

www.elsevier.nl/locate/carres

CARBOHYDRATE RESEARCH

Carbohydrate Research 319 (1999) 47-54

3,3'-Dideoxytrehaloses via dimerisation of 2-hydroxyglycal esters[☆]

Frieder W. Lichtenthaler *, Bernd Werner

Institut für Organische Chemie, Technische Universität Darmstadt, Petersenstraße 22, D-64287 Darmstadt, Germany Received 9 March 1999; accepted 27 April 1999

Abstract

The susceptibility of 2-hydroxyglycal esters i.e., 2,3,4,6-tetra-*O*-acetyl-1,5-anhydro-D-*arabino*-hex-1-enitol and its D-*lyxo* analog towards dimerization to 2,3-unsaturated α,α -trehaloses is described, smoothly occurring in moist dichloromethane or acetone solution in the presence of iodine or boron trifluoride. The reaction involves Ferrier ionization and addition of water at C-1 of the delocalized glycosyl oxocarbenium ion, followed by addition of the hex-2-enopyranose thus formed, with its 1-OH, onto the educt still present. Hydrogenation of the dimers thus formed, i.e., 2,4,6-tri-*O*-acetyl-3-deoxy- α -D-*erythro*-hex-2-enopyranosyl 2,4,6-tri-*O*-acetyl-3-deoxy- α -D-*erythro*-hex-2

Keywords: 3,3'-Dideoxy-a,a-trehaloses; 2-Hydroxyglycal esters; Allylic rearrangement; Dihydropyrans

1. Introduction

Aside from glycals, which have extensively been exploited as precursors for a variety of naturally occurring products [2], the 2-acyloxyglycals of type **1** and **2**, although just as accessible [3], have found considerably less attention. In fact, their unique enolestermasked carbonyl group at C-2 adds a structural function that considerably extends the variability of their ensuing reactions [4], and, hence, their utility as enantiopure building blocks. Thus, the C-2 carbonyl group in the D-glucose-derived 2-acetoxy- and 2-benzoyloxy-glucal esters 1 and 2 can be liberated by N-bromosuccinimide-induced methanolysis to afford ulosyl bromides of type 3 [5] that proved to be particularly efficient 'indirect' β -D-mannosyl donors [5,6]. Similar potential as 'indirect' β-D-mannosaminyl donors [7] applies to the oximinoglycosulosyl bromides 5, smoothly derived from 1 and 2 by a highyielding 3-step procedure involving hydroxylaminolysis $(\rightarrow 4 \ [8])$, benzoylation, and photobromination [9,10]. Other important hydroxyglycal ester transformations involve Lewis acid-induced allylic rearrangements to 3-deoxy-hex-2-enopyranoses by intra- or intermolecular addition at the anomeric center of an acetoxy group $(1 \rightarrow 6 [11])$, of alkoxy residues $(1 \rightarrow 7 [12])$, or of an acylperoxy group, which by fragmentation of the primary adduct leads to pyranoid enollactones ($\rightarrow 8$ [13]) (Scheme 1).

^{*} Enantiopure building blocks from sugars, Part 25. For Part 24, see Ref. [1].

^{*} Corresponding author. Tel.: + 49-6151-162-376; fax: + 49-6151-166-674.

E-mail address: fwlicht@sugar.oc.chemie.tu-darmstadt.de (F.W. Lichtenthaler)



Scheme 1.

Described herein is a useful addition to this multifaceted array of versatile building blocks, namely the Lewis acid-promoted allylic rearrangement of 2-acetoxyglucal triacetate **1** and its galactose-derived 4-epimeric analog **16** and subsequent dimerization to 2,3:2',3'-unsaturated trehalose-type dimers (**10** and **18**). This reaction, which has its analogy in the Lewis acid-induced dimerization of tri-*O*-acetyl-1,5-anhydro-2-deoxy-D-*arabino*-hex-1-enitol (tri-*O*-acetyl-D-glucal) [14], opened up the way to 3,3'-dideoxy- α , α ,-trehalose analogs that are of interest as trehalase inhibitors.

2. Results and discussion

2,3,4,6-Tetra-O-acetyl-1,5-anhydro-D-arabino-hex-1-enitol (1) in boiling acetic acid [11a] or cold acetic anhydride in the presence of zinc chloride [11b], or in benzene with catalytic amounts of boron trifluoride etherate [11c], undergoes Ferrier rearrangement to an anomeric mixture of 3-deoxyhex-2-enopyranoses in which the α anomer **6** is the preponderant product. When the solvents used are not absolutely dry, or by the deliberate addition of small amounts of water, the Ferrier rearrangement is followed by further reactions, the respective products then depending on the conditions used. Exposure of 1 in dichloromethane solution to borotrifluoride diethyl etherate in the presence of water rapidly forms a mixture of three main components, characterized as acetoxymethylfurfural (13), the α,α dimer 10, and the bis-enone 14. Their formation can be readily rationalized by 14 arising from 10 via hydrolytic cleavage of the enolester and subsequent elimination of the 4-acetoxy group, whereas 13 is an ensuing product of the intermediate hexenopyranose 9 (Scheme 2).

