

Convergent Synthesis of Potent Peptide Inhibitors of the Grb2-SH2 Domain by Palladium Catalyzed Coupling of a Terminal Alkyne

Joseph Schoepfer,* Brigitte Gay, Nicole End, Evelyne Muller, Gisela Scheffel, Giorgio Caravatti and Pascal Furet

Oncology Research, NOVARTIS Pharma AG, 4002 Basel, Switzerland

Received 22 December 2000; revised 27 February 2001; accepted 8 March 2001

Abstract—A new strategy was developed to prepare in a very efficient and convergent manner C-terminal modified tripeptides with high affinities for the Grb2-SH2 domain. Using Pd(PPh₃)₂Cl₂ as catalyst, selected naphthyl iodides and triflates were coupled to Ac-Pmp(t-Bu)₂-Ac₆c-Asn-NH(prop-2-ynyl). The resulting alkyne derivatives were hydrogenated and deprotected to afford potent Grb2-SH2 inhibitors. © 2001 Elsevier Science Ltd. All rights reserved.

The signal transduction pathways of tyrosine kinase growth factor receptors offer different possibilities of intervention in anticancer drug research. Besides targeting the kinase enzymatic activity of these receptors, 1,2 inhibiting events downstream in the signalling cascade constitutes another approach of potential interest in the search for new antitumor agents. A possible strategy in this direction is to block the interaction between the phosphotyrosine-containing activated receptors and the Src homology 2 (SH2) domain of the growth factor receptor-bound protein 2. Recent papers reported significant advances in the use of SH2 inhibitors in cancer therapy^{3–5} and osteoporosis.⁶ Phosphonopeptide 1, the corresponding prodrug,^{3,4} and the peptide 2 containing a malonic acid isostere of pTyr⁵ showed encouraging cellular activities (Fig. 1). Furthermore, combining Src-SH2 inhibition and the bone targeting properties of phosphonates, compound 3 displayed in vivo antiresorptive activity.6

The X-ray structure of the Grb2-SH2 domain in complex with a specific phosphopeptide ligand⁷ has revealed an extended hydrophobic area adjacent to the primary binding site. Therefore, hydrophobic residues were added to the terminal Asn of the minimal peptide sequence Ac-pTyr-X₊₁-Asn-NH₂ recognised by the

Continuing along this line, we have designed, synthesized, and tested C-terminal modified tripeptides [general structure Ac-Pmp-Ac₆c-Asn-NH-(3-naphthalenyl-propyl)] **4**. These compounds contain 4-(phosphonomethyl)phenylalanine (Pmp) as a stable pTyr

Figure 1. Examples of biologically profiled SH2 inhibitors.

Grb2-SH2 domain.^{8,9} This modification resulted in phosphopeptides with affinity and selectivity.

^{*}Corresponding author. Tel: +41-61-696-36-09; fax: +41-61-696-62-46; e-mail: joseph.schoepfer@pharma.novartis.com

replacement and maintain high affinities for the Grb2-SH2 domain.

Chemistry

A new strategy was developed to synthesize naphthalenyl peptides in a convergent and efficient way.

Starting from commercial BOC-Asn-OSu, the suitable alkyne amino acid **5** was obtained by reaction with propargylamine and subsequent TFA deprotection (Scheme 1). From the chloromethyl derivative **6**,¹⁰ reaction with deprotonated di-*tert*-butylphosphite and coupling of the resulting free acid with **5** afforded the common intermediate **7**.

The iodonaphthols **8b,c** were obtained by diazotation¹¹ of the commercially available aminonaphthols. The triflates **8d,e** were obtained by aromatization of the corresponding methoxy-tetralone with Pd(C) in either triisopropylbenzenze¹² or β -methylstyrene,¹³ followed by treatment with triflic anhydride in pyridine (Scheme 2).

Triflate **8f** was prepared the same way from 8-benzyloxy-naphthalen-2-ol, the major isomer obtained after alkylation of naphthalene-1,7-diol and chromatographic separation.

BOC-Asn-OSu
$$i, ii$$
 H_2N NH_2 N

Scheme 1. (i) Propargylamine, DMF, rt; (ii) TFA, 0°C; (iii) NaH, HOP(Ot-Bu)₂, DMF, rt; (iv) EDC, HOBT, NMM, 5, DMF, rt.

HO
$$\downarrow$$
NH₂
 \downarrow
NH

Scheme 2. (i) NaNO₂, HCl, KI, H₂SO₄, THF/H₂O; (ii) for precursor of 8d: Pd(C), triisopropylbenzene, 240 °C; for precursor of 8e: Pd(C), β-methylstyrene, 250 °C; (iii) benzylbromide, Cs₂CO₃, dioxan reflux; (iv) Tf₂O, pyridine, 0 °C.

The terminal alkyne of 7 was then coupled with the different naphthyl iodides or triflate using Sonogashira¹⁴ reaction conditions: Pd(PPh₃)₂Cl₂, copper iodide, and triethylamine in DMF; in the presence of tetrabutylammonium iodide¹⁵ (Scheme 3). The alkyne derivative was then hydrogenated, deprotected and purified with reverse phase medium pressure liquid chromatography.

Results and Discussion

Earlier results have already shown that targeting the hydrophobic patch was a successful strategy. 8,10 Introduction at the C-terminus of 3-naphthalenyl-propyl substituents, whose size and shape allow to bury a significant portion of the surface of the hydrophobic patch, led to high affinity peptides (Table 1). The introduction of hydroxyl and methoxy groups, in order to have electron-rich naphthols 16 afforded in the case of 4 f a compound with a significantly lower IC $_{50}$ value than 4 a.

