), 7.04 (d, J = 8.5 Hz, H-2', H-6'), 7.09 (1H, dt, J = 7.3 and 1.0 Hz, H-4"), 7.36 (2H, dt, J = 8.6, 1.2 Hz, H-3", H-5"), 7.38 (2H, d, J = 8.5 Hz, H-3', H-5'), 7.54 (2H, d, J = 8.8 Hz, H-2", H-6"), 7.57 (2H, d, J = 9.1 Hz, H-3", H-5"), 7.91 (1H, d, J = 7.8 Hz, H-6), 10.30 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  43.6, 51.2, 100.9, 115.5, 118.2, 119.4, 119.8, 121.0, 123.3, 130.0, 130.3, 132.4, 133.1, 135.0, 144.7, 151.5, 152.1, 156.1, 156.3, 157.6, 162.5, 165.4; HRMS: found m/z 598.0969; calcd for C<sub>31</sub>H<sub>24</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]+ 598.0972.

#### 4.2.4. 2-(3-(5-(4-Bromophenoxy)pentyl)-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl)acetic acid (**8**)

A mixture of 1-[5-(4-bromophenoxy)pentyl]uracil (4b) (1.42 mmol) and  $K_2CO_3$  (0.29 g, 2.10 mmol) in anhydrous DMF (10 mL) was stirred at 80 °C for 40 min. After cooling to

room temperature, ethyl bromoacetate (0.17 mL, 1.56 mmol) was added and stirring was continued for 24 h. The reaction mixture was filtered, concentrated under reduced pressure and purified by short-column flash chromatography using DCE. Crude product was dissolved in EtOH (20 mL). Then LiOH (0.20 g, 8.35 mmol) and water (10 mL) was added, and the resulting mixture was stirred at room temperature for 2 h. After adjusting pH to 2 with addition of 1 M HCl resulting precipitate was filtered and recrystallized from a mixture of hexane/i-PrOH (1:2) to give the desired product as white powder (2.68 g, 100%), Rf 0.54 (i-PrOH/EtOAc/NH4OH, 9:6:5), mp 142.5-145°C; <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>): δ 1.37  $(2H, quin, J = 7.4 Hz, CH_2), 1.55 (2H, quin, J = 7.4 Hz, CH_2),$ 1.71 (2H, quin, J = 7.4 Hz, CH<sub>2</sub>), 3.79 (2H, t, J = 7.3 Hz, NCH<sub>2</sub>), 3.92 (2H, t, / = 6.5 Hz, OCH<sub>2</sub>), 4.45 (2H, s, CH<sub>2</sub>), 5.72 (1H, d, / = 7.8 Hz, Ura-H-5), 6.89 (2H, d, / = 8.8 Hz, H-2, H-6), 7.41 (2H, d, *J* = 8.8 Hz, H-3, H-5), 7.65 (1H, d, *J* = 7.8 Hz, NH); <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>): δ 23.1, 27.1, 28.5, 50.0, 67.8, 100.4, 112.0, 117.0, 132.4, 144.9, 151.4, 158.2, 162.7, 169.8.

#### 4.2.5. N-(4-Benzoylphenyl)-2-[3-[5-(4-bromophenoxy)pentyl]-2,6-dioxo-3,6-dihydropyrimidin-1(2H)-yl]acetamide (**9**)

A mixture of acid 8 (0.70 g, 1.70 mmol) and thionyl chloride (0.15 mL, 2.06 mmol) in anhydrous DCE (10 mL) was refluxed with the exclusion of moisture for 2 h. The volatile materials were evaporated under reduced pressure and the residue was dissolved in DCE (10 mL) and cooled to -15 °C. The resulting solution was added dropwise to a stirred solution of *N*-(trimethylsilyl)-4-benzoylaniline, which was prepared *in situ* by heating 4-benzoylaniline (0.34 g, 1.72 mmol) with excess of HMDS. The reaction mixture was stirred overnight at room temperature and diluted with *i*-PrOH (8 mL). Solvents were evaporated under reduced pressure and crude product was purified with short-column flash chromatography using hexane/EtOAc (1:2) to give compound 9 as a white powder (0.57 g, 57%), mp 162.5-164 °C, R<sub>f</sub> 0.39 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 1.40 (2H, quin, *J* = 8.1 Hz, CH<sub>2</sub>), 1.66 (2H, quin, *J* = 7.1 Hz, CH<sub>2</sub>), 1.72 (2H, quin, J = 7.8 Hz, CH<sub>2</sub>), 3.76 (2H, t, J = 7.1 Hz, NCH<sub>2</sub>), 3.92 (2H, t, J = 6.3 Hz, OCH<sub>2</sub>), 4.67 (2H, s, COCH<sub>2</sub>), 5.77 (1H, d, J = 7.8 Hz, H-5), 6.87 (2H, d, J = 9.1 Hz, H-2', H-6'), 7.39 (2H, d, J = 9.0 Hz, H-3', H-5'), 7.54 (2H, t, J = 7.3 Hz, H-3" H-5"'), 7.65 (1H, dt, J = 7.6 and 1.5 Hz, H-4"'), 7.69-7.75 (6H, m, H-2", H-6", H-3", H-5", H-2"', H-6"'), 7.79 (1H, d, J = 8.0 Hz, H-6), 10.67 (1H, s, NH); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>): δ 22.6, 28.4, 43.7, 48.8, 67.8, 100.3, 112.1, 117.0, 118.6, 128.8, 129.7, 131.6, 131.8, 132.4, 132.6, 137.9, 143.2, 144.9, 151.4, 158.2, 162.5, 166.3, 194.9; HRMS: found m/z 590.1283; calcd for C<sub>30</sub>H<sub>28</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> 590.1285.

