Access to the Bicyclic Core of Isatisine, and an Investigation of Its Antibacterial Activity

Christopher J. Matthews, Mark G. Moloney,* Amber L. Thompson, Hanna Winiarska, Henry T. Winney

Department of Chemistry, Chemistry Research Laboratory, The University of Oxford, 12 Mansfield Road, Oxford OX1 3TA, UK Fax +44(1865)285002; E-mail: mark.moloney@chem.ox.ac.uk Received 15 November 2010

Abstract: A chemoselective Dieckmann ring closure using an oxazolidine derived from serine may be used to generate a tetramic acid, the further manipulation of which by reduction and ring closure leads to the bicyclic core of isatisine; depending on the nature of the ring closing electrophile, different diastereomers are obtained. None of the compounds from this sequence exhibited activity against *S. aureus* but several showed activity against *E. coli*.

Key words: heterocycles, natural products, cyclisation, antibiotics, lactams

The discovery of new antibiotics has become imperative by reason of the emergence of bacterial strains resistant to current clinically-effective drugs,¹ and the urgency of the task has recently been emphasised.² However, since there is a paucity of New Chemical Entities (NCE) entering the antibacterial pipeline,³ a number of new strategies for more efficient drug development have been elaborated in recent years⁴ and re-examination of the function and availability of natural products has been pivotal.⁵ In particular, it has recently become apparent that the physicochemical characteristics of antibacterial compounds differ sufficiently from the property space of commonly available compound libraries that re-investigation of natural products to identify novel antibacterials is fully justified⁶ and that the structural information thus obtained will be useful for the design of NCE suitable for fragment-based drug design.⁷ Libraries based upon such natural product leads have several key benefits which do not apply to combinatorially-derived systems: they will have benefited from the optimisation of bioactivity for a given receptor as a result of natural selection; they will be expected to provide an enhanced rate of positive hits for a given library size; they are more likely to provide novel structural chemotypes not currently in use in existing therapeutic regimes; they would not be immediately susceptible to resistance-conferring genes in the bacterial and DNA pools and they are likely to provide novel new target proteins and receptors.⁸ Mindful of the long-standing application of traditional Chinese medicine,9 and to develop further the compound and bioactivity space encompassed by pyrrolidin(on)es as commonly occurring core structural components of natural products,^{10–12} we examined isatisine A as an unusual lead structure. Isatisine A (1) is a structurally novel alkaloid shown to have moderate anti-HIV activity (EC₅₀ 37.8 μ M) and cytotoxicity against C8166 cells (CC₅₀ 302 μ M).¹³ This compound was first isolated from the leaves of the Chinese plant *Isatis indigotica* Fort. (cruciferae) in 2007, and the roots and leaves of *I. indigotica* have been used in traditional Chinese medicine for the treatment of bacterial and viral infections, and immunoregulatory and tumour diseases.¹⁴ The first total synthesis of (+)-isatisine A was completed earlier this year by Karadeolian and Kerr in 14 steps with an overall yield of 5.8%.¹⁵

Retrosynthetic analysis (Scheme 1) of isatisine 1 suggested that functional group simplification could give tricycle 2, which would in turn be readily generated from tetramic acid 3 by reduction and cyclisation; such systems are available from oxazolidines 4 using our previously published approaches by aldol¹⁶ or Dieckman¹⁷ cyclisation. An initial investigation of this strategy (Scheme 2) using oxazolidine 7a (prepared by acylation of oxazolidine 5 with the malonate half ester 6a) involved allylation using cinnamyl bromide to give 7b, confirmation of the relative stereochemistry being readily achieved by single crystal X-ray analysis.¹⁸ Chemoselective Dieckmann cyclisation gave tetramate 8a in 63% yield. Reduction by a wellestablished¹⁹ procedure using NaBH₄-AcOH gave epimeric alcohols 10a and 11a in a ratio of 1:1.5 and modest overall yield (relative stereochemistry assigned on the basis of nOe data; see Supporting Information). This stereochemical outcome resulted from epimerisation at the acidic C(7) position, although fully diastereoselective endo-hydride addition at C-6 was observed.²⁰ Epoxidation (MCPBA) of 10a and 11a proceeded without diastereocontrol to give 12 and 13, both in good yield, and ring closure of 12 under basic conditions gave the desired tricyclic core 14 as a mixture of four inseparable diastereomers, as shown by NMR and MS analysis. As expected, similar treatment of epimeric epoxide 13 returned unreacted starting material. This approach demonstrated that a strategy based upon cycloetherification was possible, but that improvements in yield and diastereoselectivity would be required.

