SYNTHESIS, CHARACTERIZATION AND BIOLOGICAL ACTIVITY OF RARE EARTH COMPLEXES OF 1-PHENYL-3-METHYL-5-HYDROXY-4-PYRAZOLYL PHENYL KETONE ISONICOTINOYL HYDRAZONE*

Zhengyin Yang*, Rudong Yang, Qi Li^b

Department of Chemistry, Lanzhou University, Lanzhou 730000, China and Fashen Li*

Department of Physics, Lanzhou University, Lanzhou 730000, China

ABSTRACT

The ligand, 1-phenyl-3-methyl-5-hydroxy-4-pyrazolyl phenyl ketone isonicotinoyl hydrazone (H₂L), was prepared by condensation of 1-phenyl-3-methyl-4-benzoyl-5-pyrazolone with isoniazid. Seven complexes of rare earths with H₂L have been synthesized and characterized on the basis of elemental analyses, IR, UV, ¹H NMR spectra and thermal analyses. The general formula of the complexes is RE(HL)₃·3.5H₂O (where RE = La³⁺ – Gd³⁺). It has been discovered that the ligand and its complexes of the light rare earth metals possess antioxidative activity and inhibitory action on lipid peroxidation.

Copyright © 1999 by Marcel Dekker, Inc.

^a Project supported by the China Postdoctoral Science Foundation

^b Gansu Center of Analysis and Determination, Lanzhou 730000, China

INTRODUCTION

Isoniazid is a good antituberculosis drug, because of its small size isoniazid has strong permeability and a high curing effect. It was reported that metal complexes of hydrazones derived from isoniazid and 2-oxopropionic acid possess apparent antitumor activity^{1 3} as well as scavenger effects on OH^{*} and O₂^{-*} radicals⁴. Hydrazone ligands derived from isoniazid with heterocyclic β -diones have a certain inhibitory activity on bacteria⁵, and in many cases it has been suggested that the biological activity of their complexes is superior compared to that of the ligands⁶⁻⁸. This idea led us to synthesize 1-phenyl-3-methyl-5-hydroxy-4-pyrazolyl phenyl ketone isonicotinoyl hydrazone (H₂L) and its rare earth complexes with a view to evaluating their biological activity. The ligand was prepared by the reaction of eq (1).



<u>EXPERIMENTAL</u>

<u>Materials</u>

Nitroblue tetrazolium (NBT), methionine (MET), vitamin B_2 (Vit B_2), tris (hydroxy-methyl)-aminomethane (Tris), thiobarbituric acid (TBA) and ascorbic acid were purchased from Sigma Chemical Co. 1-Phenyl-3-methyl-4-benzoyl-5-pyrazolone (PMBP), isoniazid, trichloroacetic acid (TCA), and 3-methylthiopropionaldehyde (MTPA) were products of China. All chemicals used were of analytical grade. The hydrated rare earth(III) chlorides were prepared by dissolving RE₂O₃ in concentrated HCl, then crystallizing the products. Tris-HCl and KH₂PO₄–K₂HPO₄ buffers were prepared with deionized distilled H₂O.

Physical Measurements

Carbon, hydrogen and nitrogen were analyzed on a Carlo Erba 1106 elemental analyser. The metal contents of the complexes were determined by titration with EDTA⁹. Infrared spectra (4000–400 cm⁻¹) were determined with KBr optics on a Nicolet 5DX IR spectrophotometer. Ultraviolet spectra were recorded on a Shimadzu UV-240 spectrophotometer. The thermal behavior was monitored on a PCT-2 differential thermal analyser. ¹H NMR spectra were measured on a Nicolet FT-80A NMR spectrophotometer, using TMS as a reference in DMSO-d₆. The molar conductance values were determined in ethanol on a model DSS-11A conductivity meter. Gas chromatography was performed using a Shimadzu GC-8A gas chromatographic instrument.

Preparation of the Ligand

A quanlity of 0.549 g of isoniazid (4 mmol) and 1.113 g of PMBP (4 mmol) were dissolved in ethanol (150 mL) and refluxed on a water bath for 1 h. The color of the solution changed from initial light yellow to deep red. After cooling to room temperature, a large amount of red precipitate separated out. The product was purified by recrytallizing it in anhydrous ethanol; yield, 1.256 g (79%); m.p. 244–245° C. Anal. Found: C, 69.43; H, 4.89; N, 17.08. Calc. for $C_{23}H_{19}N_5O_2(397.44)$: C, 69.50; H, 4.82; N, 17.26%. ¹H NMR (DMSO-d₆), δ (ppm): 1.50 (3H, singlet, CH₃); 7.10–7.90 (11H, multiplet, 2 phenyl,-NH-); 8.20 (2H, doublet, J = 6Hz, H³, H⁵ of pyridine ring); 8.76 (2H, broad, H², H⁶ of pyridine ring).