#### 4.2.6. 2-[3-[5-(4-Bromophenoxy)pentyl]-2,6dioxo-3,6-dihydropyrimidin-1(2H)-yl]-N-methyl-N-(4-phenoxyphenyl)acetamide (**24**)

NaH as a 60% dispersion in mineral oil (0.03 g, 0.75 mmol) was added to a solution of compound **7b** in DMF (10 mL) at 0°C followed with MeI (0.08 mL, 1.29 mmol) after 20 min. Stirring was continued for 4 h. DMF was evaporated under reduced pressure, residue was dissloved in DCE (20 mL), washed with a 5% solution of Na<sub>2</sub>SO<sub>3</sub> (80 mL), water (50 mL), and evaporated. Recrystallisation from a mixture of hexane/EtOAc (1:1) gave compound **23** as a white powder (0.33 g, 92%), mp 135.5-137 °C,  $R_f$  0.75 (DCE/ethyl acetate, 1:1); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  1.37 (2H, quin, J = 7.8 Hz, CH<sub>2</sub>), 1.63 (2H, quin, J = 7.1 Hz, CH<sub>2</sub>), 1.71 (2H, quin, J = 7.1 Hz, CH<sub>2</sub>), 3.14 (3H, s, CH<sub>3</sub>), 3.72 (2H, t, J = 7.0 Hz, NCH<sub>2</sub>), 3.93 (2H, t, J = 6.3 Hz, OCH<sub>2</sub>), 4.26 (2H, s, COCH<sub>2</sub>), 5.70 (1H, d,

*J* = 7.8 Hz, H-5), 6.88 (2H, d, *J* = 8.5 Hz, H-2", H-6"), 7.06-7.11 (4H, m, H-2', H-6', H-2"', H-6"'), 7.18 (1H, t, *J* = 7.5 Hz, H-4"'), 7.39-7.43 (6H, m, H-3', H-5", H-3", H-5", H-3"', H-5"'), 7.73 (1H, d, *J* = 7.8 Hz, H-6); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  22.6, 28.4, 37.4, 42.3, 48.7, 60.1, 67.8, 100.2, 112.1, 117.0, 119.4, 119.8, 124.3, 129.5, 130.5, 132.4, 133.0, 137.8, 144.7, 151.2, 156.7, 158.2, 162.4, 166.2; HRMS: found *m*/*z* 592.1439; calcd for C<sub>30</sub>H<sub>30</sub>BrN<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> 592.1442.

#### 4.3. Biological assays

# 4.3.1. Antiviral Activity Assays other than HIV

The compounds were evaluated against the following viruses: herpes simplex virus type 1 (HSV-1) strain KOS, thymidine kinase-deficient (TK-) HSV-1 KOS strain resistant to ACV (ACV<sup>r</sup>), herpes simplex virus type 2 (HSV-2) strains Lvons and G, human cytomegalovirus (HCMV) (strains AD-169 and Davis), varicella-zoster virus (strains OKA and YS), vaccinia virus Lederle strain, respiratory syncytial virus (RSV) strain Long, vesicular stomatitis virus (VSV), Coxsackie B4, Parainfluenza 3, Influenza virus A (subtypes H1N1, H3N2), influenza virus B, Reovirus-1, Sindbis and Punta Toro. The antiviral assays were based on inhibition of virusinduced cytopathicity or plaque formation in human embryonic lung (HEL) fibroblasts, African green monkey cells (Vero), human epithelial cells (HeLa) or Madin-Darby canine kidney cells (MDCK). Confluent cell cultures in microtiter 96well plates were inoculated with 100 CCID<sub>50</sub> of virus (1 CCID<sub>50</sub> being the virus dose to infect 50% of the cell cultures) or 10 or 100 plaque forming units (PFU) (for VZV and HCMV) in the presence of varying concentrations of the test compounds. Viral cytopathicity or plaque formation was recorded as soon as it reached completion in the control virus-infected cell cultures that were not treated with the test compounds. Antiviral activity was expressed as the EC50 or compound concentration required to reduce virus-induced cytopathogenicity or viral plaque formation by 50%.

