A Short and Simple Synthesis of the Thioglucose Analog of the Antitumor Agent Etoposide¹

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Abstract: the thioglucose-derived analogue of the antitumor agent etoposide has been synthesized from unprotected 4'-demethylepipodophyllotoxin.

Etoposide (1) is a semisynthetic podophyllotoxin derivative widely used alone ² or in combination with other drugs^{3,4}as an antitumor agent. Its clinical efficacy has stimulated the synthesis of various analogues with enhanced activity or active against etoposide-resistent tumor cells.⁵

In the course of studies aimed at the synthesis of analogues of (1), differing in the linkage between the sugar and the aglycone moieties, we have discovered a very simple route to the thioglucose-derived analogue (2).



The recent report⁶ concerning the synthesis of (2), which turns out to be also a valuable intermediate for obtaining the oxidized sulfur compounds (3 and 4), prompted us to publish our results which permit the synthesis of compound (2) in three steps starting from commercially available 2,3,4,6-tetra-O-acetyl-1 β -thio-D-glucopyranose (6) and 4'-demethylepipodophyllotoxin (5) which, under the conditions described, does not require any selective protection of the phenolic hydroxy group.

The key step for the synthesis is the simple direct glucosidation of (5) (1 eq) by reaction with (6) (1 eq) in the presence of BF₃. Et₂O (1 eq) (Scheme).

The glycosidation occurs in a short time, at room temperature and affords the C-4 β -diastereomer (7) in nearly quantitative yields (96%) and in a completely regio- and stereoselective manner.



(a) BF₃.Et₂O, CH₂Cl₂ (70 ml/mmol), 25^oC, 15 min; 96%. (b) 2 eq Zn(OAc)₂, McOH (6 ml/mmol), reflux. 40 h; 70%. (c) 21 eq MeCH(OMe)₂, 0.3 eq TsOH, MeNO₂ (12 ml/mmol), 25^oC, 2 h; 68%.

The stereochemistry of compound (7) was derived from the inspection of its ¹H-NMR spectrum (500 MHz),⁷ in particular the β -configuration at the anomeric center in a ${}^{4}C_{I}$ conformation of the pyranosidic ring, showed by a complete assignment of all pyranosidic proton signals, was evident from the axial-axial relationship of the anomeric proton and the adjacent glucosidic proton (J_{1",2"} 10 Hz) and the β -configuration at C-4 was derived from the value of the coupling constant (3.7 Hz) observed for H-3 and H-4 protons which are in a *cis* relationship⁸ [these protons show an higher (9.0-10.0 Hz) coupling constant in the 4 α configuration where are in a *trans* geometry].⁸ On the other hand the observed coupling constant values of the proton at C-3 and the adjacent protons (J_{2,3} 14.0 Hz, J_{3,11a} 10.0 Hz, J_{3,11b} 7.5 Hz) show that no epimerization of the lactonic function of (7), to afford a picropodophyllotoxin analogue, has occurred. The observed values are typical^{9,10} for podophyllotoxin series and are quite different from those observed for picropodophyllotoxin (J_{2,3} 9.0 Hz, J_{3,11a} 6.0 Hz, J_{3,11b} 1.5 Hz).

This result suggests that the condensation is a SN_1 type process that involves the attack of the glucosidic sulfur atom to the benzylic carbon ion at C-4 of the aglycone moiety generated by BF₃. Et₂O. This

Scheme

is remarkable since the glycosidation is performed on compound (5) without any protection of its phenolic hydroxy group. Previously reported glucosidations of (5) have $always^{11-13}$ carried out with selective protection of the phenolic hydroxy group, which on turn requires its successive regeneration.

Saponification of the acetate groups of (7) occurs in mild conditions without epimedization of the lactonic function and successive acetalization afford the target thioetoposide (2) in 45 % total yield from (5).

