Journal of Medicinal Chemistry

Article

Subscriber access provided by the Henry Madden Library | California State University, Fresno

Lead Optimization of a Novel series of Imidazo[1,2-*a*]pyridine Amides Leading to a Clinical Candidate (Q203) as a Multiand Extensively-Drug Resistant Antituberculosis Agent

Sunhee Kang, Ryang Yeo Kim, Min Jung Seo, Saeyeon Lee, Young Mi Kim, Mooyoung Seo, Jeong Jea Seo, Yoonae Ko, Inhee Choi, Jichan Jang, Jiyoun Nam, Seijin Park, Hwankyu Kang, Hyung Jun Kim, Jungjun Kim, Sujin Ahn, Kevin Pethe, Kiyean Nam, Zaesung No, and Jaeseung Kim *J. Med. Chem.*, Just Accepted Manuscript • DOI: 10.1021/jm5003606 • Publication Date (Web): 28 May 2014
 Downloaded from http://pubs.acs.org on May 30, 2014

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a free service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are accessible to all readers and citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



Journal of Medicinal Chemistry is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

Lead Optimization of a Novel series of Imidazo[1,2-a]pyridine Amides Leading to a Clinical Candidate (Q203) as a Multi- and Extensively-Drug Resistant Antituberculosis Agent Sunhee Kang¹, Ryang Yeo Kim², Min Jung Seo¹, Saeyeon Lee¹, Young Mi Kim¹, Mooyoung Seo¹, Jeong Jea Seo¹, Yoonae Ko¹, Inhee Choi¹, Jichan Jang², Jiyoun Nam³, Seijin Park³, Hwankyu Kang³, Hyung Jun Kim², Jungjun Kim⁴, Sujin Ahn³, Kevin Pethe², Kiyean Nam⁴, Zaesung No^{1,#,*}, Jaeseung Kim^{1*} ¹Medicinal & Bioorganic Chemistry group, Institut Pasteur Korea, 16, Daewangpango-ro 712Beon-gil, Bundang-gu, Seongnam-si, Gyeonggi-do, 463-400 Korea; ²Antibacteial Drug Discovery group, Institut Pasteur Korea, 16, Daewangpango-ro 712Beon-gil, Bundang-gu, Seongnam-si, Gyeonggi-do, 463-400 Korea; ³DMPK group, Institut Pasteur Korea, 16, Daewangpango-ro 712Beon-gil, Bundang-gu, Seongnam-si, Gyeonggi-do, 463-400 Korea; ⁴Ourient Co. Ltd, 16, Daewangpango-ro 712Beon-gil, Bundang-gu, Bundang-gu, Seongnam-si, Gyeonggi-do, 463-400 Korea; Present address: [#]Gyeonggi Biocenter, 147 Gwanggyo-ro, Youngtong-gu, Suwon-si, Gyeonggi-do, 443-270 Korea. **RECEIVED DATE** (to be automatically inserted after your manuscript is accepted if required according

to the journal that you are submitting your paper to)

ABSTRACT

A critical unmet clinical need to combat the global tuberculosis epidemic is the development of potent agents capable of reducing the time of multidrug-resistant (MDR) and extensively drug-resistant (XDR) tuberculosis therapy. In this paper, we report on the optimization of the imidazo[1,2-*a*]pyridine amide (IPA) lead compound 1, which led to the design and synthesis of Q203 (**50**). We found that the amide linker with IPA core is very important for activity against *Mycobacterium tubercu*losis H37Rv. Linearity and lipophilicity of the amine part in the IPA series play a critical role in improving *in vitro* and *in vivo* efficacy and pharmacokinetic profile. The optimized IPAs, **49** and **50** showed not only excellent oral bioavailability (80.2 and 90.7%, respectively) with high exposure of the area under curve (AUC) but also displayed significant colony-forming unit (CFU) reduction (1.52 and 3.13 log₁₀ reduction at 10 mg/kg dosing level, respectively) in the lung of mice.

INTRODUCTION

Tuberculosis (TB) is a chronic infectious disease caused by Mycobacterium tuberculosis. TB remains a serious global health problem positioned as the second leading cause of death from a single infectious agent worldwide with an incidence rate of almost 9 million cases and a fatality rate of 1.4 million in 2011¹. Notwithstanding the successful implementation of Directly Observed Treatment Short course (DOTS) in many countries, high prevalence of MDR and XDR tuberculosis has intensified the urgent need for new anti-tubercular drugs. Several new classes of compounds have been discovered for treatment of tuberculosis in the last decade²⁻⁴. A few of them are currently in clinical trials⁵⁻⁷ and one of them, bedaquiline, was recently approved for the treatment of MDR tuberculosis (Figure 1). However, given the emergence of drug resistance and the low success rate encountered in clinical development, there is still a need to develop additional drug candidates for TB treatment. We recently reported a novel chemical entity named Q203 (50) as a promising anti-TB drug candidate⁸ using phenotypic high-content screening (HCS) technology inside infected macrophages⁹. The IPA series targets QcrB^{8,10}, which encodes the b subunit of the electron transport ubiquinol cytochrome C reductase. The IPA series was reported by others as an attractive anti-TB lead series¹¹⁻¹³. Here, we report on the lead optimization that led to the design the potential clinical candidate 50.

Figure 1. Structures of bedaquiline, PA-824⁵ and delamanid.



Chemistry. General amide coupling of appropriately substituted imidazo[1,2-*a*]pyridine-3-carboxylic acid (**4a**-**4g**) with corresponding amines R3 afforded target compound in over 60 % yield (Scheme 1). One of the precursor, the imidazo[1,2-*a*]pyridine-3-carboxylic acid (**4a**-**4g**) was prepared starting from bromination of various β -keto esters except for commercially available ethyl 2-chloro-3-oxobutanoate (**2a**). Unsubstituted β -keto esters were treated with *N*-bromosuccinimide and over two equivalent of NH₄OAc in Et₂O to afford the 2-monobrominated product (**2b**)¹⁴. Alternatively, β -keto esters could be transformed to 2-bromo products by bromine (**2c-2d**) which resulted in comparable yields. The adequate monobrominated β -keto esters were condensed with the substituted 2-aminopyridines *via* imine-enamine formation in absolute ethanol at reflux temperature¹⁵ and the resulting esters (**3a-3g**) were hydrolyzed to acids (**4a-4g**).

The counter parts, methanamine derivatives, can be classified into three functional groups; i) bi-aryl, ii) 2ring system having a saturated cyclic amine, iii) 3-ring system having a saturated cyclic amine between two aryl rings. The synthetic route of the methanamine is shown in Scheme 2. Suzuki coupling of 4-chlorobenzonitrile and substituted phenylboronic acid using $PdCl_2(dppf)$ and aqueous $Na_2CO_3^{16}$ followed by reduction with

Journal of Medicinal Chemistry

lithium aluminum hydride gave biphenyl methanamines **7a-7f**. Exceptionally, biphenyl methanamines having a cyano group and *tert*-butyl group, **7f** and **7g** were synthesized by direct Suzuki coupling with 4-bromobenzylamine. The benzylamines which have saturated ring contained bis- and tris-ring were prepared in a straightforward manner as shown in Scheme 3. Commercially available 4-fluorobenzonitrile was reacted with appropriate cyclic amine and K_2CO_3 by heating in dimethyl sulfoxide and then reduced with lithium aluminum hydride to produce benzylamines **8a-8n**¹⁷.

The synthetic route for the linker modification is shown in Scheme 4. Compound 9, which has no benzylic carbon next to amide was prepared *via* acid chloride activation of 4f and *N*-methylated compound 10 was synthesized by treating sodium hydride and iodomethane from compound 1. Amide reduction of 1 in a mild condition using boron trifluoride etherate and NaBH₄ provided 11 with moderate yield.

In the case of 14 and 15, which have one more carbon between the imidazo[1,2-*a*]pyridine ring and the carbonyl group of amide bond, 4-brominated β -keto ester is required. Interestingly, the desired 4-brominated β -keto ester 12 was regioselectively synthesized by treating *N*-bromosuccinimide with 0.1 equivalent of neutral catalyst, NH₄OAc compared to the presence of over 2 equivalent of NH₄OAc for the synthesis of 2-brominated β -keto ester 2b. In this reaction, 2-brominated β -keto ester, 2b was generated initially and then the bromo group was migrated to 4-position after overnight reaction. However, the migration of the bromo group was not occurred in the presence of excess amount of NH₄OAc. 12 was condensed with 2-amino-5-chloropyridine and saponified to afford the intermediate acid 13 followed by general amide coupling afforded 14 and 15. The reverse-amide compound 17 was synthesized *via* curtius rearrangement using diphenylphosporyl azide and ACS Paragon Plus Environment

triethylamine in *t*-BuOH. Subsequent de-protection of Boc group by trifluoroacetic acid and amide coupling *via*

acid chloride activation afforded the target compound.





^{*a*}Reagents and conditions: (i^{*a*}) NBS, NH₄OAc (over 2eq.), ether, rt, 6h; (i^{*b*}) Br₂, CHCl₃, 0°C – rt, 20min; (ii) **2a-2d**, EtOH, reflux, overnight; (iii) LiOH, EtOH/H₂O (3:1, ν/ν), rt, overnight; (iv) corresponding amine, EDC, HOBt, TEA, DMF, 80°C, 2-4h







^{*a*}Reagents and conditions: (i) SOCl₂, 100 °C, 1h, then [1,1'-biphenyl]-4-amine, TEA, MC, rt, 1h; (ii) NaH, CH₃I, DMF, 0°C - rt, 1h; (iii) NaBH₄, BF₃·etherate, THF, reflux, 2h; (iv) NBS, NH₄OAc (0.1 eq.), Et₂O, rt, overnight; (v) 2-amino-5-chloropyridine, ethanol, reflux, overnight; (vi) LiOH, MeOH/H₂O (3:1, ν/ν), rt, overnight; (vii) EDC, [1,1'-biphenyl]-4-ylmethanamine, HOBt, TEA, DMF, 80°C, 3h; (viii) [1,1'-biphenyl]-4-amine, EDC, HOBt, TEA, DMF, 80°C, 3h; (viiii) DPPA, TEA, *t*-BuOH, reflux, overnight; (x) TFA, MC, rt, 1h; (xi) 2-([1,1'-biphenyl]-4-yl)acetic acid, SOCl₂, TEA, MC, rt, 1h

RESULTS AND DICUSSION

The activity of IPA derivatives was tested against *M. tuberculosis* replicating inside macrophages (intracellular MIC_{80}) and in liquid broth culture medium (extracellular MIC_{80}) (Tables 1-4), as previously described⁸. In addition, the metabolic stability of the compounds was evaluated in mouse and human liver microsomal preparations to study structure-property relationships (SPR) in order to prioritize compounds for *in vivo* pharmacokinetic evaluation. The initial structure-activity relationship (SAR) studies evaluated a set of analogues that contained replacement of the 3-carboxamide linker in an attempt to affect potency of compound 1 (Table 1). Replacement of hydrogen on NH with methyl (10) significantly decreased the activity against *M. tuberculosis* H37Rv replicating outside and inside macrophages by approximately 190 and 690-fold. Surprisingly, the modification of the length of the amide linker with one carbon (14 and 15) between the imidazo[1,2-*a*]pyridine ring and the carbonyl group of amide bond abolished the potency against *M. tuberculosis*. Introduction of *N*-phenyl group (9) at the amine position did not give any activity as well. We then

Journal of Medicinal Chemistry

investigated the contribution of the H-bonding acceptor of the carbonyl group. Modification by a reversed amide (17) or removal of the oxygen on carbonyl group (11) reduced the activity to sub-micromolar range (intracellular MIC₈₀ = 200 nM and 690 nM, respectively) compared to the parent compound 1. Accordingly, this set of modifications revealed that the carboxamide linker with the *N*-benzylic group is critical for anti-mycobacterial activity.

Table 1. Activity of linker modified IPA analogues against M. tuberculosis H37Rv



		Antimycobacterial activity a	gainst <i>M. tuberculosis</i> H37Rv
Compound	x	^a extracellular MIC ₈₀ (nM)	^b intracellular MIC ₈₀ (nM)
1	O NH	45	1.39
10	O ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	8690	970
9	O NH NH	>10000	>10000
15	M Star	>10000	>10000
14	John North	>10000	>10000
11	N H	810	200
17	N N O	1670	690

^{*a*}extracellular MIC₈₀: the inhibitory activity against *M. tuberculosis* H37Rv replicating in culture broth medium; ^{*b*}intracellular MIC₈₀: the inhibitory activity against *M. tuberculosis* H37Rv replicating inside macrophages;

 MIC_{80} is the minimum concentration required to inhibit growth of 80 %; MIC_{80} indicates average value of two independent measurement.

Next, the optimization was focused on R1, R2 and R3 modifications (Table 2). During the exploration of SAR at R2 and R3 positions, the substituent's at R1 was limited to H, 6-Cl, and 7-Cl. This is because we found early on that 6 or 7-Cl substitutions improved antibacterial activity and increased the metabolic stability compared to other substituent groups (data not shown). To investigate the hydrophobic interaction of R2 alkyl groups, the four compounds 18-21 were designed to alter the size of R2 position that might disturb positioning of the 3carboxamide. As expected, the smaller size groups, methyl and ethyl (18 and 19) showed better activity (intracellular MIC₈₀= 20 nM and 97 nM, respectively) than longer and bulky groups, propyl and iso-propyl (20) and 21). Furthermore the sterically hindered compound 21 (intracellular MIC₈₀= 3130 nM) was much less potent than the linear compound 20 (intracellular MIC_{80} = 50 nM). To confirm the improved potency of methyl and ethyl group at the R2 position, the compound 1 and 22 were prepared and evaluated. Compound 1, which possesses an ethyl group, showed approximately 30-fold greater potency against intracellular mycobacteria compared to methyl substituted compound 22. However, the methyl substitution had a better metabolic stability compared to the ethyl substitution.