Preparation of the Complexes

A quanlity of 0.79 g of the ligand H_2L (2 mmol) was added to water (40 mL), the pH of which was adjusted to 7 by the addition of an aqueous solution of NaOH (10%); the solution turned light-yellow. Then a solution of RECl₃-6H₂O (1 mmol) in water (10 mL) was added dropwise to the system. After stirring at 50° C for 0.5 h, a yellow precipitate was formed which was seperated by filtration, and washed several time with water and hot ethanol, respectively, and dried in a vacuum.

Scavenger Test of Superoxide Radicals

The superoxide radicals (O_2^{-*}) were produced by the system of MET/VitB₂/-NBT¹⁰⁻¹¹. Because NBT can be reduced quantitatively to blue formazan by O_2^{-*} , the amount of O_2^{-*} and the suppression ratio for O_2^{-*} can be calculated by measuring the absorbance at 560 nm¹⁰⁻¹². The solutions of MET, VitB₂ and NBT were prepared with 0.01 M Tris+HCl buffer (pH = 8) avoiding light. The reaction mixture (4.5 mL) included MET (13.5 μ M), NBT (1.0 μ M), VitB₂ (0.1 μ M) and the tested compound (0.056 μ M). After illuminating with a fluorescent lamp at room temperature for 10 min the absorbance (Ai) of the mixture was measured at 560 nm on a UV-240 spectrophotometer. The sample without the tested compound was used as the control, its absorbance is A₀. The suppression ratio was calculated on the basis of (A₀ - A_i)/A₀.

Scavenger Test of Hydroxyl Radicals

Hydroxyl radicals (OH^{*}) were produced by the system H_2O_2/Fe^{2^*} -EDTA/ascorbic acid¹¹, its amount was calculated by measuring the amount of ethene produced from OH^{*} reacting with MTPA^{11,12}. The reaction mixtures (4.5 mL) involved MTPA (50.0 μ M), 1:2 Fe⁺⁺ – EDTA (83.0 μ M), ascorbic acid (100 μ M), H₂O₂ (0.2 μ M) and the tested compound (0.2 μ M) and were prepared with 0.01 M of KH₂PO₄-K₂HPO₄ buffer (pH = 7.4). The tested compound was not added to the control group. The reaction was carried out at a constant 37 °C for 30 min, then the amount of ethene produced was detected by gas chromatography. The suppression ratio was calculated on the basis of (S₀ - S)/S₀, where S and S₀ are the peak areas given by the gas chromatography for the ethene content of the sample (S) and the control group (S₀).

Inhibitory Action for Lipid Peroxidation

Malondialdehyde (MDA) is one of the products formed in lipid peroxidation. The measurement of lipid peroxidation is on the basis of a TBA test¹³⁻¹⁵ based on a spectrophotometric quantitation of a red-violet ($\lambda_{max} = 532$ nm) complex formed with MDA.

1 g of fresh rabbit's liver was weighed into a glass homogenizer, 10 mL of physiological saline solution was added, and homogenized in an ice bath. The sample (3 mL) containing the tested compound (0.2 μ M) and control group without the tested compound were vibrated at a constant 37 °C for 2 h, then a 20% aqueous solution of TCA (3 mL) was immediately added to every sample and the control group, respectively. After 15 min the reaction mixtures were centrifuged at 4000 cycles per min for 10 min. The clear solution of the samples (2 mL) was mixed in a screw capped tube with 2 mL TBA reagent, and heated in a boiling water bath for 30 min, then

cooled in tap water. The absorbances (A_i, A₀) were measured at 532 nm, using 20% TCA as a reference. The inhibitory ratio was calculated on the basis of inhibition = $(A_0 - A_i)/A_0$.

RESULTS AND DISCUSSION

All complexes are yellow, soluble in DMSO and ethanol, insoluble in water, benzene and diethyl ether, and can be kept in air for a long time. The molar conductiveties of the complexes are around $2.97-10.6 \text{ ohm}^{-1} \cdot \text{cm}^2 \cdot \text{mol}^{-1}$ in ethanol (see Table I), this shows that all complexes are non-electrolytes in ethanol¹⁶. The elemental analyses (see Table I) show the formula of the complexes as RE(HL)₃. $3.5H_2O$ (RE = $La^{3+} - Gd^{3+}$), with three and a half H₂O and three ligands (HL⁻) for each complex and was confirmed by ¹H NMR and thermal analyses.

