Tetrahedron: Asymmetry 24 (2013) 699-705

Contents lists available at SciVerse ScienceDirect

Tetrahedron: Asymmetry

journal homepage: www.elsevier.com/locate/tetasy



Asymmetric synthesis of 1,2,3-trisubstituted indanes via an enantioselective copper(II)-catalyzed asymmetric nitroaldol reaction followed by an intramolecular Michael cyclization

Hongling Yuan, Junhao Hu, Yuefa Gong*

School of Chemistry and Chemical Engineering, Huazhong University of Science and Technology, 1037 Luoyu Road, Wuhan 430074, China

ARTICLE INFO

Article history: Received 21 February 2013 Accepted 7 May 2013

ABSTRACT

Herein we describe a novel approach for the synthesis of a chiral 1,2,3-trisubstituted indane, a privileged substructure in medicinal chemistry, via an enantioselective nitroaldol reaction and subsequent intramolecular Michael addition. The asymmetric copper(II)-catalyzed reactions of *ortho*-formyl cinnamates, *ortho*-formyl cinnamonitrile, or *ortho*-formyl α -benzalketones with nitromethane were carried out using the C1-symmetric chiral secondary diamine **L1** as a ligand, which afforded the nitroaldol products in high yields (up to 92%) and with good to excellent enantioselectivities (up to 97%) under mild conditions. The notable effect of the base and the dosage of nitromethane on the reaction are also discussed.

© 2013 Elsevier Ltd. All rights reserved.

1. Introduction

Indanes have found widespread application as key structural units in medicinal chemistry;¹ many natural and synthetic compounds with this skeleton have shown significant biological activities.² For example, secaloside A, which is isolated mainly from rye, exhibits potent anti-tumor activity;³ SB-209670 and SB-217242 are highly potent antagonists of endothelin receptors;⁴ indatraline [(±)-trans-3-(3,4-dichlorophenyl)-*N*-methyl-1-indanamine] is an uptake inhibitor with high affinity for dopamine, serotonin, and norepinephrine transporters.⁵ Due to the importance of these sub-

stance classes, a variety of synthetic strategies to obtain this type of compound have been developed. General synthetic methods include the reduction of indanoes,⁶ [3+2]cycloaddition of a benzyl cation with styrenes⁷ and palladium catalyzed intramolecular carboannulation reactions.⁸ These approaches often facilitate the diasteroselective synthesis of indanes, but there are only a few reports on their enantioselective synthesis.⁹ Therefore, the development of new strategies for the enantioselective synthesis of 1,2,3-trisubstituted indanes still remains an active field of research.

Recently, much effort has been devoted toward the development of catalytic asymmetric nitroaldol reactions by metal cataly-



Scheme 1. Possible reactions between 1 and nitromethane.

* Corresponding author. Fax: +86 27 8754 3632. E-mail address: gongyf@mail.hust.edu.cn (Y. Gong).



^{0957-4166/\$ -} see front matter © 2013 Elsevier Ltd. All rights reserved. http://dx.doi.org/10.1016/j.tetasy.2013.05.005

Table 1

4

5

6

7

8

9

Ligand and copper salt screening^a



a Unless otherwise stated, reactions were carried out with 1a (0.2 mmol), nitromethane (2.0 mmol, 10 equiv), ligand-Cu(II) complex (5 mol %), and DIPEA (107 µL, 0.1 equiv) for the indicated time in THF (1.0 mL) at 4 °C, then 3.0 mL of nitromethane was added and the solution was stirred continuously for another 12 h.

No additional 3.0 mL of nitromethane was added.

^c Time of nitroaldol reaction, except for entry 2.

d Isolated vield

e Determined by ¹H NMR spectroscopy of the crude products.

Enantiomeric excess was determined by HPLC analysis.

g Not determined.

sis and organocatalysis.¹⁰ Our group has also developed a series of novel chiral diamine ligands for copper-catalyzed asymmetric nitroaldol reactions.^{10a} Motivated by this work, we anticipated that the two kinds of different anions generated during the asymmetric nitroaldol reaction would undergo intramolecular Michael addition with various intramolecular electron-deficient alkenes 1 competitively to yield either oxa-heterocyclic products (path a) or carbocyclic products (path b) under the basic conditions (Scheme 1). Luzzio et al. reported on a tandem nitroaldol/oxa-Michael route to the 1,3-disubstituted-1,3-dihydrobenzo[c] furan system utilizing 1,1,3,3-tetramethylguanidine as the catalyst.¹¹ However, to the best of our knowledge, the asymmetric nitroaldol reaction followed by a carbocyclic reaction has not been reported. Herein, we report a new synthetic approach to 1,2,3-trisubstituted indanes via a highly enantioselective copper(II)-catalyzed asymmetric nitroaldol reaction followed by an intramolecular Michael cyclization.

2. Results and discussion

Our initial studies began with the copper(II)-catalyzed reaction between (E)-ethyl 3-(2-formylphenyl)acrylate 1a and nitromethane. The reaction was first performed using 5 mol % of L1 and 5 mol % of $Cu(OAc)_2 H_2O$ as the catalyst system in the presence of 0.1 equiv of diisopropylethylamine (DIPEA) in THF at 4 °C. The product analysis showed the common nitroaldol product 2a was produced predominantly rather than the expected ring-closure products 3a, and its ee value was determined by chiral HPLC (Table 1, entry 1). When an additional amount of DIPEA was added into the reaction system, the nitroaldol product 2a disappeared readily and was converted into the carbocyclic compound **3a** with three adjacent stereogenic centers. Thus, the stereochemistry of the reaction was investigated by means of chiral HPLC and ¹H NMR. Two pairs of enantiomers were clearly detected, and the ee value for the major isomer was apparently higher than that for the minor isomer. In addition, the ee value of **3a** generated under the above conditions was lower than that of **2a**, which indicates that a racemization process happened through a competitive reversible nitroaldol reaction during the intramolecular Michael reaction (entries 2 and 3). At this juncture, we found a noteworthy phenomenon that the addition of a large excess of nitromethane to the reaction mixture, instead of additional DIPEA, also led to the complete conversion of 2a into 3a. Moreover, in this case there was no detectable loss in the enantiomeric purity for the major isomer (entry 4). The major isomer can be isolated from the minor isomer by silica gel column chromatography, and their structures were identified as anti-3a and syn-3a by their spin splitting constants and NOE analysis (Scheme 2).



