

Facile and *E*-Selective Intramolecular Ring-Closing Metathesis Reactions in 3_{10} -Helical Peptides: A 3D Structural StudyAmie K. Boal,[†] Ivan Guryanov,[‡] Alessandro Moretto,[‡] Marco Crisma,[‡] Erica L. Lanni,[†]
Claudio Toniolo,[‡] Robert H. Grubbs,[§] and Daniel J. O'Leary^{*,†}

Department of Chemistry, Pomona College, Claremont, California 91711, Institute of Biomolecular Chemistry, Padova Unit, CNR, Department of Chemistry, University of Padova, 35131 Padova, Italy, and Arnold and Mabel Beckman Laboratories of Chemical Synthesis, Division of Chemistry and Chemical Engineering, California Institute of Technology, Pasadena, California 91125

Received February 16, 2007; E-mail: doleary@pomona.edu

The intramolecular ring-closing metathesis reaction (RCM) is a useful method for altering the conformational and metabolic stability of α -helical peptides.^{1–8} Prior RCM investigations have utilized tethers spanning i and $i + 4$ or $i + 7$ amino acid residues, a linkage that encompasses approximately one or two turns of an α -helical backbone and places the reactive side chains on the same side of the helix (Figure 1). This strategy has built upon earlier work with α -helices containing tethers employing salt bridges,⁹ lactams,¹⁰ disulfide bridges,¹¹ hydrophobic effects,¹² and metal ligation.¹³

Herein, we report the development of a minimal RCM constraint for the 3_{10} -helix, which is a relatively common structural motif in proteins and peptides containing C $^{\alpha}$ -tetrasubstituted α -amino acids.^{14–16} The stereochemistry of the 3_{10} -helix¹⁷ suggests that its regularity can be affected by i , $i + 3$ cross-links (Figure 1). This aspect has been investigated for the case of salt¹⁸ and lactam¹⁹ side-chain bridges. A recent theoretical study suggested that a minimal RCM constraint for a 3_{10} -helix would require two five-atom i , $i + 3$ olefinic side chains, thus producing an 18-atom macrocycle upon ring closure.²⁰

To study this proposition in greater detail, an octapeptide with the sequence Boc-Aib-Aib-Aib-L-Ser(Al)-Aib-Aib-L-Ser(Al)-Aib-OMe (Boc, *tert*-butoxy; Aib, α -aminoisobutyric acid; Al, allyl; OMe, methoxy) (**1**) was prepared using solution-phase methods.²¹ We chose this sequence because short oligopeptides containing Aib residues largely populate 3_{10} -helices.^{14,16,22} When treated with the second-generation ruthenium catalyst **4** (7 mol % of **4**, 5 mM in **1**, 40 °C, 30 min), diene **1** underwent a rapid and *E*-selective (>20:1) ring-closing reaction to yield an 18-membered macrocycle in 93% yield (Scheme 1). This result is interesting because *E*/*Z* mixtures are normally observed in RCM reactions between side chains in helical peptides.^{1–3} The olefin moiety in peptide **2** was reduced (cat. 10% Pd–C, 1 atm H₂, EtOH, 25 °C, 6 h) to provide the saturated macrocycle **3** in excellent yield.

An X-ray crystallographic analysis²³ (Figure 2) of peptides **1–3** provided a structural comparison at each stage of the modification. Each of the three peptides adopts a well-developed right-handed 3_{10} -helical structure. Peptide **1** is 3_{10} -helical for residues 1–6 and contains a type-I β -turn at the C-terminal residues 6 and 7 (a 3_{10} -helix consists of repeat type-III β -turns). This C-terminal turn behavior is also seen in peptides **2** and **3**, where the regularity of the helix is slightly disturbed at residues 4 and 5, with a deviation greater for alkene **2** than for the saturated macrocycle **3**. Despite these small differences, the structures are quite similar to one another, with rms deviations for backbone atoms of 0.996 Å