#### 4.3.2. Anti-HIV Activity Assays

Inhibition of HIV-1(III<sub>B</sub>)- and HIV-2(ROD)-induced cytopathicity in CEM cell cultures was measured in microtiter 96-well plates containing  $\sim 3 \times 10^5$  CEM cells/mL infected with 100 CCID<sub>50</sub> of HIV per milliliter and containing appropriate dilutions of the test compounds. After 4–5 days of incubation at 37 °C in a CO<sub>2</sub>-controlled humidified atmosphere, CEM giant (syncytium) cell formation was examined microscopically. The EC<sub>50</sub> (50% effective concentration) was defined as the compound concentration required to inhibit HIV-induced giant cell formation by 50%.

#### 4.3.3. Cytostatic Activity Assays

Cytotoxicity measurements were based on the inhibition of cell growth. HEL cells were seeded at a rate of 5 x 10<sup>3</sup> cells/well into 96-well microtiter plates and allowed to adhere and proliferate for 24 h. Then, medium containing different concentrations of the test compounds was added. After 3 days of further incubation at 37 °C, the cell number was determined with a Coulter counter. The cytostatic concentration was calculated as the  $CC_{50}$ , or the compound concentration required reducing cell proliferation by 50% relative to the number of cells in the untreated controls.  $CC_{50}$ values were estimated from graphic plots of the number of cells (percentage of control) as a function of the concentration of the test compounds. Alternatively, cytotoxicity of the test compounds was expressed as the minimum cytotoxic concentration (MCC) or the compound

concentration that caused a microscopically detectable alteration of cell morphology. Selectivity indexes were calculated as the ratio  $CC_{50}$  to  $EC_{50}$ .

#### Acknowledgments

The authors wish to express their gratitude to Ms. Therese Ku, Mrs. Leentje Persoons, Mrs. Frieda De Meyer, Mrs. Lies Van den Heurck, and Mrs. Lizette van Berckelaer for excellent technical assistance. This work was supported by grant of Russian Foundation for Basic Research (13-04-01391A). The biological screening was supported by KU Leuven (GOA 10/014).

#### **References and notes**

- 1. Boeckh, M.; Nichols, W. G. Blood 2004, 103, 2003.
- Fowotade, A.; Okonko, I. O.; Agbede, O. O.; Suleiman, S. T. Afr Health Sci 2015, 15, 1.
- 3. Griffiths, P.; Baraniak, I.; Reeves, M. *J Pathol* 2015, *235*, 288.
- 4. Boeckh, M. Hematol-Am Soc Hemat 2011, 305.
- 5. Sellar, R. S.; Peggs, K. S. *Expert Opin Biol Th* 2012, *12*, 1161.
- 6. *Reactions Weekly* 2015, *1536*, 106.
- 7. Boeckh, M.; Murphy, W. J.; Peggs, K. S. *Biol Blood Marrow Tr* 2015, *21*, S19.
- Skorenski, M.; Sienczyk, M. Expert Opin Ther Pat 2014, 24, 925.
- Novikov, M. S.; Babkov, D. A.; Paramonova, M. P.; Khandazhinskaya, A. L.; Ozerov, A. A.; Chizhov, A. O.; Andrei, G.; Snoeck, R.; Balzarini, J.; Seley-Radtke, K. L. *Bioorg Med Chem* 2013, 21, 4151.
- 10. Kikumoto, R.; Tobe, A.; Tonomura, S. *J Med Chem* 1981, 24, 145.
- 11. Kikumoto, R.; Tobe, A.; Fukami, H.; Egawa, M. *J Med Chem* 1983, *26*, 246.
- Wang, S.; Jin, G.; Wang, W.; Zhu, L.; Zhang, Y.; Dong, G.; Liu, Y.; Zhuang, C.; Miao, Z.; Yao, J.; Zhang, W.; Sheng, C. Eur J Med Chem 2012, 53, 292.
- Robins, M. J.; Hatfield, P. W. *Can J Chem* 1982, *60*, 547.
   Vejdělek, Z.; Holubek, J.; Bartošová, M.; Protiva, M.
- 14. Vejdělek, Z.; Holubek, J.; Bartošová, M.; Protiva, M. *Collect. Czech. Chem. Commun.* 1982, 47, 3297.
- Novikov, M. S.; Babkov, D. A.; Paramonova, M. P.; Chizhov, A. O.; Khandazhinskaya, A. L.; Seley-Radtke, K. L. *Tetrahedron Lett* 2013, 54, 576.

