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Multicomponent one-pot synthesis of 2-naphthol derivatives and evaluation of their anticancer activity

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Abstract Treatment of 2-naphthol with aldehydes and allyl tributyl stannane or anisole in the presence of AlCl₃ at 0°C to room temperature afforded its 1-alkyl derivatives in high yields (76–83%) within 4–8 h. The products were evaluated for their cytotoxic activity against four human cancer cell lines. The most potent compound (**5d**) showed IC₅₀ of 1.2 ± 1.1, 1.6 ± 1.0, 0.9 ± 0.1, and 0.8 ± 0.4 μ M against Hep G₂, A549, MDA 231, and HeLa cell lines, respectively, and its activity was found to be comparable to that of doxorubicin.

Introduction

2-naphthol derivatives are valuable pharmacologic compounds possessing various important biological properties such as antiinflammatory, antibacterial, hypotensive, and bradycardiac activities (Shen *et al.*, 1999, Huang *et al.*, 2003, Chopade *et al.*, 2010). Some of these compounds have recently been prepared by different workers (Shen

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S. K. Mamidyala · C. G. Kumar Chemical Biology Laboratory, Indian Institute of Chemical Technology, Tarnaka, Hyderabad 500 607, India *et al.*, 1999, Khodaei *et al.*, 2006, Selvam and Perumal, 2006, Das *et al.*, 2007, 2009, 2010, Srihari *et al.*, 2007, Shaterian *et al.*, 2008, Mahdavinia and Bigdeli, 2009, Zhao *et al.*, 2007). However, the multistep procedures are the disadvantages in many of these methods. Herein, we report a simple and efficient one-pot synthesis of different 2-naphthol derivatives. The anticancer activity of these compounds has also been evaluated.

Chemistry

In continuation of our work (Das *et al.*, 2007, 2009) on the development of useful synthetic methodologies, we have observed that 2-naphthol (1) when treated with aldehydes (2) and allyl tributyl stannane (3) or anisole (4) using AlCl₃ as a catalyst afforded its 1-alkyl derivatives (5 or 6) at 0°C to room temperature (Scheme 1).

Various 2-naphthol derivatives (5, 6) have been prepared using different aldehydes (2) following the above three-component one-pot synthetic method (Table 1). These derivatives included 4,4-diarylbent-1-ene compounds (5a–5g) and triaryl methanes (6a–6d). The former were formed within 4–6 h, while the latter in 6–8 h. The yields of the products were high (76–83%). The structures of these derivatives were established from their spectral (IR, ¹H, ¹³C NMR, and MS) and analytical data. All the prepared compounds were optically inactive.

The mechanism of this multicomponent synthesis involves the initial reaction of 2-naphthol (1) with aldehydes (2) in the presence of $AlCl_3$ to form the orthoquinone methide intermediate A (Van De Water and Pettus, 2002) which was subsequently attacked by the nucleophile, allyl tributyl stannane (3) or anisole (4) to afford the products (Scheme 2).



 Table 1
 Synthesis of 2-naphthol derivatives (5, 6)

Entry	R	Time (h)	Product	Yield ^c (%)	
1	C ₆ H ₅	5.0	5a ^a	79	
2	C ₆ H ₅	7.0	6a ^b	81	
3	4-Me-C ₆ H ₄	4.0	5b ^a	83	
4	4-i-Pr-C ₆ H ₄	4.5	5c ^a	81	
5	4-i-Pr-C ₆ H ₄	6.0	6b ^b	82	
6	$4-F-C_6H_4$	6.0	5d ^a	78	
7	$2-F-C_6H_4$	6.5	5e ^a	77	
8	$3-F-C_6H_4$	7.0	5f ^a	80	
9	$4-F-C_6H_4$	8.0	6c ^b	79	
10	4-Cl-C ₆ H ₄	6.0	5g ^a	76	
11	4-Cl-C ₆ H ₄	8.0	6d ^b	79	

The structures of the products were established from their spectral (IR, 1 H, 13 C NMR, and ESI–MS) data

 a 2-naphthol (1 mmol), aldehyde (1 mmol), allyl tributyl stannane (1.2 mmol), and $AlCl_3$ (10 mol %) were used

 $^{\rm b}$ 2-naphthol (1 mmol), aldehyde (1 mmol), anisole (1.2 mmol), and AlCl_3 (10 mol %) were used

^c Isolated yields after purification

The catalyst, $AlCl_3$ is easily available and less expensive. It catalyses the present multicomponent synthesis efficiently for the preparation of 2-naphthol derivatives.

