

Use of the Mitsunobu reaction in the synthesis of orthogonally protected α,β -diaminopropionic acids

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Abstract—Orthogonally protected α,β -diaminopropionic acids have been synthesised in good yields by the reaction of *N*-trityl L-serine esters with *N*-substituted sulfonamides under Mitsunobu reaction conditions (DEAD, PPh_3 , THF). The best isolated yields were obtained when *N*-Boc *p*-toluenesulfonamide was used as the nitrogen nucleophile precursor in the Mitsunobu reaction. Subsequently, the *N*-trityl group was efficiently replaced with the more stable allyloxycarbonyl (alloc) group.
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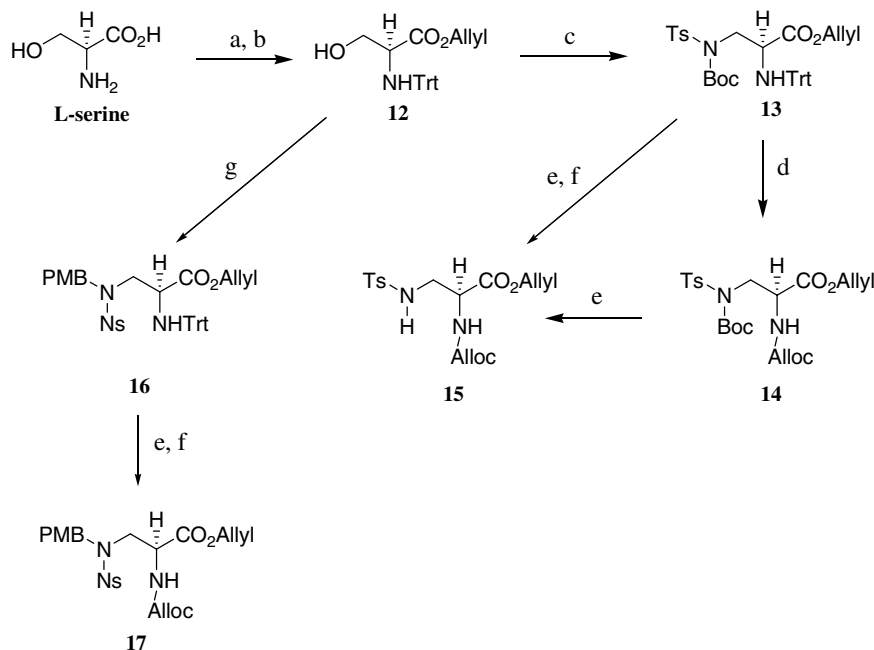
1. Introduction

Interest in the properties of α,β -diamino acids is evidenced by the large number of publications in this area. A recent review has highlighted the wide range of methods for their synthesis and also the myriad of possible biological applications.¹ A number of recent examples, in particular, demonstrate the different approaches to the preparation of these types of compounds. Lee prepared 2,3-diaminopropionates by ring opening of aziridine-2-carboxylates with azide ions and subsequent reduction of the azido group to an amine,² while Panda used the Mitsunobu reaction³ of HN_3 on *N*-protected L-serine-derived Weinreb amides.⁴ The reason for the use of the Weinreb amide derivative was to reduce the acidity of the serine α -hydrogen and thus stop the formation of a dehydroalanine (Dha) by a dehydration reaction. Pedatella prepared orthogonally protected 2,3-diamino acids by treatment of the enolate of *N,N*-dibenzylated β -amino esters with di-*tert*-butyl azodicarboxylate (DBAD). Subsequent removal of the Boc group and cleavage of the hydrazine gave the 2,3-diamino acids.⁵ Nadir prepared 2,3-diamino acids by reaction of *N*-aryl-sulfonyl aziridines with a chiral isocyanate and subsequent hydrolysis of 2-imidazolidinones.⁶

As part of a program of peptide synthesis, incorporating unusual amino acid residues, we are interested in the synthesis of orthogonally protected α,β -diaminoprop-

ionic acids for solid-phase peptide synthesis. We decided to examine the Mitsunobu reaction of L-serine derivatives with nitrogen-based nucleophiles, other than azide, for their synthesis. Our first choice was sulfonamide based nucleophiles (Ts or Ns) because they have appropriate pK_a values. In 1989 Weinreb introduced Ts–NH–Boc in the Mitsunobu reaction for the conversion of alcohols into *N*-Boc *p*-toluenesulfonamides in excellent yields.⁷ A survey of the literature shows that the correct choice of nitrogen and/or carboxyl protecting groups is critical to the positive outcome of the Mitsunobu reaction at the hydroxyl group of L-serine derivatives. In many cases, the incorrect choice of protecting group leads to elimination reactions giving Dha compounds, or cyclisations to form aziridines. Cherney and Wang showed that protection of the L-serine nitrogen with a trityl group gives excellent yields of Mitsunobu products, using phthalimide as the nitrogen nucleophile, where the carboxyl group was protected as the methyl ester.⁸ The trityl group works in two ways, by sterically preventing cyclisation of the nitrogen to form aziridines, and secondly, by reducing the acidity of the α -hydrogen compared to carbamate protecting groups, thus preventing Dha formation. We prepared *N*-trityl L-serine methyl ester (**2**) in 75% yield from L-serine methyl ester (**1**) using the method of Baldwin (Scheme 1).⁹ Subsequent treatment of **2** with the commercially available Weinreb nucleophile Ts–NH–Boc (**3**), under Mitsunobu reaction conditions (diethyl azodicarboxylate (DEAD), PPh_3 , THF), gave the orthogonally protected α,β -diaminopropionic acid **4** in 75% isolated yield. Due to the propensity for the trityl group to be easily removed, even on treatment with mild acid, we decided to replace this