Evaluation of various other catalyst/solvent combinations, e.g., zinc chloride/dichloromethane, N-iodosuccinimide or iodine/acetonitrile, eventually led to iodine/acetone in the presence of water as the most suitable conditions for the conversion $1 \rightarrow 10$:





the reaction is rapid (30 min, 0 °C) and allows the isolation of 10 in 58% yield. Saturation of the two enolester double bonds was readily effected only when hydrogenation was performed over 5% rhodium on carbon in ethyl acetate, providing the 3,3'-dideoxy-trehalose in the form of its peracetate 11 (75% isol. yield), or in its free form 12 following standard O-deacetylation. Over Pd, Pd/C and Pt/ hydrogenation of С catalysts. the 10 proceeded sluggishly and stereochemically non-uniformly, whereas the use of rhodium on carbon catalyst and educt in a 1:1 ratio led to some reductive deacetoxylation as evidenced by the isolation of 8% of the 2,3:3'-trideoxy derivative 15.

This simple 3-step sequence for the conversion of 1 into a 3,3'-dideoxy- α,α -trehalose, i.e., $1 \rightarrow 10 \rightarrow 11 \rightarrow 12$, appears to be generally applicable, since the respective D-*lyxo*-hex-1-enitol derivative 16 was also readily converted into the D-*threo*/D-*threo*-trehalose analog 18. Interestingly, the 2-acetoxy-D-galactal 16 was considerably less reactive than its D-glucal counterpart—a reactivity difference that has previously been observed [15] in the acid-induced Ferrier rearrangement to the hex-2-enopyranose tetraacetate (4-epimer of 6). Thus, exposure of 16 to iodine in aqueous acetone had no effect; under somewhat more forcing conditions, i.e., boron trifluoride in

dichloromethane containing water, however, the conversion was readily achieved to yield (53%) the nicely crystalline D-threo/D-threo dimer **18**; it was hydrogenated with remarkable stereoselectivity (84% isol. yield) to the equally crystalline D-lyxo/D-lyxo-3-3'-dideoxy trehalose hexaacetate **20** (Scheme 3).

It is interesting to note that the coupling of the α -D-hexenopyranose intermediate 17 with unreacted 16, i.e., $16 + 17 \rightarrow 18$, cannot be intercepted by other reagents, e.g., by *m*chloroperbenzoic acid to form the enollactone 20. Only when the BF₃-mediated peroxidation of 16 was effected under strictly anhydrous conditions requiring the removal of water from the commercially available MCPBA, a high yield (90%) of the enollactone could be obtained—obviously via fragmentation of the 1-peroxyacyl-hexenopyranose intermediate 19 [13].

Although configurational assignments could readily be inferred from ¹H and ¹³C NMR data, the high crystallinity of the D-galactalderived dimer **18** invited structural verification by X-ray crystallography. As depicted in Fig. 1, the molecular geometry of **18** clearly unveils the two *O*-linked dihydropyrane rings to be in a ^OH₅ halfchair conformation, in which the ring oxygen is slightly more above the plane formed by carbons 1–4 than C-5 is below (0.36 vs. 0.32 Å). The bond lengths and bond





angles are within the expected ranges [16], the dihedral angles in the two pyranoid rings differ only slightly from each other (cf. Table 1, forms A and B, respectively), and, moreover are in very close agreement to those found for the α -D-*threo*-hex-2-enopyranose analogs **22** [17] and **23** [18]. The C-4–C-5–O-5–C-1 dihedral angles are uniformly in the 64° range, evidence for an essentially identical ${}^{\rm O}H_5$ conformation of **18**, **22** and **23**.

3. Experimental

General methods.—¹H (300 MHz) and ¹³C (75 MHz) NMR spectra were recorded at 25 °C with a Bruker AC 300 spectrometer. Chemical shifts (δ) are given in ppm relative to the signal for internal Me₄Si (CDCl₃). Column chromatography was performed on Kieselgel 60 (E. Merck, 230 mesh) and fractions were monitored by TLC on Kieselgel 60 F₂₅₄ (E. Merck) by detection with UV light and then charring with H₂SO₄. Optical rotations were measured for solutions in CHCl₃ at 20 °C with a Perkin–Elmer 241 polarimeter, using a 10 cm/1 mL cell.