It works under mild reaction conditions. The reaction was conducted at 0°C to room temperature. The ether linkage was not affected.

Bioactivity

The synthetic 2-naphthol derivatives (5, 6) were evaluated for their in vitro cytotoxic activity against four human cancer cell lines including liver (Hep G₂), lung (A 549), breast (MDA 231), and cervical (HeLa) by applying MTT assay (Navarra et al., 2010). Doxorubicin was used as a positive control and DMSO as a negative control. IC_{50} values were determined for each compound (Table 2; Fig. 1). The data indicate that the 2-naphthol derivatives containing a propylene chain at the C-1 alkyl group (compounds 5) are generally more active compared with those having an aromatic ring at the same position (compounds 6). Moreover, among the products 5, the compound which contains an unsubstituted phenyl ring (5a) exhibits low activity (Table 2, entry 1). However, a compound with a substituted phenyl ring showed higher activity (entries 3, 4, 6, 8). The most active compound, 5d (entry 6) contains a 4-fluoro phenyl ring and it possess IC₅₀ of 1.2 ± 1.1 , $1.6 \pm 1.0, 0.9 \pm 0.1, \text{ and } 0.8 \pm 0.4 \ \mu\text{M}$ against Hep G₂,



Scheme 2

Table 2 $\,IC_{50}$ values ($\mu M)$ of test compounds against different cancer cell lines

Entry	Sample code	IC ₅₀ values (µM) ^a				
		HepG2	A549	MDA231	HeLa	
1	5a	53.0 ± 2.1	59.0 ± 3.2	33.0 ± 1.4	76.0 ± 3.6	
2	6a	6.2 ± 1.6	6.1 ± 2.4	5.8 ± 2.5	6.3 ± 1.7	
3	5b	5.6 ± 1.5	6.8 ± 1.2	6.0 ± 1.4	6.2 ± 1.5	
4	5c	15.0 ± 2.1	13.5 ± 3.2	16.5 ± 4.5	12.5 ± 3.2	
5	6b	44.0 ± 3.5	50.0 ± 2.1	37.8 ± 2.8	33.0 ± 4.6	
6	5d	1.2 ± 1.1	1.6 ± 1.0	$<\!\!1\pm0.1$	$<1\pm0.4$	
7	5e	9.7 ± 1.5	7.8 ± 1.9	6.6 ± 1.2	6.8 ± 2.1	
8	5f	6.5 ± 0.9	8.6 ± 1.3	9.1 ± 1.9	8.8 ± 1.1	
9	6c	14.0 ± 3.5	17.2 ± 4.1	12.5 ± 2.1	11 ± 2.0	
10	5 g	7.4 ± 1.1	9.6 ± 1.2	7.6 ± 1.1	6.4 ± 1.2	
11	6d	42.0 ± 3.7	44.6 ± 2.4	33.2 ± 2.1	29.4 ± 2.7	
12	Doxorubicin	1.2 ± 0.5	$<\!\!1\pm0.1$	${<}1\pm0.1$	$<1\pm0.2$	

No inhibition of growth was observed when 1% DMSO was used as negative control

^a All tests were performed in duplicate (n = 2) and values are represented as mean \pm standard deviations



Fig. 1 IC_{50} values of the test compounds

A 549, MDA 231, and HeLa cell lines, respectively. The activity of the compound is comparable to that of doxorubicin (Table 2). The regioisomers **5e** (containing a 2-fluoro phenyl ring) and **5f** (containing a 3-fluoro phenyl ring) of **5a** exhibited lower activity. The other three compounds **5b**, **5g**, **6a** also possess impressive cytotoxic activity.

