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Synthetic Communications: An International Journal for Rapid Communication of Synthetic Organic Chemistry

Publication details, including instructions for authors and subscription information:

http://www.tandfonline.com/loi/lsyc20

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To cite this article: Jifeng Dai & Qibing Zhou (2007) Convenient Synthesis of an N-(1-Alkoxyl-9-fluorenyl)serine Acridine Conjugate, Synthetic Communications: An International Journal for Rapid Communication of Synthetic Organic Chemistry, 37:1, 129-135, DOI: <u>10.1080/00397910600978531</u>

To link to this article: http://dx.doi.org/10.1080/00397910600978531

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Synthetic Communications[®], 37: 129–135, 2007 Copyright © Taylor & Francis Group, LLC ISSN 0039-7911 print/1532-2432 online DOI: 10.1080/00397910600978531



Convenient Synthesis of an *N*-(1-Alkoxyl-9fluorenyl)serine Acridine Conjugate

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Abstract: We report here the synthesis of an *N*-(1-alkoxyl-9-fluorenyl)serine acridine conjugate, which was achieved by a three-component assembly approach via an intra-molecular reductive amination process.

Keywords: acridine conjugate, 1-alkoxyl-9-fluorenylamine, serine

9-Fluorenylamines have recently been demonstrated as important chemophores for biological activity in a variety of organic compounds.^[1-4] For example, hydroxylated or methoxylated 9-fluorenylamino-pentanenitriles have potent anti-multidrug-resistant activity for cancer therapy,^[3] and 1-hydroxyfluorenyl dinitrophenyl hydrazone is highly cytotoxic in breast, ovary, colon, and melanoma cancer cells.^[4] The synthesis of 9-fluorenylamino derivatives has been accomplished either through reductive amination of 9-fluorenones,^[3] or nucleophilic substitution of 9-bromofluorene by amines.^[5] However, in the synthesis of α -substituted amino acid derivatives of 1-alkoxy(or hydroxyl)-fluorenylamine for potential drug candidates, we found that neither method produced desired products. Unsuccessful attempts of reductive amination were probably due to steric interaction between α -amino acids and 1-substituted fluorenones, and basic conditions for SN2type reactions were not compatible with the benzyl ester of amino acid derivatives. Thus, an alternative synthetic approach was needed. We report here a

Received in USA April 12, 2006

Address correspondence to Qibing Zhou, Department of Chemistry, 1001 West Main Street, Virginia Commonwealth University, Richmond, Virginia 23284-2006, USA. E-mail: qzhou@vcu.edu convenient synthetic approach via an intramolecular reductive amination process to obtain 1-alkoxyl-9-fluorenylaminoserine acridine, conjugate **1** (Scheme 1). In addition, this method could be easily modified to conjugate with other bioactive chemophores besides acridine which is a DNA intercalator and is used in this study for potential additional biological activity.^[6]

The synthetic design involved the coupling of serine derivative **4** and fluorenone **2** through an ether linkage to facilitate the reductive amination process (Scheme 1). Acridine **3** was conjugated at later synthetic steps. Initially, the benzyl ester of serine without glycine was used but was found not to be able to couple with fluorenone **2** in the subsequent Mitsunobo step. Thus, *t*-Boc-serine-glycine benzyl ester **4** was used instead, which was conveniently synthesized by coupling *t*-Boc-protected serine **5** and glycine benzyl ester using 1-(3-dimethylaminopropyl)-3-ethylcarbodimide hydrochloride (EDCI) and 1-hydroxybenzotriazole (HOBt) in 69% yield.

9-Fluorenone **2** was synthesized via a Suzuki coupling, followed by an acid-catalyzed acylation (Scheme 2). Boronic acid-**7** for the Suzuki coupling was obtained from bromide **6** and was used immediately in the Suzuki reaction because of its high instability.^[7,8] Triflate **8** was synthesized using reported procedures.^[8] For the Suzuki coupling of **7** and **8**, we found that a combination of Cs₂CO₃ and THF afforded a higher yield (88%) than other conditions including Na₂CO₃, K₃PO₄, dioxane, or acetonitrile (<45%). Hydrolysis of the resulting biphenyl **9** to a free acid was carried out using



Scheme 1. Synthetic design of fluorenylaminoserine acridine conjugate 1.