2,4,6-*Tri*-O-*acetyl*-3-*deoxy*-α-D-erythro-*hex*-2-enopyranosyl 2,4,6-*tri*-O-*acetyl*-3-*deoxy*-α-D-erythro-*hex*-2-enopyranoside (**10**).—A solu-

tion of 2,3,4,6-tetra-*O*-acetyl-1,5-anhydro-Darabino-hex-1-enitol (1, 3.96 g, 12 mmol) [3e] in dry acetone (8 mL) was cooled (0 °C), iodine (3.05 g, 12 mmol) and water (0.12 mL, 8 mmol) was added, and the mixture



Fig. 1. Perspective drawing of the molecular structure of 2,4,6-tri-O-acetyl-3-deoxy- α -D-*threo*-hex-2-enopyranosyl 2,4,6-tri-O-acetyl-3-deoxy- α -D-*threo*-hex-2-enopyranoside (**17**), revealing a $^{O}H_{5}$ halfchair conformation for each of the pyranoid rings.

Table 1

Selected torsion angles (°) for **18**, and the monomeric analogs methyl 4,6-di-*O*-acetyl-2,3-dideoxy- α -D-*threo*-hex-2-enopyranoside (**22**) [17] and 1,4,6-tri-*O*-acetyl-2-(*N*,*N*-diacetylamido)-2,3-dideoxy- α -D-*threo*-hex-2-enopyranose (**23**) [18]

Dihedral angels	18		Aco OAc Aco OAc OAc	
	A	В	22 ^a	23 b
Pyranoid ring O-5-C-1-C-2-C-3 C-1-C-2-C-3-C-4 C-2-C-3-C-4-C-5 C-3-C-4-C-5-O-5 C-4-C-5-O-5-C-1 C-5-O-5-C-1-C-2	+12.2 +3.6 +12.8 -44.4 +64.4 -46.1	+14.4 +1.9 +12.1 -42.3 +62.6 -46.5	+13.2 +1.2 +14.8 -46.6 +64.9 -46.8	+14.4 +6.4 +7.1 -40.5 +64.1 -49.3
Glycosidic linkage O-5–C-1–O-1–C-1'	+74.5	+76.5	+67.0	+76.8
Substituent angles C-2-C-3-C-4-O-4 O-4-C-4-C-5-C-6 O-4-C-4-C-5-O-5	-107.8 -41.3 +76.0	-108.6 -38.7 +79.6	-102.4 -53.2 +68.7	$-113.0 \\ -39.5 \\ +80.8$
Cremer–Pople puckering Q Θ Φ Conformation	parameters [20] 0.466 52.4 328.1 $^{\circ}H_{5}$	0.444 51.0 331.9 $^{\circ}H_{5}$	0.466 51.3 326.0 $^{\circ}H_{5}$	$ \begin{array}{c} 0.475 \\ 55.1 \\ 336.0 \\ ^{\rm O}H_5(\rightarrow {}^{\rm O}E) \end{array} $

^a Dihedral angles not given in Ref. [17] and the Cremer–Pople parameters were calculated by S. Immel from the original coordinates (REFC THHXPY).

^b Cremer-Pople parameters were calculated by S. Immel from the original coordinates (REFC AAXTHP).

was stirred at 0 °C for 30 min. The reaction quenched with satd. was $Na_2S_2O_3$ /satd. NaHCO₃ solution (3:1, 40 mL), extracted with CH₂Cl₂ followed by consecutive washing of the organic phase with water (30 mL), satd. NaHCO₃ $(2 \times 20 \text{ mL})$ and water (30 mL), drying $(MgSO_4)$, and evaporation to dryness. Purification of the syrupy residue by elution from a silica gel column $(3 \times 30 \text{ cm})$ with 4:1 toluene-EtOAc afforded, after removal of the solvents in vacuo, 1.95 g (58%) of 10 as a colorless syrup; $R_f 0.45$ in 1:1 toluene–EtOAc; $[\alpha]_{D}^{20} + 64.6^{\circ}$ (c 1.1, CHCl₃); ¹H NMR (CDCl₃): δ 5.83 (d, 1 H, H-3), 5.47 (ddd, 1 H, H-4), 5.42 (bs, 1 H, H-1), 4.26 (dd, 1 H, H-6b), 4.13 (m, 2 H, H-5, H-6a), 2.18, 2.10, 2.09 (three 3 H-s, AcCH₃), $J_{1,4} \sim 0.7$, $J_{3,4}$ 2.0, $J_{4.5}$ 9.5, $J_{5.6a} \sim 2.6$, $J_{5.6b}$ 12.0 Hz. ¹³C NMR $(CDCl_3)$: δ 170.6 (3 AcCO), 169.9, 167.8, 145.6 (C-2), 115.8 (C-3), 89.4 (C-1), 67.5 (C-5), 65.3 (C-4), 62.5 (C-6), 20.8, 20.8, 20.6 (3 CH₃); FD MS: m/z 581 [M⁺ + Na]. Anal.

