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Inhibitory Potency of 5-Benzyluracil, 5-Phenylcytosine and 5-Phenylpyrimidin-2one Nucleosides Against Uridine Phosphorylase from Mouse Leukemic L1210 Cells

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INHIBITORY POTENCY OF 5-BENZYLURACIL, 5-PHENYLCYTOSINE AND 5-PHENYLPYRIMIDIN-2-ONE NUCLEOSIDES AGAINST URIDINE PHOSPHORYLASE FROM MOUSE LEUKEMIC L1210 CELLS

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ABSTRACT: The inhibitory activity of a series of novel sugar-modified nucleosides derived from 5-benzyluracil, 5-phenylcytosine and 5-phenylpyrimidin-2-one against uridine phosphorylase purified from mouse leukemic L-1210 cells was investigated. Significant activity was encountered with O².2'-anhydro-5-benzylcytidine hydro-chloride, 2',3'-dideoxy-5-benzyluridine, 2',3'-dideoxy-4-thiouridine and α - and β -anomers of 5-benzyl-1-(2-deoxy-D-*arabino*-hexopyranosyl)uracil.

Introduction

Uridine phosphorylase (EC 2.4.2.3) plays a crucial role in the degradation of pyrimidine nucleosides as well as in the salvage pathways which supply precursors for the nucleic acids synthesis *de novo*. This double role in cell metabolism is strategic particularly in neoplastic tissues wherein the enzyme level can be increased by the action of cytokines and growth factors.¹ On one hand, uridine phosphorylase causes an undesirable phosphorolysis of pyrimidine nucleoside analogues used in cancer chemotherapy; thus, its inhibitors can be useful in promoting such therapy. On the other hand, the inhibition of uridine phosphorylase delays the disappearance of uridine from the cell pool and enhances the therapeutic effects of 5-fluorouracil in transformed cells by lowering its toxicity in normal tissues.² Thus, the rational design of uridine phosphorylase inhibitors has two aspects: (a) development of novel pyrimidine or pyrimidine nucleoside analogs with cytostatic activity and/or (b) potentiation of the pharmacological effect of known pyrimidine antimetabolites. One of the established

inhibitors, benzylacyclouridine (BAU, IND 039655) in combination with 5-fluorouracil, reached the clinical Phase II treatment of cancer patients.

It was also demonstrated that uridine phosphorylase has a key role in the metabolism of pathogenic protozoal parasites, e.g. *Trypanosoma* sp.,³ *Toxoplasma* sp.⁴ and/or *Schizostoma* sp.⁵ These enzymes seem to have lower restrictions for the inhibitor structure which might lead to development of inhibitors specific for the protozoal enzyme and not affecting the host cell metabolism, with a potential chemotherapeutic use.

Various structural types of uridine phosphorylase inhibitors (modified pyrimidine bases, pyrimidine nucleosides or pyrimidine acyclic derivatives) have been synthesized (for review, see ref. 6). Some of them are very specific and powerful.⁷ TdG (thymine 2'-deoxyglucoside) for example is able to distinguish between uridine phosphorylase and the frequently accompanying related enzyme, thymidine phosphorylase. The latter enzyme is an important angiogenic factor taking part in the tumor growth⁸ and its level is particularly high in certain forms of cancer (breast, ovary, bladder, colon).⁹ However, it also is the key-enzyme for tumor-specific liberation of 5-fluorouracil from its prodrugs tegafur,^{9,10} 5'-deoxy-5-fluorouridine (doxifluridine, Furtulon)¹¹ and capecitabine (5'-deoxy-5-fluoro-N⁴-pentyloxycarbonylcytidine)¹² used in cancer treatment.