The gain in activity obtained with **4f** can be ascribed to a favorable conformational restriction effect. Modeling suggests the possibility of an intramolecular hydrogen bond between the hydroxyl group of the naphthol and the C-terminal carbonyl of the compound. This internal hydrogen bond preorganizes the naphthol moiety in the conformation giving the most extended surface of contact with the hydrophobic patch, and thus burying the largest amount of surface (Fig. 2).

Conclusions

Structure base design in conjunction with a new synthetic strategy to incorporate C-terminal groups has allowed us to improve the Grb2-SH2 affinity of the Ac-Pmp-Ac₆c-Asn-NH-(3-naphthalenyl-propyl) compound class. Peptide 7 with a terminal alkyne group can be used for further exploration of the Grb2-SH2 hydrophobic patch.

Scheme 3. (i) 8, $Pd(PPh_3)_2Cl_2$, CuI, TBAI, NEt_3 , DMF, rt; (ii) Pd(C), H_2 , MeOH; (iii) TFA, $0\,^{\circ}C$.

Table 1. Starting naphthalenes and IC₅₀'s from Grb2-SH2 ELISA binding assay⁹

Compound	X = I 8a	8b X = I	X = I 8c	X = OTf 8d	X = OTf 8e	X = OTf, R = Bn 8f
Compound	X = Peptide 4a	4b	4c	4d	4e	R = H 4f
IC ₅₀ [nM]	42.9 ±2.0	24.6 ±2.9	42.7 ±2.9	129.9 ±5.9	84.4 ±5.0	11.1 ±04

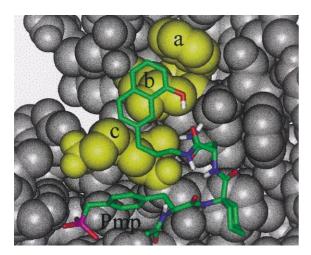


Figure 2. Compound **4f** minimized in the Grb2-SH2 binding site. Residues of the hydrophobic patch: $a = Phe \beta E3$, $b = Leu \beta D'1$, $c = Lys \beta D6$.

Acknowledgements

We gratefully acknowledge support for NMR spectroscopy from Dr. Joseph Schneider.

References and Notes

- 1. Garcia-Echeverria, C.; Traxler, P.; Evans, D. B. *Med. Res. Rev.* **2000**, *20*, 28.
- 2. Sedlacek, H. H. Drugs 2000, 59, 435.
- 3. Gay, B.; Suarez, S.; Caravatti, G.; Furet, P.; Meyer, T.; Schoepfer, J. *Int. J. Cancer* **1999**, *83*, 235.

- 4. Gay, B.; Suarez, S.; Weber, C.; Rahuel, J.; Fabbro, D.; Furet, P.; Caravatti, G.; Schoepfer, J. *J. Biol. Chem.* **1999**, 274, 23311.
- 5. Gao, Y.; Luo, J.; Yao, Z.-J.; Guo, R.; Zou, H.; Kelley, J.; Voigt, J. H.; Yang, D.; Burke, T. R. J. *J. Med. Chem.* **2000**, *43*, 911.
- 6. Shakespeare, W.; Yang, M.; Bohacek, R.; Cerasoli, F.; Stebbins, K.; Sundaramoorthi, R.; Azimioara, M.; Vu, C.; Pradeepan, S.; Metcalf, C., III; Haraldson, C.; Merry, T.; Dalgarno, D.; Narula, S.; Hatada, M.; Lu, X.; Van Schravendijk, M. R.; Adams, S.; Violette, S.; Smith, J.; Guan, W.; Bartlett, C.; Herson, J.; Iuliucci, J.; Weigele, M.; Sawyer, T. *Proc. Natl. Acad. Sci. U.S.A.* **2000**, *97*, 9373.
- 7. Rahuel, J.; Gay, B.; Erdmann, D.; Strauss, A.; Garcia-Echeverria, C.; Furet, P.; Caravatti, G.; Fretz, H.; Schoepfer, J.; Gruetter, M. G. *Nat. Struct. Biol.* **1996**, *3*, 586.
- 8. Furet, P.; Gay, B.; Caravatti, G.; Garcia-Echeverria, C.; Rahuel, J.; Schoepfer, J.; Fretz, H. J. Med. Chem. 1998, 41, 3442.
- 9. Furet, P.; Gay, B.; Garcia-Echeverria, C.; Rahuel, J.; Fretz, H.; Schoepfer, J.; Caravatti, G. *J. Med. Chem.* **1997**, *40*, 3551. 10. Schoepfer, J.; Fretz, H.; Gay, B.; Furet, P.; Garcia-Echeverria, C.; End, N.; Caravatti, G. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 221.
- 11. Snyckers, F.; Zollinger, H. Helv. Chim. Acta 1970, 53, 1294
- 12. Katritzky, A. R.; Balasubramanian, M.; Siskin, M. Org. Prep. Proced. Int. 1993, 25, 585.
- 13. JP Patent 58065235, 1983; Chem. Abstr. 1983, 99, 158060.
- 14. Sonogashira, K.; Tohda, Y.; Hagihara, N. Tetrahedron Lett. 1975, 4467.
- 15. Powell, N. A.; Rychnovsky, S. D. *Tetrahedron Lett.* **1996**, 37, 7901.
- 16. Previous work¹⁰ with indolyl analogues had suggested that electronic enrichment of the ring interacting with the hydrophobic patch was beneficial in terms of binding affinity.