Scheme 2. L1-Cu(OAc)₂ catalyzed nitroaldol reaction of 1a.

Table 2Optimization of the reaction parameters^a

Entry	Solvent	Base	Time ^b (h)	Yield ^c (%)	dr ^d	ee ^e (%)
1	THF	DIPEA	24	82 3a	79:21	87/73
2	CH_3NO_2	DIPEA	6	82 3a	79:21	35/72
3	MeCN	DIPEA	24	51 3a	78:22	31/1
4	CH_2Cl_2	DIPEA	24	62 3a	74:26	39/53
5	Toluene	DIPEA	36	43 3a	81:19	59/26
6	CH₃OH	DIPEA	36	56 3a	75:25	13/10
7	Et ₂ O	DIPEA	24	82 3a	76:24	81/76
8	THF	Et ₃ N	24	81 3a	74:26	70/59
9	THF	Piperidine	30	51 3a	76:24	16/9
10	THF	NMM	36	80 2a	-	_
11	THF	Imidazole	48	nd	-	_
12	THF	DMAP	48	nd	-	_
13 ^f	THF	DIPEA	36	91 3a	83:17	91/77

^a Unless otherwise stated, reactions were carried out with **1a** (0.2 mmol), nitromethane (2 mmol, 10 equiv), **L1**–Cu(OAC)₂·H₂O complex (5 mol %), and base (0.1 equiv) in the indicated solvent (1.0 mL) at 4 °C, then 3.0 mL of nitromethane was added and the solution was stirred continuously for 6–12 h.

^b Time for the nitroaldol reaction, except for entry 2.

^c Isolated yield.

^d Determined by ¹H NMR spectroscopy of the crude products.

^e Enantiomeric excess was determined by HPLC analysis.

^f Reaction was performed at -20 °C.

Based on the above observations, we realized that the combination of an enantioselective nitroaldol reaction and subsequent Michael addition promoted by an excess amount of nitromethane was an ideal choice for the synthesis of chiral 1,2,3-trisubstituted indanes.

The catalytic activity of other chiral diamine ligands **L2–L5** was also estimated under the above reaction conditions. When ligand **L2** derived from *D*-proline was employed, the configuration-inversed enantiomer was formed with much lower enantioselectivity (entry 5). The ligands **L3** and **L4** derived from (+)-(1*S*,2*S*,5*R*)-menthylamine showed a similar change in stereoselectivity but failed to offer better results (entries 6 and 7). Ligand **L5** prepared from

Table 3

Copper(II)-catalyzed asymmetric synthesis of 1,2,3-trisubstituted indanes^a



Ia: R=H, EWG=COOC ₂ H ₅
1b: R=H, EWG=COOCH ₃
Ic: R=H, EWG=COOC4H9-n
1d: R=H_EWG=COOC4H9-t

1e: R=4-NO₂, EWG=COOC₂H₅ **1f**: R=3,4-OCH₂O, EWG=COOC₂H₅ **1g**: R=H, EWG=PhCO

1h: R=H, EWG=4-CH₃OC₆H₄CO **1i**: R=H, EWG=4-CIC₆H₄CO **1j**: R=H, EWG=CN

e ^e (%)
1/77
3/73
5/56
7/69
6/76
7/71
8/nd ^f
1/nd
0/nd
2/nd

^a Reactions were carried out with **1** (0.2 mmol), nitromethane (2 mmol, 10 equiv), **L1**-Cu(OAc)₂·H₂O complex (5 mol %), and DIPEA (107 μL, 0.1 equiv) in THF (1.0 mL) at -20 °C. and then 3.0 mL of nitromethane was added to the solution.

^b Reaction time for each of the two steps.

^c Isolated yield.

^d Determined by ¹H NMR spectroscopy of the crude products.

^e Enantiomeric excess of the diastereomers was determined by HPLC analysis.

f Not determined.

camphor amine with L-alanine also showed inferior enantioselectivities (entries 8). Among all of the ligands tested, **L1** was identified to be the best choice in terms of both the yield and enantioselectivity of the product **3a**. The effect of some representative copper salts on the reaction was also assessed in combination with **L1** in nitromethane at 4 °C (entries 9–13). Evidently, copper acetate provided the most satisfactory results among the copper salts tested, and was used in the following experiments.

The reaction conditions were then optimized utilizing this catalyst system. Further optimization was carried out by screening solvents, bases and reaction temperature. The observed data are given in Table 2. When the reaction was carried out in nitromethane, it proceeded smoothly and furnished indane **3a** in high yield, but with rather low enantioselectivity (Table 2, entry 2). Ether solvents provided better results than other types of solvents such as nitromethane, acetonitrile, dichloromethane, toluene, and methanol; among them THF proved to be the best solvent (Table 2, entries 1–7).

Next, various organic bases were assessed. Preliminary results showed that the use of DIPEA gave the highest yield and enantioselectivity of **3a** (Table 2, entries 1, 8, and 9). Meanwhile, *N*-methylmorpholine (NMM) favored the formation of nitroaldol product **2a** (entry 10), while imidazole and DMAP as the base showed worse catalytic activity, and almost no **2a** and **3a** could be isolated in each case (Table 2, entries 11 and 12). We also found that the reaction temperature had a certain influence on the reaction. At -20 °C, the reaction time was prolonged, but the ee value was elevated to 91% (entry 13). Thus, the optimal results (91% yield, 91% ee) were achieved when the reaction of **1a** with nitromethane (10 equiv) and DIPEA (0.1 equiv) was performed in THF at -20 °C in the presence of **L1**–Cu(OAc)₂·H₂O as the catalyst.

