



Natural Product Research Formerly Natural Product Letters

ISSN: 1478-6419 (Print) 1478-6427 (Online) Journal homepage: https://www.tandfonline.com/loi/gnpl20

Inhibition of squalene synthase of rat liver by abietane diterpenes derivatives

Mariana Macías-Alonso, Lucía S. Andrés, Iván Córdova-Guerrero, Arturo Estolano-Cobián, Laura Díaz-Rubio & Joaquín G. Marrero

To cite this article: Mariana Macías-Alonso, Lucía S. Andrés, Iván Córdova-Guerrero, Arturo Estolano-Cobián, Laura Díaz-Rubio & Joaquín G. Marrero (2019): Inhibition of squalene synthase of rat liver by abietane diterpenes derivatives, Natural Product Research, DOI: <u>10.1080/14786419.2019.1678614</u>

To link to this article: https://doi.org/10.1080/14786419.2019.1678614



View supplementary material \square



Published online: 21 Oct 2019.

_	
Γ	
L	0
-	

Submit your article to this journal 🗹



View related articles 🗹



View Crossmark data 🗹

SHORT COMMUNICATION



Check for updates

Inhibition of squalene synthase of rat liver by abietane diterpenes derivatives

Mariana Macías-Alonso^a, Lucía S. Andrés^b, Iván Córdova-Guerrero^c, Arturo Estolano-Cobián^c, Laura Díaz-Rubio^c and Joaquín G. Marrero^a (b)

^aInstituto Politécnico Nacional, UPIIG, Silao de la Victoria, Guanajuato, Mexico; ^bInstituto Universitario de Bioorgánica "Antonio González", Departamento de Química Orgánica, Universidad de La Laguna, Tenerife, Spain; ^cFacultad de Ciencias Químicas e Ing, Universidad Autónoma de Baja California, Tijuana, B. C, Mexico

ABSTRACT

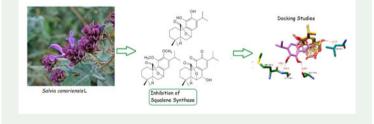
In the current study, chemical composition of cultivated Salvia canariensis L was determined. Carnosol was the main product isolated. We prepared more lipophilic derivatives from carnosol, and both isolated and semisynthetic abietane diterpenes were evaluated *in vitro* as inhibitors of squalene synthase. Among the compounds tested, carnosol was the most potent inhibitor ($IC_{50} = 17.6 \mu M$). These results highlight the great potential of this species for the production of new ingredients in nutritional supplements for the treatment of hyperlipidemia.

ARTICLE HISTORY

Received 29 July 2019 Accepted 23 September 2019

KEYWORDS

Carnosol; Hyperlipidemia; Salvia; Cholesterol; Squalene synthase



1. Introduction

Hyperlipidemia is a well-established risk factor for the development of cardio and cerebrovascular diseases. For this reason, lipid lowering is one of the major approaches used for their primary prevention. A widely used strategy to decrease plasma cholesterol concentration is the use of HMG-CoA inhibitors such as statins. However, this enzyme plays a fundamental role in the biosynthesis of many other nonsteroidal molecules, so statins have potential adverse effects (Balaz and Wolfrum 2019). The inhibition of squalene synthase (SQS), an enzyme essential in the biosynthesis of the steroids, can prevent the cholesterol biosynthesis without

CONTACT Joaquín González Marrero 🖾 jgonzalezm@ipn.mx

Supplemental data for this article can be accessed at https://doi.org/10.1080/14786419.2019.1678614.

 $\ensuremath{\mathbb{C}}$ 2019 Informa UK Limited, trading as Taylor & Francis Group

disturbing the nonsterol pathway. Several SQS inhibitors have been developed such as zaragozic acid and quinuclidine (Kourounakis et al. 2010).

The genus *Salvia* consists of some 500 species, some of which have shown interesting biological activity (Jakovljević et al. 2019). In previous studies of the aerial part of *Salvia canariensis L.*, an endemism of the Canary Islands, carnosic acid **1** and carnosol **2** were isolated as the major phenolic diterpenes (Marrero et al. 2009), which have been reported to have broad biological activities like neuroprotective and hypolipidemic properties (González 2015; Alonso et al. 2019).