Thermal Analysis

Some data of thermal analyses are listed in Table II. The DTA curves of the complexes have an endothermic peak between 83 and 93°C, the corresponding TG curves show that the weight loss is equal to three and a half water molecules. These results are in accordance with the compositions of the complexes determined by elemental analyses. Two exothermic peaks appear around 442–469°C and 501–547°C, respectively. Initial temperature of decomposition is about 450°C or so, this indicates that the complexes have a high thermal stability.

Ultraviolet Spectra

UV spectra data of complexes are listed in Table III. The free ligand exhibits a strong absorption at 298 nm and a shoulder peak at 286 nm. The complexes yield two new peaks at approximately 366 and 383 nm, respectively. This shows that there is a conjugate system formed by enolization of the ligand in the complexes. Therefore, the absorption bands of two carbonyl $\pi \rightarrow \pi^*$ transitions appear at the region observed. The complexes should be a chelate containing both five and six-membered rings. This is the reason why the complexes exhibit high thermal stability. This is consistent with the thermal analysis results.

Formula	M.w.	M.p.*	Color	Yield	Elemental	Analysis	(Calc.)	%	Λм
		(°C)		(%)	С	н	N	RE	(U) ⁶
H ₂ L C ₂₃ H ₁₉ N ₅ O ₂	397.44	244	Red	79	69.43 (69.50)	4.89 (4.82)	17.08 (17.26)		-
La(HL)3 • 3.5H2O LaC69H61N15O9 5	1390.4	458	Yellow	64	59.36 (59.57)	4.39 (4.42)	15,08 (15,10)	10.02 (9.98)	3.24
$Ce(HL)_3 \cdot 3.5H_2O$ $CeC_{69}H_{61}N_{15}O_{9.5}$	1391.4	422	Yellow	67	59.64 (59.52)	4.32 (4.42)	14.95 (15.09)	10.18 (10.06)	5.10
Pr(HL)3 • 3.5H2O PrC69H61N15O9.5	1392.4	468	Yellow	62	59.49 (59.48)	4.40 (4.41)	15.01 (15.08)	10.32 (10.11)	10.6
Nd(HL)3 • 3.5H2O NdC69H61N15O9.5	1393.4	447	Yellow	69	59.48 (59.34)	4.38 (4.40)	15.11 (15.04)	10.46 (10.33)	4.81
Sm(HL) ₃ •3.5H ₂ O SmC ₆₉ H ₆₁ N ₁₅ O _{9.5}	1403.4	449	Yellow	71	59.13 (59.08)	4.14 (4.38)	14.80 (14.98)	10.96 (10.72)	2.97
Eu(HL)3 • 3.5H2O EuC69H61N15O9.5	1404.4	473	Yellow	65	59.24 (59.02)	4.24 (4.38)	14.91 (14.96)	11.01 (10.82)	3,75
$\begin{array}{l} \mbox{Gd}(HL)_3 \bullet 3.5 H_2 O \\ \mbox{Gd}C_{69} H_{61} N_{15} O_{9.5} \end{array}$	1409.4	469	Yellow	68	58,86 (58,79)	4.32 (4.36)	14.93 (14.90)	11.21 (11.16)	8.51

Table I. Elemental Analysis and Molar Conductivity

a: M.p. for the complexes is the decomposition point; b: $U=ohm^{-1} \cdot cm^2 \cdot mol^{-1}$.

Tabl	e II.	Thermal	Analyse	s Data of	tl	he C	Comp	lexes
------	-------	---------	---------	-----------	----	------	------	-------

Complexes	Dehydra-	Process	W. loss	Decc	omp.(°C)	Process	Residue
	tion (°C)		(Calc.) %	t ₁	t ₂		
La-comp.	87	Endothermic	5.1 (4.53)	458	528	Exothermic	La_2O_3
Ce-comp.	93	Endothermic	4.9 (4.53)	422	515	Exothermic	CeO ₂
Pr-comp.	83	Endothermic	4.9 (4.52)	468	537	Exothermic	Pr ₆ O ₁₁
Nd-comp.	85	Endothermic	4.6 (4.52)	447	518	Exothermic	Nd ₂ O ₃
Sm-comp.	87	Endothermic	4.6 (4.50)	449	501	Exothermic	Sm ₂ O ₃
Eu-comp.	90	Endothermic	4.5 (4.49)	473	542	Exothermic	Eu ₂ O ₃
Gd-comp.	89	Endothermic	4.5 (4.47)	469	547	Exothermic	Gd ₂ O ₃

