

## Synthesis and antiproliferative activity of combretastatin derivatives with adamantane fragment

E. V. Nurieva,<sup>a</sup> N. A. Zefirov,<sup>a</sup> N. S. Zefirov,<sup>a,b</sup> S. A. Kuznetsov,<sup>c</sup> and O. N. Zefirova<sup>a,b\*</sup>

<sup>a</sup>M. V. Lomonosov Moscow State University, Department of Chemistry, Build. 3, 1 Leninskie Gory, 119991 Moscow, Russian Federation.

Fax: +7 (459) 939 0290. E-mail: olgaz@org.chem.msu.ru

<sup>b</sup>Institute of Physiologically Active Compounds, Russian Academy of Sciences, 1 Severnyi pr-d, 142432 Chernogolovka, Moscow Region, Russian Federation.

E-mail: kolaz92@gmail.com

<sup>c</sup>Institute of Biological Sciences, University of Rostock, 18106 Rostock, Germany.

E-mail: sergei.kuznetsov@uni-rostock.de

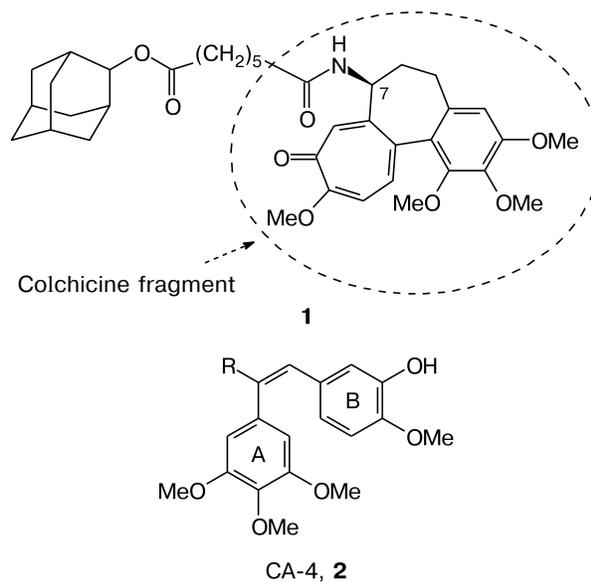
The work describes the synthesis of 2-adamantyl 7-[(2-[(2*E*)-3-(3-hydroxy-4-methoxyphenyl)-2-(3,4,5-trimethoxyphenyl)prop-2-enoyl]amino)ethyl]amino]-7-oxoheptanoate and 7-[(2-[(2*E*)-2-(3,4-dimethoxyphenyl)-3-(3-hydroxy-4-methoxyphenyl)prop-2-enoyl]amino)ethyl]amino]-7-oxoheptanoate. The latter compound exhibits moderate cytotoxicity ( $EC_{50} = 4.8 \mu\text{mol L}^{-1}$ ) against the human epithelial lung carcinoma cells A549.

**Key words:** combretastatin, adamantane, colchicine, tubuloclustin, tubulin, cytotoxicity, carcinoma A549.

Earlier,<sup>1–3</sup> in the search for compounds with anti-tumor activity we obtained C(7)-substituted derivatives of natural colchicine, among which were compounds (for example, tubuloclustin (**1**)) with very high cytotoxicity against different types of tumor cells *in vitro*.<sup>3,4</sup>

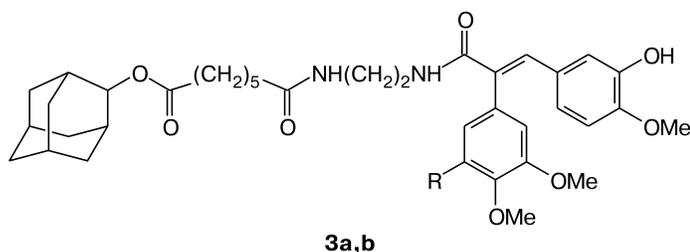
Attempted replacement of the colchicine moiety in compounds of the type **1** with other analogues possessing lower general toxicity, such as nocodazole and 2-methoxyextradiol, was unsuccessful because of either synthetic difficulties<sup>5,6</sup> or instability<sup>7</sup> and inactivity<sup>8</sup> of the compounds obtained. The purpose of the present work is the synthesis and biotesting of analogues of **1** based on the natural combretastatin A-4 (CA-4), whose antitumor activity is also related to the interaction with the colchicine binding site with its cell target protein tubulin.<sup>9</sup>

According to a pharmacophore model of the tubulin–colchicine site ligand complexes,<sup>10,11</sup> the nitrogen atom of the colchicine amide group is on a certain distance ( $\sim 3\text{--}3.5 \text{ \AA}$ ) from both carbon atoms of the combretastatin double bond. This necessitates the elongation of the linker connecting the adamantane core with CA-4, as compared to that of tubuloclustin (**1**). Among known CA-4 derivatives with substituents at the double bond carbon atoms appropriate for modification, the ability to cause the effect typical of colchicine,<sup>12</sup> *viz.*, the inhibition of tubulin polymerization approximately in the same concentrations as for colchicine, was confirmed only for com-