Further exploration of the SAR at the R3 position of the phenyl group having 6- or 7-Cl of imidazo[1,2*a*]pyridine core was conducted by substitution of various functional groups such as donating, withdrawing and carboxylic acid. For the effect of the position on the benzene ring, a *para*-chloro group (**24**) offered much more potency and metabolic stability than the *ortho*-chloro group (**23**). Encouraged by the positive effect of the *para*-**ACS Paragon Plus Environment**

Journal of Medicinal Chemistry

substitution on antibacterial effect, we focused on screening several analogues at this position to investigate the

activity and metabolic stability. Intracellular activity of all substituents such as donating and withdrawing groups (24-28), except a hydrophilic acid, were very similar within a 2-fold range of MIC_{80} = 0.4 ~1.3 nM. Even the more lipophilic and sterically hindered compound 30 showed similar intracellular activity comparable to compound 27. On the other hand, compound 29 that has a hydrophilic carboxylic acid group, gave a deleterious effect on the activity. In terms of metabolic stability, the alkyl group such as methyl (26 and 27) and *tert*-butyl (30) showed much lower microsomal stability than the other groups Cl, CN and acid (24, 25, 28 and 29) in human and mouse liver microsomes. This again suggested that the position of R3 on the benzene ring may play a role in potency and microsomal stability.

From our initial SAR study, compound **24**, **25** and **28** showed desirable extra- and intracellular potency as well as metabolic stability in human and mouse liver microsomes to perform *in vivo* pharmacokinetic (PK) experiments. However, compound **25** was too insoluble in aqueous and organic solutions caused by extension of aromatic character to conduct *in vivo* experiment. Thus, compound **24** was selected and *in vivo* PK profile was evaluated in Spraque-Dawley rat after administration by oral (p.o.) and intravenous (i.v.) routes. However, the half-life and AUC after oral administration could not be calculated because the concentration of compound detected in the plasma remained constant (or even slightly increased) up to 16 hours after dosing (supporting information, Table S1). This result suggested that compound **24** was rebounded in absorption after 4 h, could be subject to compound precipitation in gut and extended absorption due to the highly lipophilic nature of the compound **24**. Therefore, our next optimization strategy was focused on replacement of the second phenyl group with various ring systems to improve solubility and reduce lipophilicity (Table 3).



 $R_{1} \xrightarrow{6}_{N} \xrightarrow{K_{1}}_{N} R_{2}$

				An	timycobacterial a <i>M. tuberculosi</i>	ctivity against s H37Rv	Microsomal stabilit	y (t _{1/2} , min)
Cor	npound	R1	R2	R3	extracellular MIC ₈₀ (nM)	intracellular MIC ₈₀ (nM)	Human	Mouse
	18	Н	Ме	Н	250	20	^a ND	^a ND
	19	Н	Et	Н	63	97	27.1	^a ND
	20	Н	Pr	Н	1180	50	22.4	8.7
	21	Н	ⁱ Pr	Н	3130	3130	31.6	^a ND
	22	6-Cl	Ме	Н	175	42	>120	>60
	1	6-Cl	Et	Н	45	1.39	22.8	19.3
	23	6-Cl	Et	2-CI	43	9.3	38.9	13.1
	24	6-CI	Et	4-CI	0.9	0.45	83	>60
	25	6-Cl	Et	4-CN	<0.5	0.68	>120	>60
	26	6-Cl	Et	4-Me	0.7	0.43	30.5	40.6
	27	7-Cl	Et	4-Me	<0.5	1.35	25.4	23.1
	28	7-Cl	Et	4-Cl	1.3	1.01	>120	>60
	29	7-Cl	Et	4-CO ₂ H	250	217	>120	>120
	30	7-Cl	Et	4- ^t Bu	12	0.46	18.5	50.7
	Isoniazid(INH)				449	617		
	Rifampicin(RIF	.))			26.6	180		

^{*a*}ND, Not Determined.

The changes were focused on the second phenyl ring on right-hand side while keeping the 7-chloro-2ethylimidazo[1,2-a]pyridine-3-carboxamide at the left-hand side. Firstly, we introduced hetero aromatic groups that were favorable to make a salt formation to improve solubility. However, introduction of heteroaromatic ACS Paragon Plus Environment

Journal of Medicinal Chemistry

rings led to a dramatic reduction in potency (31 and 32, intracellular MIC_{80} of 140 and 740 nM, respectively). Another strategy involved the introducing of nitrogen containing saturated ring next to the first phenyl ring, such as a piperidine, azepane and piperazine. The nitrogen atom, if appropriately acidic, could potentially provide an ionizable site. When the piperidine and azepane moieties (33 and 34) were placed at the end, the cellular activity was maintained compared to heteroaromatic rings, 31 and 32. However, not only the compounds were heavily metabolized in human and mouse liver microsomes, they also had decreased antibacterial activity in macrophages compared to the lead compound 1. Furthermore we did not find any sign of improvement of microsomal stability by substitution at the 4-position on the piperidine ring (35 and 36). A series of analogues containing piperazine at the end of the phenyl ring were also synthesized and evaluated. From the previous SAR studies, it was shown that a hydrophilic character on the second ring led to decreased potency. Thus, we applied small alkyl group such as methyl and *iso*-propyl at 4-position of piperazine to protect activity loss. Unlike piperidine compounds 35 and 36, compounds 37 and 38 lost their potency against M. tuberculosis H37Rv replicating inside and outside macrophage (intracellular $MIC_{80} > 140$ nM). On the other hand, compounds 39 and 40 with the fused ring showed a potency comparable to compound 1, whether or not the fused ring had an aromaticity. The excellent potency of the lipophilic and bulky analogues 39 and 40 suggested that there is more lipophilic space and that the two-ring system could be extended to a three-ring system on the right-hand side. In addition, compound 25 having a nitrile group that is representative linearity or aromaticity next to the biphenyl ring, showed a superior potency with MIC_{80} value of less than 1 nM against M. tuberculosis H37Rv replicating outside and inside macrophages, as well as good metabolic stability in human and mouse liver microsomes (Table 2).

Table 3. Activity of IPA analogues containing two or three rings against *M. tuberculosis* H37Rv

		Antimycobacteria <i>M. tubercul</i>	l activity against o <i>sis</i> H37Rv	Metabolic stabil	ity (t _{1/2} , min)
Compound	- R	extracellular MIC ₈₀ (nM)	intracellular MIC ₈₀ (nM)	Human	Mouse
31	-	2000	140	>120	10.8
32	-§-	2220	740	28.9	^a ND
33	·§-N	16	8.2	13.2	2.3
34	-§-N	35	10	2.9	1.9
35	·ξ−N CF ₃	0.8	0.47	6.5	5.1
36	-§-NCI	19	2	11.0	5.2
37	-§-N_N-	4390	360	116.6	12.1
38	-§-N_N-<	3000	140	>120	8.9
39	-§-N	25	9.4	14.9	6.3
40	5 N	34	3.74	27.3	62.3
41	÷NN-	∕—F 5.7	0.3	>120	33.3
42	5 N	∕—F 540	0.66	6.8	18.4

^{*a*}ND, Not Determined.

With this structure-activity relationship (SAR) analysis, two analogues **41** and **42** were designed and evaluated against *M. tuberculosis* H37Rv replicating outside and inside macrophage. Compound **41** with a 4-

ACS Paragon Plus Environment

Journal of Medicinal Chemistry

1 2	fluorophenyl	piperaz	ine showed not	only dram	atically incre	eased with	MIC ₈₀ valu	ies of	0.3	nM c	ompa	ared to
3 4 5	analogue 38	(MIC ₈₀ =	=140 nM) again	st bacteria	replicating in	nside macro	ophages but	t also	appr	oxim	ately	3-fold
0 7 8 9	improved sta	bility in	mouse liver mic	crosomes. I	n the same n	nanner, the	4-fluorophe	enyl p	iperio	dine a	analog	gue 42
10 11 12	showed over	10-fold	better potency b	y incorpora	ation of phen	yl ring thar	analogue 3	33 (int	tracel	lular	MIC	₈₀ = 8.2
13 14 15	nM) with be	tter mic	rosomal stability	in mouse	. However, t	he microso	mal stabilit	ty stil	l requ	uired	to be	e more
16 17 18	improved.											
20 21 22												
23 24 25	Table 4. Act	ivity of]	IPA analogues o	containing	three ring sy	ystem agaiı	nst <i>M. tube</i>	rculos	sis H3	37Rv		
26 27 28 29 30 31						R ₂						
32 33			_	Antimycobacteri <i>M. tubercu</i>	al activity against <i>losis</i> H37Rv	Metabolicsta	bility (t _{1/2} , min)		CYP inh	ibition (I	ıC ₅₀ , uM	1)
34 35	Compound	R1	R2	extracellular MIC ₈₀ (nM)	intracellular MIC ₈₀ (nM)	Human	Mouse	3A4	2D6	1A2	2C9	2C19
36 37	41	7-Cl		5.7	0.3	>120	33.3	>40	>40	>40	>40	>40
38 39	43	6-Cl	-ş-INNF	<0.5	0.3	>120	33.3	0.49	>40	>40	1.33	1.32
40	44	7-Cl		1.8	0.11	>120	>60	>40	>40	>40	0.17	>40
41 42	45	6-CI		<0.5	0.23	>120	>60	11.54	>40	>40	0.57	>40
43	42	7-Cl	s. () () .	0.54	0.66	6.8	18.4	2.07	>40	>40	0.5	0.6
44 45	46	6-Cl	·§-N	<0.5	0.36	62.6	20.3	>40	>40	>40	>40	>40
46 47	47	7-Cl	,	1	0.46	67.3	112.0	>40	>40	>40	0.19	0.38
48	48	6-Cl	-{-N_CI	4.1	1.3	57.9	116.0	>40	>40	>40	0.26	6.77
49 50	49	7-Cl		4.0	3.7	>120	>120	>100 ^a	>100	>100	0.14	0.29
51 52	50 (Q203)	6-CI	-§-NO	CF ₃ 4.0	1.43	>120	>120	>100 ^b	>100 ^b	>100 ^b	>100 ^b	>100 ^b
53												

^aValues were determined by LC/MS method; ^bThe assay was performed using recombinant CYP enzymes and analyzed by LC/MS/MS

To improve the microsomal stability, substituents on the third aromatic ring were investigated with 7-chloro-

1
2
3
4
5
6
0
1
8
9
10
11
12
13
14
15
16
17
10
10
19
20
21
22
23
24
25
20
20
27
28
29
30
31
32
33
31
25
30
36
37
38
39
40
41
42
43
11
44 15
40
46
47
48
49
50
51
52
52
53
<u>э</u> 4
55
56
57
58
59

60

or 6-chloro-2-ethylimidazo[1,2-a]pyridine-3-carboxamide as a left-hand side. As shown in Table 4, the
analogues 47 and 48, in which the fluorine was replaced by chlorine, had a retained potency (intracellular
$MIC_{80}=$ 0.46 nM and 1.3 nM, respectively) and improved microsomal stability ($t_{1/2} > 60$ min, mouse
microsomes). Other analogues (44-45 and 49-50) bearing a trifluoromethoxy substituent showed a similar
activity and good stability in human and mouse microsome. All analogues in Table 4 displayed unprecedented
MIC ₈₀ values in single-digit nanomolar or sub-nanomolar range against <i>M. tuberculosis</i> H37Rv replicating both
outside and inside macrophage. In terms of metabolic stability, the stability-substituent correlation was followed
in order of trifluoromethoxy > chlorine > fluorine and most compounds had good metabolic stability to perform
in vivo experiment except 4-fluorophenyl piperidine analogues 42 and 46. In addition to the stability, the
incorporation of ionizable saturated ring between two aryl-rings compared to bis-phenyl IPA analogues (24-26)
resulted in improved solubility under acidic condition without activity loss (supporting information, Table S2).
All analogues were also evaluated for cytochrome P450 inhibition with 5-different isozymes in order to
prioritize compounds for in vivo PK experiments. Drug-drug interaction is a critical factor to develop an anti-
TB agent due to combination therapy with other TB drugs or HIV drugs for co-infected treatment. Based on
their overall properties, compounds 41, 49 and 50 were shortlisted for <i>in vivo</i> PK experiments.

Table 5. In vivo pharmacokinetic values in mice of selected compounds, 41, 49 and 50

_	Pharmacokinetics (i.v.)			Pharmacokinetics (p.o.)					
Compd	t _{1/2} (h)	CI (mL/min/kg)	Vd _{ss} (mL/kg)	C _{max} (ng/mL)	t _{1/2} (h)	T _{max} (h)	AUC _{0-inf} (ng [.] h/mL)	F (%)	
41	6.15	1.9	877	4450	9.4	2.0	59576	69.9	
49	62.3	3.15	14300	1987	21.3	2.0	27349	80.2	
50 ^a	16.5	4.0	5270	1490	23.4	2.0	44100	90.7	

^{*a*}PK values for compound **50** were adapted from reference 8 and presented for the sake of comparison.

The *in vivo* PK properties of compounds **41**, **49** and **50** were evaluated in mice after intravenous (i.v.) and oral (p.o.) administration of 2 and 10 mg/kg, respectively. As shown in Table 5, those compounds displayed good PK properties with long half-life, low systemic clearance and moderate to high volume of distributions. After oral dosing, all compounds reached a maximum concentration in plasma within 2 hours, their elimination half-life was favorable (9.4, 21.3 and 23.4 hours, respectively) and the area under curve (AUC) was 59,576, 27,349 and 44,100 ng h/mL, respectively. Overall, they achieved good oral exposure in systemic circulation that resulted in superior oral bioavailability (69.9, 80.2 and 90.7%, respectively).