5-Substituted pyrimidines or pyrimidine nucleosides containing a hydrophobic group (benzyl, benzyloxybenzyl) occupy an important position among the most potent inhibitors of uridine phosphorylases. Most attention has focused on 5-benzyluracil derivatives.13 In our project on uridine phosphorylase inhibitors we have synthesized four classes of base-modified nucleosides: 5-benzyluracil and 5-benzyl-4-thiouracil derivatives,¹⁴ 5-phenylcytosine derivatives¹⁵ and 5-phenylpyrimidin-2-one derivatives.¹⁶ The alteration of the sugar part of the nucleoside molecule involved the β -D-ribofuranosides (1), 5-chloro-5-deoxy- β -D-ribo- (2) and 5-chloro-5-deoxy- β -D-arabino-nucleosides (4), 5-deoxy- β -D-ribo- (3) and 5-deoxy- β -D-arabinofuranosides (5), β -D-arabinofuranosides (6), β -D-xylofuranoside (7), anomeric 2-deoxy-D-ribofuranosides (8, 9), 2,3-dideoxy- β -D-ribonucleosides (10), 2,3-dideoxy- β -D-ribofuranosides (11, 12), anomeric 3-azido-2,3-dideoxy- β -D-ribonucleosides (13, 14), anomeric 2-deoxy- β -D-arabino-hexopyranosides (15, 16) and anomeric D-arabino-2-hexeno-





pyranosides (21, 22). Furthermore, the series encompassed 2,2'-anhydronucleosides (17) and other related compounds. In this paper, we are presenting the data on the inhibitory activity of these compounds on uridine phosphorylase purified from mouse leukemic L-1210 cells, in comparison with established inhibitors TdG (23), BAU [5-benzyl-1-(2-hydroxyethoxymethyl)uracil, (24)] and 2,2'-anhydro-5-benzyluridine (17d).

Results

Chemistry. Most of the compounds were described in our previous communications.¹⁴⁻¹⁶ We report here the synthesis of the hitherto unknown α - and β -anomers of 2',3'-dideoxy-2',3'-didehydro-5-phenyluridine (Scheme 1). Methyl 5-O-benzoyl-2deoxy-3-O-mesylribofuranoside (25)^{16b} gave on reaction with 5-phenyl-2,4-bis-(trimethylsilyloxy)pyrimidine in the presence of trimethylsilyl trifluoromethanesulfonate, two anomeric mesylates 26 and 27. On treatment with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) in acetonitrile, the α -anomer 27 afforded the 5'-O-









ÒН

17a R=H, X=NH2+CIb R=Phe, X=NH2+Ci-



18











24

benzoate 29 which gave on methanolysis the elimination product 12, whilst the same reaction of the β -anomer 26 resulted in the formation of the 3',2-anhydronucleoside 28. By the action of sodium in HMPTA/THF followed by methanolysis this compound afforded 2,3-dideoxy-2,3-didehydro derivative 11a together with the 2-deoxy- β -D-*threo*-pentofuranoside 20. The structure of the ultimate products 11a, 12 and 20 was confirmed by their ¹H-NMR spectra.

Enzyme assay. The reaction mixture contained in a final volume of 100 µl: 50 mM potassium phosphate pH 7.4, 50µM [14C-U-uracil]uridine (1 MBq µmol-1), 15 µg of enzyme protein and 10 µM compound tested. Reaction mixtures were incubated for 10 min at 37 °C, and appropriate aliquots were withdrawn, spotted on W3MM paper and chromatographed in 1-butanol-acetic acid-water (10 : 1 : 3) overnight. The corresponding uracil and uridine spots were cut out and the radioactivity was determined by liquid scintillation counting. The inhibitory effect was expressed as v_r/v_o ratio at a constant concentration of the substrate (50 µM uridine) and/or compound tested (10 µM) (Table 1).

Kinetic experiments. Kinetic constants (K_m , K_i , V_{max}) were determined from the both Lineweaver-Burk and Dixon plots using five concentrations of the tested compound (including zero point) and four concentrations of the corresponding substrate (uridine). Data were evaluated by the non-linear regression method from four independent experiments. The concentration of the enzyme used was 150 μ g/ml. Reactions were carried out at 37 °C for 10 min. The results are summarized in Table 2.

Discussion

Our data (Tables 1, 2) identify six new inhibitors with the inhibitory potency which decreases in the order 15 > 17c > 16 > 10b > 10a > 1b. The activity of compound 15 relates to TdG (23) and its increased inhibitory activity in relation to the parent compound may be interpreted simply by the replacement of 5-benzyluracil for thymine. Both anomeric 2-deoxyhexopyranosides (15 and 16) are effective inhibitors of the enzyme: however, they differ by the mechanism of inhibition: while the β -anomer 15 interferes with uridine phosphorolysis *via* mixed-noncompetitive mechanism, the α -anomer 16 acts *via* competitive inhibition of the enzyme, similarly as TdG (23).¹⁷