R = H (CA-4), C(O)NH<sub>2</sub> (**2**)

pound **2**. Therefore, we decided to synthesize compounds **3a,b**, in which a substituted adamantane group is attached to the amide group of **2**. We assumed that the introduction of two additional (as compared to **1**) methylene groups into the structures **3a,b** would provide the linker elongation mentioned above. In these studies, **3b** was the target



R = H (**3a**), OMe (**3b**)

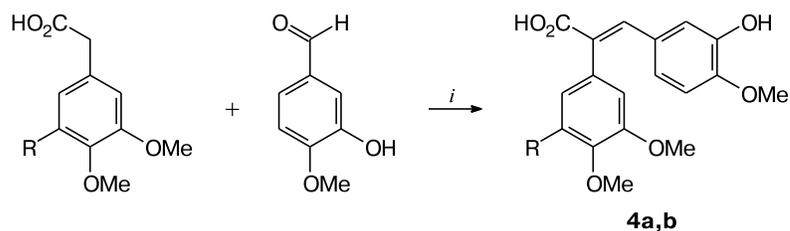
compound, whereas **3a** was used as a model for the studies of the role of trimethoxyphenyl group in **3b** in exhibiting cytotoxic properties.

The starting combretastatin derivatives **4a** and **4b** were obtained by Perkin condensation<sup>12,13</sup> from 3,4-dimethoxy- or 3,4,5-trimethoxyphenylacetic acid and 3-hydroxy-4-methoxybenzaldehyde in the presence of acetic anhydride and triethylamine (Scheme 1).

The synthesis of target compounds **3a,b** was carried out according to Scheme 2. In the first step, the reaction

of monoprotected *N*-Boc-ethylenediamine (**5**) and pimelic polyanhydride gave amide **6**. The <sup>13</sup>C NMR spectrum of this compound exhibits three signals characteristic of the carbonyl carbon atoms: the *tert*-butoxycarbonyl, the carboxy, and the amide ones at  $\delta$  157.02, 174.25, and 177.29, respectively. Then, pimelic acid monoamide **6** was esterified with adamantan-2-ol to ester **7**. In the <sup>1</sup>H NMR spectrum of compound **7**, the signal for the proton at atom C(2) of the adamantane group is downfield shifted ( $\delta$  4.89) as compared to the starting adamantan-2-ol ( $\delta$  3.83). A sim-

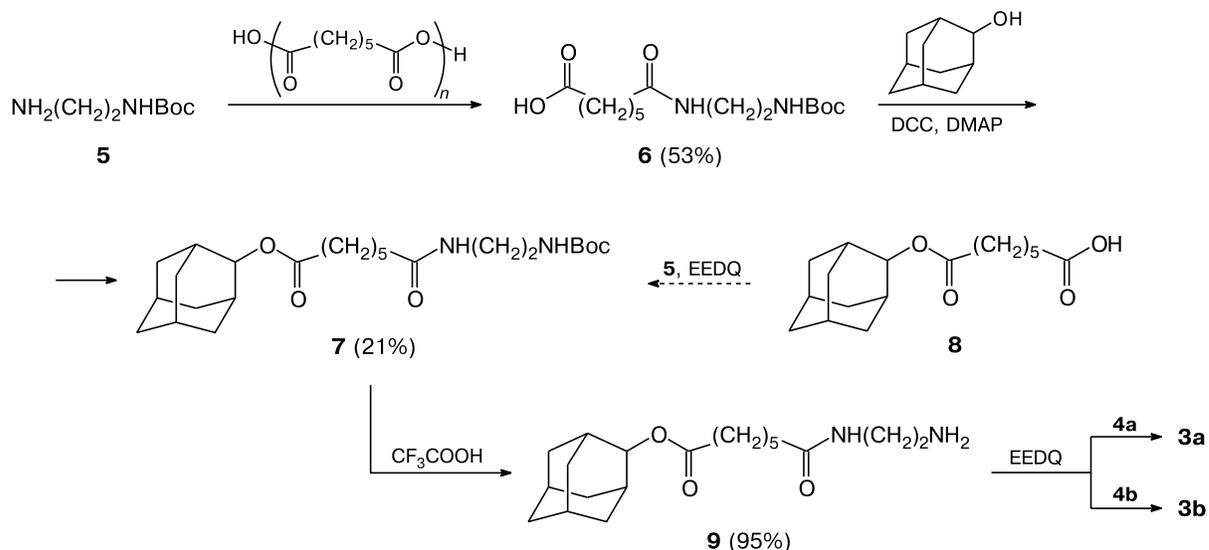
Scheme 1



R = H (**4a**, 34%), OMe (**4b**, 43%)

*i.* NEt<sub>3</sub>, Ac<sub>2</sub>O, heating.