Under the optimized conditions, the substrate scope and limitations of this process were then investigated. For this purpose, various intramolecular Michael acceptors such as enones, cinnamates, cinnamonitrile, and a nitroolefin were prepared and utilized. The observed results are listed in Table 3, and the total reaction times varied from 42 to 80 h. Cinnamates **1a–f** were ideal substrates, and



Scheme 3. Determination of the absolute configuration of 2a.

their reactions proceeded readily under the optimal conditions to give the corresponding 1,2,3-trisubstituted indanes **3a–f** with high yields (up to 92%) and with good to excellent enantioselectivities (up to 97% ee), respectively (Table 3, entries 1–6). It was clear that the structure of the ester moiety had an effect on the reaction enantioselectivity (Table 3, entries 3 and 4) with cinnamate **1d**, with a bulky *t*-butyl group giving a higher ee value than substrate **1c** with an *n*-butyl group.

the enone **1g** with nitromethane did not take place in the absence of either DIPEA or **L1**–Cu(OAc)₂·H₂O complex. These results clearly indicate that the combination catalysis of a base and a diamine ligand–Cu salt complex is necessary for this transformation. In addition, the employment of nitroolefin **1k** as an intramolecular Michael acceptor did not give the desired product **3k** under the same conditions, possibly due to its ease for oligomerization under basic conditions.



The electronic properties of the substituents on the aromatic ring were also found to have an effect on the reaction rate. The reaction of 1e with an electron-withdrawing substituent, such as an -NO₂ group, was much faster than that of 1f with an electron-donating substituent such as an -OCH₂O- group (entries 5 and 6). Nonetheless, both of them gave excellent ee values despite a slight decrease in the product yield. We next turned our attention to another typical Michael acceptor enone. Relatively lower yields and ee values were observed in the case of the enones **1g-1i** (entries 7-9), which indicated that carbonyl group had a negative effect on the nitroaldol reaction, maybe due to its competitive coordinating role to the copper(II) ion. In addition, the electronic properties of the substituents on the aromatic ring of enones had an enormous influence on the reaction enantioselectivity. The reaction of enone **1h** with an electron-donating methoxy group on the benzene ring afforded good yield and ee values, whereas that of 1i with an electron-withdrawing chloro group proceeded sluggishly, giving the desired product in only 40% yield with almost no enantioselectivity (entries 8 and 9). Next, cinnamonitrile was investigated as an intramolecular Michael acceptor. As expected, the behavior of cinnamonitrile was similar to the cinnamates, and product 3i was obtained with high yield and excellent enantioselectivity (Table 3, entry10). It is noteworthy that the reaction of

We also decided to determine the absolute configuration of *anti-***3a**. Some derivatives of *anti-***3a** such as **4a** and **5a**, were thus prepared for single crystal X-ray diffraction. However, we were unable to obtain single crystals for each of the derivatives.



As a result, we tried to determine the absolute configuration of the intermediate **2a** through derivation from the known chiral source (*S*)-**7** along the routes shown in Scheme 3. As a consequence, **2a** was assigned to have an (*S*)-configuration. This result is consistent with our previous reports.¹²

3. Conclusion

In conclusion, we have developed a novel synthetic method for 1,2,3-trisubstituted indanes via the copper(II)-catalyzed asymmetric nitroaldol reaction of *ortho*-formyl cinnamates, *ortho*-formyl cinnamonitriles, and *ortho*-formyl α -benzalketones with subsequent intramolecular Michael addition under mild conditions. The reaction provided a wide variety of 1,2,3-trisubstituted indanes in good yields (up to 92%) with good to excellent enantiose-lectivities (up to 97%). Further applications of this methodology are currently underway in our laboratory.

4. Experimental

4.1. General

Solvents were purified according to standard procedures and distilled before use. Reagents and starting materials purchased from commercial suppliers were used without further purification unless otherwise stated. For thin-layer chromatography (TLC), silica gel plates GF 254 were used, and compounds were visualized by irradiation with UV light, I₂, or by treatment with basic KMnO₄. Optical rotations were measured in the solvent indicated. Flash chromatography was carried out on silica gel 200-300 mesh. NMR spectra were measured on a 400 MHz spectrometer. ¹H NMR chemical shifts are reported in ppm with tetramethylsilane (TMS, δ 0 ppm) as the internal standard. Data for ¹H NMR are reported as follows: chemical shift (ppm), and multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad). Data for ¹³C NMR are reported in ppm. High resolution mass spectroscopy analyses (HRMS) were measured using ESI ionization. High performance liquid chromatography (HPLC) analysis was performed on chiral columns.

4.2. General procedure for the enantioselective synthesis of 1,2,3-trisubstituted indanes

Ligand **L1** (0.01 mmol, 5 mol %) and Cu(OAc)₂·H₂O (2 mg, 0.01 mmol, 5 mol %) were added to a test tube containing absolute THF (1.0 mL), and the mixture was stirred for 1 h at room temperature to afford a blue solution. To the mixture the corresponding aldehyde **1a** (41 mg, 0.2 mmol) was added, and the solution was then cooled to $-20 \,^{\circ}$ C with stirring, which was followed by the addition of nitromethane (107 µL, 5 mmol, 10 equiv) and DIPEA (3.3 µL, 0.1 equiv). Stirring was continued until all of the aldehyde was fully consumed as indicated by TLC, and then 3.0 mL of nitromethane was added to the solution. The mixture was continuously stirred at $-20 \,^{\circ}$ C and monitored by TLC until **2a** was completely converted, then 7 µL of 3 M aqueous HCl was added. The solvent was removed under reduced pressure, and the residue was directly purified by column chromatography on silica gel, eluting with petroleum ether and ethyl acetate to afford the product **3a–j**.

4.3. Spectroscopic data for the products

4.3.1. (1*R*,2*S*,3*S*)-Ethyl (3-hydroxy-2-nitroindan-1-yl)acetate *anti*-3a

Colorless oil. $[\alpha]_D^{25} = -19.1$ (*c* 2.4, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.49–7.43 (m, 1H), 7.42–7.37 (m, 2H), 7.24–7.18 (m, 1H), 5.70 (d, *J* = 5.5 Hz, 1H), 5.10 (dd, *J* = 7.1, 5.6 Hz, 1H), 4.19–4.09 (m, 3H), 3.32 (s, 1H), 3.03 (dd, *J* = 16.6, 5.3 Hz, 1H), 2.92 (dd, *J* = 16.6, 6.7 Hz, 1H), 1.24 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 171.16, 139.63, 139.26, 129.77, 128.74, 124.52, 123.34, 97.70, 78.96, 61.21, 43.25, 37.42, 14.04. HRMS-ESI (*m/z*) calcd for C₁₃H₁₅NO₅ [M+Na⁺]: 288.0848, found: 288.0851. HPLC (Chiralpak

AD-H, hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min, λ = 210 nm), t_{major} = 38.8 min, t_{minor} = 44.7 min, 91% ee. *syn*-**3a**: ¹H NMR (400 MHz, CDCl₃) δ 7.49–7.42 (m, 1H), 7.42–7.34 (m, 2H), 7.27– 7.17 (m, 1H), 5.58 (d, *J* = 6.3 Hz, 1H), 5.36 (t, *J* = 6.6 Hz, 1H), 4.19– 4.08 (m, 3H), 3.34 (s, 1H), δ 2.95 (dd, *J* = 16.0, 5.3 Hz, 3H), 2.79 (dd, *J* = 16.0, 7.5 Hz, 1H),1.22 (t, 3H).