Calcd for $C_{24}H_{30}O_{15}$ (558.5): C, 51.60; H, 5.41. Found: C, 51.41; H, 5.32.

When performing the reaction of 1 in CH₂Cl₂ (990 mg in 20 mL) with BF₃·Et₂O (0.6 mL) in the presence of water (0.1 mL) by stirring at ambient temperature for 1 h, an approximate 2:1:1 mixture of dienone 14, α, α dimer 10, and furfural 13 (R_c 0.6, 0.45 and 0.4, resp., in 1:1 toluene-EtOAc), which, on workup as described above, could be separated on a silica gel column by elution with 4:1 toluene-EtOAc. The product eluted first was secured as a colorless oil (55 mg, 11 %) to be 5-acetoxymethyl-2and proved furanaldehyde (13) by identity of its ¹H NMR data with that of an authentic sample (from Sigma–Aldrich Chemie). The fraction closely following was rich in 10 (TLC, ¹H NMR). The dienone 14, eluted last, gave upon evaporation of the appropriate fraction, 125 mg (23 %) of a colorless syrup; ¹H NMR (300 MHz, $CDCl_3$): δ 6.98 (dd, 1 H, H-4), 6.18 (dd, 1 H, H-3), 5.28 (s, 1 H, H-1), 4.83 (ddd, 1 H, H-5), 4.39 and 4.28 (two 1 H-dd, two H-6), 2.10 (s, 3 H, AcCH₃), $J_{3,4}$ 10.7, $J_{3,5}$ 2.4, $J_{4,5}$ 1.7, $J_{5,6a}$ 4.5, $J_{5,6b}$ 5.1, $J_{6,6}$ 11.8 Hz; FD MS : m/z + 377 [M + Na⁺].

Hexa-O-acetyl-3,3'-dideoxy- α,α -trehalose (2,4,6-*Tri*-O-*acetyl*-3-*deoxy*-α-D-ribo-*hexopy*ranosyl 2,4,6-tri-O-acetyl-3-deoxy- α -D-ribohexopyranoside) (11).—A solution of erythrodisaccharide 10 (340 mg, 0.6 mmol) in EtOAc (10 mL) was hydrogenated over 100 mg of 5 % rhodium on charcoal at ambient temperature for 20 h. The solution was then filtered over Celite and the residue was washed with EtOAc (80 mL), followed by removal of the solvent in vacuo. The resulting syrup was subjected to purification on a silica gel column $(2 \times 30 \text{ cm})$ with 9:1 CH₂Cl₂-EtOAc. The eluates carrying 11 (R_f 0.38 in 1:1 toluene-EtOAc) were collected and evaporated to dryness, 260 mg (75 %) of a colorless syrup of $[\alpha]_{D}^{20} + 132 (c 1, CHCl_{3}); {}^{1}H NMR (300 MHz,$ $CDCl_3$): δ 5.22 (d, 1 H, H-1), 4.96 (ddd, 1 H, H-2), 4.83 (ddd, 1 H, H-4), 4.20 and 4.08 (two 1 H-dd, two H-6), 3.98 (ddd, 1 H, H-5), 2.36 (ddd, 1 H, H-3e), 2.08 (6 H, 3 AcCH₃) and 2.07 (3 H), 2.05 (m, 1 H, H-3a); $J_{1,2}$ 3.4, $J_{2,3a}$ 12.3, $J_{2,3e}$ 4.6, $J_{3a,4}$ 10.8, $J_{3e,4}$ 4.8, $J_{4,5}$ 10.5, $J_{5,6a}$ 2.1, $J_{5,6b}$ 5.9, $J_{6,6}$ 12.1 Hz; ¹³C NMR (CDCl₃): δ 170.8 (3 C=O), 169.6, 169.6, 90.9 (C-1), 68.6 (C-5), 67.4 (C-2), 65.9 (C-4), 62.3 (C-6), 28.9 (C-3), 20.9 (3 CH₃), 20.7, 20.7; FD MS: m/z563 [MH⁺], 562 [M⁺], 273 [tri-O-acetyl-3deoxyglycosyl⁺]. Anal. Calcd for $C_{24}H_{34}O_{15}$ (562.5): C, 51.24; H, 6.09. Found: C, 51.09; H, 6.02.

