



# Catalytic enantioselective synthesis of a novel inhibitor of ceramide trafficking, (1*R*,3*R*)-*N*-(3-hydroxy-1-hydroxymethyl-3-phenylpropyl)- dodecanamide (HPA-12)

Masaharu Ueno,<sup>a,b</sup> Hidetoshi Kitagawa,<sup>a,b</sup> Haruro Ishitani,<sup>a,b</sup> Satoshi Yasuda,<sup>b,c</sup>  
Kentaro Hanada<sup>b,c</sup> and Shū Kobayashi<sup>a,b,\*</sup>

<sup>a</sup>Graduate School of Pharmaceutical Sciences, The University of Tokyo, Hongo, Bunkyo-ku, Tokyo 113-0033, Japan

<sup>b</sup>CREST, Japan Science and Technology Corporation, 4-1-8, Honchou, Kawaguchi-City, Saitama 332-0012, Japan

<sup>c</sup>Department of Biochemistry and Cell Biology, National Institute of Infectious Diseases (former National Institute of Health), Toyama, Shinjuku-ku, Tokyo 162-8640, Japan

Received 15 August 2001; revised 4 September 2001; accepted 5 September 2001

**Abstract**—A novel inhibitor of ceramide trafficking, (1*R*,3*R*)-*N*-(3-hydroxy-1-hydroxymethyl-3-phenylpropyl)dodecanamide (HPA-12), has been synthesized using a chiral zirconium-catalyzed asymmetric Mannich-type reaction as a key-step. © 2001 Elsevier Science Ltd. All rights reserved.

Sphingolipid biosynthesis is now of great interest because of its important roles in cell growth, differentiation, and apoptosis, etc.<sup>1</sup> Enzymes that catalyze sphingolipid biosynthesis are targets for chemists as well as biologists to create new drugs. Our group has accomplished total synthesis of sphingofungin B,<sup>2</sup> an inhibitor of serinepalmitoyl transferase (SPT), and more recently, khafrefungin,<sup>3</sup> an inhibitor of inositol phosphorylceramide (IPC), and the structural relationship of biological activity has been clarified. In the course of our investigations to search for a new molecule that shows

a characteristic property in sphingolipid biosynthesis, we have found that (1*R*,3*R*)-*N*-(3-hydroxy-1-hydroxymethyl-3-phenylpropyl)dodecanamide (HPA-12, **1**) is a novel inhibitor of ceramide trafficking from endoplasmic reticulum to the site of sphingomyelin (SM) synthesis. HPA-12 is the first compound of the specific inhibitor for SM synthesis in mammalian cells, and a potential drug that inhibits intracellular trafficking of sphingolipids.<sup>4</sup> In this report, we describe the first synthesis of HPA-12 using a chiral zirconium-catalyzed enantioselective Mannich reaction as a key-step.

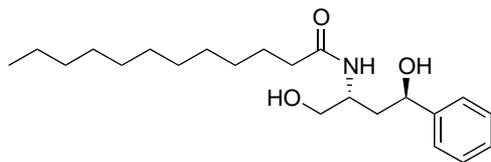


**Scheme 1.**

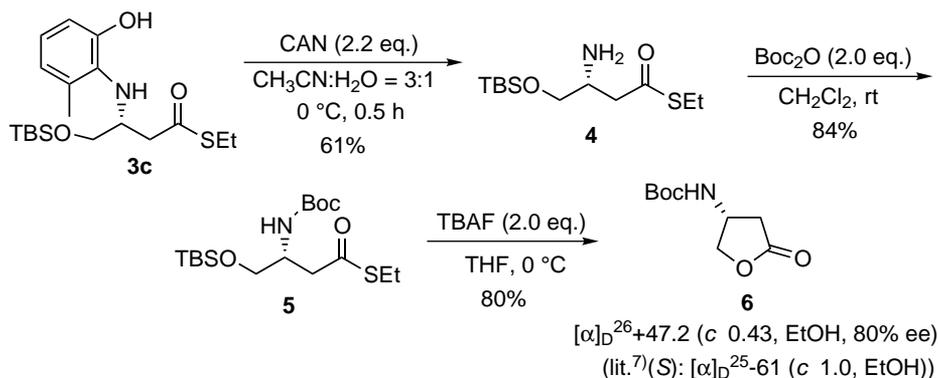
**Keywords:** HPA-12; a novel inhibitor of ceramide trafficking; Mannich-type reaction; asymmetric reaction.