4.3.2. (1*R*,2*S*,3*S*)-Methyl (3-hydroxy-2-nitroindan-1-yl)acetate *anti*-3b

Pale yellow oil. $[\alpha]_D^{25} = -23.2$ (c 2.4, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.47–7.42 (m, 1H), 7.41–7.36 (m, 2H), 7.21–7.17 (m, 1H), 5.69 (d, *J* = 5.6 Hz, 1H), 5.07 (dd, *J* = 7.3, 5.6 Hz, 1H), 4.11 (dd, *J* = 12.8, 6.5 Hz, 1H), 3.70 (s, 3H), 3.42 (s, 1H), 3.04 (dd, *J* = 16.7, 5.4 Hz, 1H), 2.93 (dd, *J* = 16.7, 6.8 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 171.66, 139.57, 139.14, 129.80, 128.77, 97.68, 78.88, 77.36, 52.09, 43.17, 37.10. HRMS-ESI (*m/z*) calcd for C₁₂H₁₃NO₅ [M+Na⁺]: 274.0691, found: 274.0687. HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min, λ = 210 nm), *t*_{major} = 42.9 min, *t*_{minor} = 49.6 min, 93% ee. *syn*-**3b**: ¹H NMR (400 MHz, CDCl₃) δ 7.61–7.55 (m, 1H), 7.54–7.49 (m, 1H), 7.27–7.21 (m, 2H), 5.87 (d, *J* = 5.9 Hz, 1H), 5.23 (dd, *J* = 7.9, 5.9 Hz, 1H), 4.30 (dd, *J* = 15.3, 7.9 Hz, 1H), 3.71 (s, 2H), 3.51 (s, 1H), δ 3.46 (dd, *J* = 31.4, 6.5 Hz, 1H), 3.11 (dd, *J* = 33.0, 6.0 Hz, 1H).

4.3.3. (1*R*,2*S*,3*S*)-*n*-Butyl (3-hydroxy-2-nitroindan-1-yl)acetate anti-3c

Colorless oil. $[\alpha]_{D}^{25} = -23.4$ (*c* 2.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.48–7.43 (m, 1H), 7.41–7.36 (m, 2H), 7.23–7.17 (m, 1H), 5.70 (d, *J* = 5.5 Hz, 1H), 5.10 (dd, *J* = 7.1, 5.5 Hz, 1H), 4.14–4.06 (m, 3H), 3.27 (s, 1H), 3.03 (dd, *J* = 16.6, 5.4 Hz, 1H), 2.93 (dd, *J* = 16.6, 6.6 Hz, 1H), 1.59 (dt, *J* = 14.6, 6.8 Hz, 2H), 1.35 (dd, *J* = 15.1, 7.5 Hz, 2H), 0.93 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 171.23, 139.68, 139.28, 129.74, 128.71, 124.52, 123.32, 97.63, 78.91, 65.07, 43.28, 37.34, 30.45, 19.04, 13.63. HRMS-ESI (*m/z*) calcd for C₁₅H₁₉NO₅ [M+Na⁺]: 316.1161, found: 316.1155. HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min, λ = 210 nm), *t*_{major} = 50.2 min, *t*_{minor} = 64.6 min, 85% ee. *syn*-**3c**: ¹H NMR (400 MHz, CDCl₃) δ 7.50–7.46 (m, 1H), 7.31–7.28 (m, 1H), 7.24–7.20 (m, 1H), 5.82 (dd, *J* = 9.7, 2.9 Hz), 4.68 (dd, *J* = 13.3, 9.7 Hz), 4.14–4.06 (m, 3H), 3.27 (s, 1H), 3.10 (dd, *J* = 14.7, 7.4 Hz), 3.03 (dd, *J* = 10.6, 5.3 Hz), 1.62–1.51 (m, 2H), 1.30 (ddd, *J* = 21.4, 14.6, 7.3 Hz, 2H), 0.96–0.86 (m, 3H).

4.3.4. (1*R*,2*S*,3*S*)-*t*-Butyl (3-hydroxy-2-nitroindan-1-yl)acetate *anti*-3d

Colorless oil. $[\alpha]_D^{25} = -7.4$ (*c* 2.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.47–7.41 (m, 1H), 7.41–7.35 (m, 2H), 7.24–7.18 (m, 1H), 5.69 (s, 1H), 5.11 (dd, *J* = 7.1, 5.6 Hz, 1H), 4.08 (dd, *J* = 12.4, 6.4 Hz, 1H), 3.42 (s, 1H), 2.94 (dd, *J* = 16.4, 5.2 Hz, 1H), 2.84 (dd, *J* = 16.4, 6.6 Hz, 1H), 1.39 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 170.41, 139.71, 139.41, 129.64, 128.61, 124.46, 123.46, 97.70, 82.00, 78.94, 43.47, 38.58, 27.89. HRMS-ESI (*m/z*) calcd for C₁₅H₁₉NO₅ [M+Na⁺]: 316.1155, found: 316.1162. HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min, λ = 210 nm), *t*_{major} = 18.0 min, *t*_{minor} = 20.0 min, 97% ee. *syn*-**3d**: ¹H NMR (400 MHz, CDCl₃) δ 7.65–7.61 (m, 1H), 7.59–7.55 (m, 2H), 7.56–7.51 (m, 1H), δ 5.87 (s, 1H), 5.21 (dd, *J* = 13.0, 7.2 Hz, 1H), 4.28 (dd, *J* = 14.9, 7.3 Hz, 1H) 3.41 (s, 1H), 3.10 (dd, *J* = 11.0, 5.4 Hz, 1H), 3.04 (dd, *J* = 14.7, 6.2 Hz, 1H), 1.35 (s, 9H).

