

## ATTACHMENT OF CHOLESTEROL TO AMINO-LNA: SYNTHESIS AND HYBRIDIZATION PROPERTIES

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□ *Here, we present our synthesis of amino-LNA with a C6-linker and hybridization studies of these. A cholesterol moiety was attached at the end of the C6-linker. This resulted in drastic drops against DNA of the modified oligonucleotide.*

**Keywords** LNA; amino-LNA; cholesterol

Oligonucleotides modified with LNA<sup>[1]</sup> monomers (Figure 1) have demonstrated an unprecedented high affinity toward complementary DNA and RNA with increases in the melting temperature ( $T_m$ ) up to 10°C per modification. The amino-LNA (Figure 1) monomers have demonstrated similar hybridization properties.<sup>[2]</sup> The secondary amino group of amino-LNA can be regarded as a handle for the attachment of various groups. The attachment of cholesterol to miRNA knockdown probes has resulted in increased activity of those.<sup>[3]</sup> We wanted to explore the opportunity of introducing several cholesterol units to knockdown probes by utilizing the handle of amino-LNA. In order to have the effect of the cholesterol unit this was introduced to the amino-LNA via a C6 linker (Figure 1).

The known nucleoside **1**<sup>[2]</sup> (Scheme 1) was alkylated with phtalimido-hexanal<sup>[4]</sup> in the presence of NaCNBH<sub>3</sub> to nucleoside **2** in 51% yield. Nucleoside **3** was obtained by protection of the primary hydroxy group with a DMT group using DMTCl in pyridine in 56% yield. Subsequently the phtalimide group was removed by treatment with hydrazine affording nucleoside **4** in 70% yield having a primary amino group ready for functionalization. The cholesterol group was introduced by formation of amide **5** in 53% yield by a chemoselective reaction with cholesteryl chloroformate in the presence of pyridine. Amide **5** was transformed into phosphoramidite **6** using standard conditions in a yield of 42%. Key intermediate **4** was also transformed into nucleoside **7** using ethyl

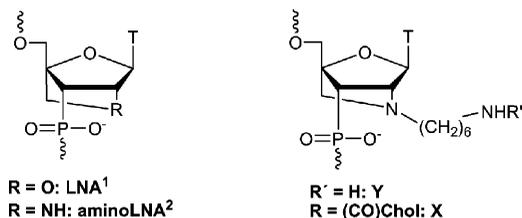
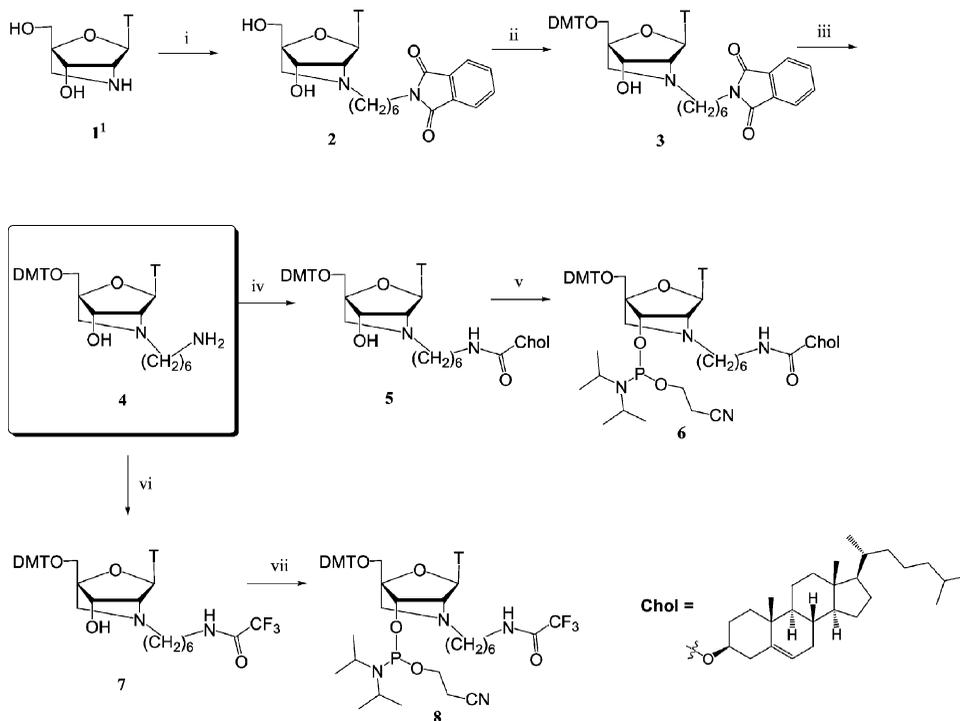


FIGURE 1 LNA and analogs.

trifluoroacetate. This was subsequently transformed into phosphoramidite **8** using the same conditions as for phosphoramidite **6**. Phosphoramidite **6** and **8** gave monomers **X** and **Y** when incorporated in oligonucleotides *vide infra*.