Scheme 2



ilar shift is also observed for atom C(2) in the  $^{13}\text{C}$  NMR spectrum ( $\delta$  76.82 for **7** and  $\delta$  74.52 for Ad-2-OH).

It should be noted that an alternative order of assembling molecule **7**, namely, the addition of *N*-Boc-ethylenediamine to monoester of adamantan-2-ol and pimelic acid **8** (obtained according to procedure<sup>3</sup>) in the presence of 2-ethoxy-*N*-ethoxycarbonyl-1,2-dihydroquinoline (EEDQ) was preparatively unsuccessful because of very low yield of product **7** (see Experimental).

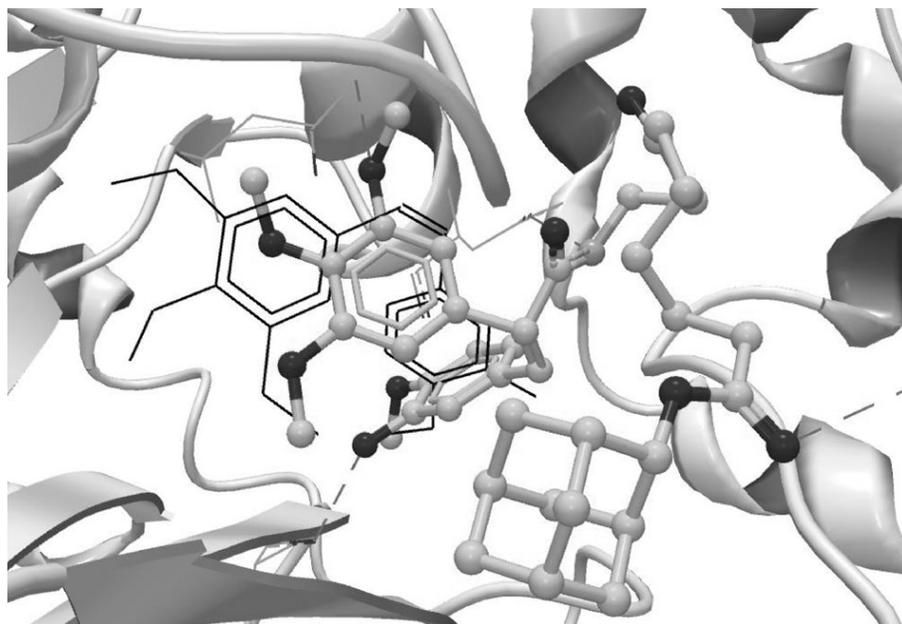
The protecting group in compound **7** was removed according to a standard procedure, the resulting amine **9** was involved in the reaction with combretastatin carboxy derivatives **4a,b** in the presence of EEDQ. The  $^{13}\text{C}$  NMR spectra of the target products exhibit the signals for three carbonyl carbon atoms in the range of  $\delta$  168–174 and the signals for the aromatic atoms of combretastatin fragment within  $\delta$  106–155. The  $^1\text{H}$  NMR spectroscopy and elemental analysis data also confirm the structure of compounds **3a** and **3b**.

The biotesting of conjugates **3a** and **3b** in the standard colorimetric MTT test<sup>14</sup> against the culture of human lung carcinoma cells A549 showed that compound **3b** possesses a noticeable, but a moderate cytotoxicity:  $\text{EC}_{50} = 4.8 \pm 0.1 \mu\text{mol L}^{-1}$  (for colchicine,  $\text{EC}_{50} = 0.04 \pm 0.09 \mu\text{mol L}^{-1}$ ). Compound **3a** is low active ( $\text{EC}_{50} > 50 \mu\text{mol L}^{-1}$ ).

Based on the assumption that the obtained values of cytotoxicity are determined only by the binding with protein (without considering metabolism, the ease of penetration into the cell, *etc.*), we attempted to explain the

results using computer molecular modeling. The results of the automatic docking of ligand **3b** into the 3D model of tubulin in the colchicine binding site<sup>10</sup> using the CLC Drug Discovery Workbench program showed that in one of the most favorable variants of arrangement of structure **3b** in the colchicine site (among several different variants with similar to each other and to compound **1** scoring functions), the combretastatin group of **3b** was placed into the region similar to that occupied by free CA-4 (Fig. 1).

The binding of **3b** with protein involves both methoxy groups in *meta*-positions of the aromatic ring. The oxygen atom of one of them forms a hydrogen bond with the hydrogen atom of the amide group of the main chain  $\beta\text{Leu255}$ , whereas the methyl group of the other one is involved in hydrophobic interactions with the side chains  $\beta\text{Ala354}$ ,  $\beta\text{Ala316}$ ,  $\beta\text{Val318}$  and the alkyl fragment of the side chain  $\beta\text{Lys352}$ . This explains a decrease in the activity for analogue **3b** without one *meta*-methoxy substituent in the aromatic ring (**3a**). A decrease in cytotoxicity of conjugate **3b** as compared to tubuloclastin (**1**) can be explained by a noticeable difference in the position of free CA-4 and that in conjugate **3b** (see Fig. 1), apparently, caused either by too high conformational "rigidity" or too long diamide chain of **3b**. Therefore, the further search for the tubulin ligands of the type under consideration should be focused on the analogues of **3b** with more "flexible" and shorter group between the CA-4 fragment and the substituted adamantane core. The approaches to the synthesis of such compounds are currently under study.