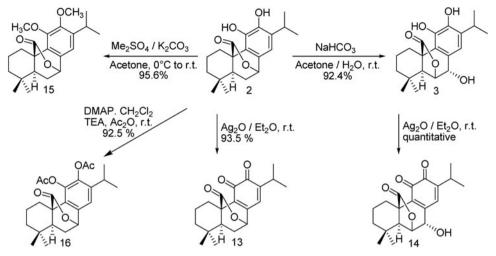
As a continuation of our work on the isolation and synthesis of diterpenes to identify bioactive components, and due to the great importance to discover potential SQS inhibitors, the purpose of this study was to provide the chemical profile of cultivated *S. canariensis*, obtain synthetic derivatives from the isolated products and evaluate their ability to inhibit SQS *in vitro*. These results make this plant to be considered as a crop that could have commercial importance as useful ingredient in nutritional supplements.

2. Results and discussion

The acetonic extract of cultivated *S. canariensis* was fractionated and purified following the protocol designed by Marrero et al. (2009). The analysis of the extract revealed carnosol **2** as the main product. The concentration of rosmanol **3** was also significant, with 7α methoxyrosmanol **4**, 7α -ethoxyrosmanol **5** and galdosol **6** as minority constituents (Table S1). These compounds have been previously obtained from *S. canariensis* (Marrero et al. 2009). We also reported here, for the first time from this species, the isolation of rosmadial **7**, epirosmanol **8** and isorosmanol **9** previously found in *Rosmarinus officinalis* (Nakatani and Inatani 1984); isogaldosol **10** and demetilsalvicanol **11** previously isolated from *Salvia mellifera* (González et al. 1992); and 7β -methoxyrosmanol **12** identified previously in *Salvia dorrii* (Ahmed et al. 2006). On the other hand, in our study we could not detect carnosic acid **1**, probably due to the harvest time of the plant (Luis and Johnson, 2005). Although 16-acetoxycarnosol was isolated from *in vitro* culture of *S. canariensis* (Luis et al. 1992), we could not detect any abietane derivative oxidized at C-16.

All compounds were tested for SQS inhibitory activity at a concentration of 10 μ M (Amin *et al*, 1992) (Table S1). Compounds **8**, **10**, **11** and **12** were not evaluated due to the small amount isolated. Most compounds exhibited weak inhibition activity of SQS, except carnosol **2** (IC₅₀ = 17.6 μ M). At this point, we decided to prepare some semisynthetic derivatives of carnosol **2**, where we change the hydroxyl groups for more lipophilic substituents (Scheme 1). The biological evaluation of these derivatives showed that 11,12-di-O-methylcarnosol **15** was weaker than carnosol **2**. However, the oxidation of rosmanol **3** to rosmaquinone **14**, improves the biological activity (Table S1).

To obtain more information about the mechanism as well as the mode of interactions of these phytochemicals with SQS, the three compounds with the best *in vitro* results were selected to perform a docking analysis (Table S2). Regularly, squalene inhibitors have been found interacting with Ala176, Met207, Gly208 and Leu76 residues and the ones nearby, mainly by Van der Waals and hydrophobic interactions, present in a large



Scheme 1. Semisynthesis of Carnosol derivatives.

hydrophobic pocket that the SQS receptor possess (Figure S3) (Ichikawa et al. 2012). The hydroxyl groups of carnosol and rosmaquinone may not interact with the R group of the residues, but with their skeletons. This would be the case for one of carnosol phenolic hydroxyl with the skeletal N of Gly208; a weak hydrogen or dipole interaction among these atoms could be possible, as their distance is 2.5 Å. This same hydroxyl has a 3.08 Å distance to the carbonyl of Met207. The second phenolic hydroxyl also has a 2.96 Å distance to the carbonyl of Ala176, so similar interactions can take part (Figure S4). The hydroxyl of rosmaguinone has Leu76 carbonyl at a distance of 4.35 Å. Thus, we can assume a weak dipole interaction from this group may be favouring rosmaguinone. These additional interactions, other than the hydrophobic ones, would partially explain why 11,12-dimethoxycarnosol (which have none of them) has the lowest docking score, even though it shares a very similar position in space with rosmaguinone. Even though carnosol and rosmaquinone have the best docking results, in vitro we see how carnosol has the lowest IC₅₀ (17.6 μ M), followed by 11,12-di-O-methylcarnosol (18.4 μ M). Although this can look as a disagreement, we can observe that the docked poses give the correct insight for the in silico-in vitro relationship. The docked compounds are very close to each other in the cavities where is well known inhibitors bind (Ichikawa et al. 2012), but carnosol can be seen deeper in such cavity. Different studies for SQS inhibitors have made the observation that, the better the *in vitro* inhibition shown, the deeper such compound goes interacting in the hydrophobic cavity in the docking analysis (Kourounakis et al. 2010). In such way, the docking poses allows to explain why carnosol has a better interaction with the receptor by means of its spatial position.