- 16. Colla, L.; Busson, R.; De Clercq, E.; Vanderhaeghe, H. *Eur J Med Chem* 1982, *17*, 569.
- 17. Farr, R. A.; Bey, P.; Sunkara, P. S.; Lippert, B. J. *J Med Chem* 1989, *32*, 1879.
- Kanao, M.; Hashizume, T.; Ichikawa, Y.; Irie, K.; Isoda, S. J Med Chem 1982, 25, 1358.
- 19. Cahiez, G.; Chaboche, C.; Jezequel, M. *Tetrahedron* 2000, 56, 2733.
- 20. Hanessian, S.; Delorme, D.; Dufresne, Y. *Tetrahedron Lett* 1984, 25, 2515.
- 21. Hill, A. J.; Keach, D. T. J Am Chem Soc 1926, 48, 257.
- Novikov, M. S.; Ozerov, A. A.; Orlova, Y. A.; Buckheit, R. W. Chem Heterocycl Compd 2005, 41, 625.
- Babkov, D. A.; Valuev-Elliston, V. T.; Paramonova, M. P.; Ozerov, A. A.; Ivanov, A. V.; Chizhov, A. O.; Khandazhinskaya, A. L.; Kochetkov, S. N.; Balzarini, J.; Daelemans, D.; Pannecouque, C.; Seley-Radtke, K. L.; Novikov, M. S. *Bioorg Med Chem* 2015, 23, 1069.
- Dworkin, R. H.; Johnson, R. W.; Breuer, J.; Gnann, J. W.; Levin, M. J.; Backonja, M.; Betts, R. F.; Gershon, A. A.; Haanpaa, M. L.; McKendrick, M. W.; Nurmikko, T. J.; Oaklander, A. L.; Oxman, M. N.; Pavan-Langston, D.; Petersen, K. L.; Rowbotham, M. C.; Schmader, K. E.; Stacey, B. R.; Tyring, S. K.; van Wijck, A. J. M.; Wallace, M. S.; Wassilew, S. W.; Whitley, R. J. *Clin Infect Dis* 2007, 44, S1.
- Oien, N. L.; Brideau, R. J.; Hopkins, T. A.; Wieber, J. L.; Knechtel, M. L.; Shelly, J. A.; Anstadt, R. A.; Wells, P. A.; Poorman, R. A.; Huang, A.; Vaillancourt, V. A.; Clayton, T. L.; Tucker, J. A.; Wathen, M. W. *Antimicrob Agents Chemother* 2002, *46*, 724.
   Falardeau, G.; Lachance, H.; St-Pierre, A.; Yannopoulos,
  - Falardeau, G.; Lachance, H.; St-Pierre, A.; Yannopoulos, C. G.; Drouin, M.; Bedard, J.; Chan, L. *Bioorg Med Chem Lett* 2005, *15*, 1693.
- Larsen, S. D.; Zhang, Z. J.; DiPaolo, B. A.; Manninen, P. R.; Rohrer, D. C.; Hageman, M. J.; Hopkins, T. A.; Knechtel, M. L.; Oien, N. L.; Rush, B. D.; Schwende, F. J.; Stefanski, K. J.; Wieber, J. L.; Wilkinson, K. F.; Zamora, K. M.; Wathen, M. W.; Brideau, R. J. *Bioorg Med Chem Lett* 2007, *17*, 3840.
- Schnute, M. E.; Brideau, R. J.; Collier, S. A.; Cudahy, M. M.; Hopkins, T. A.; Knechtel, M. L.; Oien, N. L.; Sackett, R. S.; Scott, A.; Stephan, M. L.; Wathen, M. W.; Wieber, J. L. *Bioorg Med Chem Lett* 2008, *18*, 3856.
- Nieman, J. A.; Nair, S. K.; Heasley, S. E.; Schultz, B. L.; Zerth, H. M.; Nugent, R. A.; Chen, K.; Stephanski, K. J.; Hopkins, T. A.; Knechtel, M. L.; Oien, N. L.; Wieber, J. L.; Wathen, M. W. *Bioorg Med Chem Lett* 2010, 20, 3039.