4.3.5. (1*R*,2*S*,3*S*)-Ethyl (3-hydroxy-2,5-dinitroindan-1-yl)acetate *anti*-3e

White solid. Mp 92.5–94.2 °C. $[\alpha]_D^{25} = -9.9$ (*c* 3.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.31 (s, 1H), 8.25 (dd, *J* = 8.4, 2.1 Hz, 1H), 7.37 (d, *J* = 8.4 Hz, 1H), 5.74 (d, *J* = 5.4 Hz, 1H), 5.20 (dd, *J* = 7.1, 5.5 Hz, 1H), 4.14 (dd, *J* = 7.1, 2.9 Hz, 3H), 3.59 (s, 1H), 3.04

(dd, *J* = 5.6, 3.7 Hz, 2H), 1.24 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 170.68, 148.71, 146.32, 141.70, 125.20, 124.27, 120.38, 96.90, 77.80, 61.55, 43.14, 36.63, 14.02. HRMS-ESI (*m*/*z*) calcd for C₁₃H₁₄N₂O₇ [M+Na⁺]: 333.0699, found: 333.0690. HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 80:20, flow rate: 0.5 mL/min, λ = 210 nm), t_{major} = 19.5 min, t_{minor} = 21.0 min, 96% ee. *syn*-**3e**: ¹H NMR (400 MHz, CDCl₃) δ 8.33 (dt, *J* = 8.4, 1.7 Hz, 1H), 8.26 (s, 1H), 7.50 (dd, *J* = 18.1, 8.4 Hz, 1H), 5.88 (ddd, *J* = 8.9, 5.9, 3.0 Hz, 1H), 5.62–5.49 (m, 1H), 4.10–3.86 (m, 3H), 3.57 (s, 1H), 3.01–2.78 (m, 2H), 1.31 (t, *J* = 7.2 Hz, 3H).

4.3.6. (1*R*,2*S*,3*S*)-Ethyl (7-hydroxy-6-nitro-6,7-dihydro-5*H*-ind-eno[5,6-*d*][1,3]dioxol-5-yl) acetate *anti*-3f

Pale yellow solid. Mp 93.6–94.0 °C. $[\alpha]_D^{25} = -69.7$ (*c* 1.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 6.86 (s, 1H), 6.63 (s, 1H), 6.00 (d, J = 1.0 Hz, 2H), 5.55 (d, J = 4.2 Hz, 1H), 5.06 (dd, J = 6.4, 4.9 Hz, 1H), 4.18–4.11 (m, 2H), 3.99 (q, J = 6.1 Hz, 1H), 3.28 (s, 1H), 2.95 (dd, J = 16.6, 5.4 Hz, 1H), 2.86 (dd, J = 16.6, 6.6 Hz, 1H), 1.26 (t, J = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 171.13, 149.43, 148.49, 132.91, 132.79, 104.65, 103.54, 101.65, 97.72, 78.68, 61.19, 43.12, 37.70, 14.03. HRMS-ESI (m/z) calcd for C14H15NO7 [M+Na⁺]: 332.0746, found: 332.0749. HPLC (Chiralpak AD-H, hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min, λ = 210 nm), t_{max} $t_{ior} = 62.9 \text{ min}, t_{minor} = 72.7 \text{ min}, 97\%$ ee. syn-**3f**: ¹H NMR (400 MHz, CDCl₃) δ 7.13 (s, 1H), 7.04 (s, 1H), 6.05 (dd, J = 5.3, 1.3 Hz, 2H), 5.84 (dd, J = 9.5, 2.7 Hz, 1H), 4.51 (dd, J = 13.6, 9.6 Hz, 1H), 4.33-4.25 (m, 2H), & 4.19-4.10 (m, 1H), 3.19 (s, 1H), & 2.95 (dd, J = 16.7, 5.3 Hz, 1H), 2.86 (dd, J = 16.7, 6.6 Hz, 1H), 1.36 (t, I = 7.1 Hz, 3H).

4.3.7. (1*R*,2*S*,3*S*)-2-(3-Hydroxy-2-nitroindan-1-yl)-1-phenylethanone *anti*-3g

Pale yellow solid. Mp 98.6–99.1 °C. $[\alpha]_D^{25} = +8.4$ (*c* 1.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.05–7.97 (m, 2H), 7.68–7.61 (m, 1H), 7.51 (dd, *J* = 15.5, 8.0 Hz, 3H), 7.37 (td, *J* = 13.1, 7.4 Hz, 2H), 7.16 (d, *J* = 7.3 Hz, 1H), 5.79 (d, *J* = 5.2 Hz, 1H), 5.06 (dd, *J* = 7.0, 5.3 Hz, 1H), 4.36 (dd, *J* = 12.5, 6.1 Hz, 1H), 3.70 (d, *J* = 5.8 Hz, 1H), 3.20 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 197.48, 140.32, 139.73, 136.27, 133.77, 129.80, 128.81, 128.56, 128.13, 124.51, 123.53, 97.94, 79.04, 42.81, 42.10. HRMS-ESI (*m/z*) calcd for C₁₇H₁₅NO₄ [M+Na⁺]: 320.0899, found: 320.0897. HPLC (Chiralpak AS-H, hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min, λ = 210 nm), *t*_{major} = 68.5 min, *t*_{minor} = 86.4 min, 58% ee.

4.3.8. (1*R*,2*S*,3*S*)-2-(3-Hydroxy-2-nitroindan-1-yl)-1-(4-meth-oxyphenyl)ethanone *anti*-3h

Pale yellow solid. Mp 99.5–101.0 °C. [α]₂₅²⁵ = +5.5 (*c* 2.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.95 (d, *J* = 8.9 Hz, 1H), 7.46 (d, *J* = 7.0 Hz, 1H), 7.38–7.30 (m, 1H), 7.14 (d, *J* = 7.1 Hz, 1H), 6.95 (d, *J* = 8.9 Hz, 1H), 5.75 (d, *J* = 4.3 Hz, 1H), 5.05 (dd, *J* = 6.8, 5.2 Hz, 1H), 4.31 (q, *J* = 6.1 Hz, 1H), 3.88 (s, 3H), 3.62 (d, *J* = 5.8 Hz, 2H), 3.29 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 195.91, 164.00, 140.55, 139.83, 130.46, 129.75, 129.39, 128.48, 124.51, 123.53, 113.94, 98.02, 79.16, 77.33, 77.22, 77.01, 55.56, 53.43, 43.05, 41.65. HRMS-ESI (*m*/*z*) calcd for C₁₈H₁₇NO₅ [M+Na⁺]: 350.1004, found: 350.1010. HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min, λ = 210 nm), t_{major} = 20.4 min, t_{minor} = 24.5 min, 81% ee.