On the basis of the promising *in vivo* PK profile, we conducted *in vivo* efficacy studies for compound **49** in an established mouse model under the same condition as previously described⁸. BALB/c mice were infected with 2 $\times 10^2$ to 2 $\times 10^3$ CFU of *M. tuberculosis* H37Rv by the intranasal route. Compound treatment was initiated three weeks after infection. Compound **49** or the reference drug isoniazid (INH) was administered by oral gavage for 28 days, five times per week. Bacterial load in the lungs of infected mice was determined by CFU enumeration as shown in Table 6. Both compounds displayed potential efficacy results in a dosedependent manner. The compound **49** was active and promoted a significant reduction of the bacterial burden in the lungs of infected animals. The reduction in CFU was proportional to the administered dose, but was less pronounced than for isoniazid despite better pharmacodynamic indices. The compound **49** (1.52 \log_{10} CFU reduction at 10 mg/kg) was also less efficacious than **50** (3.13 \log_{10} CFU reduction at 10 mg/kg)⁸, the drug candidate that was selected for clinical evaluation.

Table 6. In vivo efficacy of compound 49 against M. tuberculosis in an established mouse model

Compd.	Dose (mg/kg)	CFU (Log ₁₀)/lung
49	2	6.23 ± 0.30
	10	5.72 ± 0.40
	50	5.65 ± 0.40
INH	15	5.01 ± 0.14
Untreated		7.24 ± 0.17

CONCLUSIONS

In this study, we report on the optimization of the lead compound **1** that led to **50** including SAR studies to improve an activity against *M. tuberculosis* H37Rv replicating inside and outside macrophage assay and SPR studies for development potential. We found that 3-carboxamide linker of this series played a critical role in anti-tuberculosis activity. The issue of accumulation in plasma of bi-phenyl analogue **24** from *in vivo* PK study was investigated by replacement of the last phenyl ring with heterocyclic groups. Further SAR investigations with three ring systems, which had a linear and long hydrophobic groups revealed that the incorporation of a piperidine or piperazine group in middle of two phenyl rings showed enhanced potency (intracellular MIC₈₀ < 1

nM), improved metabolic stability ($t_{1/2} > 60$ min) and no inhibitory profile against 5 cytochrome P450 isozymes (IC₅₀ > 40 μ M). Ultimately, the optimization process led to compounds **49** and **50** that were performed *in vivo* PK and efficacy in a mouse model and **50** was selected as a final candidate for further evaluation as a clinical candidate based on its overall properties and high potency in the mouse model of tuberculosis. Preclinical study and target engagement study of **50** will be reported in due course.

EXPERIMENTAL SECTION

Chemistry. All reactions were carried out under an argon atmosphere in oven-dried glassware with magnetic stirring and the reaction solvents were purified by passage through a bed of activated alumina. Purification of reaction products was carried out by flash chromatography using silica gel 60 (Merck, 230-400 mesh). Analytical thin layer chromatography was performed on 0.25 mm silica gel $60-F_{254}$ plates (Merck). Visualization was accomplished with 254 nm of UV light and PMA or potassium permanganate staining followed by heating. ¹H-NMR (at 400 MHz), ¹³C-NMR (at 100 MHz) and ¹⁹F-NMR (at 376 MHz) spectra were reported on a Varian 400 MHz spectrometer. ¹H-NMR spectra (CDCl₃ at 7.26 ppm) and ¹³C-NMR spectra (CDCl₃ at 77.2 ppm) were recorded in ppm using solvent as an internal standard. ¹⁹F-NMR spectra were recorded in ppm using α, α, α -trifluorotoluene as an external standard (at -64.72 ppm). Data are reported as (ap = apparent, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad; coupling constant(s) in Hz; integration). LC/MS data were obtained using a Waters 2695 LC and Micromass ZQ spectrometer. The purity of all biologically tested compounds was ≥ 95 % by HPLC. Yields refer to purified products and are not optimized.

Minimum inhibitory concentration determination, metabolic stability, CYP inhibition assay (Fluorescence method), in vivo pharmacokinetics and in vivo efficacy were performed as previously descibed⁸.

Compound 50 has been tested using recombinant CYP enzymes to confirm its inhibitory activities.

Recombinant CYP inhibition assay:

 μ L test compound/positive controls working solution are incubated with 100 μ L substrate and recombinant CYP enzyme mixture working solution in the absence and presence of 98 μ L cofactor solutions for 10 minutes for CYP1A2 and CYP2C9, 20 min for CYP2C19 and CYP2D6 and 3 min for CYP3A4. The final incubations are terminated by 200 μ L cold IS-fortified (100 ng/mL tolbutamide) stop solution and the samples analyzed by LC-MS/MS. The reaction mixtures (200 μ L final volume) contained approximately 100 mM potassium phosphate buffer (pH 7.4), 3.3 mM MgCl₂, 1 mM NADPH and 5 pmol/mL recombinant CYP enzymes.

General Procedure for the Preparation of 2b-d. Method A: *Ethyl 2-bromo-3-oxopentanoate (2b)*. To a stirred solution of ethyl 3-oxopentanoate (13.8 mmol) in Et₂O (70 mL) were added ammonium acetate (41.4 mmol) and *N*-bromosuccinimide (13.8 mmol) and the reaction mixture was stirred at room temperature for 6 hours. The reaction mixture was diluted with Et₂O (20 mL) and washed with water (50 mL \times 2). The organic phase was dried over anhydrous MgSO₄ and concentrated in vacuo to give a title a compound as a clear oil that was used for next reaction without further purification.

Journal of Medicinal Chemistry

Method B: Ethyl 2-bromo-3-oxohexanoate (2c) and ethyl 2-bromo-4-methyl-3-oxopentanoate (2d). To a stirred solution of ethyl 3-oxohexanoate (6.3 mmol) in chloroform (5 mL) was added a solution of bromine (6.3 mmol) in chloroform (30 mL) dropwise under ice-bath and the resulting solution was allowed to room temperature and stirred for 20 min. The reaction was quenched with saturated NaHCO₃ (aq. 10 mL) and extracted with chloroform. The resulting organic phase was washed with brine (20 mL), dried MgSO₄ and concentrated in vacuo to give a title compound 2c. The resulting crude residue was used for next reaction without further purification. In a similar manner, 2d was synthesized according to method B. General Procedure for the Preparation of 3a-3g. To a solution of ethyl 2-bromo-3-oxopentanoate (2b, 12.9 mmol) in EtOH (25 mL) was added 2-amino-5-chloropyridine (12.9 mmol). The mixture was stirred at reflux temperature for overnight. After cooling, the reaction mixture was concentrated. The resulting dark residue was dissolved in EtOAc (20 mL) and washed with water (20 mL). The organic phase was washed with brine (20 mL), dried MgSO₄ and concentrated in vacuo. The crude residue was purified by flash column chromatography (*n*-hexane: EtOAc = 4:1) to give **3f** as a pale yellow solid. *Ethyl 2-methylimidazo*[1,2-a]pyridine-3-carboxylate (**3a**). ¹H NMR (400 MHz, CDCl₃); δ 1.29 (t, J = 7.2 Hz, 3H), 2.56 (s, 3H), 4.27 (q, J = 7.2 Hz, 2H), 6.77 – 6.81 (m, 1H), 7.17 – 7.22 (m, 1H), 7.42 – 7.45 (m, 1H), 9.11 -9.13 (m, 1H). *Ethyl 2-ethylimidazo*[1,2-*a*]*pyridine-3-carboxylate* (**3b**). ¹H NMR (400 MHz, CDCl₃); δ 1.36 (t, J = 7.6 Hz, 3H), 1.43 (t, J = 7.2 Hz, 3H), 3.12 (q, J = 7.6 Hz, 2H), 4.43 (q, J = 7.2 Hz, 2H), 6.95 - 6.98 (m, 1H), 7.35 - 7.39 (m, 1H), 7.63 – 7.65 (m, 1H), 9.31 – 9.33 (m, 1H).

Ethyl 2-propylimidazo[1,2-a]*pyridine-3-carboxylate* (3c). ¹H NMR (400 MHz, CDCl₃); δ 1.02 (t, J = 7.6 Hz, 3H), 1.44 (t, J = 7.2 Hz, 3H), 1.76 – 1.85 (m, 2H), 3.08 (t, J = 7.6 Hz, 2H), 4.43 (q, J = 7.2 Hz, 2H), 6.95 – 6.99 (m, 1H), 7.35 – 7.39 (m, 1H), 7.62 – 7.64 (m, 1H), 9.32 – 9.34 (m, 1H). *Ethyl 2-isopropylimidazo*[1,2-*a*]*pyridine-3-carboxylate* (3*d*). ¹H NMR (400 MHz, CDCl₃); δ 1.38 (d, *J* = 6.8 Hz, 6H), 1.44 (t, J = 7.2 Hz, 3H), 3.80 - 3.87 (m, 1H), 4.43 (t, J = 7.2 Hz, 2H), 6.94 - 6.97 (m, 1H), 7.34 - 7.38(m, 1H), 7.66 – 7.69 (m, 1H), 9.32 – 9.34 (m, 1H). *Ethyl* 6-chloro-2-methylimidazo[1,2-a]pyridine-3-carboxylate (3e). ¹H NMR (400 MHz, CDCl₃); δ 1.43 (t, J = 7.2 Hz, 3H), 2.69 (s, 3H), 4.42 (q, J = 7.2 Hz, 2H), 7.34 (dd, J = 9.2, 2.0 Hz, 1H), 7.54 (d, J = 9.2 Hz, 1H), 9.38 (d, J = 2.0 Hz, 1H). *Ethyl* 6-chloro-2-ethylimidazo[1,2-a]pyridine-3-carboxylate (3f). ¹H NMR (400 MHz, CDCl₃); δ 1.35 (t, J = 7.6 Hz, 3H), 1.44 (t, J = 7.2 Hz, 3H), 3.11 (q, J = 7.6 Hz, 2H), 4.44 (q, J = 7.2 Hz, 2H), 7.35 (dd, J = 9.6, 2.0 Hz, 1H), 7.58 (d, J = 9.6 Hz, 1H), 9.42 (d, J = 2.0 Hz, 1H).

Ethyl 7-*chloro-2-ethylimidazo*[1,2-*a*]*pyridine-3-carboxylate* (**3***g*). ¹H NMR (400 MHz, CDCl₃); δ 1.34 (t, *J* = 7.6 Hz, 3H), 1.43 (t, *J* = 7.2 Hz, 3H), 3.09 (q, *J* = 7.6 Hz, 2H), 4.43 (q, *J* = 7.2 Hz, 2H), 6.95 (dd, *J* = 7.6, 2.0 Hz, 1H), 7.62 (d, *J* = 2.0 Hz, 1H), 9.26 (d, *J* = 7.6 Hz, 1H).

General Procedure for the Preparation of 4a-4g. To a solution of 3f (4.3 mmol) in EtOH (30 mL) was added an aqueous solution of lithium hydroxide (13.0 mmol in 10 mL of water) and the mixture was stirred at room temperature for overnight. The organic solvent was evaporated and 1N HCl was added until pH was reached to 4. The residual pale solid was collected by filtration, washed with water and dried to give 4f as a white solid.

In similar manner, **4a-4e** and **4g** were synthesized.

General Procedure for the Preparation of 7a-7g. Method A (7a-7e): To a solution of 4-chlorobenzonitrile (3.63 mmol) in dimethoxyethane (9 mL) were added 4-chlorophenylboronic acid (4.36 mmol), 1,1'bis(diphenylphosphino)ferrocene)-dichloropalladium(II) (0.11 mmol), Na₂CO₃ (7.26 mmol in 3 mL of water) and the mixture was stirred at 150°C. After 1h, the mixture was cooled to room temperature, then the mixture was extracted with EtOAc (20 mL), washed with sat. NaHCO₃ (aq. 15 mL) and brine (15 mL) and dried over MgSO₄ and concentrated. The resulting residue was purified by flash column chromatography (nhexane:EtOAc = 10:1) to give **6b** as a white solid (67% yield). To a solution of **6b** (2.38 mmol) in THF (24 mL) was added lithium aluminum hydride (7.14 mmol) under ice-bath and then the resulting mixture was refluxed for an hour. The reaction mixture was cooled to room temperature, quenched with water, added sat. Na₂CO₃ (aq. 15 mL) and extracted with EtOAc (30 mL \times 2). The combined organic layers were washed with brine (20 mL), dried over MgSO₄ and concentrated in vacuo to give **7b** as a pale yellow solid (89%). The resulting residue was used for next reaction without further purification.

In a similar manner, **7a** and **7c-7e** were synthesized according to method A.

Method B (**7f-7g**): A solution of 4-bromobenzyl amine (0.32 mmol) in dimethoxyethane (1 mL) were added (4-(tert-butyl)phenyl)boronic acid (0.39 mmol), 1,1'-bis(diphenylphosphino)ferrocene)-dichloropalladium(II) (0.01 mmol), Na₂CO₃ (0.64 mmol in 350 uL of water) and the mixture was stirred at 150 °C. After 1h, the mixture was cooled to room temperature, then the mixture was extracted with EtOAc (10 mL), washed with saturated NaHCO₃ (aq. 10 mL) and brine (10 mL), dried over MgSO₄ and concentrated in vacuo to give crude **7f**. The resulting residue was used for part resetion without further purification.

7f. The resulting residue was used for next reaction without further purification.

In a similar manner, **7g** was synthesized according to method B.