To develop this approach further, allylation of oxazolidine **7a** proved to be possible, but the yield was only 36%, and we found that the synthesis of the required allyl derivative **7c** could be better achieved in 58% yield by acylation of oxazolidine **5** with ethyl allylmalonate **6b** directly (prepared by partial hydrolysis of diethyl allylmalonate) using

SYNLETT 2011, No. 3, pp 0378–0382 Advanced online publication: 19.01.2011 DOI: 10.1055/s-0030-1259330; Art ID: D30410ST © Georg Thieme Verlag Stuttgart · New York



Scheme 1

DCCI-DMAP, conditions which we found to be superior to a similar recently reported coupling using MsCl- Et_3N ²¹ Dieckmann ring closure of oxazolidine 7c to give allyl tetramate 8b proceeded smoothly and in excellent yield (92%), but column chromatographic purification required the use of alumina, since silica gel led to epimerisation at C(7); the relative stereochemistry was determined by an NOE experiment (see Supporting Information). Moreover, we found that variable quantities of 9 were also obtained from this cyclisation, presumably as a result of adventitious water leading to ester hydrolysis and decarboxylation, and this material could not be separated from the major product 8b; careful exclusion of water successfully prevented this side reaction, as has been noted previously.¹⁷ Application of the above reduction procedure using excess NaBH₄-AcOH led to formation of alcohols 10b and 11b (ratio 2:1), along with significant recovery of starting material 8b (18-41%). Alternative conditions were investigated (2-15 equiv of NaBH₄ with addition of CeCl₃ or CaCl₂; use of LiBH₄), but selective formation of 10b could not be achieved. The relative stereochemistries of 10b and 11b were readily established by NOE experiments (see Supporting Information). Attempted conversion of 11b to 10b by deprotonation (LDA, -78 °C) and low temperature quench with ammonium chloride gave a 49% recovery of 11b, along with some of the desired alcohol **10b** (13% isolated yield).

With the desired allylpyroglutamate 10b in hand, investigation of electrophilic tetrahydrofuran ring closure was investigated (Scheme 2). Treatment with I₂-NaHCO₃ resulted in 5-exo-tet ring closure giving diastereomers 15a and 16a in a ratio of 10:3 (61% yield). The two epimers were inseparable by column chromatography, but careful recrystallisation (CH2Cl2-n-heptane) provided access to the pure major epimer 15a, whose structure was determined by an NOE experiment and single crystal X-ray analysis (see Supporting Information and Figure 1).¹⁸ Reaction of the 15a/16a mixture with potassium p-nitrobenzoate and a catalytic amount of 18-crown-6 gave inseparable benzoates 15b/16b in 31% overall yield, and another separable by-product 17 (24%), presumably arising by elimination from 15b/16b followed by double bond equilibration. Hydrolysis of the mixture of esters 15b/16b proceeded smoothly under basic conditions and furnished alcohols **15c/16c** in quantitative yield. Unsurprisingly, epimeric alcohol 11b did not ring close under these conditions, since this would have led to a *trans*-fused bicyclic ring system.



Figure 1 Thermal ellipsoid plot of 15a with the heteroatoms labelled and drawn at 50% probability

With the expectation that this route might be shortened by one step, it was of interest to apply our recently published protocol for plumbolactonisation,²² a modification of which was found to be also suitable for plumboetherification;²³ we found that cyclisation of tetramate **10b** using lead tetrabenzoate or lead tetra(thiophene-2-carboxylate) gave the desired products, but only as the unwanted diastereomers 16d and 16e, respectively (yields of 21% and 99%) with no trace of the other stereoisomers 15d and 15e; relative stereochemistry being again established by NOE experiments (see Supporting Information). We have observed that the thiophene-2-carboxylate system gives better yields and is also much more reactive than other arylcarboxylates in these cyclisations²²⁻²⁴ and believe that the change in stereochemical outcome in this case arises because the reaction proceeds by initial co-ordination of the C(6) hydroxy function rather than the alkene to the lead(IV) followed by syn-oxyplumbylation.²⁵ Deprotection of 16e using Corey-Reichard conditions²⁶ gave the bicyclic product 18a in excellent yield (75% yield) and this material was readily converted to diester 18b by DCC-DMAP coupling. Once again, epimeric alcohol 11b did not ring close under these conditions, but of interest is that tetramate **8b** cyclised to give tricycle **19**, whose structure was established by NMR analysis, although it was formed in only 12% yield.