Scheme 1

Compound	v _i /v _o	Reference	Compound	v _i /v _o	Reference
1a	0.83	14	11a	1.14	a
1b	0.33	14	11b	1.08	15
1c	1.07	14	12	0.94	а
1 d	1.00	14	13	0.79	16b
2a	1.20	14	14	1.16	16b
2b	1.31	15	15	0.02	14
3a	1.13	14	16	0.14	14
3b	0.91	15	17 a	1.04	20
4	1.19	15	17b	1.17	15
5	1.03	15	17c	0.03	14
6a	1.01	15	17d	0.05	18
6b	0.87	14	18	0.92	16a
7	0,78	16 a	19	0.94	16 a
8a	0.99	14	20	1.21	а
8b	0.82	16a	21	0.93	14
9a	1.15	14	22	0.95	14
9b	0.84	16 a	23	0.16	21
10a	0.12	14	24	0.06	19
10b	0.13	14			
10c	1.05	14			
10d	0.91	15			
10e	1.20	14			
10f	1.23	16b			

TABLE 1. Inhibition of uridine phosphorylase from L1210 cells.

a This paper.

The second most active inhibitor of the series, compound 17c belongs to the uridine phosphorylase inhibitors with the 2,2'-anhydronucleoside structure. Neither 2,2'-anhydrouridine nor 2,2'-anhydrocytidine (17a) inhibit the enzyme, however, 2,2'-anhydro-5-benzyluridine (17d) is a powerful inhibitor.¹⁸ Contrary to the 5-benzyluracil nucleosides which regularly exhibit inhibitory activity disregarding the character of the sugar residue, none of their 5-benzylcytosine counterparts was active. Compound 17c is the only exception. Comparison of the inhibitory activity of 17c with the activity of 2,2'-anhydro-5-benzyluridine (17d) on uridine phosphorylase from L-1210 cells showed that the uridine derivative 17d is approximately one order of magnitude more potent compared to its 5-benzylcytidine congener (Table 2). However, it cannot be excluded that compound 17c might behave *in vivo* as a prodrug liberating 2,2'-anhydro-5-benzyl-

Compound	K _i (μM)	K _i /K _m	
<i>b</i> 1b	2.27 ± 0.14	0.051	-
10b	0.478 ± 0.062	0.011	
10a	0.575 ± 0.035	0.013	
c 15	0.097 ± 0.011	0.002	
16	0.401 ± 0.031	0.009	
17c	0.152 ± 0.012	0.003	
17d	0.0124 ± 0.0018	0.0003	
23	0.579 ± 0.044	0.013	
d 24	0.113 ± 0.009	0.003	

TABLE 2. Kinetic characteristics of uridine phosphorylase (L1210 cells) inhibitors a

^a K_m for uridine: $44.3 \pm 1.4 \mu$ M, V_{max} = $72.6 \pm 2.6 \mu$ mol/min/mg. ^b Compound 1b is a substrate for uridine phosphorylase, its phosphorolysis was confirmed by HPLC analysis. ^c Mixed-noncompetitive type of inhibition. ^d Partially-competitive type of inhibition.

uridine (17d). It might be worthwhile to examine its influence on uridine levels in the cellular pool and in plasma following its application in an animal study

The comparatively high inhibitory activity of the two 2',3'-dideoxynucleosides 10 is interesting from the viewpoint of their relation to the antiretroviral dideoxynucleosides. The uracil derivative 10a has been already described¹⁹ and found inactive against HIV, however, in our hands it turned out to be a potent inhibitor of uridine phosphorylase.

Also the related 4-thiouracil compound 10b is an interesting structural lead: Another 4-thiouracil derivative, 5-benzyl-4-thiouridine (1b) inhibits the enzyme as well, furthermore, it is a substrate for uridine phosphorylase (at 50 μ M concentration, 57% of 1b is phosphorolyzed by the addition of 0.5 mg/ml of the enzyme). Comparison with 5substituted 2-pyrimidinone derivatives shows that the 4-oxo (thioxo) group at the pyrimidine ring is indispensable for the inhibition of uridine phosphorylase and that the replacement of the 5-benzyl group by 5-phenyl results in the loss of activity.