Complexes λ_{max} (nm) and $\varepsilon \times 10^{-3}$ (L·mol ⁻¹ ·cm ⁻¹)										
	λι	£¦	λα	٤u	λш	٤ _D	λιν	ειν	λν	٤v
H ₂ L	198	1.23	286	5.81	298	6.34				
La-comp.	200	4.49	290	6.10	297	30.17	365	12.57	391	7.82
Ce-comp.	208	4.87	289	31.01	306	27.21	369	11.46	395	6.49
Pr-comp.	210	4.32	297	24.8	310	25.91	376	8.10	397	9.54
Nd-comp.	200	5.40	283	24.18	298	25.12	366	10.33	383	7.98
Sm-comp.	200	5.67	287	30.01	298	31.43	365	12.76	382	10.63
Eu-comp.	199	6.81	286	31.36	297	32.55	366	13.61	383	10.95
Gd-comp.	198	7.14	285	29.36	297	30.55	368	14.28	383	11.50

Table III. Ultraviolet Spectral Data

Infrared Spectra

The main stretching frequencies of the IR spectra of the ligand (H₂L) and its complexes are tabulated in Table IV. The aqueous $\upsilon(OH)$ for the complexes appears at 3350 cm⁻¹, $\upsilon(C^7=O)$ and $\upsilon(C^5=O)$ vibration of the free ligand are at 1616 and 1560 cm⁻¹, respectively; for the complexes these peaks shift to 1602, 1434 cm⁻¹ or so, $\Delta \upsilon$ is equal to 14–16, and 119–126 cm⁻¹. The band at 460 cm⁻¹ is assigned to $\upsilon(M-O)^{17}$. This demonstrates that the oxygen of the carbonyl has formed a coordinative bond with the rare earth ions, one of which (C⁵=O) is in the enolic form. The band at 1588 cm⁻¹ for the free ligand is assigned to the $\upsilon(C=N)$ stretch, it shifts to 1571 cm⁻¹ or so for its complexes, weak bands at 410 cm⁻¹ are assigned to $\upsilon(M-N)$. This further confirms that the nitrogen of the imino-group bonds to the rare earth ions. All of these data confirm the fact that a conjugate chelate ring formed by ligand enolazition exists in the complexes.

¹H Nuclear Magnetic Resonance Spectra

The ¹H NMR spectra of La(HL)₃·3.5H₂O only are discussed in the paper. La(HL)₃·3.5H₂O, δ (ppm): 1.18 (3H, singlet, CH₃), 1.36 (3H, singlet, CH₃), 8.13 (4H, doublet, J = 6Hz, 2H³, 2H⁵ of pyridine ring), 8.51 (4H, broad, 2H², 2H⁶ of pyridine ring), 6.80 - 7.86 (20 H, multiplet, 4 phenyl), 3 - 3.5 (H₂O). Two methyls of the

Compounds	υ(H ₂ O)	υ(NH)	υ(C ⁷ =O)	υ(C=N)	δ(NNH)	υ(C ⁵ =O)	δ(H ₂ O)	υ(M-O)	υ(M-N)
H₂L		3018 m	1616 s	1588 s	1532 m	1560 s	1356 m		
La-comp.	3353 m	3051 m	1602 s	1572 s	1539 s	1434 s	1638 s	456 w	405 w
Ce-comp.	3353 m	3051 m	1602 s	1572 s	1546 s	1441 s	1631 s	456 w	405 w
Pr-comp.	3353 m	3051 m	1602 s	1572 s	1546 s	1434 s	1645 s	456 w	410 w
Nd-comp.	3353 m	3068 m	1602 s	1568 s	1546 s	1441 s	1638 s	456 w	410 w
Sm-comp.	3360 m	3065 m	1602 s	1574 s	1546 s	1434 s	1638 s	456 w	410 w
Eu-comp.	3360 m	3058 m	1602 s	1574 s	1546 s	1434 s	1638 s	456 w	410 w
Gd-comp.	3353 m	3058 m	1602 s	1574 s	1546 s	1434 s	1638 s	456 w	405 w

Table IV. Some Main IR Data of Ligand and its Complexes (cm⁻¹)



Fig.1. The Probable Structure of the Complexes

complexes, compared with the free ligand, shift upfield by 0.06, 0.24 ppm, respectively. The hydrogen atoms of the pyrazole ring are not seen in the ¹H NMR spectra of the complexes, implying that the 5-pyrazolone group in H₂L exists in the enolic form in the complexes. The probable structure of the complexes is shown in Fig.1 and is derivefrom the above results of IR, UV, and ¹H NMR.