This document was downloaded for personal use only. Unauthorized distribution is strictly prohibited.



Scheme 2 Reagents and conditions: (i) EDAC or DCC, DMAP, **5**; (ii) NaOMe, MeOH, r.t., 3 d; (iii) NaBH₄ (15 equiv), AcOH (10%), CH₂Cl₂, r.t.; (iv) MCPBA, CH₂Cl₂; (v) K₂CO₃, MeOH; (vi) I₂, NaHCO₃, MeCN; (vii) KO₂CC₆H₄NO₂, 18-C-6, MeCN; (viii) Na₂CO₃, H₂O, EtOH; (ix) (PhCO₂)₄Pb, F₃CPh; (x) (C₄H₃SCO₂)₄Pb, F₃CPh; (xi) HS(CH₂)₃SH, F₃CCH₂OH, r.t.; (xii) DCC, DMAP, C₄H₃SC(O)Cl.

In those reactions in which tetramate **9** was formed along with **8b** (Scheme 2), reduction of the mixture permitted isolation of alcohol **20** (Scheme 3) and plumboetherification gave tricycle **21** (59% yield). Assignment of the formation of the five-membered ring and the stereochemistry was made by comparison of the chemical shifts with those of product **16e** [C(8)H, the two C(9)H and the two C(12)H have very similar chemical shifts in both compounds] and deprotection to lactam **22** proceeded efficiently (72% yield).

Of interest is that bioassay²⁷ against *S. aureus* and *E. coli* indicated that no compounds were active against the

former, but that several were active against the latter, typically with a potency of about 0.05–0.07% of the cephalosporin standard; this outcome is of interest given that Gram-negative bacteria are intrinsically more resistant to antibacterials as a result of a more impermeable outer cell membrane, and particularly efficient efflux pumps.⁶ Bioactivity and chemical informatics²⁸ analysis is of interest (see Table in Supplementary Information). Although oxazolidines **7c**, tetramates **8a,8b**, pyroglutamates **10a,b**, **11a,b**, **12**, **13** and **18b** are active against *E. coli*, tricycles **15a**, **16a**, **16d** and **16e** (which might be considered most closely to mimic the core ring system of isatisine) are totally inactive against both organisms. For the active mol-



Scheme 3 Reagents and conditions: (i) NaBH₄, AcOH (10%), CH₂Cl₂, r.t.; (ii) (C₄H₃SCO₂)₄Pb, F₃CPh; (iii) HS(CH₂)₃SH, F₃CCH₂OH, r.t.

ecules, the van der Waals molecular surface area is in the range 452–587 $Å^{2}$,²⁹ and the CMR value, which is a measure of the van der Waals attractive forces that act in drugreceptor interactions³⁰ is in the range 74–107; this is to be expected from the common structural skeleton of these compounds. The active molecules have ClogD, %PSA and CMR average values of 2.5±0.76, 14.5±1.3 and 91.6±12.9, respectively. The polar surface area parameter (PSA), which correlates the presence of polar atoms with membrane permeability and therefore gives an indication of drug transport properties,³¹ has been reported to have an optimal value of $70 < PSA < 120 \text{ Å}^2$ for a non-CNS orally absorbable drug,³² and of interest is that the compounds active against E. coli all lie at the lower end of this range, with the exception of diester 18b. The use of this approach, guided by bioactive natural product structures, is likely to be valuable for the characterisation of chemical space³³ and may provide a useful start point for the identification of novel chemotypes for fragment-based drug design, especially where target identity or structure is not known.34

We have recently demonstrated the low intrinsic antibacterial activity of simple pyroglutamates¹² and tetramates,¹¹ but that modest manipulation of the skeletal functionality^{10,35} or homologation to longer chain sideunits³⁶ can restore activity. It may be that similar adjustment of the tricyclic skeletons synthesised in this work will be required before antibacterial bioactivity is observed; however, it is noted that no antibacterial activity of isatisine **1** *per se*, as opposed to the plant extracts, has been reported,¹³ and it is possible that the reported antibacterial plant extract activity may not result from this compound alone.