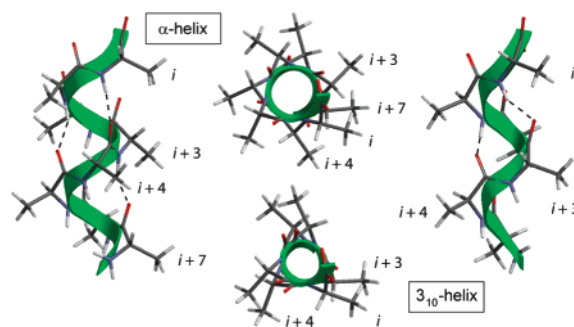
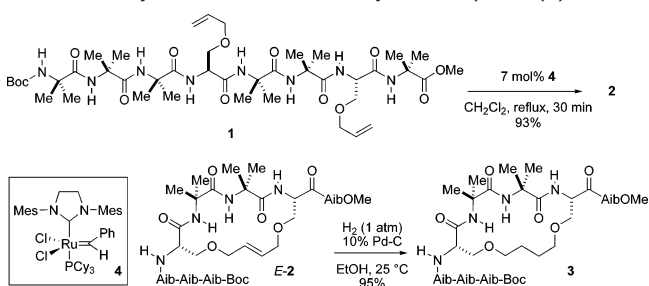


Figure 1. Molecular models for (L-Ala)_n α - and 3_{10} -helices. Intramolecular hydrogen bonds are indicated with dashed lines.

Scheme 1. Synthesis of RCM Macrocyclized Peptides (*E*)-**2** and **3**

between peptides **1** and **2** and 0.624 Å between peptides **1** and **3**. With the exception of the C-terminal residue 8, which is helical in **1** and **2** while semi-extended in **3**, most of the backbone ϕ , ψ torsion angle values of corresponding residues in **2** and **3** do not differ by more than 10° if compared to **1**. For **2**, the largest ϕ , ψ deviations are observed at Ser(4) and Ser(7) [$|\Delta\phi|$, $|\Delta\psi|$ = 22°, 39° and 14°, 16°, respectively]. For **3**, deviations within 10–16° are found for ψ_2 , ψ_5 , ϕ_6 , and ψ_6 . As commonly found,^{14,16,22} all internal Aib residues exhibit ϕ , ψ torsion angles typical of helical residues. In alkene **2**, the 3_{10} -helical H-bonding pattern is interrupted by the lack of the intramolecular H-bond between N6 and O3, as each of these two atoms is intermolecularly H-bonded to a co-crystallized solvent molecule. In **3**, the N6...O3 separation, 3.573(4) Å, is only slightly above the upper limit for a C=O...H–N H-bond. To the best of our knowledge, this is the first X-ray diffraction 3D structural comparison of a helical peptide before and after installation of a side-chain cross-link, RCM-derived or otherwise.

We note that in methanol solution peptides **1–3** exhibited circular dichroism (CD) spectra consistent with 3_{10} -helical structures²⁴ (Figure 3). This helix is characterized by a strong negative maximum near 205 nm and a much weaker (60–75% less intense) negative maximum at 222–232 nm.

[†] Pomona College.[‡] University of Padova.[§] California Institute of Technology.

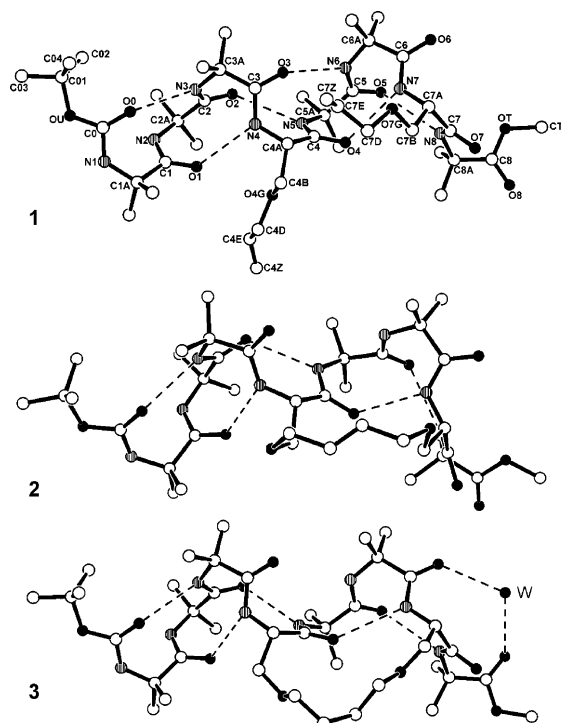


Figure 2. X-ray crystal structures of octapeptides 1–3. Hydrogen atoms have been omitted for clarity. Dashed lines represent intramolecular N–H···O=C hydrogen bonds. In 3, the co-crystallized water molecule (W) is also shown.