**Fig. 1.** One of the most energetically favorable variants of the arrangement of structure of **3b** in the tubulin dimer (the  $\beta$ -subunit is on the left) according to the results of automatic docking (CLC Drug Discovery Workbench). The molecule of free combretastatin A-4 is shown for comparison, hydrogen bonds are depicted in dashed lines (hydrogen atoms are omitted).

## Experimental

Automatic docking into the 3D model of the tubulin complex with *N*-deacetyl-*N*-(2-mercaptoacetyl)colchicine (a predetermined radius 16 Å) was performed using the CLC Drug Discovery Workbench program (Version 1.5): Evaluation license (2014).

Reaction progress and purity of compounds were monitored by TLC on Silufol-UV254 plates. Chromatographic separation was carried out on a column with Acros silica gel (40–60 μm). <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Bruker Avance 400 spectrometer (400 and 100 MHz, respectively) at 28 °C. Chemical shifts are given relative to the residual signals of the solvent. Elemental analysis was performed on a Vario micro cube CHN-analyzer. IR spectra were recorded on a IR-200 ThermoNicolet spectrophotometer in KBr pellets.

**(2*E*)-3-(3-Hydroxy-4-methoxyphenyl)-2-(3,4-dimethoxyphenyl)acrylic acid (4a).** 3,4-Dimethoxyphenylacetic acid (1.73 g, 8.82 mmol) was added to a solution of 3-hydroxy-4-methoxybenzaldehyde (0.67 g, 4.4 mmol) in Ac<sub>2</sub>O (4 mL, 42.4 mmol) and NEt<sub>3</sub> (2 mL, 14.29 mmol). The mixture was stirred for 6 h at 120 °C, poured onto ice (20 g), and acidified with concentrated HCl. A precipitate was filtered, washed with cold CH<sub>2</sub>Cl<sub>2</sub>, and recrystallized from EtOH to obtain compound **4a** (0.5 g, 34%), yellow crystals, m.p. 173–175 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>), δ: 3.83 (s, 3 H, Me); 3.88 (s, 3 H, Me); 3.94 (s, 3 H, Me); 6.69 (s, 1 H, C<sub>Ar</sub>(2)H); 6.70 (m, 2 H, C<sub>Ar</sub>(5)H and C<sub>Ar</sub>(6)H); 6.77 (d, 1 H, C<sub>Ar</sub>(2)H, *J* = 1.7 Hz); 6.83 (dd, 1 H, C<sub>Ar</sub>(5)H, *J* = 8.4 Hz, *J* = 1.7 Hz); 6.93 (d, 1 H, C<sub>Ar</sub>(6)H, *J* = 8.4 Hz); 7.83 (s, 1 H, C=CH). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>), δ: 55.87 (Me); 55.93 (Me); 55.98 (Me); 111.99, 112.31, 113.68, 117.61, 122.23 (C<sub>Ar</sub>(6)); 123.26 (C<sub>Ar</sub>(6)); 127.74 (C(2)); 129.34 (C<sub>Ar</sub>(1)); 130.77 (C<sub>Ar</sub>(1)); 139.39 (C(3)); 146.29 (C<sub>Ar</sub>(3)); 146.78 (C<sub>Ar</sub>(3)); 148.63 (C<sub>Ar</sub>(4)); 149.18 (C<sub>Ar</sub>(4)); 169.29 (C=O). IR (KBr), ν/cm<sup>-1</sup>: 808, 1024, 1136–1144, 1255, 1439, 1514, 1603, 1666 (C=O), 2837, 2953, 3417 (OH). Found (%): C, 65.52; H, 5.51. C<sub>18</sub>H<sub>18</sub>O<sub>6</sub>. Calculated (%): C, 65.45; H, 5.49.