\* Corresponding author.

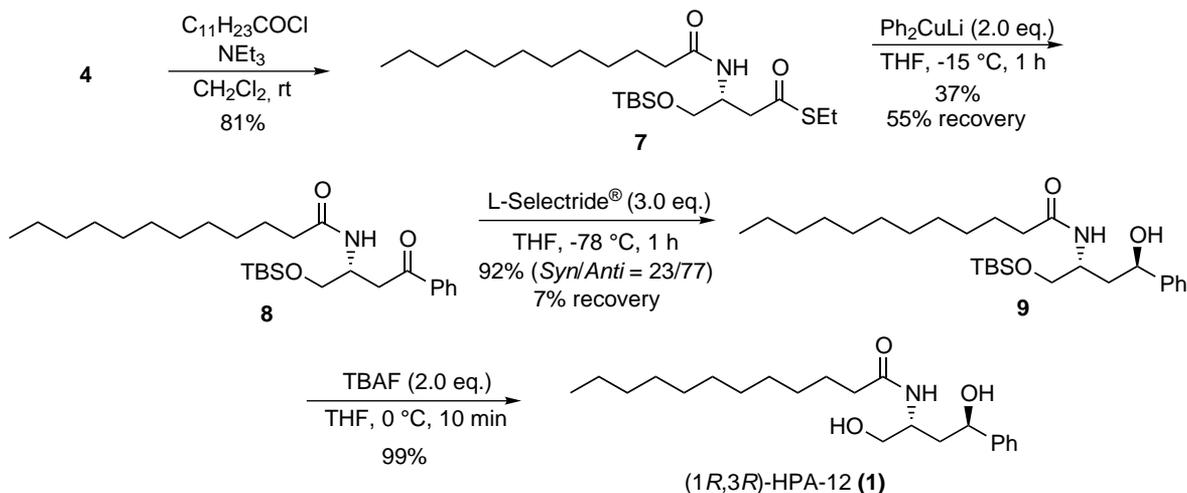
In the initial and key-step, we examined the three-component reaction of an  $\alpha$ -alkoxy aldehyde, 2-amino-*m*-cresol, and 1-ethylthio-1-trimethylsilyloxyethene in the presence of a catalytic amount of a chiral zirconium complex prepared from zirconium *tert*-butoxide, (*R*)-6,6'-Br<sub>2</sub>BINOL, and *N*-methylimidazole (Scheme 1).<sup>5</sup> It was found that the alkoxy part of the aldehyde significantly influenced the enantioselectivity of the product, and that a high level of selectivity was obtained when the *tert*-butyldimethylsilyloxy group was used. The benzyloxy group and more bulky *tert*-butyldiphenylsilyloxy group gave much lower selectivity.

(1*R*,3*R*)-HPA-12 (**1**)

The absolute configuration of the product **3c**<sup>6</sup> was determined after converting to literature-known lactone **6** as shown in Scheme 2. Treatment of **3c** with cerium ammonium nitrate (CAN) in acetonitrile–water (3:1) gave amine-free adduct **4** in 61% yield. *t*-Butoxycarbonyl (Boc) protection of the amino group followed by



Scheme 2.