4,6-Di-O-acetyl-2,3-dideoxy- α -D-erythrohexopyranosyl 2,4,6-tri-O-acetyl-3-deoxy- α -Dribo-hexopyranoside (15).—When using larger amounts of catalyst in the hydrogenation of 10 as described above, e.g., a 1:1 ratio in mg, a minor product was detectable in the reaction mixture ($R_c 0.43$ in 1:1 toluene–EtOAc) which could be eluted separately from a column and was obtained as a colorless syrup (28 mg, 8 %), which proved to be the 4-deoxygenation product **15**; $[\alpha]_D^{20} + 114^\circ$ (*c* 1.3, CHCl₃); ¹H NMR (300 MHz, CDCl₃): (a) erythro part: δ 5.16 (broadened s, 1 H, H-1), 4.73 (ddd, 1 H, H-4), 4.20 and 4.06 (1 H-dd each, two H-6), 3.95 (ddd, 1 H, H-5), 2.06–1.90 (7 H-m, 2 AcCH₃, H-3b), 1.83 (3 H-m, two H-2, H-3a),

 $J_{1,2} \leq 0.5, J_{4,5} = 10.0, J_{5,6} = 2.1 \text{ and } 6.1, J_{6,6} = 11.7$ Hz; (b) *ribo* part: $\delta = 5.24$ (d, 1 H, H-1), 4.96 (ddd, 1 H, H-2), 4.86 (ddd, 1 H, H-4), 4.24 and 4.10 (two 1 H-dd, two H-6), 3.89 (ddd, 1 H, H-5), 2.32 (ddd, 1 H, H-3b), 2.05 (10 H-m, H-3a, 3 AcCH₃); $J_{1,2} = 3.5, J_{2,3a} = 4.7, J_{2,3b} = 12.4, J_{3a,4} = 4.9, J_{3b,4} = 10.8, J_{4,5} = 10.1, J_{5,6a} = 2.4, J_{5,6b} = 5.2, J_{6,6} = 11.8$ Hz; FD MS: m/z = 505 [MH⁺], 504 [M⁺].

3,3' - Dideoxy - α, α - trehalose (3 - deoxy - α - Dribo-hexopyranosyl 3-deoxy- α -D-ribo-hexopyranoside) (12).—To a solution of hexaacetate 11 (170 mg, 0.3 mmol) in MeOH (5 mL) was added 0.33 mL of N methanolic MeONa and the mixture was kept at ambient temperature for 1 h, then quenched with AcOH (1 mL), and evaporated to dryness. Purification of the residue by elution from a silica gel column (1.5 \times 20 cm) with 2:1 CHCl₃–MeOH gave, after evaporation of the appropriate fraction, 83 mg (86 %) of 12 as a chromatographically uniform solid; R_f 0.34 in 2:1 CHCl₃–MeOH; $[\alpha]_{D}^{20}$ + 161° (*c* 1.3, MeOH); ¹H NMR (300 MHz, MeOH- d_4): δ 5.08 (d, 1 H, H-1), 3.8–3.5 (unresolved m, 5 H, H-2, H-4, H-5, and two H-6), 2.09 (m, 1 H, H-3b), 1.94 (m, 1 H, H-3a), $J_{1,2}$ 3.4 Hz; ¹³C NMR (75.5 MHz, MeOH- d_4): δ 92.7 (C-1), 70.4 (C-5), 67.4 (C-2), 65.3 (C-4), 61.8 (C-6), 35.3 (C-3); FD MS: m/z 333 [M⁺ + Na], 310 [M⁺], 293 [M⁺-OH]. Anal. Calcd for C₁₂H₂₂O₉ (310.3): C, 46.45; H, 7.15. Found: C, 46.35; H, 7.03.

2,4,6-Tri-O-acetyl-3-deoxy- α -D-threo-hex-2-enopyranosyl 2,4,6-tri-O-acetyl-3-deoxy-α-D-threo-hex-2-enopyranoside (18).—A solution of 2,3,4,6-tetra-O-acetyl-1,5-anhydro-Dlyxo-hex-1-enitol (16, 660 mg, 2 mmol) [3e,15,19] in CH₂Cl₂ (10 mL) was stirred with BF_3 ·Et₂O (803 µL, 20 mmol) for about 5 min at room temperature. Water (360 µL, 20 mmol) was then added, followed by stirring for another 5 min and quenching the reaction by pouring into satd aq NaHCO₃ (20 mL). The mixture was diluted with CH₂Cl₂ (50 mL), and the organic phase was separated, dried (MgSO₄), evaporated to dryness, and the syrupy residue was purified by elution from a silica gel column $(1.5 \times 25 \text{ cm})$ with 9:1 CH₂Cl₂-EtOAc. Removal of the solvents gave a syrup, which crystallized from t-BuOMe/npentane: 290 mg (53%) of 18 as colorless prisms; mp 156.0–156.5 °C; $[\alpha]_D^{20}$ –194.8° (c 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 6.06 (d, 1 H, H-3), 5.53 (s, 1 H, H-1), 5.31 (ddd, 1 H, H-4), 4.39–4.32 (m, 2 H, H-5, H-6b), 4.22 (ddd, 1 H, H-6a), 2.24 (3 s, 3 H each, 3 AcCH₃), 2.14, 2.12; $J_{3,4}$ 6.1, $J_{4,5}$ 2.2, $J_{4,6a} \sim 1.0$, $J_{5,6a}$ 3.6, $J_{6a,b}$ 9.8 Hz; ¹³C NMR (75.5 MHz, CDCl₃): δ 170.4, 170.1, 167.7 (3 AcCO), 148.6 (C-2), 112.4 (C-3), 88.6 (C-1), 67.0 (C-5), 64.0 (C-4), 62.2 (C-6), 20.8 (3 AcCH₃), 20.7, 20.6; FI MS: m/z 558 [M⁺]. Anal. Calcd for C₂₄H₃₀O₁₅ (558.49): C 51.60; H 5.41. Found: C 51.58; H 5.35.