**(2*E*)-3-(3-Hydroxy-4-methoxyphenyl)-2-(3,4,5-trimethoxyphenyl)acrylic acid (4b)** was obtained according to the procedure described earlier.<sup>12</sup> The yield was 43%, yellow crystals, m.p. 238–240 °C (*cf.* Ref. 13: m.p. 237–239 °C). Spectral data agree with those given in the literature.<sup>12</sup>

**7-[2-(*tert*-Butoxycarbonylamino)ethylamino]-7-oxoheptanoic acid (6).** Amine **5** (0.64 g, 4 mmol) was added to a solution of pimelic anhydride (0.715 g, 5 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL). After stirring the mixture for 24 h at 20 °C, it was diluted with Et<sub>2</sub>O (15 mL) and washed with water. The organic fraction was dried with MgSO<sub>4</sub> and concentrated, the residue was subjected to chromatography (eluent ethyl acetate–light petroleum ether (40–60 °C), 1 : 1; then methanol–dichloromethane, 1 : 10) to obtain compound **6** (0.638 g, 53%), a colorless oily liquid. <sup>1</sup>H NMR (CDCl<sub>3</sub>), δ: 1.35 (m, 2 H, C<sub>γ</sub>H<sub>2</sub>); 1.41 (s, 9 H, Bu<sup>t</sup>); 1.62 (m, 4 H, C<sub>β</sub>H<sub>2</sub>); 2.19 (m, 2 H, C<sub>α</sub>H<sub>2</sub>); 2.32 (m, 2 H, C<sub>α</sub>H<sub>2</sub>); 3.29 (m, 2 H, CH<sub>2</sub>NHCO); 3.32 (m, 2 H, CH<sub>2</sub>NHBoc); 5.38 (br.s, 1 H, NH); 6.80 (br.s, 1 H, NH). <sup>13</sup>C NMR (CDCl<sub>3</sub>), δ: 24.37, 25.25, 28.36 (Me); 28.52, 33.83 (C<sub>o</sub>); 36.02 (CH<sub>2</sub>CONH); 36.21 (CH<sub>2</sub>NHCO); 40.16 (CH<sub>2</sub>NHBoc); 79.62 (CMe<sub>3</sub>); 157.02 (CO<sub>2</sub>Bu<sup>t</sup>); 174.25 (CO<sub>2</sub>H); 177.29 (CONH). Found (%): C, 55.60; H, 8.69; N, 9.25. C<sub>14</sub>H<sub>26</sub>N<sub>2</sub>O<sub>5</sub>. Calculated (%): C, 55.61; H, 8.67; N, 9.26.

**2-Adamantyl 7-[2-(*tert*-butoxycarbonylamino)ethylamino]-7-oxoheptanoate (7).** Method *A*. 1,3-Dicyclohexylcarbodiimide

(0.494 g, 2.4 mmol), adamantan-2-ol (0.357 g, 2.3 mmol), and a catalytic amount of DMAP were added to a solution of acid **6** (0.638 g, 2.1 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL). The reaction mixture was stirred for 20 h at 20 °C, concentrated, diluted with ethyl acetate (10 mL), and allowed to stand for 3 h at 0–5 °C. The crystals formed were filtered off, the solution was concentrated, the residue was subjected to chromatography (eluent ethyl acetate–light petroleum ether (40–60 °C), 1 : 1) to obtain compound **7** (0.187 g, 21%), a colorless oily liquid.

**Method B.** A solution of 7-(2-adamantylloxy)-7-oxoheptanoic acid **8** (0.298 g, 0.001 mol), amine **5** (0.131 mL, 0.83 mmol), EEDQ (0.329 g, 1.33 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was stirred for 24 h at 20 °C; the mixture was concentrated, the residue was subjected to chromatography (eluent ethyl acetate–light petroleum ether (40–60 °C), 1 : 2; then methanol–dichloromethane, 1 : 15) to obtain compound **7** (0.020 g, 5%). <sup>1</sup>H NMR (CDCl<sub>3</sub>), δ: 1.38 (m, 2 H, C<sub>γ</sub>H<sub>2</sub>); 1.46 (s, 9 H, Bu<sup>t</sup>); 1.56–1.59 (m, 2 H, Ad); 1.64–1.72 (m, 4 H, C<sub>β</sub>H<sub>2</sub>); 1.75–1.80 (m, 4 H, Ad); 1.85 (m, 4 H, Ad); 1.89–2.03 (m, 4 H, Ad); 2.16 (t, 2 H, C<sub>α</sub>H<sub>2</sub>, <sup>3</sup>J<sub>α,β</sub> = 7.8 Hz); 2.31 (t, 2 H, C<sub>α</sub>H<sub>2</sub>, <sup>3</sup>J<sub>α,β</sub> = 7.4 Hz); 3.25 (m, 2 H, BocNH–CH<sub>2</sub>); 3.33 (m, 2 H, CH<sub>2</sub>NHC=O); 4.89 (m, 1 H, C<sub>Ad</sub>(2)H); 5.21 (br.s, 1 H, NH); 6.49 (br.s, 1 H, NH). <sup>13</sup>C NMR (CDCl<sub>3</sub>), δ: 24.76, 25.28, 26.98, 27.21, 28.35 (Me); 28.71 (γ-CH<sub>2</sub>); 31.77 (C<sub>Ad</sub>); 31.86 (C<sub>Ad</sub>); 34.61 (CH<sub>2</sub>CO<sub>2</sub>); 36.31 (C<sub>Ad</sub>); 36.39 (CH<sub>2</sub>CONH); 37.36, 40.29 (CH<sub>2</sub>NHCO); 40.68 (CH<sub>2</sub>NHBoc); 76.82 (C<sub>Ad</sub>(2)); 79.60 (CMe<sub>3</sub>); 158.18 (CO<sub>2</sub>Bu<sup>t</sup>); 173.10 (C=O); 173.63 (C=O). Found (%): C, 66.09; H, 9.19; N, 6.40. C<sub>24</sub>H<sub>40</sub>N<sub>2</sub>O<sub>5</sub>. Calculated (%): C, 66.03; H, 9.23; N, 6.42.