General Procedure for the Preparation of 8a-8n. A mixture of 4-fluorobenzonitrile (3.0 mmol), 4-(4-trifluoromethoxy)piperidine (3.3 mmol) and K₂CO₃ (6.0 mmol) in DMSO (5 mL) was heated to 120° C for 4 hours. After the cooling, the mixture was poured to the water and then generating solid was filtered, washed with water and dried. The resulting crude product (2.1 mmol) was dissolved in THF (10 mL) and then lithium aluminum hydride (6.2 mmol) was added slowly. After the refluxing for an hour, the reaction mixture was cooled to room temperature, quenched with water and filtered off the insoluble aggregates using of cellite. The filtrate was basified with saturated Na₂CO₃ (aq. 20 mL) and then extracted with EtOAc (20 mL \times 2). The combined organic layers were washed with brine (15 mL), dried over MgSO₄ and concentrated in vacuo to give **8m** as a pale yellow solid (64%, 2 steps).

In a similar manner, **8a-8l** and **8n** were synthesized according to procedure above.

General Procedure for amide coupling. To a stirred solution of 6-chloro-2-ethylimidazo[1,2-a]pyridine-3carboxylic acid (**4f**, 2.83 mmol) in anhydrous DMF (10 mL) was added 1-(3-dimethylaminopropyl)-3ethylcarbodiimide hydrochloride (3.84 mmol), 1-hydroxybenzotriazole (1.54 mmol), triethylamine (5.12 mmol) and 4-(4-(4-(trifluoromethoxy)phenyl)piperidin-1-yl)phenyl)methanamine (**8n**, 2.56 mmol) at room temperature, then the resulting solution was heated to 70 °C with stirring. After 2 hours, the reaction mixture was cooled to room temperature and evaporated. Water (50 mL) was added into the crude residue, the resulting solid was collected by filtration, the filter cake was washed with water (50 mL) and dried to afford crude product. The resulting crude compound was purified by flash column chromatography (*n*-hexane:EtOAc:methylene chloride

= 1:1:1), then recrystallized from EtOAc to give title compound, 50 as a white solid.

6-Chloro-2-ethyl-N-(4-(4-(trifluoromethoxy)phenyl)piperidin-1-yl)benzyl)imidazo[1,2-a]pyridine-3-

	<i>carboxamide</i> (50). mp = 164.0 °C; ¹ H NMR (400 MHz, CDCl ₃); δ 1.37 (t, J = 7.6 Hz, 3H), 1.82 – 1.97 (m, 4H),
	2.64 - 2.70 (m, 1H), $2.80 - 2.87$ (m, 2H), 2.93 (q, $J = 7.6$ Hz, 2H), $3.80 - 3.83$ (m, 2H), 4.61 (d, $J = 5.2$ Hz, 2H),
)	6.00 (brt, J = 5.2 Hz, 1H), 6.96 - 6.99 (m, 2H), 7.15 (d, J = 8.0 Hz, 2H), 7.24 - 7.30 (m, 5H), 7.52 (dd, J = 9.6, 3.0 Hz)
- 3 1 5	0.8 Hz, 1H), 9.53 (dd, <i>J</i> = 2.0, 0.8 Hz, 1H); 13C NMR (100 MHz, CDCl3) δ 13.3, 23.6, 33.4, 42.0, 43.3, 50.4,
, ; ;	115.4, 117.0, 121.2, 121.6, 121.9, 126.3, 128.2, 128.3, 128.7, 128.9, 144.5, 144.7, 147.7, 151.4, 151.5, 161.2;
))	¹⁹ F NMR (376 MHz, CDCl ₃) δ 58.31 (s, 3F); LCMS (ESI) m/z 557 $[M + H]^+$; HRESIMS calcd for
2 3 1	$C_{29}H_{29}ClF_3N_4O_2 [M + H]^+ 557.1926$, found 557.1918.
5	N-([1,1'-Biphenyl]-4-ylmethyl)-6-chloro-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (1). ¹ H NMR (400
3))	MHz, CDCl ₃) δ 1.41 (t, <i>J</i> = 7.6 Hz, 3H), 2.98 (q, <i>J</i> = 7.6 Hz, 2H), 4.74 (d, <i>J</i> = 5.6 Hz, 2H), 6.15 (brs, 1H), 7.29
 2 3	(dd, <i>J</i> = 9.6, 2.0 Hz, 1H), 7.35 - 7.37 (m, 1H), 7.43 - 7.47 (m, 4H), 7.55 (d, <i>J</i> = 9.2 Hz, 1H), 7.58 - 7.62 (m, 4H),
4 5 6	9.56 (d, $J = 2.0$ Hz, 1H); LCMS (ESI) m/z 390 [M + H] ⁺ .
7 3 9	<i>N-([1,1'-Biphenyl]-4-ylmethyl)-2-methylimidazo[1,2-a]pyridine-3-carboxamide</i> (18). ¹ H NMR (400 MHz,
) <u>2</u>	DMSO- d_6) δ 2.62 (s, 3H), 4.57 (d, $J = 6.0$ Hz, 2H), 7.00 (dd, $J = 6.8$, 6.8 Hz, 1H), 7.33 – 7.40 (m, 2H), 7.45 –
3 1 5	7.48 (m, 4H), 7.56 (d, $J = 8.8$ Hz, 1H), 7.64 – 7.66 (m, 4H), 8.38 (brt, $J = 6.0$ Hz, 1H), 9.04 (d, $J = 6.8$ Hz, 1H);
) 7 }	MS (ESI) m/z 342 $[M + H]^+$.
,) 	<i>N-([1,1'-Biphenyl]-4-ylmethyl)-2-ethylimidazo[1,2-a]pyridine-3-carboxamide</i> (19). ¹ H NMR (400 MHz,
- 3 4 5	CDCl ₃) δ 1.42 (t, J = 7.6 Hz, 3H), 3.02 (q, J = 7.6 Hz, 2H), 4.75 (d, J = 5.6 Hz, 2H), 6.19 (brs, 1H), 6.92 (dd, J
5 7 8	= 6.4, 6.4 Hz, 1H), 7.32 (d, J = 7.6 Hz, 1H), 7.35 (d, J = 7.2 Hz, 1H), 7.45 (d, J = 8.0 Hz, 4H), 7.58 – 7.59 (m,
)	5H), 9.41 (d, $J = 6.8$ Hz, 1H); LCMS (ESI) m/z 356 [M + H] ⁺ .
	ACS Faragon Flus Environment

N-([1,1'-Biphenyl]-4-ylmethyl)-2-propylimidazo[1,2-a]pyridine-3-carboxamide (20). ¹H NMR (400 MHz, $CDCl_3$) δ 0.98 (t, J = 7.4 Hz, 3H), 1.80 – 1.89 (m, 2H), 2.93 (t, J = 7.8 Hz, 2H), 4.73 (d, J = 5.6 Hz, 2H), 6.29 (t, J = 7.4 Hz, 3H), 1.80 – 1.89 (m, 2H), 2.93 (t, J = 7.8 Hz, 2H), 4.73 (d, J = 5.6 Hz, 2H), 6.29 (t, J = 7.4 Hz, 3H), 1.80 – 1.89 (m, 2H), 2.93 (t, J = 7.8 Hz, 2H), 4.73 (d, J = 5.6 Hz, 2H), 6.29 (t, J = 7.4 Hz, 3H), 1.80 – 1.89 (m, 2H), 2.93 (t, J = 7.8 Hz, 2H), 4.73 (d, J = 5.6 Hz, 2H), 6.29 (t, J = 7.8 Hz, 2H), 4.73 (d, J = 5.6 Hz, 2H), 6.29 (t, J = 7.8 Hz, 2H), 4.73 (t, J = 7.8 Hz, 2H), 6.29 (t, J = 7.8 Hz, 2H), 4.73 (t, J = 7.8 Hz, 2H), 6.29 (t, J = 7.8 Hz, 2H), 7.8 J = 5.6 Hz, 1H), 6.89 (dd, J = 6.8, 1.2 Hz, 1H), 7.27 – 7.37 (m, 2H), 7.42 – 7.46 (m, 4H), 7.56 – 7.61 (m, 5H), 9.35 (d, J = 6.8 Hz, 1H).; LCMS (ESI) m/z 370 [M + H]⁺. *N*-([1,1'-Biphenyl]-4-ylmethyl)-2-isopropylimidazo[1,2-a]pyridine-3-carboxamide (21). ¹H NMR (400 MHz, $CDCl_3$) δ 1.44 (d, J = 6.4 Hz, 6H), 3.34 – 3.41 (m, 1H), 4.76 (d, J = 5.6 Hz, 2H), 6.16 (m, 1H), 6.90 (dd, J = 7.2, 7.2 Hz, 1H), 7.29 - 7.37 (m, 2H), 7.42 - 7.47 (m, 4H), 7.60 - 7.64 (m, 5H), 9.32 (d, J = 7.2 Hz, 1H); LCMS $(ESI) m/z 370 [M + H]^+$. *N*-([1,1'-Biphenyl]-4-ylmethyl)-6-chloro-2-methylimidazo[1,2-a]pyridine-3-carboxamide (22). ¹H NMR (400 MHz, DMSO- d_6) δ 2.61 (s, 3H), 4.56 (d, J = 5.6 Hz, 2H), 7.32 - 7.34 (m, 1H), 7.41 - 7.46 (m, 5H), 7.62 - 7.64 (m, 5H), 8.47 (brt, J = 5.6 Hz, 1H), 9.13 – 9.14 (m, 1H); LCMS (ESI) m/z 376 [M+H]⁺. 6-Chloro-N-((2'-chloro-[1,1'-biphenyl]-4-yl)methyl)-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (23). ¹H NMR (400 MHz, CDCl₃) δ 1.44 (t, J = 7.6 Hz, 3H), 3.02 (q, J = 7.6 Hz, 2H), 4.77 (d, J = 6.0 Hz, 2H), 6.18 (m, 1H), 7.27 - 7.35 (m, 4H), 7.43 - 7.48 (m, 5H), 7.56 (d, J = 9.6 Hz, 1H), 9.56 (d, J = 2.0 Hz, 1H); LCMS (ESI) $m/z 424 [M + H]^+$. 6-Chloro-N-((4'-chloro-[1,1'-biphenyl]-4-yl)methyl)-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (24). ¹H NMR (400 MHz, CDCl₃) δ 1.43 (t, J = 7.6 Hz, 3H), 3.01 (q, J = 7.6 Hz, 2H), 4.71 (d, J = 6.0 Hz, 2H), 6.13 (m, 1H), 7.31 (dd, J = 9.6, 2.0 Hz, 1H), 7.41 (d, J = 8.8 Hz, 2H), 7.45 (d, J = 8.0 Hz, 2H), 7.51 (d, J = 8.4 Hz, 2H), 7.54 – 7.58 (m, 3H), 9.55 (d, J = 2.0 Hz, 1H); LCMS (ESI) m/z 424 [M+H]⁺; HRESIMS calcd for $C_{23}H_{20}Cl_2N_3O [M + H]^+ 424.0978$, found 424.0989. ACS Paragon Plus Environment

1 2 3	6-Chloro-N-((4'-cyano-[1,1'-biphenyl]-4-yl)methyl)-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (25). ¹ H
4 5 6	NMR (400 MHz, CDCl ₃) δ 1.42 (t, J = 7.6 Hz, 3H), 2.992 (q, J = 7.2 Hz, 2H), 4.75 (d, J = 5.6 Hz, 2H), 6.18
7 8 9	(brt, <i>J</i> = 5.6 Hz, 1H), 7.30 (dd, <i>J</i> = 9.6, 2.0 Hz, 1H), 7.49 (d, <i>J</i> = 8.0 Hz, 2H), 7.56 (d, <i>J</i> = 9.6 Hz, 1H), 7.59 (d, <i>J</i>
10 11 12	= 8.0 Hz, 2H), 7.67 (d, <i>J</i> = 8.4 Hz, 2H), 7.72 (d, <i>J</i> = 8.4 Hz, 2H), 9.55 (d, <i>J</i> = 2.0 Hz, 1H); LCMS (ESI) m/z 415
13 14 15	$\left[\mathrm{M}+\mathrm{H}\right]^{+}$.
16 17 18	6-Chloro-2-ethyl-N-((4'-methyl-[1,1'-biphenyl]-4-yl)methyl)imidazo[1,2-a]pyridine-3-carboxamide (26). ¹ H
19 20 21	NMR (400 MHz, CDCl ₃) δ 1.42 (t, J = 7.6 Hz, 3H), 3.00 (q, J = 7.6 Hz, 2H), 2.40 (s, 3H), 4.74 (d, J = 5.6 Hz,
22 23 24	2H), 6.16 (m, 1H), 7.25 (d, <i>J</i> = 7.2 Hz, 2H), 7.30 (dd, <i>J</i> = 9.6, 2.0 Hz, 1H), 7.44 (d, <i>J</i> = 8.0 Hz, 2H), 7.49 (d, <i>J</i> =
25 26 27	8.0 Hz, 2H), 7.54 (d, <i>J</i> = 9.6 Hz, 1H), 7.59 (d, <i>J</i> = 8.4 Hz, 2H), 9.55 (d, <i>J</i> = 2.0 Hz, 1H); LCMS (ESI) m/z 404
28 29 30	$[M + H]^+$.
31 32 33	7-Chloro-2-ethyl-N-((4'-methyl-[1,1'-biphenyl]-4-yl)methyl)imidazo[1,2-a]pyridine-3-carboxamide (27). 1 H
34 35 36	NMR (400 MHz, CDCl ₃) δ 1.41 (t, J = 7.6 Hz, 3H), 2.40 (s, 3H), 2.99 (q, J = 7.6 Hz, 2H), 4.73 (s, 2H), 6.91 (dd, 2H))
37 38 39	J = 7.6, 2.0 Hz, 1H), 7.25 (d, $J = 8.4$ Hz, 2H), 7.43 (d, $J = 8.0$ Hz, 2H), 7.48 (d, $J = 8.0$ Hz, 2H), 7.58 – 7.60 (m,
40 41 42	3H), 9.38 (d, $J = 7.2$ Hz, 1H) ; LCMS (ESI) m/z 404 [M + H] ⁺ .
43 44 45	7-Chloro-N-((4'-chloro-[1,1'-biphenyl]-4-yl)methyl)-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (28). 1 H
46 47 48	NMR (400 MHz, CDCl ₃) δ 1.42 (t, <i>J</i> = 7.6 Hz, 3H), 2.99 (q, <i>J</i> = 7.6 Hz, 2H), 4.73 (s, 2H), 6.15 (brs, 1H), 6.91
49 50 51 52	(dd, J = 7.6, 2.0 Hz, 1H), 7.40 (d, J = 8.4 Hz, 2H), 7.44 (d, J = 8.0 Hz, 2H), 7.51 (d, J = 8.4 Hz, 2H), 7.56 (d, J
53 54 55	= 8.0 Hz, 2H), 7.60 (d, J = 1.6 Hz, 1H), 9.38 (d, J = 7.2 Hz, 1H) ; LCMS (ESI) m/z 424 [M + H] ⁺ ; HRESIMS
56 57 58 59 60	calcd for $C_{23}H_{20}Cl_2N_3O [M + H]^+ 424.0978$, found 424.0983.