Conclusions

In conclusion, we have developed an efficient method for the conversion of 2-naphthol into its derivatives by reaction with aldehydes and allyl tributyl stannane or anisole in the presence of $AlCl_3$ at 0°C to room temperature. The mild reaction conditions, high yields, operational simplicity, and one-pot conversion are the advantages of the method. The evaluation of cytotoxic property of these 2-naphthol derivatives identified a compound having activity comparable to that of doxorubicin.

Experimental

General

All commercially available reagents were used directly without further purification unless otherwise stated. The solvents used were all of AR grade and were distilled under a positive pressure of dry nitrogen atmosphere wherever necessary. The progress of the reactions was monitored by analytical thin layer chromatography (TLC) performed on Merck Silica Gel 60 F₂₅₄ plates. Column chromatography was carried out using silica gel 60-120 mesh (Qingdao Marine Chemical, China). IR spectra were recorded on a Perkin-Elmer RX1 FT-IR spectrophotometer and mass spectra on VG-Autospec micromass. NMR spectra were recorded on Gemini 200 MHz spectrometer with tetramethylsilane as internal standard using CDCl₃. The chemical shifts are expressed as δ values in parts per million (ppm) and the coupling constants (J) are given in Hertz (Hz). Yields were of purified compounds and were not optimized. Optical rotations were measured with JASCO DIP 300 digital polarimeter at 25°C.

General experimental procedure

AlCl₃ (10 mol %) was added to a solution of a 2-naphthol (1 mmol), aldehyde (1 mmol), and allyl tributyl stannane or anisole (1.2 mmol) in MeCN (5 ml) at 0°C under N₂ atmosphere. The mixture was stirred at room temperature and the reaction was monitored by TLC. After completion, the solvent was removed from the reaction mixture, and water and EtOAc (10 ml each) were added. The organic layer was separated and concentrated. The residue was subjected to column chromatography (silica gel, hexane–EtOAc) to obtain a pure product (**5** or **6**).

The spectral and analytical data of the unknown products

1-[1-(4-isopropylphenyl)but-3-enyl]naphthalen-2-ol (5c)

IR: 3449, 1637, 1462, 1219 cm⁻¹; ¹H NMR (200 MHz, CDCl₃): δ 8.02, (1H, d, J = 8.0 Hz, Ar–H), 7.71 (1H, d, J = 8.0 Hz, Ar–H), 7.60 (1H, d, J = 8.0 Hz, Ar–H), 7.40

(1H, *t*, *J* = 8.0 Hz, Ar–H), 7.31–7.20 (4H, m, Ar–H), 7.12 (2H, d, *J* = 8.0 Hz, Ar–H), 6.93 (1H, d, *J* = 8.0 Hz, Ar–H), 5.69 (1H, m, H-2), 5.10–4.92 (2H, m, H₂-1), 4.87 (1H, brd, *J* = 8.0 Hz, H-4), 3.13 (1H, m, CHMe₂), 3.01–2.81 (2H, m, H₂-3), 1.22 (6H, *t*, *J* = 7.0 Hz, CHMe₂); ¹³C NMR (50 MHz, CDCl₃): δ 152.2 (Ar–C), 147.0 (Ar–C), 143.4 (Ar–C), 137.0 (Ar–C), 134.6 (C-2), 129.9 (Ar–C), 128.8 (Ar–C), 128.2 (Ar–C), 127.9 (Ar–C), 127.1 (Ar–C), 126.8 (Ar–C), 123.2 (Ar–C), 122.1 (Ar–C), 119.4 (Ar–C), 117.9 (Ar–C), 116.4 (C-1), 40.3 (C-4), 36.2 (C-3), 34.2 (CHMe₂), 24.1 (CHMe₂); ESIMS: *m*/z 317 [M + H]^{+,}; Anal. Calc for C₂₃H₂₄O: C 87.34, H 7.60%; Found: C 87.45, H 7.58%.