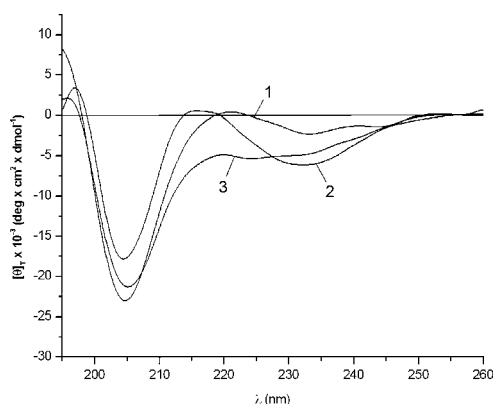


Figure 3. CD spectra of peptides 1–3 (1 mM in MeOH) at 25 °C.

Concerning the highly *E*-selective RCM reactivity of octapeptide diene **1**, we note that rapid RCM reactions and 12:1 *E*-selectivity are observed in a shorter sequence, the hexapeptide Boc-Aib-L-Ser(Al)-Aib-Aib-L-Ser(Al)-Aib-OMe (**5**). We have also investigated the RCM reaction in a heptapeptide with the sequence Boc-Val-Ser(Al)-Leu-Aib-Ser(Al)-Val-Leu-OMe (**6**).²⁵ When treated with the second-generation ruthenium catalyst **4** (10 mol % of **4**, 5 mM in **6**, 40 °C, 3 h), diene **6** formed an 18-membered macrocycle in quantitative yield with 7:1 *E/Z*-selectivity. The origin of the higher *E*-selectivity in the Aib-rich peptides may be due to ϕ/ψ conformational restrictions imposed by the C $^{\alpha}$ -tetrasubstituted α -amino residues. CD curves in 2,2,2-trifluoroethanol solution comparable to those of Figure 3 have been also obtained for the RCM macrocyclic products derived from both hexapeptide **5** and heptapeptide **6** (spectra not shown).

In conclusion, we have shown that an RCM-derived 18-membered macrocycle can be used to cross-link the side chains of *i* and *i* + 3 amino acids in short 3₁₀-helical peptide sequences. The intramolecular RCM reactions are efficient and highly *E*-selective,

especially in peptides with high Aib content. In an Aib-rich octapeptide, this macrocyclization does not significantly disturb 3₁₀-helicity, as judged by an X-ray diffraction study of acyclic diene **1**, *E*-olefin RCM product **2**, and its hydrogenated derivative **3**. While other sequences (also including C $^{\alpha}$ -tetrasubstituted α -amino acids with allyl side chains) and tether lengths remain to be studied, it is apparent from these studies that a minimal, RCM-derived, macrocyclic constraint can be readily incorporated into 3₁₀-helical peptides.

Acknowledgment. D.J.O. thanks the Mellon Foundation and Susan and David Hirsch for financial support. Work at Caltech was supported by the National Institutes of Health. We thank Lawrence M. Henling and Dr. Michael W. Day (Caltech) for providing the X-ray crystallographic analysis of peptide **2**, and Professor Helen Blackwell (University of Wisconsin) for helpful discussions.

Supporting Information Available: Preparative procedures and characterization data, including X-ray crystal structure coordinates and files in CIF format. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References