4'-((7-Chloro-2-ethylimidazo[1,2-a]pyridine-3-carboxamido)methyl)-[1,1'-biphenyl]-4-carboxylic acid (29). ¹H NMR (400 MHz, DMSO- d_6) δ 1.26 (t, J = 7.6 Hz, 3H), 2.98 (q, J = 7.6 Hz, 2H), 4.58 (d, J = 6.0 Hz, 2H), 7.08 (dd, J = 7.2, 2.4 Hz, 1H), 7.48 (d, J = 8.4 Hz, 2H), 7.72 (d, J = 8.0 Hz, 2H), 7.78 - 7.80 (m, 3H), 8.00 (d, J = 8.0 Hz, 2H), 7.80 (m, 3H), 8.00 (m, 3H), 8. $= 8.4 \text{ Hz}, 2\text{H}, 8.51 \text{ (brt, } J = 6.0 \text{ Hz}, 1\text{H}), 8.97 \text{ (d, } J = 7.2 \text{ Hz}, 1\text{H}); \text{LCMS (ESI) m/z } 434 \text{ [M + H]}^{+}.$ N-((4'-(tert-Butyl)-[1,1'-biphenyl]-4-yl)methyl)-7-chloro-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (30). ¹H NMR (400 MHz, CDCl₃) δ 1.36 (s, 9H), 1.41 (t, J = 7.6 Hz, 3H), 2.99 (q, J = 7.6 Hz, 2H), 4.73 (d, J = 5.6 Hz, 2H), 6.13 (brs, 1H), 6.91 (dd, J = 7.2, 2.0 Hz, 1H), 7.43 (d, J = 8.0 Hz, 2H), 7.47 (d, J = 8.4 Hz, 2H), 7.53 $(d, J = 8.4 \text{ Hz}, 2\text{H}), 7.59 - 7.61 \text{ (m, 3H)}, 9.38 \text{ (d, } J = 7.2 \text{ Hz}, 1\text{H}); LCMS \text{ (ESI) m/z 446 [M + H]}^+.$ *N*-(4-(1*H*-Pyrrol-2-yl)benzyl)-7-chloro-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (**31**). ¹H NMR (400 MHz, CDCl₃) δ 1.37 (t, J = 7.6 Hz, 3H), 2.94 (q, J = 7.6 Hz, 2H), 4.67 (d, J = 6.0 Hz, 2H), 6.10 (brs, 1H), 6.29 -6.32 (m, 1H), 6.52 - 6.54 (m, 1H), 6.86 - 6.88 (m, 1H), 6.89 (dd, J = 7.2, 2.0 Hz, 1H), 7.35 (d, J = 8.0 Hz, 2H), 7.47 (d, J = 8.4 Hz, 2H), 7.58 (d, J = 2.0 Hz, 1H), 8.51 (brs, 1H), 9.35 (d, J = 7.2 Hz, 1H); LCMS (ESI) m/z 379 $[M + H]^+$. 7-Chloro-2-ethyl-N-(4-(pyridin-4-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (32). ¹H NMR (400 MHz, $CDCl_3$) δ 1.42 (t, J = 7.6 Hz, 3H), 3.01 (q, J = 7.6 Hz, 2H), 4.76 (d, J = 5.6 Hz, 2H), 6.20 (brs, 1H), 6.91 (dd, J = 7.6, 2.0 Hz, 1H), 7.26 – 7.51 (m, 4H), 7.61 (d, J = 2.0 Hz, 1H), 7.65 (d, J = 7.6 Hz, 2H), 8.65 (brs, 2H), 9.37 $(d, J = 7.6 \text{ Hz}, 1\text{H}); \text{ LCMS (ESI) } \text{m/z } 391 \text{ [M + H]}^+.$

7-Chloro-2-ethyl-N-(4-(piperidin-1-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (33). ¹H NMR (400 MHz, CDCl₃) δ 1.35 (t, J = 7.6 Hz, 3H), 1.55 – 1.57 (m, 2H), 1.66 – 1.70 (m, 4H), 2.91 (q, J = 7.6 Hz, 2H),
3.12 – 3.15 (m, 4H), 4.56 (d, J = 5.6 Hz, 2H), 6.07 (brs, 1H), 6.86 (dd, J = 7.6, 2.0 Hz, 1H), 6.90 (d, J = 8.4 Hz, ACS Paragon Plus Environment

1 2 3	2H), 7.22 (d, <i>J</i> = 8.4 Hz, 2H), 7.54 (d, <i>J</i> = 2.0 Hz, 1H), 9.30 (d, <i>J</i> = 7.6 Hz, 1H) ; LCMS (ESI) m/z 397 [M +
4 5 6	H] ⁺ .
7 8 9	<i>N-(4-(Azepan-1-yl)benzyl)-7-chloro-2-ethylimidazo[1,2-a]pyridine-3-carboxamide</i> (34). ¹ H NMR (400 MHz,
10 11 12	CDCl ₃) δ 1.38 (t, <i>J</i> = 7.6 Hz, 3H), 1.52-1.55 (m, 4H), 1.78 (m, 4H), 2.94 (q, <i>J</i> = 7.6 Hz, 2H), 3.45 (t, <i>J</i> = 6.0 Hz,
13 14 15	4H), 4.56 (d, <i>J</i> = 5.2 Hz, 2H), 5.95 (brs, 1H), 6.67 (d, <i>J</i> = 8.8 Hz, 2H), 6.89 (dd, <i>J</i> = 7.6, 2.4 Hz, 1H), 7.20 (d, <i>J</i>
16 17 18	= 8.8 Hz, 2H), 7.57 (d, J = 2.4 Hz, 1H), 9.36 (d, J = 7.6 Hz, 1H); LCMS (ESI) m/z 411 [M+H] ⁺ .
19 20 21	7-Chloro-2-ethyl-N-(4-(4-(trifluoromethyl)piperidin-1-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (35).
22 23 24 25	mp = 209.4 °C; ¹ H NMR (400 MHz, CDCl ₃); δ 1.34 (t, J = 7.6 Hz, 3H), 1.68 – 1.78 (m, 2H), 1.94 – 1.98 (m,
25 26 27 28	2H), 2.11 – 2.20 (m, 1H), 2.66 – 2.73 (m, 2H), 2.90 (q, <i>J</i> = 7.6 Hz, 2H), 3.73 – 3.77 (m, 2H), 4.58 (d, <i>J</i> = 5.2 Hz,
20 29 30 31	2H), 6.03 (brt, <i>J</i> = 5.2 Hz, 1H), 6.86 (dd, <i>J</i> = 7.6, 2.4 Hz, 1H), 6.91 (d, <i>J</i> = 8.8 Hz, 2H), 7.25 (d, <i>J</i> = 8.8 Hz, 2H),
32 33 34	7.56 (d, $J = 2.4$ Hz, 1H), 9.32 (d, $J = 7.6$ Hz, 1H); LCMS (ESI) m/z 465 [M + H] ⁺ .
35 36 37	7-Chloro-N-(4-(4-chloropiperidin-1-yl)benzyl)-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (36). ¹ H NMR
38 39 40	(400 MHz, CDCl ₃); δ 1.36 (t, J = 7.2 Hz, 3H), 1.99 – 2.04 (m, 2H), 2.17 – 2.20 (m, 2H), 2.92 (q, J = 7.2 Hz,
41 42 43	2H), 3.05 – 3.10 (m, 2H), 3.50 – 3.52 (m, 2H), 4.20 – 4.23 (m, 1H), 4.59 (d, J = 5.6 Hz, 2H), 5.99 (brs, 1H),
44 45 46	$6.89 - 6.94 \text{ (m, 3H)}, 7.26 - 7.27 \text{ (m, 2H)}, 7.58 \text{ (s, 1H)}, 9.35 \text{ (d, } J = 6.8 \text{ Hz}, 1\text{H}); LCMS \text{ (ESI) m/z 431 [M + H]}^+.$
47 48 49	7-Chloro-2-ethyl-N-(4-(4-methylpiperazin-1-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (37). ¹ H
50 51 52	NMR (400 MHz, CDCl ₃) δ 1.37 (t, <i>J</i> = 7.6 Hz, 3H), 2.35 (s, 3H), 2.57 – 2.59 (m, 4H), 2.94 (q, <i>J</i> = 7.6 Hz, 2H),
53 54 55	3.20 – 3.23 (m, 4H), 4.59 (d, <i>J</i> = 5.2 Hz, 2H), 6.00 (brs, 1H), 6.88 – 6.94 (m, 3H), 7.27 (d, <i>J</i> = 8.4 Hz, 2H), 7.58
56 57 58 59	(d, $J = 2.0$ Hz, 1H), 9.35 (d, $J = 7.6$ Hz, 1H); LCMS (ESI) m/z 412 [M + H] ⁺ .

 ^{1}H 7-Chloro-2-ethyl-N-(4-(4-isopropylpiperazin-1-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (38). NMR (400 MHz, CDCl₃) δ 1.09 (d, J = 6.0 Hz, 6H), 1.35 (t, J = 7.6 Hz, 3H), 2.65 – 2.75 (m, 4H), 2.91 (q, J = 7.6 Hz, 2H), 3.18 - 3.27 (m, 4H), 4.59 (d, J = 5.6 Hz, 2H), 5.99 (brs, 1H), 6.88 (dd, J = 7.6, 2.0 Hz, 1H), 6.91 (d, J = 8.4 Hz, 2H), 7.26 – 7.28 (m, 2H), 7.58 (d, J = 2.0 Hz, 1H), 9.36 (d, J = 7.6 Hz, 1H); LCMS (ESI) m/z 440 $[M + H]^{+}$. 7-Chloro-2-ethyl-N-(4-(octahydro-2H-isoindol-2-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (**39**). ¹H NMR (400 MHz, CDCl₃); δ 1.36 (t, J = 7.6 Hz, 3H), 1.40 – 2.03 (m, 8H), 2.29 – 2.34 (m, 2H), 2.92 (q, J = 7.2) Hz, 2H), 3.16 (dd, J = 9.2, 5.2 Hz, 2H), 3.29 (dd, J = 8.8, 6.8 Hz, 2H), 4.55 (d, J = 5.2 Hz, 2H), 5.97 (brs, 1H), 6.49 (d, J = 8.4 Hz, 2H), 6.88 (dd, J = 7.6, 2.4 Hz, 1H), 7.21 (d, J = 8.4 Hz, 2H), 7.56 (d, J = 2.4 Hz, 1H), 9.33(d, J = 7.6 Hz, 1H); LCMS (ESI) m/z 437 $[M + \text{H}]^+$. 7-Chloro-2-ethyl-N-(4-(4,5,6,7-tetrahydro-2H-isoindol-2-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (40). ¹H NMR (400 MHz, CDCl₃); δ 1.39 (t, J = 7.6 Hz, 3H), 1.74 – 1.77 (m, 4H), 2.63 (m, 4H), 2.97 (q, J = 7.6 Hz, 3H), 1.74 – 1.77 (m, 4H), 2.63 (m, 4H), 2.97 (q, J = 7.6 Hz, 3H) Hz, 2H), 4.68 (d, J = 6.0 Hz, 2H), 6.14 (brs, 1H), 6.78 (s, 2H), 6.91 (dd, J = 7.6, 2.0 Hz, 1H), 7.32 (d, J = 8.4 Hz, 2H), 7.38 (d, J = 8.4 Hz, 2H), 7.59 (d, J = 2.0 Hz, 1H), 9.36 (d, J = 7.2 Hz, 1H); LCMS (ESI) m/z 433 [M + H]⁺. 7-Chloro-2-ethyl-N-(4-(4-(4-fluorophenyl)piperazin-1-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (41). mp = 212 - 213 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.39 (t, J = 7.6 Hz, 3H), 2.95 (q, J = 7.6 Hz, 2H), 3.24 - 3.26 (m, 4H), 3.33 - 3.36 (m, 4H), 4.62 (d, J = 5.6 Hz, 2H), 6.01 (brs, 1H), 6.89 - 7.02 (m, 7H), 7.30 (d, J = 8.4 Hz, 10.10 Hz)2H), 7.59 (d, J = 2.0 Hz, 1H), 9.37 (d, J = 7.2 Hz, 1H); LCMS (ESI) m/z 492 [M + H]⁺; HRESIMS calcd for $C_{27}H_{28}CIFN_5O [M + H]^+ 492.1961$, found 492.1964. ACS Paragon Plus Environment