The crystal of 18, subjected to X-ray analysis, had the dimensions $0.55 \times 0.375 \times 0.175$ mm, was monoclinic, space group $P2_1$ with a = 7.859 (4), b = 22.401 (7), and c = 8.092 (4) Å, $\alpha = \gamma = 90^{\circ}$, $\beta = 108.16$ (4)°, V = 1353.1 Å³, Z = 2, T = 298 K, μ (Mo K α) = 0.11 mm⁻¹, and Dc = 1.37 g cm⁻³. A total of 2444 reflections were collected on a Enraf-Nonius CAD4 diffractometer, equipped with a graphite-monochromated Mo K α ($\lambda = 0.71093$ A) radiation using 1928 independent reflections to refine the complete structure, i.e., R(F) 0.0357 for 1860 reflections with $I \ge 2\sigma I$, $wR(F^2)$, and 0.0961 for all 1928 reflections $(w = 1/(\sigma^2(F_0^2) + (0.0725)^2 + 0.248P); P = (F_0^2)^2$ $(-2F_c^2)/3$. All non-hydrogen atoms were refined anisotropically. Hydrogen atoms on the molecule were considered in calculated positions with the $1.2 \times U_{eq}$ value of the corresponding atom. Data reduction was done by a Stoe-REDU4 program, structural resolution and refinement by SHELXS-86 and SHELXL-93 [21]. For atomic coordinates and further details, see Section 4.

2,4,6-*Tri*-O-*acetyl*-3-*deoxy*- α -D-lyxo-*hex*opyranosyl 2,4,6-tri-O-acetyl-3-deoxy- α -Dlyxo-*hexopyranoside* (21).—In а hydrogenation vessel, rhodium on carbon (5 %, 30 mg) was added to an EtOAc solution of *threo*/ threo-disaccharide 18 (140 mg, 0.25 mmol, in 15 mL), and the mixture was hydrogenated over H_2 for 16 h. Subsequent filtration through Kieselgur, thorough washing with EtOAc and removal of the solvent in vacuo left a syrup which crystallized from 2propanol: 120 mg (84 %) of 20 as colorless, waddy needles; mp 201 °C; $[\alpha] + 48.9^{\circ}$ (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 5.21 (bs, 1 H, H-1), 4.90 (bs, 1 H, H-4), 4.75 (bs, 1 H, H-2), 4.26–4.13 (m, 3 H, H-5, two 6-H), 2.38 (brm,

1 H, H-3b), 2.16 (m, 1 H, H-3a), 2.11, 2.10, 2.05 (three 3 H-s, 3 AcCH₃); $J_{1,2} < 0.5$ Hz; ¹³C NMR (CDCl₃): δ 170.6, 170.3, 169.9 (3 AcCO), 91.6 (C-1), 67.5 (C-5), 66.0 (C-2), 64.8 (C-4), 63.0 (C-6), 26.7 (C-3), 21.0, 21.0, 20.6 (3 AcCH₃); FD MS: m/z 562 [M⁺-H]. Anal. Calcd for C₂₄H₃₄O₁₅ (562.5): C, 51.24; H, 6.09. Found: C, 51.19; H, 6.15.