Biological Activity

The data of biological activity are listed in Table V. The results show that H_2L and its light rare earth complexes possess a certain scavenger effect on OH[•] and O_2 ^{••}

Complexes	Dose	Average suppres	sion ratio (%)	Average inhibitory ratio (%)	
	(mg)	for OH'	for O_2^{-1}	for lipid peroxidation	
H ₂ L	0.079	38.5	45.2	41.5	
La-comp.	0.186	75.4	73.9	80.1	
Ce-comp.	0.186	72.8	58.4	66.8	
Pr-comp.	0.186	46.7	57.6	62.0	
Nd-comp.	0.187	41.1	48.3	50.1	
Sm-comp.	0.188	59.8	54.1	72.9	
Eu-comp.	0.190	49.2	58.0	65.2	
Gd-comp.	0.190	68.6	61.2	77.3	
Cu(HSal)2	3.10 ^a		59		
Na2Cu(Sal)2	3.10 ^a		30		

Table V. Biological Activity Data

a: μ mol of Cu (see reference 18).

and can inhibit lipid peroxidation. The biological activity of the ligand is less than that of its rare earth complexes. It is clear that coordination enhances the biological activity, and that the scavenger effects on OH^{*} and O_2^{-*} as well as the inhibitory action on lipid peroxidation of the lanthanum and gadolinium complexes are the strongest in all complexes studied. Perhaps this is related to the valence electron arrangement of La(III)4t⁰ and Gd(III)4t⁷, the scavenger effects on O_2^{-*} of the two complexes is superior compared to that of the transition metal Cu(II) salicylate complexes¹⁸ (see Table V).

REFERENCES

1. Z. Y. Yang, L. F. Wang, J. Q. Wu, X. Y. Li, *Chinese J. App. Chem.*, 9, 31 (1992); Chem. Abstr., 117, 102982y (1992).

- 2. L. F. Wang, Z. Y. Yang, Z. R. Pen, G. Q. Cheng, J. Coord. Chem., 28, 167 (1993).
- 3. Z. Y. Yang, L. F. Wang, J. Q. Wu, K. W. Yang, Acta Chem., 51, 115 (1993).
- 4. S. Y. Yu, S. X. Wang, Q. H. Luo, L. F. Wang, Z. R. Peng, X. Gao, Polyhedron, 12, 1093 (1993)
- 5. Z. Y. Yang, L. F. Wang, J. Q. Wu, X. Y. Li, Sci. Bull., 9, 813 (1992).

- 6. J. G. Wu, R. W. Deng, Z. N. Chen, Trans. Met. Chem., 18, 23 (1993).
- 7. K. N. Thimmaiah, W. D. Lioyd, G. T. Chan, Trans. Met. Chem., 10, 94 (1985).
- 8. R. Srivastava, J. Inorg. Nucl. Chem., 4, 1526 (1980).
- 9. C. N. Reilley, R. W. Schmid and F. S. Sadek, J. Chem. Educ., 36, 555 (1959).
- V. Ponti, M. U. Dianzani, K. Cheeseman and T. F. Slater, Chem-Biol.Interact, 23, 281 (1978).
- 11. C. C. Winterbourn, Biochem. J., 198, 125 (1981).
- 12. C. C. Winterbourn, Biochem. J., 182, 625 (1979).
- 13. A. Schmedes and G. Holmer, J. Am. Oil Chem. Soc., 66, 813 (1989).
- 14. H. Ohkawa, N. Ohishi and K. Yagi, Anal. Biochem., 95, 351 (1979).
- 15. T. Asakawa, S. Matsushita, Lipids, 14, 401 (1979).
- 16. W. J. Geary, Coord. Chem. Rev., 7, 81 (1971).
- 17. R. Malhotra, J. P. Singh, M. Dudeja, K. S. Dhindsa, J. Inorg. Biochem., 46, 119 (1992).
- 18. C. C. Young and S. J. Lippard, J. Am. Chem. Soc., 102, 4920 (1980).

Received:	31 December 1997	Referee I:	G. N. Holder
Accepted:	17 August 1998	Referee II:	L. C. Nathan