7-Chloro-2-ethyl-N-(4-(4-(4-fluorophenyl)piperidin-1-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (42). mp = 182.7 °C; ¹H NMR (400 MHz, CDCl₃); δ 1.35 (t, J = 7.6 Hz, 3H), 1.79 – 1.95 (m, 4H), 2.59 – 2.67 (m, 1H), 2.78 - 2.85 (m, 2H), 2.91 (q, J = 7.6 Hz, 2H), 3.79 - 3.82 (m, 2H), 4.59 (d, J = 5.6 Hz, 2H), 6.03 (brt, J = 5.6 Hz, 2H), 5.03 (brt, J = 5.6 Hz, J = 5.5.6 Hz, 1H), 6.87 (dd, J = 7.6, 2.4 Hz, 1H), 6.96 – 7.01 (m, 4H), 7.17 – 7.21 (m, 2H), 7.26 (d, J = 8.8 Hz, 2H), 7.57 (d, J = 2.4 Hz, 1H), 9.33 (d, J = 7.6 Hz, 1H); LCMS (ESI) m/z 491 [M + H]⁺; HRESIMS calcd for $C_{28}H_{29}CIFN_4O [M + H]^+ 491.2008$, found 491.2003. 6-Chloro-2-ethyl-N-(4-(4-(4-fluorophenyl)piperazin-1-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (43). mp = 212 - 213 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.39 (t, J = 7.6 Hz, 3H), 2.95 (q, J = 7.6 Hz, 2H), 3.24 -3.27 (m, 4H), 3.29 - 3.63 (m, 4H), 4.62 (d, J = 5.6 Hz, 2H), 6.03 (brs, 1H), 6.89 - 7.02 (m, 7H), 7.30 (d, J = 8.4)Hz, 2H), 7.59 (d, J = 2.0 Hz, 1H), 9.37 (d, J = 7.2 Hz, 1H); LCMS (ESI) m/z 492 [M + H]⁺; HRESIMS calcd for $C_{27}H_{28}ClFN_5O [M + H]^+ 492.1961$, found 492.1958. 7-Chloro-2-ethyl-N-(4-(4-(trifluoromethoxy)phenyl)piperazin-1-yl)benzyl)imidazo[1,2-a]pyridine-3*carboxamide* (44). mp = 216.3 – 217.0 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.39 (t, J = 7.6 Hz, 3H), 2.95 (q, J = 7.6 Hz, 2H), 3.30 - 3.40 (m, 8H), 4.62 (d, J = 5.6 Hz, 2H), 6.01 - 6.02 (m, 1H), 6.90 (dd, J = 7.2, 2.0 Hz, 1H), 6.94 (d, J = 9.2 Hz, 2H), 6.98 (d, J = 8.8 Hz, 2H), 7.14 (d, J = 8.8 Hz, 2H), 7.31 (d, J = 8.8 Hz, 2H), 7.59 (d, J = 8.81.6 Hz, 1H), 9.37 (d, J = 7.6 Hz, 1H); LCMS (ESI) m/z 558 [M + H]⁺; HRESIMS calcd for C₂₈H₂₈ClF₃N₅O₂ [M + H]⁺ 558.1878, found 558.1885. 6-Chloro-2-ethyl-N-(4-(4-(4-(trifluoromethoxy)phenyl)piperazin-1-yl)benzyl)imidazo[1,2-a]pyridine-3*carboxamide* (45). mp = 206.5 – 207.0 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.40 (t, J = 7.6 Hz, 3H), 2.96 (q, J = 7.6 Hz, 2H), 3.30 - 3.40 (m, 8H), 4.63 (d, J = 5.2 Hz, 2H), 6.03 - 6.04 (m, 1H), 6.95 (d, J = 9.2 Hz, 2H), 6.98

(u, J = 0.4 Hz, 211), 7.14 (u, J = 0.0 Hz, 211), 7.27 = 7.52 (m, 511), 7.54 (u, J = 9.0 Hz, 111), 9.55 = 9.54 (m, 111), 7.54 (u, J = 9.0 Hz, 111), 9.55 = 9.54 (m, 111), 9.55
LCMS (ESI) m/z 558 $[M + H]^+$; HRESIMS calcd for $C_{28}H_{28}ClF_3N_5O_2 [M + H]^+$ 558.1878, found 558.1881.
6-Chloro-2-ethyl-N-(4-(4-(4-fluorophenyl)piperidin-1-yl)benzyl)imidazo[1,2-a]pyridine-3-carboxamide (46).
mp = 164.0 °C; ¹ H NMR (400 MHz, CDCl ₃); δ 1.35 (t, J = 7.6 Hz, 3H), 1.76 – 1.95 (m, 4H), 2.60 – 2.66 (m,
1H), $2.78 - 2.85$ (m, 2H), 2.92 (q, $J = 7.6$ Hz, 2H), $3.79 - 3.82$ (m, 2H), 4.60 (d, $J = 5.2$ Hz, 2H), 6.03 (brt, $J = 5.2$ Hz, 2H), 5.03 (brt, $J = 5.2$ Hz, 5.2 Hz,
5.2 Hz, 1H), 6.96 – 7.01 (m, 4H), 7.17 – 7.21 (m, 2H), 7.26 – 7.29 (m, 3H), 7.51 (d, <i>J</i> = 9.6 Hz, 1H), 9.52 (d, <i>J</i>
= 1.6 Hz, 1H); LCMS (ESI) m/z 491 $[M + H]^+$; HRESIMS calcd for C ₂₈ H ₂₉ ClFN ₄ O $[M + H]^+$ 491.2008, found
491.1996.
7-Chloro-N-(4-(4-(4-chlorophenyl)piperidin-1-yl)benzyl)-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (47).
mp = 177.0 °C; ¹ H NMR (400 MHz, CDCl ₃) δ 1.40 (t, J = 7.4 Hz, 3H), 1.83 – 1.95 (m, 4H), 2.60 – 2.67 (m,
1H), 2.79 - 2.86 (m, 2H), 2.96 (q, <i>J</i> = 7.4 Hz, 2H), 3.80 – 3.83 (m, 2H), 4.62 (q, <i>J</i> = 5.2 Hz, 2H), 6.02 (brs, 1H),
6.98 (d, J = 8.8 Hz, 2H), 7.18 (d, J = 8.4 Hz, 2H), 7.26 – 7.31 (m, 5H), 7.54 (d, J = 9.6 Hz, 1H), 9.30 (d, J = 7.6
Hz, 1H); LCMS (ESI) m/z 507 $[M + H]^+$; HRESIMS calcd for $C_{28}H_{29}Cl_2N_4O [M + H]^+$ 507.1713, found
507.1709.
6-Chloro-N-(4-(4-(4-chlorophenyl)piperidin-1-yl)benzyl)-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (48).
¹ H NMR (400 MHz, CDCl ₃) δ 1.39 (t, J = 7.6 Hz, 3H), 1.80 – 1.96 (m, 4H), 2.60 – 2.66 (m, 1H), 2.79 – 2.86 (m, 2H), 2.89 (m, 2H
2H), 2.95 (q, J = 7.6 Hz, 2H), 3.79 – 3.83 (m, 2H), 4.61 (q, J = 5.2 Hz, 2H), 6.00 (brs, 1H), 6.90 (dd, J = 7.6,
2.0 Hz, 1H), 6.98 (d, J = 8.4 Hz, 2H), 7.18 (d, J = 8.4 Hz, 2H), 7.26 – 7.29 (m, 4H), 7.59 (d, J = 2.0 Hz, 1H),
9.30 (d, $J = 7.6$ Hz, 1H); LCMS (ESI) m/z 507 [M + H] ⁺ ; HRESIMS calcd for C ₂₈ H ₂₉ Cl ₂ N ₄ O [M + H] ⁺
507.1713, found 507.1711.

7-*Chloro-2-ethyl-N-(4-(4-(trifluoromethoxy)phenyl)piperidin-1-yl)benzyl)imidazo[1,2-a]pyridine-3carboxamide* (**49**). ¹H NMR (400 MHz, CDCl₃); δ 1.36 (t, *J* = 7.6 Hz, 3H), 1.82 – 1.96 (m, 4H), 2.64 – 2.70 (m, 1H), 2.79 – 2.86 (m, 2H), 2.91 (q, *J* = 7.6 Hz, 2H), 3.80 – 3.83 (m, 2H), 4.59 (d, *J* = 5.6 Hz, 2H), 6.04 (brs, 1H), 6.87 (dd, *J* = 1.6, 7.2 Hz, 1H), 6.97 (d, *J* = 8.4 Hz, 2H), 7.14 (d, *J* = 8.4 Hz, 2H), 7.24 – 7.28 (m, 4H), 7.57 (d, *J* = 1.6 Hz, 1H), 9.34 (d, *J* = 7.2 Hz, 1H); LCMS (ESI) m/z 558 [M + H]⁺; HRESIMS calcd for C₂₉H₂₉ClF₃N₄O₂ [M + H]⁺ 557.1926, found 557.1912.

Synthesis of N-([1,1]-biphenyl]-4-yl)-6-chloro-2-ethylimidazo[1,2-a]pyridine-3-carboxamide (9). A mixtureof**4f**(0.13 mmol) and thionyl chloride (2 mL) was heated to 100°C for an hour. The reaction mixture wasconcentrated and dried in vacuo. The resulting residue was dissolved in methylene chloride (5 mL) and then[1,1]-biphenyl]-4-amine (0.16 mmol) and triethylamine (0.40 mmol) were added. The reaction mixture wasstirred for 2 hours, diluted with methylene chloride (10 mL) and washed with water (10 mL) and brine (10 mL).The organic phase was dried over MgSO₄ and concentrated in vacuo. The resulting crude residue was purifiedby flash column chromatography (*n*-hexane:EtOAc = 1:1 to methylene chloride:MeOH = 20:1) to give a targetcompound as a white solid (60%). ¹H NMR (400MHz, DMSO-*d* $₆) <math>\delta$ 1.25 – 1.29 (m, 3H), 3.01 – 3.05 (m, 2H), 7.31 – 7.35 (m, 1H), 7.42 – 7.46 (m, 2H), 7.46 – 7.51 (m, 1H), 7.65 – 7.71 (m, 5H), 7.80 (d, *J* = 8.8 Hz, 2H), 8.96 (d, *J* = 2.0 Hz, 1H), 10.17 (s, 1H); LCMS (ESI) m/z 376 [M + H]⁺.

Synthesis of N-([1,1'-biphenyl]-4-ylmethyl)-6-chloro-2-ethyl-N-methylimidazo[1,2-a]pyridine-3-carboxamide (10). To a stirred solution of 1 (0.26 mmol) in DMF (5 mL) was added NaH (60% dispersion in paraffin, 0.38 mmol) under ice-bath. After 20 min, iodomethane (0.32 mmol) was added and the reaction mixture was allowed to ambient temperature and further stirred for an hour. The mixture was diluted with EtOAc (20 mL), washed ACS Paragon Plus Environment

2
2
3
4
5
6
7
1
8
9
10
10
11
12
12
13
14
15
16
47
17
18
19
20
20
21
22
23
20
24
25
26
20
21
28
29
20
30
31
32
33
24
34
35
36
27
51
38
39
40
14
41
42
43
44
45
45
46
47
10
40
49
50
51
50
5Z
53
54
55
55
56
57
58
50
29
~~

with water (15 mL) and brine (15 mL), dried over MgSO₄ and concentrated in vacuo. The resulting residue was purified by flash column chromatography (*n*-hexane:EtOAc = 1:2) to give a target compound as a white solid (81%). ¹H NMR (400MHz, DMSO- d_6) δ 1.26 (t, J = 7.6 Hz, 3H), 2.73 (q, J = 7.6 Hz, 2H), 2.96 (s, 3H), 4.72 (2H), 7.36 - 7.48 (m, 6H), 7.63 - 7.67 (m, 5H), 8.51 (d, J = 2.0 Hz, 1H); LCMS (ESI) m/z 404 [M + H]⁺. of 1-([1,1'-biphenyl]-4-yl)-N-((6-chloro-2-ethylimidazo[1,2-a]pyridin-3-yl)methyl)methanamine Synthesis (11). To a stirred solution of 1 (0.050 mmol) in THF (10 mL) under ice-bath was added sodium borohydride (0.25 mmol) and boron trifluoride etherate (0.25 mmol) was then added dropwise. The mixture was heated to 70 °C for 2 hours. After cooling, the mixture was diluted with EtOAc (10 mL), washed with brine (10 mL), dried over MgSO₄ and concentrated in vacuo. The resulting crude residue was purified by flash column chromatography (*n*-hexane:EtOAc = 1:1 to 1:2) to give a title compound as a pale yellow solid (22%). ¹H NMR $(400 \text{ MHz}, \text{ acetone-} d_6) \delta 1.26 \text{ (t, } J = 7.6 \text{ Hz}, 3\text{H}), 2.71 \text{ (q, } J = 7.6 \text{ Hz}, 2\text{H}), 3.85 \text{ (s, } 2\text{H}), 4.17 \text{ (s, } 2\text{H}), 7.17 \text{ (dd, } J = 7.6 \text{ Hz}, 3\text{H}), 3.85 \text{ (s, } 2\text{H}), 4.17 \text{ (s, } 2\text{H}), 7.17 \text{ (dd, } J = 7.6 \text{ Hz}, 3\text{H}), 3.85 \text{ (s, } 2\text{H}), 4.17 \text{ (s, } 2\text{H}), 7.17 \text{ (dd, } J = 7.6 \text{ Hz}, 3\text{H}), 3.85 \text{ (s, } 2\text{H}), 4.17 \text{ (s, } 2\text{H}), 7.17 \text{ (dd, } J = 7.6 \text{ Hz}, 3\text{H}), 3.85 \text{ (s, } 2\text{H}), 3.85 \text{ (s, } 2\text{H$ = 9.6, 1.2 Hz, 1H, 7.34 - 7.36 (m, 1H), 7.42 - 7.47 (m, 5H), 7.60 (d, J = 8.0 Hz, 2H), 7.65 (d, J = 8.0 Hz, 2H),8.53 (d, J = 1.6 Hz, 1H); LCMS (ESI) m/z 376 [M + H]⁺.