Supporting Information for this article is available online at http://www.thieme-connect.com/ejournals/toc/synlett.

References and Notes

- (a) Kotra, L. P.; Golemi, D.; Vakulenko, S.; Mobashery, S. Chem. Ind. (London) 2000, 341. (b) Niccolai, D.; Tarsi, L.; Thomas, R. J. Chem. Commun. 1997, 2333. (c) Peet, N. P. Drug Discovery Today 2010, 15, 583.
- (2) (a) Morel, C.; Mossialos, E. Br. Med. J. 2010, 340, 1115.
 (b) So, A. D.; Gupta, N.; Cars, O. Br. Med. J. 2010, 340, 1091.
- (3) Newman, D. J.; Cragg, G. M. J. Nat. Prod. 2007, 70, 461.

- (4) (a) Danishefsky, S. *Nat. Prod. Rep.* 2010, 27, 1114.
 (b) Cheng, C. C.; Shipps, G. W.; Yang, Z.; Sun, B.; Kawahata, N.; Soucy, K. A.; Soriano, A.; Orth, P.; Xiao, L.; Mann, P.; Black, T. *Bioorg. Med. Chem. Lett.* 2009, *19*, 6507. (c) Galloway, W. R. J. D.; Bender, A.; Welch, M.; Spring, D. R. *Chem. Commun.* 2009, 2446. (d) Cordier, C.; Morton, D.; Murrison, S.; Nelson, A.; O'Leary-Steele, C. *Nat. Prod. Rep.* 2008, *25*, 719. (e) Newman, D. J. *J. Med. Chem.* 2008, *51*, 2589.
- (5) (a) vonNussbaum, F.; Brands, M.; Hinzen, B.; Weigand, S.; Habich, D. Angew. Chem. Int. Ed. 2006, 45, 5072. (b) Nören-Müller, A.; Reis-Correa, I.; Prinz, H.; Rosenbaum, C.; Saxena, K.; Schwalbe, H. J.; Vestweber, D.; Cagna, G.; Schunk, S.; Schwarz, O.; Schiewe, H.; Waldmann, H. Proc. Natl. Acad. Sci. U.S.A. 2006, 103, 10606. (c) Baltz, R. H. J. Ind. Microbiol. Biotechnol. 2006, 33, 507. (d) Gullo, V. P.; McAlpine, J.; Lam, K. S.; Baker, D.; Petersen, F. J. Ind. Microbiol. Biotechnol. 2006, 33, 523. (e) Koch, M. A.; Waldmann, H. Drug. Discovery Today 2005, 10, 471. (f) Koehn, F. E.; Carter, G. T. Nat. Rev. Drug Discovery 2005, 4, 206. (g) Shang, S.; Tan, D. S. Curr. Opin. Chem. Biol. 2005, 9, 248. (h) Ganesan, A. Curr. Opin. Biotechnol. 2004, 15, 584. (i) Njardarson, J. T.; Gaul, C.; Shan, D.; Huang, X.-Y.; Danishefsky, S. J. J. Am. Chem. Soc. 2004, 126, 1038. (j) Rouhi, A. M. Chem. Eng. News 2003, 81, 77. (k) Breinbauer, R. B.; Vetter, I. R.; Waldmann, H. Angew. Chem. Int. Ed. 2002, 41, 2878. (1) Hemkens, P. H. H.; Ottenheijm, H. C. J.; Rees, D. C. Tetrahedron 1997, 53.5643
- (6) O'Shea, R.; Moser, H. E. J. Med. Chem. 2008, 51, 2871.
- (7) (a) Hajduk, P. J. J. Med. Chem. 2006, 49, 6972. (b) Rees,
 D. C.; Congreve, M.; Murray, C. W.; Carr, R. Nat. Rev. Drug Discovery 2004, 3, 660.
- (8) (a) Balamurugan, R.; Dekkerab, F. J.; Waldmann, H. *Mol. BioSyst.* 2005, *1*, 36. (b) Koch, M. A.; Wittenberg, L. O.; Basu, S.; Jeyaraj, D. A.; Gourzoulidou, E.; Reinecke, K.; Odermatt, A.; Waldmann, H. *Proc. Natl. Acad. Sci. U.S.A.* 2004, *101*, 16721.
- (9) Zhou, J.; Xie, G.; Yan, X. *Traditional Chinese Medicines*; Ashgate: England, **2004**.
- (10) Anwar, M.; Cowley, A. R.; Moloney, M. G. *Tetrahedron: Asymmetry* **2010**, *21*, 1758.
- (11) Jeong, Y.-C.; Moloney, M. G. Synlett 2009, 2487.
- (12) (a) Chandan, N.; Moloney, M. G. Org. Biomol. Chem. 2008,
 6, 3664. (b) Hill, T.; Kocis, P.; Moloney, M. G. Tetrahedron Lett. 2006, 47, 1461.
- (13) Liu, J.-F.; Jiang, Z.-Y.; Wang, R.-R.; Zheng, Y.-T.; Chen, J.-J.; Zhang, X.-M.; Ma, Y.-B. *Org. Lett.* 2007, *9*, 4127.
- (14) Communication from Shanghai Innovative Research Center of Traditional Chinese Medicine.
- (15) Karadeolian, A.; Kerr, M. A. Angew. Chem. Int. Ed. 2010, 49, 1133.
- (16) Andrews, M. D.; Brewster, A. G.; Moloney, M. G. Synlett 1996, 612.