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Scheme 2. Reagents and conditions: (a) trityl chloride, trichloromethylsilane, Et₃N, DCM, rt, 48%; (b) (i) Cs₂CO₃, MeOH; (ii) allyl bromide, DMF, rt, 90%; (c) **3**, DEAD, PPh₃, THF, rt, 72%; (d) (i) 5% TFA in CHCl₃, (ii) allyl chloroformate, NaHCO₃, H₂O, 1,4-dioxane, rt, 19% from **13**; (e) 50% TFA in CHCl₃, rt; (f) allyl chloroformate, NaHCO₃, H₂O, rt; (g) **11**, DEAD, PPh₃, THF, rt, 51%.

other reactions only the starting materials were re-isolated. As before, **16** was efficiently converted to the N-alloc protected compound **17** (Scheme 2).

In conclusion, we have prepared orthogonally protected α,β -diaminopropionic acids in good yields from protected L-serines using the Mitsunobu reaction of sulfonamide-derived nitrogen nucleophiles. Currently we are studying the chemistry of these compounds, for example, the clean removal of the individual protecting groups, and their incorporation into peptide structures using solid-phase peptide synthesis. We are also examining further functionalisation reactions of N–H sulfonamide compounds **6** and **15**. The results of these studies will be reported in due course.

2. Typical procedure for Mitsunobu reaction, exemplified by the synthesis of **13**

To a solution of *N*-(*tert*-butoxycarbonyl)-*p*-toluenesulfonamide **3** (0.16 g, 0.68 mmol) in dry THF (3 ml) was added PPh₃ (0.34 g, 1.4 mmol), followed by the addition of **12** (0.16 g, 0.46 mmol) and DEAD (0.19 g, 1.2 mmol). The resulting mixture was allowed to stir at room temperature, under a nitrogen atmosphere, for 10 h. The solvent was removed in vacuo giving an orange oil, which was purified by flash column chromatography on silica gel, in petroleum ether/ethyl acetate (10:1), to give a white solid (0.30 g, 72%). Mp: 143–145 °C. *R*_f: 0.80, petroleum ether–ethyl acetate (2:1). IR (KBr) cm⁻¹: 3433, 3066, 2924, 1734, 1595, 1234, 1139. ¹H NMR (CDCl₃, 300.4 MHz) δ ppm, 7.77 (d, 2H, *J* = 12.3 Hz, *ortho* tosyl), 7.54 (d, 6H, *J* = 12.9 Hz, *ortho* trityl), 7.27–7.23 (d, 2H, *J* = 12.3 Hz, *meta* tosyl and m, 9H, *para* and *meta* trityl), 5.56 (m, 1H, vinyl CH), 5.11

(m, 2H, vinyl CH₂), 4.24 (dd, 1H, *J* = 8.4 and 8.6 Hz, allyl CH₂), 4.11 (m, 1H, allyl CH₂), 3.92 (dd, 1H, *J* = 5.3 and 6.0 Hz, α -CH), 3.80 (m, 2H, β -CH₂), 2.86 (d, 1H, *J* = 11.1 Hz, NH), 2.37 (s, 3H, tosyl CH₃), 1.25 (s, 9H, *t*-butyl). ¹³C NMR (CDCl₃, 75.45 MHz) δ ppm, 172.6 (ester C=O), 150.8 (Boc C=O), 145.7 (*ipso* trityl), 144.1 (*para* tosyl), 137.4 (*ipso* tosyl), 131.8 (vinyl CH), 129.2 (*meta* trityl), 128.8 (*meta* tosyl), 128.1 (*ortho* trityl), 127.8 (*ortho* tosyl), 126.4 (*para* trityl), 118.3 (vinyl CH₂), 84.4 (C_q *t*-butyl), 70.9 (allyl CH₂), 65.9 (C(Ph)₃), 56.0 (α -CH), 50.4 (β -CH₂), 27.8 (CH₃ *t*-butyl), 21.6 (CH₃ tosyl). Mass Spec: expected [M+1] 641.2685, observed [M+1] 641.2690.

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12. Compounds **8** and **10** were prepared according to Kurosawa, W.; Kan, T.; Fukuyama, T.; Cheung, M.; Kan, T. *Synlett* **1999**, 1301, and compound **9** was prepared by an adaptation of this method. Compounds **11a** and **11b** were prepared according to the method of Kurosawa, W.; Kan, T.; Fukuyama, T. *Org. Synth.* **2002**, *79*, 186. Note: Analytical data for **9**: Mp: 121–122 °C. R_f : 0.69, methanol–chloroform (1:2). IR (KBr) cm^{-1} : 3308, 3021, 1540, 1364, 1158. ^1H NMR (CDCl_3 , 300.4 MHz) δ ppm, 8.05 (d, 2H, $J = 8.9$ Hz, *ortho* nosyl), 7.83 (d, 2H, $J = 8.9$ Hz, *meta* nosyl), 7.54 (d, 2H, $J = 8.2$ Hz, *ortho* tosyl), 7.03 (d, 2H, $J = 7.8$ Hz, *meta* tosyl), 2.31 (s, 3H, tosyl). ^{13}C NMR (CDCl_3 , 75.45 MHz) δ ppm, 149.1 (*para* nosyl), 143.3 (*ipso* nosyl), 141.8 (*para* tosyl), 139.3 (*ipso* tosyl), 129.5 (*meta* tosyl), 128.6 (*ortho* nosyl), 127.9 (*ortho* tosyl), 123.1 (*meta* nosyl), 21.4 ($-\text{CH}_3$, tosyl).
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