Synthesis of ethyl 4-bromo-3-oxohexanoate (12). To a flask containing ethyl 3-oxohexanoate (9.48 mmol) in diethyl ether (50 mL) was added ammonium acetate (0.95 mmol). Here, 0.1 equivalent of ammonium acetate is essential to give 4-bromo product. After 2 hours of pre-stirring, *N*-bromosuccinimide (10.43 mmol) was then added and the resulting mixture was stirred for overnight. The reaction mixture was diluted with diethyl ether (20 mL), washed with brine (50 mL), dried over MgSO₄ and concentrated in vacuo to give a title compound as a pale yellow oil. The resulting crude compound was used for next reaction without further purification.

1 2 3	Synthesis of 2-(6-chloro-2-ethylimidazo[1,2-a]pyridin-3-yl)acetic acid (13). A mixture of 12 (9.48 mmol) and
4 5 6	2-amino-5-chloroaniline (9.48 mmol) in EtOH (20 mL) was heated to reflux temperature. After overnight, the
7 8 0	solution was cooled to room temperature and concentrated under reduced pressure. The resulting residue was
10 11	dissolved in EtOAc (50 mL), washed with sat. NaHCO ₃ (aq. 40 mL) and brine (40 mL), dried over MgSO ₄ and
12 13 14 15	concentrated in <i>vacuo</i> . The crude residue was purified by flash column chromatography (n -hexane:EtOAc = 5:1
16 17 18	to 2:1). The obtained ester (2.63 mmol) was dissolved in MeOH (9 mL) and aqueous lithium hydroxide (7.89
19 20 21	mmol in 3mL H ₂ O) was then treated. The resulting mixture was stirred at ambient temperature for overnight.
22 23 24	The organic solvent was removed and remained aqueous solution was acidified with 1N HCl until pH reach
25 26 27	around 5. The generated pale yellow solid was filtered and dried under reduced pressure to give a title
28 29 30	compound as a pale yellow solid. ¹ H NMR (400MHz, MeOH- d_4) δ 1.30 (t, J = 7.6 Hz, 3H), 3.08 (q, J = 7.6 Hz,
31 32 33	2H), 4.03 (s, 2H), 7.90 – 7.98 (m, 2H), 9.00 (s, 1H).
34 35 36	The target compounds (14-15) were synthesized according to general amide coupling procedure which was
37 38 39	described above.
40 41 42	N-([1,1'-biphenyl]-4-ylmethyl)-2-(6-chloro-2-ethylimidazo[1,2-a]pyridin-3-yl)acetamide (14). ¹ H NMR
43 44 45	(400MHz, CDCl ₃) δ 1.22 (t, <i>J</i> = 7.6 Hz, 3H), 2.91 (q, <i>J</i> = 7.6 Hz, 2H), 3.78 (s, 2H), 4.50 (d, <i>J</i> = 5.6 Hz, 2H),
46 47 48	7.12 – 7.14 (m, 1H), 7.28 – 7.53 (m, 10H), 7.55 (brs, 1H), 7.93 – 7.94 (m, 1H); LCMS (ESI) m/z 404 [M + H] ⁺ .
49 50 51 52	N-([1,1'-biphenyl]-4-yl)-2-(6-chloro-2-ethylimidazo[1,2-a]pyridin-3-yl)acetamide (15). ¹ H NMR (400MHz,

CDCl₃) δ 1.26 (t, *J* = 7.6 Hz, 3H), 2.94 (q, *J* = 7.6 Hz, 2H), 3.86 (s, 2H), 7.19 – 7.20 (m, 1H), 7.38 – 7.39 (m, 1H), 7.40 – 7.41 (m, 2H), 7.51 – 7.62 (m, 7H), 7.97 – 7.98 (m, 1H), 9.70 (s, 1H); LCMS (ESI) m/z 390 [M +

 $H]^+$.

Synthesis of tert-butyl (6-chloro-2-ethylimidazo[1,2-a]pyridin-3-yl)carbamate (16). To a stirred solution of 4f (1.33 mmol) in t-BuOH (8 mL) were added diphenylphosphoryl azide (1.60 mmol) and triethyl amine (2.00 mmol). The resulting mixture was heated to 95 $^{\circ}$ C with stirring for overnight. The organic solvent was removed and the resulting residue was dissolved in methylene chloride (20 mL). The organic solution was washed with water (15 mL) and brine (15 mL), dried over MgSO₄ and concentrated in vacuo. The crude residue was purified by flash column chromatography (*n*-hexane:EtOAc = 2:1 to 1:1) to give a title compound as a white solid (51%). ¹H NMR (400MHz, acetone- d_6) $\delta 1.25$ (t, J = 7.6 Hz, 3H), 1.41 (s, 9H), 2.63 (q, J = 7.6 Hz, 2H), 7.21 (dd, J = 2.0, 9.2 Hz, 1H), 7.46 (d, J = 9.2 Hz, 1H), 8.09 (d, J = 2.0 Hz, 1H), 8.15 (brs, 1H, NH). Synthesis of 2-([1,1'-biphenyl]-4-yl)-N-(6-chloro-2-ethylimidazo[1,2-a]pyridin-3-yl)acetamide (17). To a stirred solution of 16 (0.64 mmol) in methylene chloride (2 mL) was treated trifluroacetic acid (2 mL) and the reaction mixture was stirred for an hour at room temperature. The mixture was evaporated and then sticky residue was dissolved in methylene chloride (15 mL) again. The organic solution was washed with saturated Na₂CO₃ (aq. 10 mL), dried over MgSO₄ and concentrated in *vacuo* to give the amine intermediate as a yellow solid (88%). A mixture of 2-([1,1'-biphenyl]-4-yl)acetic acid (0.76 mmol) and thionyl chloride (6 mL) was heated to 100 °C for an hour. The reaction mixture was concentrated and dried in *vacuo*. The resulting residue was dissolved in methylene chloride (5 mL) and then amine intermediate (0.64 mmol) and triethylamine (1.92 mmol) were added. The reaction mixture was stirred for 2 hours, diluted with methylene chloride (10 mL) and washed with water (10 mL) and brine (10 mL). The organic phase was dried over MgSO₄ and concentrated in *vacuo*. The resulting crude residue was purified by flash column chromatography (*n*-hexane:EtOAc = 1:1 to methylene chloride:MeOH = 20:1) to give a title compound (12%) as a white solid. ¹H NMR (400MHz, **ACS Paragon Plus Environment**

1 2 3	acetone- d_6) δ 1.18 (t, J = 7.6 Hz, 3H), 2.60 (q, J = 7.6 Hz, 2H), 3.93 (s, 2H), 7.16 – 7.20 (dd, J = 9.2, 2.0 Hz,
4 5 6	1H), 7.32 – 7.38 (m, 1H), 7.40 – 7.49 (m, 3H), 7.57 – 7.68 (m, 2H), 7.65 – 7.69 (m, 4H), 7.98 (d, <i>J</i> = 2.0 Hz,
7 8 9 10 11 12	1H), 9.14 (s, 1H); LCMS (ESI) m/z 390 [M + H] ⁺ .
13 14 15 16	ASSOCIATED CONTENT
17 18 19 20	Supporting Information
21 22 23	In vivo PK results of compound 24; kinetic solubility values of compound 24-26, 43, 45-46 and 48-50; HPLC
24 25 26	purity of compounds 41 – 50; ¹ H NMR spectra of compounds 3a – 3g, 1, 9-11 and 13-50; ¹³ C and ¹⁹ F NMR
27 28 29 30	spectra of compound 50 . This material is available free of charge via the Internet at http://pubs.acs.org.
31 32 33	AUTHOR INFORMATION
34 35 36	Corresponding Author
37 38 39	*Jaeseung Kim (Jaeseung K.); Phone: +82-10-4262-3528, Fax.: +82-31-8018-8015, E-mail:
40 41 42 43 44	silanediol@gmail.com; *Zaesung No (Z. N.); Phone: +82-31-888-6006, E-mail: jsnoh@gstep.re.kr
45 46 47	Author Contributions
48 49 50	K.P., J.J., H.J.K. and R.K., designed and performed growth inhibition experiments, S.K., M.J.S, S.L., Y.M.K.,
51 52 53	M.S., J.J.S., Y.K., I.C., and Jaeseung K., designed and synthesized the compounds, S.A., S.P., J.N., Jung. K.,
54 55 56	H.K. and K.N. performed and designed <i>in vivo</i> pharmacokinetic and efficacy experiments, S.K. and Jaeseung K.
57 58 59	wrote the manuscript with contributions from other authors. Z. N. and Jaeseung K. supervised the project.
60	The authors declare no competing financial interest. ACS Paragon Plus Environment

ACKNOWLEDGEMENTS

This work was supported by the National Research foundation of Korea (NRF) grant funded by the Korea government (MSIP) (No. 2007-00559), Gyeonggi-do, Qurient Inc. and KISTI.

Abbreviations

AUC, area under curve; Boc, *tert*-butyloxycarbonyl; CFU, colony-forming unit; EDC, *N*-(3-dimethylaminopropyl)-*N* ´-ethylcarbodiimide hydrochloride; HCS, high-content screening; HOBt, 1-hydroxybenzotriazole; INH, isoniazid; IPA, imidazo[1,2-*a*]pyridine amide; MDR, multidrug-resistant; XDR, extensively drugresistant;

REFERENCES

1. WHO. Global Tuberculosis Control WHO Report 2012; WHO/ HTM/TB/2012.6, 2012.

Stover, C.K.; Warrener, P.; VanDevanter, D.R.; Sherman, D.R.; Arain, T.M.; Langhorne, M.H.;
 Anderson, S.W.; Towell, J.A.; Yuan, Y.; McMurray, D.N.; Kreiswirth, B.N.; Barry, C.E.; Baker, W.R. A small-molecule nitroimidazopyran drug candidate for the treatment of tuberculosis. *Nature* 2000, *405*, 962–966.
 Andries, K.; Verhasselt, P.; Guillemont, J.; Göhlmann, H.W.; Neefs, J.M.; Winkler, H.; Van Gestel, J.;
 Timmerman, P.; Zhu, M.; Lee, E.; Williams, P.; de Chaffoy, D.; Huitric, E.; Hoffner, S.; Cambau, E.; Truffot-Pernot, C.; Lounis, N.; Jarlier, V. A diarylquinoline drug active on the ATP synthase of *Mycobacterium*

tuberculosis. Science **2005**, 307, 223–227.

Journal of Medicinal Chemistry

1 2 3	4. Makarov, V.; Manina, G.; Mikusova, K.; Möllmann, U.; Ryabova, O.; Saint-Joanis, B.; Dhar, N.; Pasca,
4 5 6	M.R.; Buroni, S.; Lucarelli, A.P.; Milano, A.; De Rossi, E.; Belanova, M.; Bobovska, A.; Dianiskova, P.;
7 8 9	Kordulakova, J.; Sala, C.; Fullam, E.; Schneider, P.; McKinney, J.D.; Brodin, P.; Christophe, T.; Waddell, S.;
10 11 12	Butcher, P.; Albrethsen, J.; Rosenkrands, I.; Brosch, R.; Nandi, V.; Bharath, S.; Gaonkar, S.; Shandil, R.K.;
13 14 15	Balasubramanian, V.; Balganesh, T.; Tyagi, S.; Grosset, J.; Riccardi, G.; Cole, S.T. Benzothiazinones kill
16 17 18 19	Mycobacterium tuberculosis by blocking arabinan synthesis. Science 2009, 324, 801–804.
20 21 22	5. Diacon, A.H.; Dawson, R.; Hanekom, M.; Narunsky, K.; Maritz, S.J.; Venter, A.; Donald, P.R.; van
23 24 25 26	Niekerk, C.; Whitney, K.; Rouse, D.J.; Laurenzi, M.W.; Ginsberg, A.M.; Spigelman, M.K. Early bactericidal
20 27 28	activity and pharmacokinetics of PA-824 in smear-positive tuberculosis patients. Antimicrob. Agents Chemother.
29 30 31	2010 , <i>54</i> , 3402–3407.
32 33 34 35 36	6. Diacon, A.H.; Donald, P.R.; Pym, A.; Grobusch, M.; Patientia, R.F.; Mahanyele, R.; Bantubani, N.;
37 38 39	Narasimooloo, R.; De Marez, T.; van Heeswijk, R.; Lounis, N.; Meyvisch, P.; Andries, K.; McNeeley, D.F.
40 41 42	Randomized pilot trial of eight weeks of bedaquiline (TMC207) treatment for multidrug-resistant tuberculosis:
43 44 45	long-term outcome, tolerability, and effect on emergence of drug resistance. Antimicrob. Agents Chemother.
46 47 48 49	2012 , <i>56</i> , 3271–3276.
50 51 52	7. Gler, M.T.; Skripconoka, V.; Sanchez-Garavito, E.; Xiao, H.; Cabrera-Rivero, J.L.; Vargas-Vasquez,
53 54 55 56	D.E.; Gao, M.; Awad, M.; Park, S.K.; Shim, T.S.; Suh, G.Y.; Danilovits, M.; Ogata, H.; Kurve, A.; Chang, J.;
57 58 59	Suzuki, K.; Tupasi, T.; Koh, W.J.; Seaworth, B.; Geiter, L.J.; Wells, C.D. Delamanid for multidrug-resistant
60	pulmonary tuberculosis. <i>N. Engl. J. Med.</i> 2012 , <i>366</i> , 2151–2160. ACS Paragon Plus Environment