4.3.9. (1R,2S,3S)-1-(4-Chlorophenyl)-2-(3-hydroxy-2-nitroindan-1-yl)ethanone *anti*-3i

Pale yellow solid. Mp 105.2–106.2 °C. $[\alpha]_D^{25} = +0.40$ (*c* 1.4, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.96–7.92 (m, 2H), 7.51–7.47 (m, 3H), 7.37 (dd, *J* = 7.3, 5.6 Hz, 2H), 7.15 (d, *J* = 7.3 Hz, 1H), 5.78 (d, *J* = 5.3 Hz, 1H), 5.03 (dd, *J* = 7.1, 5.4 Hz, 1H), 4.35 (dd, *J* = 13.0, 6.4 Hz, 1H), 3.66 (d, *J* = 5.9 Hz, 2H), 3.25 (S, 1H). ¹³C NMR

(100 MHz, CDCl₃) δ 196.33, 140.36, 140.08, 139.65, 134.60, 129.85, 129.55, 129.18, 128.66, 124.55, 123.48, 97.88, 78.95, 42.67, 42.10. HRMS-ESI (*m*/*z*) calcd for C₁₇H₁₄ClNO₄ [M+Na⁺]: 354.0509, found: 354.0498. HPLC (Chiralpak OD-H, hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min, λ = 210 nm), t_{major} = 36.2 min, t_{minor} = 53.0 min, 0% ee.

4.3.10. (1R,2S,3S)-(3-Hydroxy-2-nitroindan-1-yl)acetonitrile anti-3j

Pale yellow solid. Mp 94.0–95.1 °C. $[\alpha]_D^{25} = +35.54$ (*c* 1.8, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.55–7.46 (m, 3H), 7.40 (dd, *J* = 8.0, 4.7 Hz, 1H), 5.77 (d, *J* = 6.3 Hz, 1H), 4.90 (dd, *J* = 8.2, 6.4 Hz, 1H), 4.08–3.95 (m, 1H), 3.23–3.00 (m, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 139.22, 136.15, 130.22, 129.69, 124.68, 123.21, 116.66, 95.84, 77.38, 42.34, 20.88. HRMS-ESI (*m/z*) calcd for C₁₁H₁₀N₂O₃ [M+Na⁺]: 241.0589, found: 241.0579. HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min, $\lambda = 210$ nm), $t_{major} = 23.8$ min, $t_{minor} = 30.0$ min, 92% ee.

4.3.11. Procedure for the synthesis of the compound 6a

4.3.11.1. Path A: preparation of (E)-ethyl 3-(2-(1-hydroxy-2nitroethyl)phenyl)acrylate 2a. Ligand L1 $(4.8 \, \text{mg})$ 0.02 mmol, 5 mol %) and Cu(OAc)2·H2O (4 mg, 0.02 mmol, 5 mol%) were added to a test tube containing absolute THF (2.0 mL), and the mixture was stirred for 1 h at room temperature to afford a blue solution. To the mixture the corresponding aldehyde 1a (82 mg, 0.4 mmol) was added with stirring, then the solution was cooled to -20 °C, after which nitromethane (214 μ L, 4 mmol, 10 equiv) and DIPEA (3.3 µL, 0.1 equiv) were added. The stirring was continued until the aldehyde was fully consumed as indicated by TLC, then 14 µL of 3 M HCl aqueous was added, and the mixture was concentrated and directly purified by column chromatography on silica gel, eluting with petroleum ether and ethyl acetate to afford product **2a** (96 mg, 91%). ¹H NMR (400 MHz, CDCl₃) δ 8.04 (d, J = 15.7 Hz, 1H), 7.62 (d, J = 7.2 Hz, 1H), 7.55 (d, *J* = 7.7 Hz, 1H), 7.45 (td, *J* = 7.6, 1.1 Hz, 1H), 7.37 (dt, *I* = 7.5, 3.8 Hz, 1H), 6.35 (d, *I* = 15.7 Hz, 1H), 5.85 (dd, *I* = 9.6, 2.9 Hz, 1H), 4.56 (dd, /=13.3, 9.6 Hz, 1H), 4.46 (dd, /=13.3, 3.0 Hz, 1H), 4.24 (q, I = 7.1 Hz, 2H), 1.33 (t, I = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) & 166.79, 140.36, 137.25, 132.24, 130.55, 128.99, 127.26, 126.64, 121.77, 80.59, 67.61, 61.02, 14.20.

Preparation of ethyl 2-((4S)-1,2,3,4-tetrahydro-4-hydroxyisoquinolin-1-yl)acetate **6a**. At first, Zn powder (390 mg, 6 mmol) was added to a solution of 2a (0.3 mmol) in ethanol/H₂O (10:3, v/v) with rigorous stirring, and then concentrated HCl (2 mL) was added. The mixture was stirred at room temperature for 4 h. The solution was then neutralized by dilute aqueous NaOH until pH = 10, and extracted with ethyl acetate. The organic phase was dried over anhydrous Na2SO4, and evaporated under reduced pressure. The crude product was purified by column chromatography eluting with dichloromethane and methanol to afford the product **6a** as a pale yellow solid (46 mg, 65% yield). $[\alpha]_{D}^{25} = +73.3$ (c 2.0, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 7.41 (ddd, J = 8.9, 6.5, 2.9 Hz, 1H), 7.34-7.27 (m, 2H), 7.18-7.06 (m, 1H), 4.56 (dt, J = 4.8, 2.9 Hz, 1H), 4.43 (ddd, J = 11.8, 9.7, 3.0 Hz, 1H), 4.26–4.14 (m, 2H), 3.28 (td, J = 12.5, 3.0 Hz, 1H), 3.12-2.99 (m, 2H), 2.84-2.55 (m, 3H), 1.29 (dt, J = 13.3, 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 172.34, 172.11, 137.50, 137.21, 130.03, 129.40, 128.17, 127.95, 127.18, 127.00, 126.32, 125.14, 66.36, 66.02, 60.78, 60.71, 52.86, 52.12, 49.53, 46.11, 40.95, 39.91, 14.24, 14.18. HRMS-ESI (*m*/*z*) calcd for C₁₃H₁₇NO₃ [M+Na⁺]: 258.1106, found: 258.1109.