3. Conclusions

We describe the majority diterpenes present in the extract of cultivated *S. canariensis*, which was confirmed to be a taxon strongly characterized by the presence of carnosol **2**. We isolated eleven abietane-type compounds. The present study portrays the first report of the isolation of six of these compounds from this species. We prepared

more lipophilic derivatives from carnosol, and both isolated and synthetic compounds were evaluated as SQS inhibitors. It was observed that carnosol **2**, rosmaquinone **14** and 11,12-di-O-methylcarnosol **15** showed *in vitro* SQS inhibitory effect. In order to obtain data about the interactions between these molecules and SQS, we performed *in silico* docking studies. To our knowledge, this is the first report of the inhibition of SQS by abietanes. Application of the findings from this docking study could led to the development of more potent SQS inhibitors aimed against atherosclerosis. We think that this species could be cultivated and used as a new useful ingredient in nutritional supplements with antioxidant and anti-atherosclerosis function.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This work was supported by CONACYT and Research and Postgraduate Secretary of the National Polytechnic Institute (SIP- IPN) are gratefully acknowledged.

ORCID

Joaquín G. Marrero (D) http://orcid.org/0000-0003-3720-4528

References

- Ahmed AA, Mohamed A, Karchesy J, Asakawa Y. 2006. Salvidorol, a nor-abietane diterpene with a rare carbon skeleton and two abietane diterpene derivatives from Salvia dorrii. Phytochemistry 67(5):424–428.
- Alonso MM, Sancén SR, Marrero JG. 2019. Carnosic acid and its derivatives: Diterpenes of biological interest. Biomed J Sci & Tech Res. 16(4):12172–12174.
- Amin D, Cornell SA, Gustafson SK, Needle SJ, Ullrich JW, Bilder GE, Perrone MH. 1992. Bisphosphonates used for the treatment of bone disorders inhibit squalene synthase and cholesterol biosynthesis. J Lipid Res. 33(11):1657–1663.

Balaz M, Wolfrum C. 2019. Statins: benefits and risks revisited. Aging 11(13):4300-4302.

- González MA. 2015. Aromatic abietane diterpenoids: their biological activity and synthesis. Nat Prod Rep. 32(5):684–704.
- González AG, Andrés LS, Aguiar ZE, Luis JG. 1992. Diterpenes from *Salvia mellifera* and their biogenetic significance. Phytochemistry 31(4):1297–1305.
- Ichikawa M, Ohtsuka M, Ohki H, Haginoya N, Itoh M, Sugita K, Usui H, Suzuki M, Terayama K, Kanda A. 2012. Discovery of novel tricyclic compounds as Squalene synthase inhibitors. Bioorg Med Chem. 20(9):3072–3093.
- Jakovljević M, Jokić S, Molnar M, Jašić M, Babić J, Jukić H, Banjari I. 2019. Bioactive Profile of Various Salvia officinalis L. preparations. Plants (Basel). 8(3):55.
- Kourounakis AP, Matralis AN, Nikitakis A. 2010. Design of more potent Squalene synthase inhibitors with multiple activities. Bioorg Med Chem. 18(21):7402–7412.
- Luis JG, González AG, Andrés LS, Mederos S. 1992. Diterpenes from in vitro-grown Salvia canariensis. Phytochemistry 31(9):3272–3273.
- Luis JC, Johnson CB. 2005. Seasonal variations of rosmarinic and carnosic acids in rosemary extracts. Analysis of their in vitro antiradical activity. Span J Agric Res. 3(1):106–112.

- Marrero JG, Moujir L, Andrés LS, Montaño NP, Araujo L, Luis JG, 2009. Semisynthesis and biological evaluation of abietane-type diterpenes. Revision of the structure of rosmaquinone. J Nat Prod. 72(8):1385–1389.
- Nakatani N, Inatani R. 1984. Two antioxidative diterpenes from rosemary (Rosmarinus officinalis) and a revised structure for rosmanol. Agric Biol Chem. 48:2081–2085.