The synthesis of monomers **X** and **Y** was achieved. The hybridization studies of ONs modified with monomer **Y** show that this modification induced an increased towards complementary DNA resulting in  $\Delta T_m/\text{mod}$ . between 1 and 9°C (Table 1). These results are similar to those of amino-LNA. Introduction of the cholesterol group did, however, result in dramatic decreases in  $T_m$  of the modified ONs when hybridized toward



**SCHEME 1** i) 6-Phtalimido-hexanal, NaCNBH<sub>3</sub>, MeOH; ii) DMTCl, pyridine; iii) H<sub>2</sub>NNH<sub>2</sub>, EtOH, pyridine, acetic acid; iv) Cholesteryl chloroformate, CH<sub>2</sub>Cl<sub>2</sub>, pyridine; v) ((iPr)<sub>2</sub>N)<sub>2</sub>PO(CH<sub>2</sub>)<sub>2</sub>CN, DCl, CH<sub>2</sub>Cl<sub>2</sub> vi) CF<sub>3</sub>COOEt, Et<sub>3</sub>N; vii) ((iPr)<sub>2</sub>N)<sub>2</sub>PO(CH<sub>2</sub>)<sub>2</sub>CN, DCl, CH<sub>2</sub>Cl<sub>2</sub>.

**TABLE 1** Thermal denaturation temperatures measured as the maximum of the first derivative of the melting curve ( $A_{260}$  versus temperature; 5°C to 80°C with an increase of 1°C/minute) recorded in medium salt buffer (100 mM NaCl, 10 mM  $\text{NaH}_2\text{PO}_4$  0.2 mM EDTA, pH 7.0)

	5'-d(GATAGCGAAGA)	
	$T_m$ °C	$\Delta T_m/\text{mod.}$ °C
5'-d(TCTTCGCTATC)	34.2	ref.
5'-d(TCTTCGCTA <u>X</u> C)	32.6	-1.6
5'-d(TCTTCGCTA <u>Y</u> C)	38.5	+4.3
5'-d( <u>X</u> CTTCGCTATC)	30.2	-4.0
5'-d( <u>Y</u> CTTCGCTATC)	35.4	+1.2
5'-d(TC <u>X</u> TCGCTATC)	30.2	-4.0
5'-d(TC <u>Y</u> TCGCTATC)	37.6	+3.4
5'-d(TCTTCG <u>CX</u> ATC)	32.2	-2.0
5'-d(TCTTCG <u>CY</u> ATC)	43.1	+8.9
5'-d(TCT <u>X</u> CG <u>CX</u> ATC)	<10	>-12.1
5'-d(TCT <u>Y</u> CG <u>CY</u> ATC)	44.8	+5.3
5'-d( <u>X</u> CT <u>X</u> CG <u>CX</u> ATC)	<10	>-8.1
5'-d( <u>Y</u> CT <u>Y</u> CG <u>CY</u> ATC)	44.8	+3.5
5'-d( <u>X</u> CTTCGCTA <u>X</u> C)	<10	>-12,1
5'-d( <u>Y</u> CTTCGCTA <u>Y</u> C)	38.6	+2.2
5'-d( <u>X</u> CT <u>X</u> CGCTATC)	<10	>-12,1
5'-d( <u>Y</u> CT <u>Y</u> CGCTATC)	36.1	+1.0

complementary DNA. The incorporation of more than 2 **X** monomers in an 11-mer led to  $T_m$ 's lower than 10°C. This effect can be contributed to a steric effect of the cholesterol groups, having the cholesterol groups interfering with the nucleobases. We, therefore, conclude that this construct was unsuited for the use in knock-down probes.

## REFERENCES

1. Koshkin, A.A.; Singh, S.K.; Nielsen, P.; Rajwanshi, V.K.; Kumar, R.; Meldgaard, M.; Olsen, C.E.; Wengel, J. LNA (Locked Nucleic Acids): Synthesis of the adenine, cytosine, guanine, 5-methylcytosine, thymine and uracil bicyclonucleoside monomers, oligomerisation, and unprecedented nucleic acid recognition. *Tetrahedron* **1998**, 3607–3630.
2. Sorensen, M.D.; Petersen, M.; Wengel, J. Functionalized LNA (locked nucleic acid): high-affinity hybridization of oligonucleotides containing *N*-acylated and *N*-alkylated 2'-amino-LNA monomers. *Chem. Commun.* **2003**, 2130–2131.
3. Krützfeldt, J.; Rajewsky, N.; Braich, R.; Rajeev, K.G.; Tuschl, T.; Manoharan, M.; Stoffel, M. Silencing of microRNAs in vivo with 'antagomirs.' *Nature* **2005**, 438, 685–689.
4. Agathocleous, D.C.; Page, P.C.B.; Cosstick, R.; Galpin, I.J.; McLennan, A.G.; Prescott, M. Synthesis of bridged dinucleosides. *Tetrahedron* **1990**, 46 (6), 2047–2058.

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