- Blackwell, H. E.; Grubbs, R. H. *Angew. Chem., Int. Ed.* **1998**, *37*, 3281.
- Blackwell, H. E.; Sadowsky, J. D.; Howard, R. J.; Sampson, J. N.; Chao, J. A.; Steinmetz, W. E.; O'Leary, D. J.; Grubbs, R. H. *J. Org. Chem.* **2001**, *66*, 5291.
- Schafmeister, C. E.; Po, J.; Verdine, G. L. *J. Am. Chem. Soc.* **2000**, *122*, 5891.
- Walensky, L. D.; Kung, A. L.; Escher, I.; Malia, T. J.; Barbuto, S.; Wright, R. D.; Wagner, G.; Verdine, G. L.; Korsmeyer, S. J. *Science* **2004**, *305*, 1466.
- Chapman, R. N.; Dimartino, G.; Arora, P. S. *J. Am. Chem. Soc.* **2004**, *126*, 12252.
- Wang, D.; Chen, K.; Kulp, J. L., III; Arora, P. S. *J. Am. Chem. Soc.* **2006**, *128*, 9248.
- Wang, D.; Chen, K.; Dimartino, G.; Arora, P. S. *Org. Biomol. Chem.* **2006**, *4*, 4074.
- Walensky, L. D.; Pitter, K.; Morash, J.; Oh, K. J.; Barbuto, S.; Fisher, J.; Smith, E.; Verdine, G. L.; Korsmeyer, S. J. *Mol. Cell* **2006**, *24*, 199.
- Scholtz, J. M.; Qian, H.; Robbins, V. H.; Baldwin, R. L. *Biochemistry* **1993**, *32*, 9668 and references cited within.
- Phelan, J. C.; Skelton, N. J.; Braisted, A. C.; McDowell, R. S. *J. Am. Chem. Soc.* **1997**, *119*, 455 and references cited within.
- Jackson, D. Y.; King, D. S.; Chmielewski, J.; Singh, S.; Schultz, P. G. *J. Am. Chem. Soc.* **1991**, *113*, 9391.
- Albert, J. S.; Hamilton, A. D. *Biochemistry* **1995**, *34*, 984.
- Kelso, M. J.; Hoang, H. N.; Oliver, W.; Sokolenko, N.; March, D. R.; Appleton, T. G.; Fairlie, D. P. *Angew. Chem., Int. Ed.* **2003**, *42*, 421 and references cited within.
- Karle, I. L.; Balam, P. *Biochemistry* **1990**, *29*, 6747.
- Toniolo, C.; Benedetti, E. *Trends Biochem. Sci.* **1991**, *16*, 350.
- Toniolo, C.; Crisma, M.; Formaggio, F.; Peggion, C. *Biopolymers* **2001**, *60*, 396 and references cited within.
- Relative to the α -helix ($\phi = -63^\circ$, $\psi = -42^\circ$, *i*, *i* + 4 hydrogen bonds), the 3₁₀-helix ($\phi = -57^\circ$, $\psi = -30^\circ$, *i*, *i* + 3 hydrogen bonds) is more tightly wound, contains a different pattern of intramolecular hydrogen bonds, and possesses a triangular shape when viewed down the long axis.¹⁵ The 3₁₀-helix is characterized by three amino acids per turn and 10 atoms in the pseudo-ring formed by an intramolecular *i*, *i* + 3 hydrogen bond (a type-III β -turn).
- Yokum, T. S.; Bursavich, M. G.; Gauthier, T.; Hammer, R. P.; McLaughlin, M. L. *Chem. Commun.* **1998**, 1801.
- Schievano, E.; Pagano, K.; Mammi, S.; Peggion, E. *Biopolymers* **2005**, *80*, 294 and references cited within.
- Saviano, M.; Benedetti, E.; Vitale, R. M.; Kaptein, B.; Broxterman, Q. B.; Crisma, M.; Formaggio, F.; Toniolo, C. *Macromolecules* **2002**, *35*, 4204.
- Full experimental details are provided in the Supporting Information.
- Marshall, G. R.; Hodgkin, E. E.; Langs, D. A.; Smith, G. D.; Zabrocki, J.; Leplawy, M. T. *Proc. Natl. Acad. Sci. U.S.A.* **1990**, *87*, 487.
- Peptide **1** was crystallized from ethyl acetate, peptide **2** from slow evaporation of a 2:1 dichloromethane/isopropanol solution, and peptide **3** from moist acetonitrile. The structures were solved by standard methods, and the atomic coordinates have been deposited with the Cambridge Crystallographic Data Centre.
- Toniolo, C.; Polese, A.; Formaggio, F.; Crisma, M.; Kamphuis, J. *J. Am. Chem. Soc.* **1996**, *118*, 2744.
- The chiral amino acids have the L configuration; this sequence is a permutation of that used in refs 1 and 2.

JA071148M