Scheme 2. Conditions and reagents: (i) nBuLi, THF, -78° C then B(O-*i*-Pr)₃; (ii) triflate **8**, Pd(PPh₃)₄, KBr, Cs₂CO₃, THF-H₂O, reflux for 16 h; (iii) (a) NaOH, THF-H₂O; (b) TFA, TFAA, 0°C; (iv) PPh₃, **4**, DIAD, THF, sonication; (v) (a) TFA; (b) silica gel, reflux in CH₃CN, 16 h; (c) NaBH₃CN; (vi) (a) 5% HCOOH in MeOH, Pd/C; (b) acridine **3**, HBTU, DIPEA, DMF, 3 h.

NaOH in a mixed THF-water solution. Acid-catalyzed acylation was achieved with trifluoroacetic acid and anhydride (2:1 ratio) at 0°C, affording fluorenone **2** as the major isomer in 72% yield. The structure of fluorenone **2** was confirmed by the observed coupling pattern of a singlet and four doublets in the aromatic region of the ¹H NMR spectrum.

Initial attempts to couple serine **4** and fluorenone **2** under normal Mitsunobu reaction conditions^[9] with diethyl azodicarboxylate (DEAD) or diisopropyl azodicarboxylate (DIAD) produced no desired product **10**. Recently, Lepore and He reported that under sonication, Mitsunobu reactions of steric hindered alcohols proceeded smoothly at high concentrations.^[10] Indeed, we found that the coupling was successful only under this specific condition with serine derivative **4** in 40% yield. The subsequent intramolecular reductive amination of 9-fluorenone **10** was accomplished first by the removal of the *t*-Boc protecting group at 0°C, then the formation of imine through refluxing with silica gel, and finally the reduction with NaBH₃CN.^[11] A comparable yield was obtained when 3-Å molecular sieves were used as the dehydrating agent.

For incorporation of the acridine moiety, the benzyl protecting groups of 9-aminofluorene **11** were removed by Pd/C in a 5% HCOOH solution. Acridine **3** was coupled through an amide linkage using O-benzotriazol-1-yl-1,1,3,3-tetramethyluronium hexafluorophosphate (HBTU) and N,N-diiso-propylethylamine (DIPEA) in DMF (Scheme 2). The progress of the reaction was monitored by high performance liquid chromatography (HPLC) analysis. Conjugate **1** was finally isolated in 24% yield by HPLC separation using a gradient condition of 10–70% CH₃CN in a triethylamine acetate buffer (pH 6) over 30 min. Conjugate **1** was confirmed by both ¹H and ¹³C NMR analyses and showed a molecular ion of 574.00 m/z for $M + H^+$ (calculated for C₃₄H₃₂N₅O₄, 574.25) in the ESI-MS analysis.

In conclusion, the synthesis of 1-alkoxyl-9-fluorenylaminoserine acridine was accomplished by a three-component assembly approach through an intramolecular reductive amination process. We are currently investigating the potential biological activity of this novel fluorenylamine compound.

EXPERIMENTAL PROCEDURES

Compound 4: DIPEA (314 mg, 2.43 mmol), *t*-butoxycarbonylserine (500 mg, 2.39 mmol), HOBt (395 mg, 2.92 mmol), and EDCI (489 mg, 2.55 mmol) were added to **5** (863 mg, 2.43 mmol) in 20 mL of CH₂Cl₂. The resulting reaction was stirred under N₂ for 16 h and worked up with CH₂Cl₂. Upon concentration, an analytically pure product (587 mg) was obtained in 69% yield as a colorless oil. ¹H NMR (CDCl₃, 300 MHz): δ 7.42–7.34 (m, 5H), 7.20 (br s, 1H), 5.63 (d, *J* = 7.2 Hz, 1H), 5.16 (s, 2H), 4.22 (br s, 1H), 4.15–**08** (m, 3H), 3.79–3.67 (m, 1H), 1.44 (s, 9H). ESI-MS calcd. for C₁₇H₂₅N₂O₆ (M + H⁺) 353.17; found 353.03.