1 2 3	8.	Pethe, K.; Bifani, P.; Jang, J.; Kang, S.; Park, S.; Ahn S.; Jiricek, J.; Jung, J.; Jeon, H.; Cechetto, J.;
4 5 6	Christo	phe, T.; Lee, H.; Kempf, M.; Jackson, M.; Lenaerts, A.J.,; Pham, H.; Jones, V.; Seo, M.J.; Kim, Y.; Seo,
7 8 9	M.; Sec	o, J.; Park, D.; Ko, Y.; Choi, I.; Kim, R.; Kim, S.; Lim, S.; Yim, S.; Nam, J.; Kang, H.; Kwon, H.; Oh,
10 11 12	C.; Cho	o, Y.; Jang, Y.; Kim, J.; Chua, A.; Tan, B.H.; Nanjundappa, M.B.; Rao, S.P.; Barnes, W.S.; Wintjens, R.;
13 14 15	Walker	, J.R.; Alonso, S.; Lee, S.; Kim, J.; Oh, S.; Oh, T.; Nehrbass, U.; Han, S-J.; No, Z.; Lee, J.; Brodin, P.;
16 17 18	Cho, S.	; Nam, K.; Kim, Jaeseung. Discovery of Q203, a potent clinical candidate for the treatment of
19 20 21	tubercu	losis. Nature Medicine 2013, 19(9), 1157-1160.
22 23 24 25	9.	Christophe, T.; Jackson, M.; Jeon, H.K.; Fenistein, D.; Contreras-Dominguez, M.; Kim, J.; Genovesio,
26 27 28	A.; Car	ralot, J.P.; Ewann, F.; Kim, E.H.; Lee, S.Y.; Kang, S.; Seo, M.J.; Park, E.J.; Skovierová, H.; Pham, H.;
29 30 31	Riccard	li, G.; Nam, J.Y.; Marsollier, L.; Kempf, M.; Joly-Guillou, M.L.; Oh, T.; Shin, W.K.; No, Z.; Nehrbass,
32 33 34	U.; Bro	sch, R.; Cole, S.T.; Brodin, P. High content screening identifies decaprenyl-phosphoribose 2' epimerase
35 36 37 38	as a tar	get for intracellular antimycobacterial inhibitors. PLoS Pathog. 2009, 5, e1000645
39 40 41 42	10.	Abrahams, K. A.; Cox, J. A. G.; Spivey, V. L.; Loman, N. J.; Patten, M.J.; Constantinidoou, C.;
43 44 45	Fernand	dex, R.; Alemparte, C.; Remuinan, M. J.; Barros, D.; Ballell, L.; Besra, G.S. Identification of novel
46 47 48 49	imidazo	o[1,2-a]pyridine inhibitors targeting <i>M. tuberculosis</i> QcrB. <i>PLoS One</i> 2012 , 7, e52951.
50 51 52 53	11.	Moraski, G. C.; Markley, L. D.; Hipskind, P. A.; Boshoff, H.; Cho, S.; Franzblau, S. G.; Miller, M. J.
54 55 56	Advent	of imidazo[1,2-a]pyridine-3carboxamides with potent multi- and extended drug resistant
57 58 59 60	antitubo	erculosis activity. ACS Med. Chem. Lett. 2011, 2, 466–470.

Journal of Medicinal Chemistry

	12.	Ollinger, J.; Bailey, M-A; Moraski, G. C.; Casey, A.; Florio, S.; Alling, T.; Miller, M. J.; Parish, T. A
	dual rea	ad-out assay to evaluate the potency of compounds active against Mycobacterium tuberculosis. PLoS
	One, 2 ()13 , 8, e60531.
0 1 2 3	13.	Mak, P. M.; Rao, S. P. S.; Tan, MP.; Lin, X.; Chyba, J.; Tay, J.; Ng, SH.;Tan,BH.;Cherian,J.;
4 5 6	Duraisv	wamy,J.; Bifani,P.;Vim,V.;Lee,B.H.; Ma, NL.; Beer, D.; Thayalan, P.; Kuhen, K.; Chatterjee, A.;
7 8 9	Supek,	F.; Glynne, R.; Zheng, J.; Boshoff, H. I.; Barr, C. E.; Dick, T.; Pethe, K.; Camacho, L. R. A high-
0 1 2	through	nput screen to identify inhibitors of ATP homeostasis in non replicating Mycobacterium tuberculosis.
3 4 5	ACS Cl	hem. Biol. 2012 , 7, 1190–1197.
6 7 8 9	14.	Tanemura, K.; Suzuki, T.; Nishida, Y.; Satsumabayashi, K.; Horaguchi, T. A mild and efficient
1 2	procedu	are for α -brominatio of ketones using N-bromosuccinimide catalysed by ammonium acetate. <i>Chem</i> .
5 4 5 6	Commu	un. 2004, <i>4</i> , 470-471.
7 8 9 0	15.	Ribeiro, I. G.; da Silva, K. C. M.; Parrini, S. C.; de Miranda, A. L. P.; Fraga, C. A. M.; Barreiro, E. J.
1 2 3	Synthes	sis and antinociceptive properties of new structurally planned imidazo[1,2-a]pyridine 3-
4 5 6 7	acylary	lhydrazone derivatives. Eur. J. Med. Chem. 1998, 33, 225-235.
7 8 9 0 1	16.	Michellys, P. Y.; D'Arrigo, J.; Grese, T. A.; Karanewsky, D. S.; Leibowitz, M. D.; Mais, D. A.; Mapes,
2 3 4	C. M.; 1	Reifel-Miller, A.; Rungta, D.; Boehm, M. F. Design, synthesis and structure-activity relationship of
5 6 7	novel R	XR-selective modulators. Bioorg. Med. Chem. Lett. 2004, 14, 1593-1598.
8 9 0		

17. Perner, R. J.; DiDomenico, S.; Koenig, J. R.; Gomtsyan, A.; Bayburt, E. K.; Schmidt, R. G.; Drizin, I.;

Zheng, G. Z.; Turner, S. C.; Jinkerson, T.; Brown, B. S.; Keddy, R. G.; Lukin, K.; McDonald, H. A.; Honore,

P.; Mikusa, J.; Marsh, K. C.; Wetter, J. M.; George, K. S.; Jarvis, M. F.; Faltynek, C. R.; Lee, C. H. In vitro

structure-activity relationship and in vivo characterization of 1-(aryl)-3-(4-(amino)benzyl)urea transient

receptor potential vanilloid 1 antagonists. J. Med. Chem. 2007, 50, 3651-3660.

18. Koul, A., Arnoult, E., Lounis, N., Guillemont, J. & Andries, K. The challenge of new drug discovery for tuberculosis. *Nature* **2011**, *469*, 483–490.









^{*a*}Reagents and conditions: (i^{*a*}) NBS, NH₄OAc (over 2eq.), ether, rt, 6h; (i^{*b*}) Br₂, CHCl₃, 0°C – rt, 20min; (ii) **2a-2d**, EtOH, reflux, overnight; (iii) LiOH, EtOH/H₂O (3:1, v/v), rt, overnight; (iv) corresponding amine, EDC, HOBt, TEA, DMF, 80°C, 2-4h





^aReagents and conditions: (i) Pd(dppf)Cl₂, Na₂CO₃, DME/H₂O (3:1, v/v), 150°C, 1h-3h; (ii) LAH,

THF, 0°C - reflux, 1h



Scheme 4. Synthetic scheme for linker modification^a



^{*a*}Reagents and conditions: (i) SOCl₂, 100 °C, 1h, then [1,1'-biphenyl]-4-amine, TEA, MC, rt, 1h; (ii) NaH, CH₃I, DMF, 0°C - rt, 1h; (iii) NaBH₄, BF₃-etherate, THF, reflux, 2h; (iv) NBS, NH₄OAc (0.1 eq.), Et₂O, rt, overnight; (v) 2-amino-5-chloropyridine, ethanol, reflux, overnight; (vi) LiOH, MeOH/H₂O (3:1, ν/ν), rt, overnight; (vii) EDC, [1,1'-biphenyl]-4-ylmethanamine, HOBt, TEA, DMF, 80°C, 3h; (viii) [1,1'-biphenyl]-4-amine, EDC, HOBt, TEA, DMF, 80°C, 3h; (viiii) DPPA, TEA, *t*-BuOH, reflux, overnight; (x) TFA, MC, rt, 1h; (xi) 2-([1,1'-biphenyl]-4-yl)acetic acid, SOCl₂, TEA, MC, rt, 1h





		Antimycobacterial activity a	gainst <i>M. tuberculosis</i> H37Rv
Compound	x	^a extracellular MIC ₈₀ (nM)	^b intracellular MIC ₈₀ (nM)
1	NH NH	45	1.39
10	0 ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	8690	970
9	O NH NH	>10000	>10000
15	N St	>10000	>10000
14	Jun N Je	>10000	>10000
11	N H	810	200
17	H N O	1670	690



			_	Antimycobacterial activity against <i>M. tuberculosis</i> H37Rv		Microsomal stability (t _{1/2} , min	
Compound	R1	R2	R3	extracellular MIC ₈₀ (nM)	intracellular MIC ₈₀ (nM)	Human	Mouse
18	Н	Ме	Н	250	20	^a ND	^a ND
19	Н	Et	Н	63	97	27.1	^a ND
20	Н	Pr	Н	1180	50	22.4	8.7
21	Н	ⁱ Pr	Н	3130	3130	31.6	^a ND
22	6-CI	Ме	Н	175	42	>120	>60
1	6-CI	Et	Н	45	1.39	22.8	19.3
23	6-CI	Et	2-Cl	43	9.3	38.9	13.1
24	6-CI	Et	4-Cl	0.9	0.45	83	>60
25	6-CI	Et	4-CN	<0.5	0.68	>120	>60
26	6-CI	Et	4-Me	0.7	0.43	30.5	40.6
27	7-CI	Et	4-Me	<0.5	1.35	25.4	23.1
28	7-CI	Et	4-Cl	1.3	1.01	>120	>60
29	7-CI	Et	4-CO ₂ H	250	217	>120	>120
30	7-CI	Et	4- ^t Bu	12	0.46	18.5	50.7
Isoniazid(IN	NH)			449	617		
Rifampicin(RIF)				26.6	180		





	A	Antimycobacteria <i>M. tuberculo</i>	l activity against sis H37Rv	Metabolic stab	ility (t _{1/2} , min)
Compound	R	extracellular MIC ₈₀ (nM)	intracellular MIC ₈₀ (nM)	Human	Mouse
31	HZ -	2000	140	>120	10.8
32	-§- \ N	2220	740	28.9	^a ND
33	·§-N	16	8.2	13.2	2.3
34	-ξ-N	35	10	2.9	1.9
35	-{−NCF3	0.8	0.47	6.5	5.1
36	-§-NCI	19	2	11.0	5.2
37	-{-N_N-	4390	360	116.6	12.1
38	-§-N_N-<	3000	140	>120	8.9
39	-§-N	25	9.4	14.9	6.3
40	S N	34	3.74	27.3	62.3
41	₹NN-	—F 5.7	0.3	>120	33.3
42	ξN	—F ⁵⁴⁰	0.66	6.8	18.4

Table 4. Activity of IPA analogues containing three ring system against *M. tuberculosis* H37Rv



		/	Antimycobacteria <i>M. tubercu</i> i	al activity against <i>losi</i> s H37Rv	Metabolic sta	bility (t _{1/2} , min)		CYP inh	nibition (IC ₅₀ , uN	N)
Compound	R1		extracellular MIC ₈₀ (nM)	intracellular MIC ₈₀ (nM)	Human	Mouse	3A4	2D6	1A2	2C9	2C19
41	7-CI		5.7	0.3	>120	33.3	>40	>40	>40	>40	>40
43	6-CI	-§-NNF	<0.5	0.3	>120	33.3	0.49	>40	>40	1.33	1.32
44	7-CI		1.8	0.11	>120	>60	>40	>40	>40	0.17	>40
45	6-CI	R2 -\$-N_N-~-F \$-N_N-~-F -\$-NF -\$-NF -\$-NCI -\$-NCI	⁻³ <0.5	0.23	>120	>60	11.54	>40	>40	0.57	>40
42	7-CI		0.54	0.66	6.8	18.4	2.07	>40	>40	0.5	0.6
46	6-CI	-g-INF	<0.5	0.36	62.6	20.3	>40	>40	>40	>40	>40
47	7-CI		1	0.46	67.3	112.0	>40	>40	>40	0.19	0.38
48	6-CI	-{-NCI	4.1	1.3	57.9	116.0	>40	>40	>40	0.26	6.77
49	7-CI		4.0	3.7	>120	>120	>100 ^a	>100	>100	0.14	0.29
50 (Q203)	6-CI	-§-NOCF	⁻³ 4.0	1.43	>120	>120	>100 ^b	>100 ^b	>100 ^b	>100 ^t	'>100 ^b

^aValues were determined by LC/MS method; ^bThe assay was performed using recombinant CYP enzymes

and analyzed by LC/MS/MS

	Pł	narmacokinetic	s (i.v.)		Ph	armacokine	tics (p.o.)	
Compd	t _{1/2} (h)	CI (mL/min/kg)	Vd _{ss} (mL/kg)	C _{max} (ng/mL)	t _{1/2} (h)	T _{max} (h)	AUC _{0-inf} (ngˈh/mL)	F (%)
41	6.15	1.9	877	4450	9.4	2.0	59576	69.9
49	62.3	3.15	14300	1987	21.3	2.0	27349	80.2
50 ^a	16.5	4.0	5270	1490	23.4	2.0	44100	90.7

Compd.	Dose (mg/kg)	CFU (Log10)/lung
49	2	6.23 ± 0.30
	10	5.72 ± 0.40
	50	5.65 ± 0.40
INH	15	5.01 ± 0.14
Untreated		7.24 ± 0.17