Experimental

Melting points were determined on a Kofler block and are uncorrected. ¹H NMR spectra (δ , ppm; *J*, Hz) were recorded with a Varian UNITY 500 instrument in hexa-deuteriodimethyl sulfoxide with tetramethylsilane as internal standard. Column chrom-

atography was performed on 30–60 μ m silica gel (Service Laboratories of the Institute) and thin-layer chromatography (TLC) on Silufol UV254 foils (Kavalier, Votice). If not stated otherwise, the solutions were evaporated at 2 kPa and bath temperature 30–60 °C; compounds were dried at 13 Pa.

1-(5-O-Benzoyl-2-deoxy-3-O-methanesulfonyl-β-D-erythro-pentofuranosyl)-5phenyluracil (26) and 1-(5-O-benzoyl-2-deoxy-3-O-methanesulfonyl-a-D-erythropentofuranosyl)-5-phenyluracil (27). A suspension of 5-phenyluracil (2.63 g, 14 mmol) and ammonium sulfate (20 mg) in hexamethyldisilazane (100 ml) was heated at 150 °C until it dissolved, and then for additional 2 h. The solvent was evaporated and the residue was codistilled with xylene (100 ml), leaving the silyl derivative. Trimethylsilyl trifluoromethanesulfonate (4.5 ml) was added to a solution of the silyl 5-O-benzoyl-2-deoxy-3-O-methanesulfonyl-a-L-erythroderivative and methvl pentofuranoside (25)16b (4.59 g, 13.9 mmol) in acetonitrile (100 ml) cooled to 0 °C. The mixture was stirred at 0 °C for 20 min and the reaction was quenched with saturated aq. NaHCO₃ (100 ml). After stirring at room temperature for 10 min, the product was taken up in ethyl acetate (3 x 300 ml). The organic layer was dried over Mg₂SO₄, the solvent was evaporated and the residue was chromatographed on a column of silica gel (500 g) in toluene-ethyl acetate (1 : 2) to give 2.09 g (31%) of the β anomer 26 and 0.67 g (10%) of α -anomer 27 as solid foams.

β-anomer 26: ¹H NMR: δ 2.62 (ddd, 1 H, $J_{2a',1'} = 5.2$, $J_{2a',2b'} = 14.6$, $J_{2a',3'} = 2.6$, H-2a'), 2.77 (ddd, 1 H, $J_{2b',1'} = 7.8$, $J_{2b',3'} = 6.9$, H-2b'), 3.33 (s, 3 H, OSO₂CH₃), 4.51 (dd, 1 H, $J_{5a',4'} = 5.1$, $J_{5a',5b'} = 14.2$, H-5a'), 4.52 (m, 1 H, H-4'), 4.59 (dd, 1 H, $J_{5b',4'} = 8.3$, H-5b'), 5.84 (dt, 1H, $J_{3',4'} = 2.6$, H-3'), 6.27 (dd, 1 H, H-1'), 7.24, 7.39, 7.47, 7.65, 7.95 (H-arom.), 7.75 (s, 1 H, H-6), 11.65 (s, 1 H, H-3). Anal Calcd for C₂₃H₂₂N₂O₈S (486.5): C 56.78; H 4.56, N 5.76; S 6.59; Found: C 56.99; H 4.78; N 5.53; S 6.39.

α-anomer 27: ¹H NMR: δ 2.60 (brdt, 1 H, $J_{2a',1'} = 2.5$, $J_{2a',2b'} = 15.4$, H-2a'), 3.00 (brdt, 1 H, $J_{2b',1'} = 6.8$, H-2b'), 3.29 (s, 3 H, OSO₂CH₃), 4.39 (dd, 1 H, $J_{5a',4'} = 5.2$, $J_{5a',5b'} = 12.2$, H-5a'), 4.42 (dd, 1 H, $J_{5b',4'} = 4.5$, H-5b'), 5.03 (brtd, 1 H, $J_{4',3'} = 1.4$, H-4'), 5.49 (ddd, 1 H, $J_{3',2a'} = 2.2$, $J_{3',2b'} = 6.3$, H-3'), 6.32 (dd, 1 H, H-1'), 7.31 (7.37, 7.56, 7.69, 8.10, H-arom.), 7.79 (s, 1 H, H-6), 11.60 (s, 1 H, H-3). Anal Calcd for C₂₃H₂₂N₂O₈S (486.5.: C 56.78; H 4.56; N 5.76; S 6.59. Found: C 57.02; H 4.67; N 5.49; S, 6.34.