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Compound 9: Compound 7 (387 mg, 1.69 mmol), Pd(PPh₃)₄ (90 mg, 0.08 mmol), KBr (146 mg, 1.22 mmol), and Cs₂CO₃ (590 mg, 1.81 mmol) were added to **8** (395 mg, 1.21 mmol) in dry THF (18 mL). The mixture was stirred for 15 min, and then 0.5 mL of water was added. The resulting mixture was refluxed for 16 h. The reaction was worked up with brine and ether. The desired product **9** (383 mg) was purified by a flash column in 88% yield as a colorless oil. ¹H NMR (CDCl₃, 300 MHz): δ 7.63 (t, J = 7.7 Hz, 1H), 7.58 (d, J = 6.8 Hz, 2H), 7.52 (t, J = 7.2 Hz, 2H), 7.45 (t, J = 7.7 Hz, 2H), 7.14–7.05 (m, 5H), 5.21 (s, 2H), 1.91 (s, 6H). ESI-MS calcd. for C₂₃H₂₁O₄ (M + H⁺) 361.14; found 361.07.

Compound 2: NaOH (70 mg) and 0.5 mL water were added to 9 (94 mg, 0.26 mmol) in 10 mL of THF. The reaction mixture was refluxed for 16 h and then acidified to pH 4–5 with diluted HCl. The reaction was worked up with ether. Upon concentration, the resulting residue was dissolved in 5 mL of CH₂Cl₂. Trifluoroacetic acid (2 mL) and trifluoroacetic anhydride (1 mL) were then added at 0°C. After 2 h, the reaction was worked up with ether. The desired product 2 (56.4 mg) was purified by a flash column in 72% yield as a yellow solid (mp 103–104°C). ¹H NMR (CDCl₃, 300 MHz): δ 8.48 (s, 1H), 7.58 (d, J = 8.3 Hz, 1H), 7.46–7.31 (m, 6H), 7.10 (s, 1H), 6.98 (d, J = 7.2 Hz, 1H), 6.81 (d, J = 7.8 Hz, 1H), 6.76 (d, J = 8.6 Hz, 1H), 5.16 (s, 2H). ESI-MS calcd. for C₂₀H₁₅O₃ (M + H⁺) 303.10; found 302.99.

Compound 10: Serine 4 (300 mg, 0.85 mmol) and PPh₃ (260 mg, 0.99 mmol) were added to 2 (200 mg, 0.66 mmol) in dry THF (0.8 mL). The mixture was sonicated for 15 min, and then DIAD (143 μ L, 0.73 mmol) was added under sonication. The reaction mixture was sonicated for another 40 min and worked up with CH₂Cl₂. The desired product (168 mg) was purified by a flash column in 40% yield as a yellow oil. ¹H NMR (CDCl₃, 300 MHz): δ 8.40 (br s, 1H), 7.52 (d, J = 8.2 Hz, 1H), 7.47–7.34 (m, 6H), 7.26–7.22 (m, 5H), 7.12–7.09 (m, 2H), 6.81 (d, J = 2.2 Hz, 1H), 6.78 (d, J = 8.4 Hz, 1H), 5.90 (br s, 1H), 5.16 (s, 2H), 5.06 (d, J = 4.1 Hz, 2H), 4.62 (br s, 2H), 4.25 (dd, $J_1 = 6.7$ Hz, $J_2 = 17.6$ Hz, 1H), 4.10 (dd, $J_1 = 5.30$ Hz, $J_2 = 17.6$ Hz, 1H), 3.82 (br t, J = 8.6 Hz, 1H), 1.46 (s, 9H). ESI-MS calcd. for C₃₇H₃₇N₂O₈ (M + H⁺) 637.25; found 637.00.