1-[1-(4-fluorophenyl)but-3-enyl]naphthalen-2-ol (5d)

IR: 3448, 1630, 1509, 1461, 1270, 1224 cm⁻¹; ¹H NMR (200 MHz, CDCl₃): δ 7.91 (1H, d, J = 8.0 Hz, Ar–H), 7.70 (1H, d, J = 8.0 Hz, Ar–H), 7.58 (1H, d, J = 8.0 Hz, Ar–H), 7.39–7.02 (4H, m, Ar–H), 6.98–6.82 (3H, m, Ar– H), 5.69 (1H, m, H-2), 5.10–4.98 (2H, m, H₂-1), 4.82 (1H, brd, J = 7.0 Hz, H-4), 3.19–3.01 (2H, m, H₂-3); ¹³C NMR (50 MHz, CDCl₃): δ 160.2 (d, J = 280.0 Hz, Ar–C–F), 151.9 (Ar–C), 137.1 (Ar–C), 133.5 (C-2), 131.1 (Ar–C), 131.0 (Ar–C), 129.9 (Ar–C), 128.8 (Ar–C), 126.8 (Ar–C), 124.3 (Ar–C), 123.2 (Ar–C), 123.0 (Ar–C), 122.0 (Ar–C), 119.1 (Ar–C), 119.0 (d, J = 8.0 Hz, Ar–C), 116.2 (C-1), 40.5 (C-4), 36.9 (C-3); ESIMS: m/z 293 [M + H]⁺; Anal. Calc for C₂₀H₁₇FO: C 82.19, H 5.82%; Found: C 82.28, H 5.78%.

1-[1-(2-fluorophenyl)but-3-enyl]naphthalen-2-ol (5e)

IR: 3450, 1632, 1514, 1465, 1231 cm⁻¹; ¹H NMR (200 MHz, CDCl₃): δ 7.89 (1H, d, J = 8.0 Hz, Ar–H), 7.72 (1H, d, J = 8.0 Hz, Ar–H), 7.58 (1H, d, J = 8.0 Hz, Ar–H), 7.55 (1H, t, J = 8.0 Hz, Ar–H), 7.36–6.92 (3H, m, Ar–H), 6.89–6.80 (3H, m, Ar–H), 5.71 (1H, m, H-2), 5.17– 4.92 (2H, m, H₂-1), 4.87 (1H, brd, J = 7.0 Hz, H-4), 3.20– 3.04 (2H, m, H₂-3); ¹³C NMR (50 MHz, CDCl₃): δ 160.5 (d, J = 280.0 Hz, Ar–C–F), 151.7 (Ar–C), 132.8 (C-2), 131.2 (Ar–C), 131.4 (d, J = 8.0 Hz, Ar–C), 129.5 (Ar–C), 128.7 (Ar–C), 128.0 (Ar–C), 127.2 (Ar–C), 126.5 (Ar–C), 125.2 (Ar–C), 119.1 (Ar–C), 118.6 (d, J = 8.0 Hz, Ar–C), 116.2 (C-1), 40.3 (C-4), 36.5 (C-3); ESIMS: m/z 293 [M + H]⁺; Anal. Calc for C₂₀H₁₇FO: C 82.19, H 5.82%; Found: C 82.12, H 5.84%.

1-[1-(3-fluorophenyl)but-3-enyl]naphthalen-2-ol (5f)

IR: 3447, 1628, 1512, 1464, 1218 cm⁻¹; ¹H NMR (200 MHz, CDCl₃): δ 7.87 (1H, d, J = 8.0 Hz, Ar–H), 7.68 (1H, d, J = 8.0 Hz, Ar–H), 7.54 (1H, d, J = 8.0 Hz,