2,3'-Anhydro-1-(5-O-benzoyl-2-deoxy-\beta-D-threo-pentofuranosyl)-5-phenyluracil (28). A solution of the 3'-mesyl derivative 26 (973 mg, 2 mmol) and 1,8-diazabicyclo-[5.4.0]undec-7-ene (0.45 ml, 3 mmol) in acetonitrile (45 ml) was refluxed for 40 min. The solution was cooled and taken down *in vacuo*. Chromatography of the residue on a silica gel column (120 g) in ethyl acetate-acetone-ethanol-water (18 : 3 : 2 : 1) afforded 500 mg (64%) of crystalline anhydro derivative 28, m.p. 228.5-230 °C; ¹H NMR: δ 2.60 (ddd, 1 H, $J_{2a',1'} = 4.0$, $J_{2a',2b'} = 12.9$, $J_{2a',3'} = 3.0$, H-2a'), 2.71 (brdd, 1 H, $J_{2b',1'} =$ 0.5, $J_{2b',3'} = 1.5$, H-2b'), 4.44 (dd, 1 H, $J_{5a',4'} = 5.6$, $J_{5a',5b'} = 11.6$, H-5a'), 4.60 (dd, 1 H, $J_{5b',4'} = 5.1$, H-5b'), 4.64 (brtd, 1 H, $J_{4',3'} = 2.2$, H-4'), 5.49 m, 1 H, H-3'), 6.06 (brd, 1 H, H-1'), 7.31, 7.37, 7.47, 7.57, 7.64, 7.92, (H-arom.), 8.01 (s, 1 H, H-6). Anal Calcd for $C_{22}H_{18}N_2O_5$ (390.40): C 67.68; H 4.65; N 7.18% N; Found: C 67.52; H 4.67; N 7.39.

1-(2,3-Dideoxy- β -D-glycero-pent-2-enofuranosyl)-5-phenyluracil (11a) and 1-(2deoxy- β -D-threo-pentofuranosyl)-5-phenyluracil (20). Tetrahydrofuran (20 ml) and anhydro derivative 28 (390 mg, 1 mmol) were added to a solution of sodium (46 mg, 2 mmol) in hexamethylphosphorous triamide, HMPTA) (1 ml) and the mixture was refluxed for 4 h. Then another portion of sodium solution (46 mg) in HMPTA (1 ml) was added and the mixture was refluxed for 3 h. The mixture was cooled, diluted with methanol (20 ml) and stirred for 20 h. The solution was neutralized with Dowex 50, H⁺), the resin was filtered off and solvents were evaporated. The residue was chromatographed on a silica gel column (100 g) in ethyl acetate-acetone-ethanol-water (36 : 6 : 1 : 1) to give 150 mg (36%) of compound 11a and 115 mg (26%) of compound 20.

Compound 11a: M.p. 312.5–314.5 °C. ¹H NMR: δ 3.65 (dd, 2 H, $J_{5',4'} = 2.5$, $J_{5',OH} = 5.0$, 2 x H-5'), 4.83 (m, 1 H, H-4'), 5.13 t, 1 H, 5'-OH), 5.98 (dt, 1 H, $J_{3',1'} = J_{3',4'} = 1.5$, $J_{3',2'} = 6.0$, H-3'), 6.41 (dt, 1 H, $J_{2',1'} = J_{2',4'} = 1.5$, H-2'), 6.90 (dt, 1 H, $J_{1',4'} = 3.2$, H-1'), 7.28, 7.34,7.52, H-arom.), 8.10 (s, 1 H, H-6), 11.50 (s, 1 H, H-3). Anal Calcd for C₁₅H₁₄N₂O₄ (286.3): C 62.93; H 4.93; N 9.79; Found: C 62.70; H 4.93; N 9.87.