Compound 11: Trifluoroacetic acid (1 mL) was added to 10 (90 mg, 0.14 mmol) in dry CH₂Cl₂ (2 mL) at 0°C. After 3 h, the solvent was removed by vacuum. The resulting brown residue was dissolved in a mixture of 3 mL of CH₂Cl₂ and 9 mL of CH₃CN, followed by the addition of 460 mg of silica gel. The resulting mixture was refluxed for 16 h under N₂. After cooling to room temperature, NaBH₃CN (96 mg, 1.5 mmol) was added. The reaction was stirred for 2 h, and acetic acid (170 mg) was added. After 3 h, the reaction was worked up with CH₂Cl₂. The desired product (34 mg) was purified by a flash column in 47% yield as a yellow oil. ¹H NMR (CDCl₃, 300 MHz): δ 7.94 (br s, 1H), 7.58 (d, *J* = 8.2 Hz, 1H), 7.48–7.26 (m, 13 H), 6.95 (d, *J* = 7.4 Hz, 2H), 5.27 (d, *J* = 13.2 Hz, 1H), 5.17 (s, 2H), 5.15 (s, 2H), 4.85 (s, 1H), 4.17–4.10 (m, 3H), 3.64 (br t,

J = 12.0 Hz, 1H). ESI-MS calcd. for $C_{32}H_{29}N_2O_5$ (M + H⁺), 521.21; found 520.97.

Compound 1: To 11 (20 mg, 0.038 mmol) in 5% HCOOH in MeOH (2 mL), 10% Pd/C (80 mg) was slowly added under N₂. After 20 min, the solid was removed with a 50 µm filter, and then 0.5 mL of trifluoroacetic acid was added to the filtrate. Upon concentration, the free acid (11 mg) was obtained. For the coupling, acridine 3 (14 mg, 0.04 mmol), HBTU (41 mM in DMF, 1.1 mL), and DIPEA (41 mM in DMF, 2.6 mL) were added to the resulting free acid (5 mg, 0.015 mmol) in DMF (1 mL). After 3 h, the desired product 1 (5.3 mg) was purified by a HPLC separation using a Microsorb MV C18 column (10×250 mm, 8 µm) from Varian (Walnut Creek, CA) in 24% yield as a yellow oil. ¹H NMR (MeOH-d4, 300 MHz): δ 8.37 (d, J = 6.6 Hz, 2H), 7.83 (d, J = 6.3 Hz, 2H), 7.74 (t, J = 6.6 Hz, 2H), 7.50 (d, J = 6.0 Hz, 1H), 7.41 (t, J = 5.7 Hz, 2H), 7.35 (d, J = 6.0 Hz, 1Hz, 1Hz, 1Hz, 1Hz), 7.35 (d, J = 6.0 Hz, 1(d, J = 5.4 Hz, 1H), 7.24 (t, J = 5.7 Hz, 1H), 7.12 (s, 1H), 6.84 (d, J = 5.7 Hz, 1H), 6.74 (d, J = 6.0 Hz, 1H), 4.97 (d, J = 10.5 Hz, 1H), 4.81 (s, 1H), 4.07 (d, J = 5.4 Hz, 1H), 3.98 (t, J = 4.8 Hz, 2H), 3.87 (d, J = 4.2 Hz, 2H), 3.74 - 3.65 (m, 1H), 3.43 - 3.37 (m, 2H), 2.29 - 1.96(m, 2H). ESI-MS calcd. for $C_{34}H_{32}N_5O_4$ (M + H⁺) 574.25; found 574.00.

ACKNOWLEDGMENT

We thank the Thomas F. and Kate Miller Jeffress Memorial Trust (J-723) for the financial support.

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