Ar–H), 7.36–6.92 (4H, m), 6.88–6.78 (3H, m, Ar–H), 5.62 (1H, m, H-2), 5.12–4.96 (2H, m, H₂-1), 4.83 (1H, brd, J = 7.0 Hz, H-4), 3.17–3.02 (2H, m, H₂-3); ¹³CNMR (50 MHz, CDCl₃): δ 162.7 (d, J = 280.0 Hz, Ar–C–F), 151.2 (Ar–C), 143.8 (Ar–C), 134.1 (C-2), 131.3 (Ar–C), 131.1 (Ar–C), 129.5 (Ar–C), 128.7 (Ar–C), 126.4 (Ar–C), 124.2 (Ar–C), 124.1 (Ar–C), 123.7 (Ar–C), 122.8 (Ar–C), 121.6 (Ar–C), 118.8 (Ar–C), 118.3 (d, J = 8.0 Hz, Ar–C), 116.7 (C-1), 114.2 (d, J = 8.0 Hz, Ar–C), 41.0 (C-4), 36.7 (C-3); ESIMS: m/z 293 [M + H]⁺; Anal. Calc for C₂₀H₁₇FO: C 82.19, H 5.82%; Found: C 82.31, H 5.86%.

1-[1-(4-chlorophenyl)but-3-enyl]naphthalen-2-ol (5g)

IR: 3448, 1629, 1491, 1462, 1366, 1237 cm⁻¹; ¹H NMR (200 MHz, CDCl₃): δ 7.91 (1H, d, J = 8.0 Hz, Ar–H), 7.70 (1H, d, J = 8.0 Hz, Ar–H), 7.58 (1H, d, J = 8.0 Hz, Ar–H), 7.40–7.04 (6H, m, Ar–H), 6.93 (1H, d, J = 8.0 Hz, Ar–H), 5.68 (1H, m, H-2), 5.08–4.92 (2H, m, H₂-1), 4.82 (1H, brd, J = 7.0 Hz, H-4), 3.18–2.98 (2H, m, H₂-3); ¹³C NMR (50 MHz, CDCl₃): δ 151.8 (Ar–C–Cl), 142.2 (Ar–C), 137.0 (Ar–C), 135.3 (Ar–C), 133.4 (C-2), 129.9, (Ar–C), 129.1 (Ar–C), 128.2 (Ar–C), 126.4 (Ar–C), 124.2 (Ar–C), 118.9 (Ar–C), 116.2 (C-1), 40.7 (C-4), 36.2 (C-3); ESIMS: m/z 309, 311 [M + H]⁺; Anal. Calc for C₂₀H₁₇ClO: C 77.80, H 8.15%; Found: C 77.89, H 8.09%.

1-[(4-fluorophenyl)(4-methoxyphenyl)methyl]naphthalen-2-ol (**6**c)

IR: 3468, 1626, 1603, 1508, 1463, 1248 cm⁻¹; ¹H NMR (200 MHz, CDCl₃): δ 7.86 (1H, d, J = 8.0 Hz, Ar–H), 7.73–7.61 (2H, m, Ar–H), 7.55 (1H, d, J = 8.0 Hz, Ar–H), 7.38–7.12 (4H, m, Ar–H), 7.06 (2H, d, J = 8.0 Hz, Ar–H), 7.02–6.91 (2H, m, Ar–H), 6.80 (2H, d, J = 8.0 Hz, Ar–H), 6.27 (1H, s, Ar–H), 5.22 (1H, brs, CHAr₃), 3.72 (3H, s, OMe); ¹³C NMR (50 MHz, CDCl₃): δ 156.1, (d, J = 280.0 Hz Ar–C), 154.0 (Ar–C), 152.9 (Ar–C), 137.8 (Ar–C), 132.2 (Ar–C), 130.8 (Ar–C), 130.1 (Ar–C), 130.0 (Ar–C), 129.1 (Ar–C), 127.9 (Ar–C), 126.9 (Ar–C), 126.6 (Ar–C), 123.2 (Ar–C), 122.8 (Ar–C), 120.0 (Ar–C), 118.1 (Ar–C), 116.2, (d, J = 10.0 Hz, Ar–C), 114.7 (Ar–C), 54.9 (OMe), 47.1 (CHAr₃),; ESIMS: m/z 381 [M + Na]⁺; Anal. Calc for C₂₄H₁₉FO₂: C 80.45, H 5.31%; Found: C 80.58, H 5.27%.