4.3.11.2. Path B: (*S*)-1-(2-Bromophenyl)-2-nitroethanol 7. Ligand L1 (12 mg, 0.05 mmol, 5 mol %) and Cu(OAc)₂·H₂O (10 mg, 0.05 mmol, 5 mol %) were added to a test tube containing absolute THF (5 mL), and the mixture was stirred for 1 h at room temperature to afford a blue solution. To the mixture the corresponding aldehyde **1** (185 mg, 1 mmol) was added with stirring, and the solution was then cooled to -20 °C, after which nitromethane (535 µL, 10 mmol, 10 equiv) and DIPEA (165 µL, 1.0 equiv) were added. After 350 µL of 3 M aqueous HCl was added, the mixture was concentrated and directly purified by column chromatography on silica gel, eluting with petroleum ether and ethyl acetate to afford product **7** as a yellow oil (234 mg, 95% yield), $[\alpha]_D^{25} = -37.8$ (*c* 2.1, CH₂Cl₂). Lit.^{10c} data: $[\alpha]_D^{25} = -27.3$ (CH₂Cl₂) 73% ee.

4.3.11.3. (S)-2-Amino-1-(2-bromophenyl)ethanol. At first, Zn powder (1.17 g, 18 mmol) was added to a solution of **7** (0.9 mmol) in ethanol/H₂O (12:3, v/v), followed by concentrated HCI (2.5 mL). The mixture was stirred at room temperature for 4 h. Then, the solution was neutralized by 5 M aqueous NaOH until the pH = 10, and extracted with ethyl acetate. The organic phase was dried over anhydrous Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by column chromatography eluting with dichloromethane and methanol to afford the product as a pale yellow solid (185 mg, 95% yield).

(S)-5-(2-Bromophenyl)-4,5-dihydro-2-methyloxaz-4.3.11.4. ole. To a stirred milky solution of ethyl acetimidate hydrochloride (113 mg, 0.9 mmol) in dry CCl₄ (2 mL) at 0 °C, a solution of (S)-2-amino-1-(2-bromophenyl)ethanol (94 mg, 0.7 mmol) in dry CCl₄ (3 ml) was added dropwise. The resulting mixture was stirred overnight at room temperature and poured into 5 mL of 2% aqueous NaHCO₃, and the organic layer was separated and the aqueous layer extracted with CH₂Cl₂. The combined organic extracts were dried over anhydrous Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by column chromatography to give 151 mg (90% yield) of a colorless oil. ¹H NMR (400 MHz, CDCl₃) & 7.59-7.50 (m, 1H), 7.35-7.30 (m, 2H), 7.15 (ddd, J = 8.4, 5.6, 3.6 Hz, 1H), 5.72 (dd, J = 10.2, 7.5 Hz, 1H), 4.39 (ddd, J = 14.3, 10.3, 1.5 Hz, 1H), 3.60 (ddd, J = 14.3, 7.5, 1.4 Hz, 1H), 2.11 (t, I = 1.4 Hz, 3H), ¹³C NMR (101 MHz, CDCl₃) δ 164.55, 140.75, 132.76, 129.17, 127.66, 125.87, 120.69, 79.62, 62.33, 13.99.

(2E)-Ethyl-3-(2-((S)-4,5-dihydro-2-methyloxazol-5-4.3.11.5. yl)phenyl)acrylate. A mixture of (S)-5-(2-bromophenyl)-4,5dihydro-2-methyloxazole (150 mg, 0.62 mmol), ethyl acrylate $(530 \,\mu\text{L})$, Pd(OAc)₂ (15 mg), PPh₃ (35 mg), and Et₃N (4 mL) was stirred at 80 °C for 16 h. The reaction was then cooled to room temperature, diluted with H₂O and extracted with CH₂Cl₂. The organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated, and the crude material was purified by flash chromatography eluting with petroleum ether and ethyl acetate to afford the coupling product as a yellow oil (78 mg, 50% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.84 (d, J = 15.7 Hz, 1H), 7.58 (d, J = 7.7 Hz, 1H), 7.45– 7.37 (m, 2H), 7.37-7.31 (m, 1H), 6.36 (d, J = 15.7 Hz, 1H), 5.81 (dd, J = 10.3, 8.1 Hz, 1H), 4.28 (q, J = 7.2 Hz, 2H), 3.64 (ddd, J = 14.1, 8.1, 1.4 Hz, 1H), 2.12 (t, J = 1.3 Hz, 3H), 1.35 (t, J = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 166.42, 164.79, 140.55, 140.00, 131.91, 130.26, 128.23, 127.10, 125.24, 121.28, 77.96, 62.80, 60.67, 14.29, 13.94.

4.3.11.6. Ethyl 2-((4S)-1,2,3,4-tetrahydro-4-hydroxyisoquinolin-1-yl)acetate 6a. To a stirred solution of the above coupling product (60 mg, 0.25 mmol) in ethanol was added 3 M HCl (84 µL, 1.0 equiv). After stirring overnight, the solvent was neutralized with 1 M NaOH aqueous until pH = 10 and extracted with ethyl acetate. The organic phase was dried over anhydrous Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by column chromatography eluting with dichloromethane and methanol to afford product **6a** as a pale yellow solid (40 mg, 70%). $[\alpha]_D^{25} = +99.2$ (*c* 2.1, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 7.41 (ddd, *J* = 8.9, 6.5, 2.9 Hz, 1H), 7.34–7.27 (m, 2H), 7.18–7.06 (m, 1H), 4.56 (dt, *J* = 4.8, 2.9 Hz, 1H), 4.43 (ddd, *J* = 11.8, 9.7, 3.0 Hz, 1H), 4.26–4.14 (m, 2H), 3.28 (td, *J* = 12.5, 3.0 Hz, 1H), 3.12–2.99 (m, 2H), 2.84–2.55 (m, 3H), 1.29 (dt, *J* = 13.3, 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 172.34, 172.11, 137.50, 137.21, 130.03, 129.40, 128.17, 127.95, 127.18, 127.00, 126.32, 125.14, 66.36, 66.02, 60.78, 60.71, 52.86, 52.12, 49.53, 46.11, 40.95, 39.91, 14.24, 14.18. HRMS-ESI (*m*/*z*) calcd for C₁₃H₁₇NO₃ [M+Na⁺]: 258.1106, found: 258.1110.