Compound 20: M.p. 205 - 206.5 °C; ¹H NMR: δ 2.00 (brd, 1 H, $J_{2a',3'} = 1.0$, $J_{2a',2b'} = 13.9$, H-2a'), 2.60 (ddd, 1 H, $J_{2b',1'} = 8.3$, $J_{2b',3'} = 5.1$, H-2b'), 3.64 (brdt, 1 H, $J_{5a',4'} = 6.8$, $J_{5a',5b'} = 11.7$, H-5a'), 3.73 (brdt, 1H, $J_{5b',4'} = 4.9$, H-5b'), 3.87 (m, 1 H, H-4'), 4.27 (m, 1 H, H-3'), 4.74 (t, 1 H, $J_{OH,5'} = 5.6$, 5'-OH), 5.37 (d, 1 H, $J_{3',OH} = 3.0$, 3'-OH),

6.16 (dd, 1 H, $J_{1',2a'}$ = 1.5, H-1'), 7.30 9, 7.38, 7.52, H-arom.), 8.29 (s, 1 H, H-6), 11.47 (s, 1 H, H-3). Anal Calcd for C₁₅H₁₆N₂O₅ (304.3): C 59.21; H 5.30; N 9.21; Found: C 59.10; H 5.34; N 9.12.

1-(2,3-Dideoxy-α-D-glycero-pent-2-enofuranosyl)-5-phenyluracil (12). 1,8-Diazabicyclo[5,4,0]undec-7-ene (0.23 ml, 1.5 mmol) was added to a solution of mesvl deriv-ative 27 (487 mg, 1 mmol) in acetonitrile (15 ml) and the solution was refluxed for 12 h. The solvent was evaporated and the residue was partitioned between ethyl acetate (15 ml) and water (5 ml). The organic layer was washed with water (5 ml), dried (Mg₂SO₄) and the solvent evaporated. A solution of the residue (29) in methanolic ammonia (30 ml) was set aside at room temperature for 2 days and then taken down *in vacuo*. Chromato-graphy of the residue on silica gel (50 g) in tolueneacetone-triethylamine (15:15:1) afforded 170 mg (59%) of compound 12 as solid foam. ¹H NMR: δ 3.46 (dd, 1 H, $J_{5a',4'} = 4.6$, $J_{5a',5b'} = 11.5$, H-5a'), 3.51 (dd, 1 H, $J_{5b',4'} = 4.4$, H-5b'), 4.85 (brs, 1 H (5'-OH), 5.10 m, 1 H, H-4'), 6.02 (dt, 1 H, $J_{3',1'} = 1.4$ $J_{3',4'} = 1.7, J_{3',2'} = 6.0, H-3'$), 6.45 dt, 1 H, $J_{2',1'} = J_{2',4'} = 1.7, H-2'$), 6.93 (dt, 1 H, $J_{1',4'}$ = 5.2, H-1', 7.31, 7.37, 7.47, H-arom.), 7.46 (s, 1 H, H-6), 11.(s, 1 H, H-3).) Anal Calcd for C15H14N2O4 (286.3.: C 62.93; H 4.93; N 9.79; Found: C 63.05; H 5.11; N 9.66.

Enzyme source. Mouse leukemic L1210 cells were inoculated (i.p., 10⁵ cells per animal) in DBA/2 inbred female mice (25 g) and after 8 days ascites were released from peritoneum and washed twice in phosphate-buffered saline (PBS).

Purification of uridine phosphorylase (2.4.2.3). Cells were resuspended in 20 mM potassium phosphate pH 8.0 supplemented with 1 mM DTT, 1 mM EDTA (buffer A) and protease inhibitors (Sigma, 40 μ g/ml bestatin, 2 μ g/ml pepstatin, 2 μ g/ml leupeptin and 2 μ g/ml aprotinin). The cell suspension was then freeze-thawed (three times) and homogenized in a Dounce tissue grinder (Wheaton, pestle A, 15 strokes). The crude extract was centrifuged at 100 000 g for 100 min and the supernatant was fractionated by ammonium sulfate. The enzyme proteins were salted out between 25 - 55% of (NH₄)₂SO₄ saturation, dissolved in buffer A and desalted on Sephadex PD-10 columns (Pharmacia LKB) in the same buffer. The resulting eluent was then applied onto the column (4.5 x 2.1 cm) of DEAE-Sephacel (Pharmacia LKB, equilibrated in buffer A) and chromatography was performed in a concentration gradient of KC1 (0 - 0.25 M) in

buffer A. The enzyme activity was detected as a single peak between 0.1 - 0.25 M KCl. The most active fractions were then pooled and used as a partially purified uridine phosphorylase.

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