2,4,6-Tri-O-acetyl-3-deoxy-D-threo-hex-2enono-1,5-lactone [(5R,6R)-3,5-bis(acetoxy)-6-acetoxymethyl-5,6-dihydro-2H-pyran-2-one] (20).—A solution of 1.0 g (3 mmol) of 16 [15] in CH_2Cl_2 (10 mL) was stirred for 20 min at ambient temperature, then cooled to -20 °C and followed by the addition of BF₃·Et₂O (267 μ L, 2.2 mmol) and of a solution of anhydrous, 85 % *m*-chloroperbenzoic acid¹ in CH₂Cl₂ (600 mg, 3.5 mmol, in 6 mL). After vigorous stirring for 15 min at -20 °C, the mixture was quenched by pouring into satd. NaHCO₃ solution (15 mL) containing 10-20 mg of $Na_2S_2O_3$ (i.e., without prior warming to ambient temperature). The organic phase is removed, and the aqueous layer is extracted with CH_2Cl_2 (2 × 15 mL), followed by concentration of the combined organic phases in vacuo to a syrup, which was purified by elution from a silica gel column (2×25 cm) with 2:1 toluene-EtOAc. Removal of the solvents from the eluates with R_f 0.30 (TLC in 2:1) toluene-EtOAc) gave 786 mg (90 %) of enelactone **20** as a colorless syrup; $[\alpha]_{D}^{20}$ -151.5° (c 1, CHCl₃); lit. [22] : $[\alpha]_{D}^{25}$ -150° (c 0.5, CHCl₃); ¹H NMR (CDCl₃): δ 6.64 (d, 1 H, H-3), 5.44 (dd, 1 H, H-4), 4.93 (dt, 1 H, H-5), 4.36 (d, 2 H, 7-H₂), 2.20 (three 3 H-s, 3 AcC H_3), 2.04, 2.03, $J_{4,5}$ 6.4, $J_{5,6}$ 2.6, $J_{6,7}$ 6.3 Hz; ¹³C NMR (CDCl₃): δ 170.5, 169.8, 168.1

¹ The *m*-chloroperbenzoic acid used was the commercially available anhydrous 85% grade, i.e., containing 15% mchlorobenzoic acid. Since the overall reaction requires the BF₃-induced generation of an allylcarboxonium ion intermediate subsequently undergoing peroxidation, conditions have to be strictly anhydrous (e.g., dry CH₂Cl₂ as the solvent) in order to prevent hydrolysis of the Lewis acid. By consequence, it is essential that lower grade MCPBA (e.g., 55% with 30-35% of water and 10-15% 3-chlorobenzoic acid) is freed from water before use. This is readily effected by dissolving 55% MCPBA in CH₂Cl₂ (5 g in 100 mL of freshly distilled solvent to exclude the presence of small amounts of HCl), separating the aqueous layer, drying of the organic phase (Na_2SO_4) , and removal of the solvent in vacuo (bath temp. below 35 °C) to yield a fluffy solid that can directly be used.

(3 AcCO), 157.6 (C-1), 141.7 (C-2), 124.6 (C-3), 76.2 (C-5), 63.0 (C-4), 61.3 (C-6), 20.6, 20.4, 20.3 (3 AcCH₃); FI MS: m/z = 287 [MH⁺], 286 [M⁺]. Anal. Calcd for C₁₂H₁₄O₈ (286.2): C, 50.37; H, 4.91. Found: C, 50.33; H, 4.96.

4. Supplementary material

Tables of atomic coordinates, bond lengths and bond angles have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication no. CCDC-121452. Copies of this data can be obtained, free of charge, from CCDC, 12 Union Road, Cambridge, CB2 1EZ, UK (fax: +44-1223-336-033 or e-mail: deposit@ccdc.cam.ac.uk).

Acknowledgements

The authors are indebted to the Fonds der Chemischen Industrie for financial support and to Mrs S. Foro and Professor Dr H.J. Lindner for determining the X-ray structure of 18.

References

- [1] F.W. Lichtenthaler, Carbohydr. Res., 313 (1998) 69-90.
- [2] S.J. Danishefsky, M.T. Bilodeau, Angew. Chem., Int. Ed. Engl. 35 (1996) 1380-1419, and lit. cited therein.
- [3] (a) K. Maurer, Ber. Dtsch. Chem. Ges., 62 (1929) 332–338. (b) K. Maurer, W. Petsch, Ber. Dtsch. Chem. Ges., 66 (1933) 995–1000. (c) M.G. Blair, Methods Carbohydr. Chem., 2 (1963) 411–414. (d) R.J. Ferrier, G.H. Sankey, J. Chem. Soc. C, (1966) 2339–2345. (e) D.R. Rao, L.M. Lerner, Carbohydr. Res., 22 (1972) 345–350.
 [4] For a review see: F.W. Lichtenthaler, in R. Scheffold
- [4] For a review see: F.W. Lichtenthaler, in R. Scheffold (Ed.), *Modern Synthetic Methods*, Vol. 6, VCH, Weinheim, 1992, pp. 273–376.
- [5] (a) F.W. Lichtenthaler, U. Kläres, M. Lergenmüller, S. Schwidetzky, *Synthesis*, (1992) 179–184. (b) F.W. Lichtenthaler, T. Schneider-Adams, *J. Org. Chem.*, 59 (1994) 6728–6734.
- [6] F.W. Lichtenthaler, T. Schneider-Adams, S. Immel, J. Org. Chem., 59 (1994) 6735–6738. (b) F.W. Lichtenthaler, U. Kläres, Z. Szurmai, B. Werner, Carbohydr. Res., 305 (1998) 293–303.