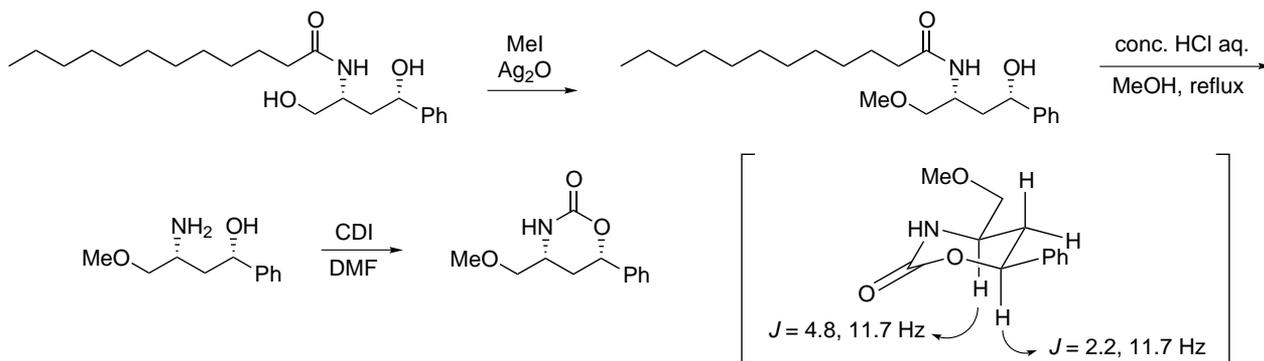


Scheme 3.

deprotection of the *t*-butyldimethylsilyloxy group gave the desired lactone **6** in high yield. Comparison of the optical rotation of **6** with that in literature<sup>7</sup> revealed that the absolute configuration of Mannich adduct **3c** was *R*.

The preparation of HPA-12 from **3c** was performed according to the transformations shown in Scheme 3. The adduct **4** was acylated under standard conditions to give amide **7** in 81% yield. Conversion of **7** to ketone **8** was performed using diphenylcopper lithium in THF at  $-15^\circ\text{C}$  for 1 h. While the yield of **8** was moderate (37%), 55% of the starting material (**7**) was recovered (82% conversion yield). Anti-selective reduction of **8** proceeded using L-Selectride<sup>®</sup> in THF at  $-78^\circ\text{C}$  (92%, *syn/anti* = 23/77).<sup>8</sup> The use of lithium borohydride instead of L-Selectride<sup>®</sup> gave lower selectivity (82%, *syn/anti* = 54/46). These selectivity would be explained by the preferential conformation of **8**.<sup>9</sup> Finally, deprotection of the *tert*-butyldimethylsilyloxy group of **9** using tetrabutylammonium fluoride gave HPA-12 in 99% yield. After recrystallization from ether/hexane, HPA-12 was obtained in 96% ee.<sup>10</sup>

Thus, HPA-12, a novel inhibitor of ceramide trafficking, has been synthesized using a chiral zirconium-catalyzed Mannich-type reaction as a key-step. Based on



Scheme 4.

this synthetic scheme, we have synthesized all four stereoisomers of HPA-12, and confirmed that only the (1*R*,3*R*)-isomer showed high activity.<sup>4</sup> Further investigations to search for more active compounds as well as to clarify biological aspects of the inhibition are now in progress.

### Acknowledgements

This work was partially supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science, Sports, and Culture, Japan. M.U. thanks the JSPS fellowship for Japanese Junior Scientists.