1-[(4-chlorophenyl)(4-methoxyphenyl)methyl]naphthalen-2-ol (*6d*)

IR: 3468, 1627, 1603, 1509, 1463, 1246 cm⁻¹; ¹H NMR (200 MHz, CDCl₃): δ 7.78 (1H, d, J = 8.0 Hz, Ar–H), 7.63 (1H, d, J = 8.0 Hz, Ar–H), 7.61 (1H, d, J = 8.0 Hz,

Ar–H), 7.30–7.12 (4H, m, Ar–H), 7.10 (2H, d, J = 8.0 Hz, Ar–H), 7.01 (2H, d, J = 8.0 Hz, Ar–H), 6.92 (1H, d, J = 8.0 Hz, Ar–H), 6.72 (2H, d, J = 8.0 Hz, Ar–H), 6.19 (1H, s, Ar–H), 5.09 (1H, brs, CHAr₃), 3.69 (3H, s, OMe); ¹³C NMR (50 MHz, CDCl₃): δ 159.0 (Ar–C), 152.9 (Ar–C), 140.4 (Ar–C), 133.1 (Ar–C), 133.0 (Ar–C), 130.5 (Ar–C), 130.0(Ar–C), 129.9 (Ar–C), 128.9 (Ar–C), 128.0 (Ar–C), 127.1 (Ar–C), 124.7 (Ar–C), 124.4 (Ar–C), 123.5 (Ar–C), 122.9 (Ar–C), 119.9 (Ar–C), 119.3 (Ar–C), 114.9 (Ar–C), 55.2 (OMe), 47.1 (CHAr₃; ESIMS: m/z 375, 377 [M + H]⁺; Anal. Calc for C₂₄H₁₉ClO₂: C 76.90, H 5.07%; Found: C 76.82, H 5.13%.

Evaluation of cytotoxic activity

Cell lines used for testing in vitro cytotoxicity included HepG2 derived from human hepatocellular liver carcinoma cells (ATCC No. HB-8065), A549 derived from human lung adenocarcinoma epithelial cells (ATCC No. CCL-185), MDA231 derived from human breast adenocarcinoma cells (ATCC No. HTB-26) and HeLa derived from human cervical cancer cells (ATCC No. CCL-2). These cell lines were obtained from American Type Culture Collection, Manassas, VA, USA.

All tumor cell lines were maintained in a Modified Eagle's medium (Sigma-Aldrich, USA) supplemented with 10% fetal bovine serum (Sigma), along with 1% nonessential amino acids without L-glutamine (Sigma), 0.2% sodium bicarbonate, 1% sodium pyruvate (Sigma), and 1% of antibiotic mixture (10,000 units penicillin and 10 mg streptomycin per ml, Sigma). The cells were washed and resuspended in the above medium and 100 µl of this suspension was seeded in 96-well flat bottom plates. The cells were maintained at 37°C in a humidified 5% CO2 incubator (Model 2406 Shellab CO₂ incubator, Sheldon manufacturing, Cornelius, OR, USA). After 24 h of incubation, the cells were treated for 2 days with eleven test compounds at concentrations ranging from 0.1 to 100 µM in DMSO (1% final concentration) and were assayed at the end of the second day. Each assay was performed with two internal controls: (1) an IC₀ with cells only, (2) an IC₁₀₀ with media only. After 48 h of incubation, the cells were subjected to MTT colorimetric assay (5 mg ml⁻¹ MTT). The effect of the different test compounds on the viability of tumor cell lines was measured at the wavelength of 540 nm on a multimode reader (Infinite[®] M200, Tecan, Switzerland). Dose-response curves were plotted for the test compounds and controls after correction by subtracting the background

absorbance from that of the blanks. The IC_{50} values (50% inhibitory concentration) were calculated from the plotted absorbance data for the dose–response curves. IC_{50} values (in μ M) are expressed as the average of two independent experiments.

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