**2-Adamantyl 7-(2-aminoethylamino)-7-oxoheptanoate (9).** Trifluoroacetic acid (0.5 mL, 6.52 mmol) was added to protected amine **7** (0.187 g, 0.429 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2.5 mL). The mixture was stirred for 3 h at 20 °C and concentrated. The residue was diluted with CH<sub>2</sub>Cl<sub>2</sub> (10 mL), washed with saturated aqueous NaHCO<sub>3</sub>, dried with MgSO<sub>4</sub>, and concentrated to obtain compound **9** (0.155 g, 95%), a colorless oily liquid. <sup>1</sup>H NMR (CDCl<sub>3</sub>), δ: 1.36 (m, 2 H, C<sub>γ</sub>H<sub>2</sub>); 1.38–1.48 (br.s, 2 H, NH<sub>2</sub>); 1.55 (m, 2 H, Ad); 1.65 (m, 4 H, 2 C<sub>β</sub>H<sub>2</sub>); 1.72–1.76 (m, 4 H, Ad); 1.82 (m, 4 H, Ad); 1.96–2.00 (m, 4 H, Ad); 2.19 (t, 2 H, CH<sub>2</sub>CONH, <sup>3</sup>J<sub>α,β</sub> = 7.6 Hz); 2.33 (m, 2 H, CH<sub>2</sub>CO<sub>2</sub>); 2.84 (t, 2 H, CH<sub>2</sub>NH<sub>2</sub>, <sup>3</sup>J = 5.5 Hz); 3.30 (m, 2 H, CH<sub>2</sub>NHCO, <sup>3</sup>J = 5.5 Hz); 4.90 (m, 1 H, C<sub>Ad</sub>(2)H); 6.09 (br.s, 1 H, NH). <sup>13</sup>C NMR (CDCl<sub>3</sub>), δ: 24.77, 25.53, 26.97, 27.20, 28.70, 31.76 (Ad); 31.87 (Ad); 34.6, 36.31 (Ad); 36.41, 37.36, 41.24 (CH<sub>2</sub>NH<sub>2</sub>); 41.55 (CH<sub>2</sub>NH); 76.83 (C<sub>Ad</sub>(2)H); 173.14 (C=O); 173.39 (C=O). Found (%): C, 67.76; H, 9.64; N, 8.39. C<sub>19</sub>H<sub>32</sub>N<sub>2</sub>O<sub>3</sub>. Calculated (%): C, 67.82; H, 9.59; N, 8.33.

**2-Adamantyl 7-{2-[(2*E*)-2-(3,4-dimethoxyphenyl)-3-(3-hydroxy-4-methoxyphenyl)prop-2-enoylamino]ethylamino}-7-oxoheptanoate (3a)** Acid **4a** (0.030 g, 0.09 mmol) and EEDQ (0.15 g, 0.607 mmol) were added to a solution of amine **9** (0.123 g, 0.366 mmol) in CH<sub>2</sub>Cl<sub>2</sub>. The mixture was stirred for 24 h at 20 °C and concentrated, the residue was subjected to chromatography (eluent ethyl acetate–light petroleum ether (40–60 °C), 1 : 1; then methanol–dichloromethane, 1 : 10) to obtain compound **3a** (0.050 g, 52%), a white solid substance, m.p. 130–135 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>), δ: 1.35 (m, 2 H, C<sub>γ</sub>H<sub>2</sub>); 1.53–1.56 (m, 2 H, Ad); 1.62–1.68 (m, 4 H, 2 C<sub>β</sub>H<sub>2</sub>); 1.73–1.77 (m, 4 H, Ad); 1.82 (m, 4 H, Ad); 1.97–2.00 (m, 4 H, Ad); 2.17 (t, 2 H, C<sub>α</sub>H<sub>2</sub>, <sup>3</sup>J<sub>α,β</sub> = 7.6 Hz); 2.32 (t, 2 H, C<sub>α</sub>H<sub>2</sub>, <sup>3</sup>J<sub>α,β</sub> = 7.6 Hz); 3.34 (m, 2 H, BocNHCH<sub>2</sub>); 3.43 (m, 2 H, CH<sub>2</sub>NHC=O); 3.83 (s, 6 H, 2 OMe); 3.95 (s, 3 H, OMe); 4.90 (m, 1 H, C<sub>Ad</sub>(2)H); 5.99 (s, 1 H,