- (17) Andrews, M. D.; Brewster, A. G.; Crapnell, K. M.; Ibbett, A. J.; Jones, T.; Moloney, M. G.; Prout, K.; Watkin, D. J. Chem. Soc., Perkin Trans. 1 1998, 223.
- (18) The diffraction data for **7b** and **15a** were collected at 150 K^{37} using an Enraf-Nonius KCCD diffractometer.³⁸ Structures were solved using SIR92,39 and refined using the CRYSTALS software suite⁴⁰ as per the supplementary information (CIF file). The Flack x parameter ⁴¹ for **7b** refined to -0.2 (8), however analysis of the Bijvoet pairs gave a Hooft y parameter⁴² of 0.0(3) giving a 99.2% probability that the structure is of the correct handedness (assuming full enantiopurity).43 In the absence of a strong anomalous signal, the Friedel pairs were merged for the final refinement. The Flack x parameter for **15a** refined to -0.02(3). Crystallographic data (excluding structure factors) have been deposited with the Cambridge Crystallographic Data Centre [CCDC 800835 (7b), CCDC 800836 (15a)] and copies of these data can be obtained via www.ccdc.cam.ac.uk/data_request/cif.
- (19) Galeotti, N.; Poncet, J.; Chiche, L.; Jouin, P. J. Org. Chem. 1993, 58, 5370.
- (20) Andrews, M. D.; Brewster, A. G.; Moloney, M. G. J. Chem. Soc., Perkin Trans. 1 2002, 80.
- (21) Ling, T.; Macherla, V. R.; Manam, R. R.; McArthur, K. A.; Potts, B. C. M. Org. Lett. 2007, 9, 2289.
- (22) Cottrell, I. F.; Cowley, A. R.; Croft, L. J.; Hymns, L.; Moloney, M. G.; Nettleton, E. J.; Smithies, H. K.; Thompson, A. L. *Tetrahedron* 2009, *65*, 2537.
- (23) Cottrell, I. F.; Moloney, M. G.; Smithies, H. K. *Tetrahedron Lett.* 2009, *50*, 1097.
- (24) Moloney, M. G.; Nettleton, E.; Smithies, K. *Tetrahedron Lett.* **2002**, *43*, 907.
- (25) (a) Moloney, M. G. *Main Group Met. Chem.* 2001, *24*, 653.
 (b) Buston, J. E. H.; Claridge, T. D. W.; Moloney, M. G. *J. Chem. Soc., Perkin Trans.* 2 1995, 639.
- (26) Corey, E. J.; Reichard, G. A. J. Am. Chem. Soc. 1992, 114, 10677.
- (27) Bioassay of Compounds:⁴⁴ Microbiological assays were performed by the hole-plate method with the test organism *Staphylococcus aureus* N.C.T.C. 6571 or *E. coli* X580. Solutions (100 mL) of the compounds to be tested (4 mg/ mL) were loaded into wells in bioassay plates, and incubated overnight at 37 °C. The diameters of the resultant inhibition zones were measured (±1 mm).