In conclusion our synthetic route to the thioanalogue of etoposide (2) represent the first direct regioand stereoselective glycosidation of the podophyllum lignan 4'-demethylepipodophylloloxin (5) and complements the reported⁶ approach which also obviates the use of a protecting group for the phenolic hydroxy group by starting from the unstable⁶ 4-bromo-4-deoxy-4'-demethylepipodophyllotoxin. Our method has the merit of a relative simplicity, almost quantitative yields in the key glycosidation, and utilizes the readily available and stable compound (5).^{14,15}

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- All new compounds have satisfactory elemental analyses and spectroscopic data confirming the 7. assigned structure. (2): mp 180-186 °C (dec); $[\alpha]_D^{27} = -114^\circ$ (c 1, CHCl₃); ¹H-NMR 500 MHz, CDCl₃: δ 6.89 (1 H, s, H-5), 6.40 (1 H, s, H-8), 6.25 (2 H, s, H-2' and H-6'), 5.94, 5.92 (2 H, 2 × d, O-CH₂-O), 5.43 (1 H, s, 4'-OH), 4.71 (1 H, q, J 4.9 Hz, CH₃-CHO₂), 4.53-4.49 (3 H, overlapping, H-1", H-1 and H-11a), 4.43 (1 H, d, J 4.2 Hz, H-4), 4.12 (1 H, dd, J 6.3 and J 8.4 Hz, H-11b), 4.12 (1 H. dd, J 4.9 and J 10.5 Hz, H-6"a), 3.75 (6 H, s, 3'- and 5'-OCH₃), 3.71 (1 H, ddd, J 2.8, J 9.1, and J 9.1 Hz, H-3"), 3.51 (1 H, dd, J 10.5 and J 10.5 Hz, H-6"b), 3.42 (1 H, ddd, J 2.8, J 9.1, and J 9.1 Hz, H-2"), 3.32 (1 H, dd, J 9.1 and J 9.1 Hz, H-4"), 3.26 (1 H, ddd, J 4.9, J 9.8 and J 10.5 Hz, H-5"), 3.15-3.06 (2 H, ov erlapping, H-2 and H-3), 3.04 (1 H, d, J 2.8 Hz, 3"-OH), 3.02 (1 H, d, J 2.8 Hz, 2"-OH), and 1.35 (3 H, d, J 4.9 Hz, CH₃CHO₂). (7): mp 140-150 °C (dec); $[\alpha]_D^{27} = -70^\circ$ (c 1, CHCl₃); ¹H-NMR 500 MHz, CDCl₃: δ 6.81 (1 H, s, H-5), 6.42 (1 H, s, H-8), 6.26 (2 H, s, H-2' and H-6'), 5.95, 5.93 (2 H, 2 x d, O-C \underline{H}_2 -O), 5.39 (1 H, s, 4'-OH), 5.22 (1 H, dd, J 10.0 and J 10.0 Hz, H-3"), 5.03 (1 H, dd, J 10.0 and J 10.0 Hz, H-4"), 4.96 (1 H, dd, J 10.0 and J 10.0 Hz, H-2"), 4.64 (1 H, d, J 10.0 Hz, H-1"), 4.52 (1 H, d, 5.0 Hz, H-1), 4.52 (1 H, dd, J 7.0 and J 10.0 Hz, H-11a), 4.41 (1 H, d, J 3.7 Hz, H-4), 4.24 (1 H, dd, J 7.0 and J 7.5 Hz, H-11b), 4.17 (1 H, dd, J 3.5 and J 12.5 Hz, H-6"a), 4.13 (1 H, dd, J 5.5 and J 12.5 Hz, H-6"b), 3.75 (6 H, s, 3'- and 5'-OCH₃), 3.62 (1 H, ddd, J 3.5, J 5.5, and J 10.0 Hz, H-5"), 3.11 (1 H, dddd, J 3.7, J 7.0, J 10.0, and J 14.0 Hz, H-3), 3.07 (1 H, dd, J 5.0 and J 14.0 Hz, H-2), 2.07, 2.06, 2.02 and 1.99 (12 H, 4 x s, 4 x CH₃COO). Hydrolisys of (6) afforded the corresponding alcohol: mp 263-266 °C; $[\alpha]_D^{27} =$ - 110° (c 0.5, MeOH); ¹H-NMR 500 MHz, CD₃OD (structurally diagnostic signals): 4.66 (1 H, dd, J 9.0 and J 9.5 Hz, H-11a), 4.63 (1 H, d, J 3.5 Hz, H-4), 4.55 (1 H, d, J 9.5 Hz, H-1"), 4.51 (1 H, d, J 5.5 Hz, H-1), and 4.41 (1 H, dd, J 7.0 and J 9.0 Hz, H-11b).
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