- [7] (a) F.W. Lichtenthaler, E. Kaji, S. Weprek, J. Org. Chem., 50 (1985) 3505–3515. (b) E. Kaji, F.W. Lichtenthaler, T. Nishino, Y. Yamane, S. Zen, Bull. Chem. Soc. Jpn., 61 (1988) 1291–1297. (c) E. Kaji, F.W. Lichtenthaler, Y. Osa, K. Takahashi, S. Zen, Chem. Lett., (1992) 707–710; Bull. Chem. Soc. Jpn., 68 (1995) 2401–2408. (d) E. Kaji, Y. Osa, K. Takahashi, M. Hirooka, S. Zen, F.W. Lichtenthaler, Bull. Chem. Soc. Jpn., 67 (1994) 1130–1140.
- [8] (a) F.W. Lichtenthaler, E.S.H. El Ashry, V.H. Göckel, *Tetrahedron Lett.*, 21 (1980) 1429–1432. (b) P. Jarglis, F.W. Lichtenthaler, *Tetrahedron Lett.*, 21 (1980) 1425– 1428; *Angew. Chem.*, 94 (1982) 140–141; *Angew. Chem.*, *Int. Ed. Engl.*, 21 (1982) 141–142; *Angew. Chem. Suppl.*, (1982) 175–183.
- [9] F.W. Lichtenthaler, P. Jarglis, W. Hempe, *Liebigs Ann. Chem.*, (1983) 1959–1972.
- [10] Reviews: (a) E. Kaji, F.W. Lichtenthaler, *Trends Glycosci. Glycotechnol.*, 5 (1993) 121–142; (b) F. Barresi, O. Hindsgaul, in S.H. Khan, R.A. O'Neill (Eds.), *Modern Methods in Carbohydrate Synthesis*, Harwood, Amsterdam, 1996, pp. 251–276.
- [11] (a) R.J. Ferrier, W.G. Overend, G.H. Sankey, J. Chem. Soc. C, (1965) 2830–2836. (b) R.U. Lemieux, D.R. Lineback, M.L. Wolfrom, F.B. Moody, E.G. Wallace, F. Komitsky, Jr., J. Org. Chem., 30 (1965) 1092–1096. (c) R.J. Ferrier, N. Prasad, G.H. Sankey, J. Chem. Soc. C, (1968) 974–977.
- [12] (a) R.J. Ferrier, N. Prasad, G.H. Sankey, J. Chem. Soc. C, (1969) 587–591. (b) S. Hanessian, P.C. Tyler, Y. Chapleur, *Tetrahedron Lett.*, 22 (1981) 4583–4586. (c) Y. Ichikawa, M. Isobe, H. Masaki, T. Kawai, T. Goto, C. Katayama, *Tetrahedron*, 43 (1987) 4759–4766. (d) O. Varela, G.M. de Fina, R.M. de Lederkremer, *Carbohydr. Res.*, 167 (1987) 187–196.
- [13] F.W. Lichtenthaler, S. Rönninger, P. Jarglis, *Liebigs Ann. Chem.*, (1989) 1153–1161.
- [14] (a) H.H. Baer, L. Siemsen, J. Defaye, K. Burak, *Carbohydr. Res.*, 134 (1984) 49–61. (b) H.P. Wessel, G. Englert, *J. Carbohydr. Chem.*, 14 (1995) 179–196.
- [15] R.J. Ferrier, G.H. Sankey, J. Chem. Soc. C, (1966) 2339–2345.
- [16] G.A. Jeffrey, A.D. French, *Mol. Struct. Diffr. Methods*, 6 (1978) 183–206.
- [17] J.W. Krajewski, Z. Urbanczyk-Lipkowska, P. Gluzinski, J. Bleidelis, A. Kemme, *Acta Cryst.*, *Sect. B*, 35 (1979) 2625–2629.
- [18] B. Kojic-Prodic, Z. Ruzic-Toros, Acta Cryst., Sect. B, 32 (1976) 1833–1838.
- [19] (a) K. Maurer, H. Mahn, Ber. Dtsch. Chem. Ges., 60 (1927) 1316–1320. (b) K. Maurer, A. Müller, Ber. Dtsch. Chem. Ges., 63 (1930) 2069–2073.
- [20] (a) D. Cremer, J.A. Pople, J. Am. Chem. Soc., 97 (1975) 1354–1358. (b) G.A. Jeffrey, R. Taylor, Carbohydr. Res., 81 (1980) 182–183.
- [21] G.M. Sheldrick, SHELX-86 and SHELX-93 Programs for Crystal Structure Determination, University of Göttingen, 1986 and 1993.
- [22] J. Kovács, I. Pintér, M. Kajtár-Peredy, L. Somsák, *Te-trahedron*, 53 (1997) 15041–15050.