Acknowledgements

This work was supported by the National Science Foundation of China (No. 21172082). The Analysis and Testing Centre of Huazhong University of Science and Technology is acknowledged for the characterization of new compounds.

References

- (a) Biswas, A.; Sarkar, S. D.; Fröhlich, R.; Studer, A. Org. Lett. 2011, 13, 4966–4969; (b) Kirchberg, S.; Fröhlich, R.; Studer, A. Angew. Chem., Int. Ed. 2010, 49, 6877–6880; (c) Ulmschneider, S.; Müller-Vieira, U.; Klein, C. D.; Antes, I.; Lengauer, T.; Hartmann, R. W. J. Med. Chem. 2005, 48, 1563–1575; (d) Camps, P.; Formosa, X.; Galdeano, C.; Gómez, T.; Muñoz-Torrero, D.; Scarpellini, M.; Viayna, E.; Badia, A.; Clos, M. V.; Camins, A.; Pallàs, M.; Bartolini, M.; Mancini, F.; Andrisano, V.; Estelrich, J.; Lizondo, M.; Bidon-Chanal, A.; Luque, F. J. J. Med. Chem. 2008, 51, 3588–3598.
- (a) Sass, D. C.; Lucca, E. C., Jr; Barbosa, J. D. S.; Oliveria, K. T. D. *Tetrahedron Lett.* 2011, 52, 5371–5374; (b) Arnaiz, S. R.; Arnaiz, D. O. J. Org. Chem. 1992, 57, 5937–5947; (c) Kunstmann, R.; Lerch, U.; Gerhards, H.; Leven, M.; Schacht, U. J. Med. Chem. 1984, 27, 432–439; (d) Cross, B. E.; Galt, R. H. B.; Norton, K. Tetrahedron 1968, 24, 231–237.
- (a) Lantaño, B.; Aguirre, J. M.; Ugliarolo, E. A.; Torviso, R.; Pomilio, N.; Moltrasio, G. Y. *Tetrahedron* **2012**, 68, 913–921; (b) Jaun, B.; Martinoni, B.; Marazza, F.; Bertolin, M. J. Nat. Prod. **1997**, 60, 361–365.
- (a) Clark, W. M.; Tickner-Eldridge, A. M.; Huang, G. K.; Pridgen, L. N.; Olsen, M. A.; Mill, R. J.; Lantos, I.; Baine, N. H. *J. Am. Chem. Soc.* **1998**, *120*, 4550–4551; (b) Elliott, J. D.; Lago, M. A.; Cousins, R. D.; Gao, A.; Leber, J. D.; Erhard, K. F.; Nambi, P.; Elshourbagy, N. A.; Kumar, C.; Lee, J. A.; Bean, J. W.; Debrosse, C. W.; Eggleston, D. S.; Brooks, D. P.; Feuerstein, G.; Ruffolo, R. R., Jr; Weinstock, J.; Gleason, J. G.; Peishoff, C. E.; Ohlstein, E. H. *J. Med. Chem.* **1994**, *37*, 1553–1557.
- Yu, H.; Kim, I. J.; Folk, J. E.; Tian, X.; Rothman, R. B.; Baumann, M. H.; Dersch, C. M.; Flippen-Anderson, J. L.; Parrish, D.; Jacobson, A. E.; Rice, K. C. J. Med. Chem. 2004, 47, 2624–2634.
- (a) Nishiyama, Y.; Hamanaka, S.; Ogawa, A.; Kambe, N.; Sonoda, N. J. Org. Chem. 1998, 53, 1326–1329; (b) Khan, Z. A.; Iwaoka, M.; Wirth, T. Tetrahedron 2010, 66, 6639–6646.
- (a) Angle, S. R.; Arnaiz, D. O. J. Org. Chem. 1992, 57, 5937–5947; (b) Marcuzzi, F.; Melloni, G.; Modena, G. J. Org. Chem. 1979, 44, 3022–3028.
- Larock, R. C.; Song, H.; Baker, B. E.; Gong, W. H. Tetrahedron Lett. 1988, 29, 2919–2922.
- Belmessieri, D.; Morrill, L. C.; Simal, C.; Slawin, A. M. Z.; Smith, A. D. J. Am. Chem. Soc. 2011, 133, 2714–2720.
- (a) Zhou, Y.; Dong, J.; Zhang, F.; Gong, Y. J. Org. Chem. 2011, 76, 588-600; (b) Gualandi, A.; Cerisoli, L.; Toeckli-Evans, H.; Savoia, D. J. Org. Chem. 2011, 76, 3399-3408; (c) Reddy, B. V. S.; Reddy, S. M.; Manisha, S.; Madan, C. Tetrahedron: Asymmetry 2011, 22, 530-535; (d) Dhahagani, K.; Rajesh, J.; Kannan, R.; Rajagopal, G. Tetrahedron: Asymmetry 2011, 22, 857-865; (e) Wei, Y.; Yao, L.; Zhang, B.; He, W.; Zhang, S. Tetrahedron 2011, 67, 8552-8558; (f) Palomo, C.; Oiarbide, M.; Laso, A. Eur. J. Org. Chem. 2007, 2561-2574; (g) Boruwa, J.; Gogoi, N.; Saikia, P. P.; Barua, N. C. Tetrahedron: Asymmetry 2006, 17, 3315-3326; (h) Palomo, C.; Oiarbide, M.; Mielgo, A. Angew. Chem., Int. Ed. 2004, 43, 5442-5444.
- 11. Luzzio, F. A.; Okoromoba, O. E. Tetrahedron Lett. **2011**, *52*, 6530–6533.
- (a) Lu, D.; Zhou, Y.; Li, Y.; Yan, S.; Gong, Y. J. Org. Chem. 2011, 76, 8869–8878;
 (b) Gu, L.; Zhou, Y.; Zhang, J.; Gong, Y. Tetrahedron: Asymmetry 2012, 23, 124–129.