OH); 6.02 (br.s, 1 H, NH); 6.53 (br.s, 1 H, NH); 6.57 (dd, 1 H, Ar,  $J = 8.7$  Hz,  $J = 1.7$  Hz); 6.65 (d, 1 H, Ar,  $J = 8.7$  Hz); 6.67 (d, 1 H, Ar,  $J = 1.8$  Hz); 6.72 (d, 1 H, Ar,  $J = 1.7$  Hz); 6.80 (dd, 1 H, Ar,  $J = 8.2$  Hz,  $J = 1.7$  Hz); 6.96 (d, 1 H, Ar,  $J = 8.2$  Hz); 7.72 (s, 1 H, C=CHAR).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ),  $\delta$ : 24.81, 25.31, 26.95, 27.19, 28.73, 31.75, 31.83, 34.61, 36.29, 36.36, 37.35, 39.83 (NH— $\text{C}_\alpha\text{H}_2$ ), 40.41 (NH— $\text{C}_\beta\text{H}_2$ ), 55.78 (Me); 55.87 (Me), 55.99 (Me), 76.79 ( $\text{C}_{\text{Ad}}(2)$ ), 110.36, 112.12, 112.60; 116.82, 122.14, 123.26, 127.74, 128.10, 131.63, 137.24, 145.24, 147.51, 149.18, 149.91, 169.12 (C=O), 173.11 (C=O), 173.6 (C=O). Found (%): C, 68.45; 7.38; N, 4.35.  $\text{C}_{37}\text{H}_{48}\text{N}_2\text{O}_8$ . Calculated (%): C, 68.50; H, 7.46; N, 4.32.

**2-Adamantyl 7-{2-[(2E)-3-(3-hydroxy-4-methoxyphenyl)-2-(3,4,5-trimethoxyphenyl)prop-2-enoylamino]ethylamino}-7-oxoheptanoate (3b)** was obtained similarly to compound **3a** from amine **9** (0.123 g, 0.366 mmol), acid **4b** (0.16 g, 0.444 mmol), and EEDQ (0.15 g, 0.607 mmol). The yield was 0.037 g (16%), a white wax-like mass.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ),  $\delta$ : 1.33–1.38 (m, 2 H,  $\text{C}_\gamma\text{H}_2$ ); 1.53–1.56 (m, 2 H, Ad); 1.62–1.65 (m, 4 H,  $\text{C}_\beta\text{H}_2$ ); 1.73–1.77 (m, 4 H, Ad); 1.82 (m, 4 H, Ad); 1.92–2.0 (m, 4 H, Ad); 2.15–2.18 (t, 2 H,  $\text{C}_\alpha\text{H}_2$ ,  $^3J_{\alpha,\beta} = 7.5$  Hz); 2.31 (t, 2 H,  $\text{C}_\alpha\text{H}_2$ ,  $^3J_{\alpha,\beta} = 7.1$  Hz); 3.34 (m, 2 H, BocNH— $\text{C}_\beta\text{H}_2$ ); 3.44 (m, 2 H,  $\text{CH}_2$ —NHC=O); 3.81 (s, 6 H, OMe); 3.84 (s, 3 H, OMe); 3.93 (s, 3 H, OMe); 4.90 (m, 1 H,  $\text{C}_{\text{Ad}}(2)\text{H}$ ); 6.05 (br.s, 2 H, 2 NH); 6.45 (s, 2 H); 6.52 (m, 1 H); 6.55 (dd, 1 H, Ar,  $J = 8.4$  Hz,  $J = 1.9$  Hz); 6.65 (d, 1 H, Ar,  $J = 8.4$  Hz); 6.69 (d, 1 H, Ar,  $J = 1.9$  Hz); 7.7 (s, 1 H, C=CHAR).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ),  $\delta$ : 24.8, 25.3, 26.97, 27.2, 28.74, 31.76, 36.3, 37.37, 39.9 (NH— $\text{C}_\alpha\text{H}_2$ ); 40.45 (NH— $\text{C}_\beta\text{H}_2$ ); 55.81 (Me); 56.27 (2 Me), 61.03 (Me); 76.81 ( $\text{C}_{\text{Ad}}(2)$ ); 106.56, 110.37, 116.81, 123.26, 127.92, 131.13, 131.80, 137.22, 138.18, 145.26, 147.54, 154.33, 168.72 (C=O); 173.09 (C=O); 173.57 (C=O). IR (KBr),  $\nu/\text{cm}^{-1}$ : 1128, 1240, 1257, 1412, 1452, 1512, 1581, 1651, 1725 (C=O); 2854, 2927, 3369 (OH and NH). Found (%): C, 67.20; H, 7.38; N, 4.07.  $\text{C}_{38}\text{H}_{50}\text{N}_2\text{O}_9$ . Calculated (%): C, 67.24; H, 7.42; N, 4.13.