### References

- Review: (a) Kolter, T.; Sandhoff, K. *Angew. Chem., Int. Ed. Engl.* **1999**, *38*, 1532. See also: (b) Dickson, R. C. *Annu. Rev. Biochem.* **1998**, *67*, 27.
- (a) Kobayashi, S.; Furuta, T.; Hayashi, T.; Nishijima, M.; Hanada, K. *J. Am. Chem. Soc.* **1998**, *120*, 908; (b) Kobayashi, S.; Hayashi, T.; Iwamoto, S.; Furuta, T.; Matsumura, M. *Synlett* **1996**, 672.
- (a) Wakabayashi, T.; Mori, K.; Kobayashi, S. *J. Am. Chem. Soc.* **2001**, *123*, 1372; (b) Kobayashi, S.; Mori, K.; Wakabayashi, T.; Yasuda, S.; Hanada, K. *J. Org. Chem.* **2001**, *66*, 5580.
- Yasuda, S.; Kitagawa, H.; Ueno, M.; Ishitani, H.; Fukasawa, M.; Nishijima, M.; Kobayashi, S.; Hanada, K. *J. Biol. Chem.*, in press.
- (a) Ishitani, H.; Ueno, M.; Kobayashi, S. *J. Am. Chem. Soc.* **1997**, *119*, 7153; (b) Ishitani, H.; Ueno, M.; Kobayashi, S. *J. Am. Chem. Soc.* **2000**, *122*, 8180.
- Compound 3c**:  $[\alpha]_D^{21} +8.26$  (*c* 1.23, CHCl<sub>3</sub>, 80% ee); IR (neat): 3358, 2928, 2856, 1680, 1589, 1476, 1254, 836 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  0.00 (s, 6H), 0.85 (s, 9H), 1.18 (t, 3H,  $J=7.4$  Hz), 2.18 (s, 3H), 2.66 (dd, 1H,  $J=4.7, 13.9$  Hz), 2.82 (q, 2H,  $J=7.5$  Hz), 2.85 (m, 1H), 3.40 (dd, 1H,  $J=3.6, 8.2$  Hz), 3.45–3.56 (m, 2H), 6.56 (dd, 1H,  $J=0.7, 7.2$  Hz), 6.68 (dd, 1H,  $J=1.3, 7.2$  Hz), 6.79 (t, 1H,  $J=7.7$  Hz); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  -5.6, -5.5, 14.6, 17.6, 18.3, 23.6, 25.9, 46.1, 55.7, 64.0, 113.5, 121.6, 124.8, 131.5, 132.4, 152.1, 199.7; HPLC: Daicel Chiralpak AD, hexane/<sup>i</sup>PrOH=200/1, flow rate=1.0 ml/min: <sup>t</sup>R=16.5 min (3*R*), <sup>t</sup>R=20.3 min (3*S*); HRMS: calcd for C<sub>19</sub>H<sub>33</sub>NO<sub>3</sub>SSi (M<sup>+</sup>) 383.1950, found 383.1932.
- Nitta, H.; Ueda, I.; Hatanaka, M. *J. Chem. Soc., Perkin Trans. 1* **1997**, 1973.
- The stereochemical assignment of the *syn*-adduct was performed by <sup>1</sup>H NMR analysis after converting to the 3,4,5,6-tetrahydro-1,3-oxazin-2-one (Scheme 4).
- Pilli, D. A.; Russowsky, D.; Dias, L. C. *J. Chem. Soc., Perkin Trans. 1* **1990**, 1213.
- HPA-12 (1)**: mp 75.5–77.0°C;  $[\alpha]_D^{22} -35.1$  (*c* 0.8, CHCl<sub>3</sub>, 96% ee); IR (neat): 3293, 2919, 2849, 1643, 1551, 1493, 1054, 746 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  0.88 (t, 3H,  $J=6.8$  Hz), 1.26 (bs, 16H), 1.59 (bt, 2H,  $J=7.2$  Hz), 1.92 (ddd, 1H,  $J=6.6, 8.5, 15.1$  Hz), 2.03 (ddd, 1H,  $J=3.6, 5.7, 14.6$  Hz), 2.15 (t, 2H,  $J=7.7$  Hz), 3.65 (ddd, 1H,  $J=4.1, 11.3, 15.5$  Hz), 4.01–4.08 (m, 1H), 4.79 (dd, 1H,  $J=3.4, 8.8$  Hz), 6.48 (d, 1H,  $J=6.8$  Hz), 7.23–7.36 (m, 5H); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  14.1, 22.7, 25.7, 29.3, 29.3, 29.3, 29.5, 29.6, 29.6, 31.9, 36.8, 40.7, 50.4, 65.5, 71.8, 125.5, 127.7, 128.5, 144.2, 174.3; HPLC: Daicel Chiralpak AD, hexane/<sup>i</sup>PrOH=19/1, flow rate=1.0 ml/min: <sup>t</sup>R=11.4 min (1*S*,3*S*), <sup>t</sup>R=15.0 min (1*R*,3*R*); HRMS: calcd for C<sub>22</sub>H<sub>37</sub>NO<sub>3</sub> (M<sup>+</sup>) 363.2273, found 363.2279.