- (28) Marvin was used for drawing, displaying and structure property prediction and calculation (ClogD_{7.4}, PSA, MSA and CMR calculations), Marvin 5.2.1, 2009, ChemAxon (http://www.chemaxon.com).
- (29) Ferrara, P.; Apostolakis, J.; Caflisch, A. Proteins: Struct., Funct., Bioinf. 2002, 46, 24.
- (30) (a) Padrón, J. A.; Carrasco, R.; Pellón, R. F. *J. Pharm. Pharmaceut. Sci.* **2002**, *5*, 258. (b) Ghose, A. K.; Crippen, G. M. *J. Chem. Inf. Comput. Sci.* **1987**, *27*, 21.
 (c) Viswanadhan, V. N.; Ghose, A. K.; Reyankar, G. R.; Robins, R. K. *J. Chem. Inf. Comput. Sci.* **1989**, *29*, 163.
- (31) Ertl, P.; Rohde, B.; Selzer, P. J. Med. Chem. 2000, 43, 3714.
 (32) Ertl, P. 'Polar Surface Area', in Molecular Drug Properties; Mannhold, R., Ed.; Wiley-VCH: Weinheim, 2007, 111–126.
- (33) Reymond, J.-L.; van Deursen, R.; Blum, L. C.; Ruddigkeit, L. MedChemComm 2010, 1, 30.
- (34) Hajduk, P. J.; Greer, J. Nat. Rev. Drug Discovery 2007, 211.
- (35) Anwar, M.; Moloney, M. G. *Tetrahedron Lett.* **2007**, *48*, 7259
- (36) (a) Bagwell, C. L.; Moloney, M. G.; Thompson, A. L. Bioorg. Med. Chem. Lett. 2008, 18, 4081. (b) Bagwell, C. L.; Moloney, M. G.; Yaqoob, M. Bioorg. Med. Chem. Lett. 2010, 20, 2090.
- (37) Cosier, J.; Glazer, A. M. J. Appl. Crystallogr. 1986, 19, 105.
- (38) Otwinowski, Z.; Minor, W. *Methods Enzymol.* **1997**, 276, 307.
- (39) Altomare, A.; Cascarano, G.; Giacovazzo, G.; Guagliardi, A.; Burla, M. C.; Polidori, G.; Camalli, M. J. Appl. Crystallogr. 1994, 27, 435.
- (40) Betteridge, P. W.; Carruthers, J. R.; Cooper, R. I.; Prout, K.; Watkin, D. J. J. Appl. Crystallogr. 2003, 36, 1487.
- (41) (a) Flack, H. D. *Acta Crystallogr., Sect. A* 1983, *39*, 876.
 (b) Flack, H. D.; Bernardinelli, G. *J. Appl. Crystallogr.* 2000, *33*, 1143.
- (42) Hooft, R. W. W.; Straver, L. H.; Spek, A. L. J. Appl. Crystallogr. 2008, 41, 96.
- (43) (a) Thompson, A. L.; Watkin, D. J.; Gal, Z. A.; Jones, L.; Hollinshead, J.; Jenkinson, S. F.; Fleet, G. W. J.; Nash, R. J. *Acta Crystallogr., Sect. C* 2008, 64, o649. (b) Thompson, A. L.; Watkin, D. J. *Tetrahedron: Asymmetry* 2009, 20, 712.
- (44) (a) Smith, B.; Warren, S. C.; Newton, G. G. F.; Abraham, E. P. *Biochem. J.* **1967**, *103*, 877. (b) Baldwin, J. E.; Coates, J. B.; Halpern, J.; Moloney, M. G.; Pratt, A. J. *Biochem. J.* **1989**, *261*, 197. (c) Baldwin, J. E.; Pratt, A. J.; Moloney, M. G. *Tetrahedron* **1987**, *43*, 2565.

Copyright of Synlett is the property of Georg Thieme Verlag Stuttgart and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.