**MTT-test on cytotoxicity.** The human epithelial lung carcinoma cells (line A-549, CCL-185) were seeded in 96-well plates (~3000 cells per well) in DMEM culture medium (200  $\mu\text{L}$ ) at 37 °C. Solutions of tested compounds (or colchicine as a positive control) in DMSO were added into the wells in the concentration range 0.005–50  $\mu\text{mol L}^{-1}$  (eight wells for each concentration) and this was matured for 24 h. A solution of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) (5 mg  $\text{mL}^{-1}$ ) was prepared in a phosphate buffer, filtered through a 0.22-mm filter, and 2 h before the end of maturing the tested compound, the solution of MTT (20  $\mu\text{L}$ ) was added into each well so that the final concentration would be 0.45 mg  $\text{mL}^{-1}$ . After removal of the cell medium, a solution of DMSO containing 10% of sodium dodecylsulfate and 0.6% of acetic acid (100  $\mu\text{L}$ ) was added into each well, the crystals of formazane formed were solubilized by stirring, using a shaker. Optical density was measured at 590 nm with a 690-nm reference filter on a EL808 Ultra Microplate Reader instrument (Bio-Tek Instruments, USA).

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## References

1. O. N. Zefirova, E. V. Nurieva, H. Lemcke, A. A. Ivanov, D. V. Shishov, D. G. Weiss, S. A. Kuznetsov, N. S. Zefirov, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 5091.
2. O. N. Zefirova, E. V. Nurieva, H. Lemcke, A. A. Ivanov, N. V. Zyk, D. G. Weiss, S. A. Kuznetsov, N. S. Zefirov, *Mendelev Commun.*, 2008, **18**, 183.
3. O. N. Zefirova, E. V. Nurieva, D. V. Shishov, I. I. Baskin, F. Fuchs, H. Lemcke, F. Schröder, D. G. Weiss, N. S. Zefirov, S. A. Kuznetsov, *Bioorg. Med. Chem.*, 2011, **19**, 5529.
4. O. N. Zefirova, H. Lemcke, M. Lantow, E. V. Nurieva, B. Wobith, G. E. Onishchenko, A. Hoenen, G. Griffiths, N. S. Zefirov, S. A. Kuznetsov, *ChemBioChem.*, 2013, **14**, 1444.
5. E. V. Nurieva, A. A. Beloglazkina, D. V. Shishov, V. V. Gogol, Ya. S. Glazkova, B. Wobith, N. S. Zefirov, S. A. Kuznetsov, O. N. Zefirova, *Moscow University Chem. Bull.*, 2013, **68**, 37 [*Vestn. Mosk. un-ta. Ser. 2. Khim.*, 2013, **54**, 45].
6. N. S. Zefirov, Ya. S. Glazkova, I. V. Kuznetsova, E. V. Nurieva, O. N. Zefirova, *Moscow University Chem. Bull.*, 2015, **70**, 69 [*Vestn. Mosk. un-ta. Ser. 2. Khim.*, 2015, **56**, 85].
7. O. N. Zefirova, E. V. Nurieva, Ya. S. Glazkova, N. A. Zefirov, A. V. Mamaeva, B. Wobith, V. I. Romanenko, N. A. Lesnaya, E. M. Treschalina, S. A. Kuznetsov, *Pharm. Chem. J. (Engl. Transl.)*, 2014, **48**, 373 [*Khim. Farm. Zh.*, 2014, **48**, 19].
8. O. N. Zefirova, Ya. S. Glazkova, E. V. Nurieva, N. A. Zefirov, A. V. Mamaeva, B. Wobith, N. S. Zefirov, S. A. Kuznetsov, *Russ. Chem. Bull. (Int. Ed.)*, 2014, **63**, 1126 [*Izv. Akad. Nauk, Ser. Khim.*, 2014, 1126].
9. N. H. Nam, *Curr. Med. Chem.*, 2003, **10**, 1697.
10. T. L. Nguyen, C. McGrath, A. R. Hermone, J. C. Burnett, D. W. Zaharevitz, B. W. Day, P. Wirf, E. Hamel, R. Gussio, *J. Med. Chem.*, 2005, **48**, 6107.
11. O. N. Zefirova, A. G. Diikov, N. V. Zyk, N. S. Zefirov, *Russ. Chem. Bull. (Int. Ed.)*, 2007, **56**, 680 [*Izv. Akad. Nauk, Ser. Khim.*, 2007, 655].
12. C. Borrel, S. Thoret, X. Cachet, D. Guénard, F. Tillequin, M. Koch, S. Michel, *Bioorg. Med. Chem.*, 2005, **13**, 3853.
13. K. Gaukroger, J. A. Hadfield, L. A. Hepworth, *J. Org. Chem.*, 2001, **66**, 8135.
14. T. Mosmann, *J. Immunol. Methods*, 1983, **65**, 55.

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