Accepted Manuscript

Piperazine-Azole-Fluoroquinolone Hybrids: Conventional and Microwave Irradiated Synthesis, Biological Activity Screening and Molecular Docking Studies

Arif Mermer, Ozlem Faiz, Ahmet Demirbas, Neslihan Demirbas, Manikandan Alagumuthu, Sivakumar Arumugam

PII: DOI: Reference:	S0045-2068(18)30841-1 https://doi.org/10.1016/j.bioorg.2019.01.009 YBIOO 2713
To appear in:	Bioorganic Chemistry
Received Date:	6 August 2018
Revised Date:	1 January 2019
Accepted Date:	6 January 2019



Please cite this article as: A. Mermer, O. Faiz, A. Demirbas, N. Demirbas, M. Alagumuthu, S. Arumugam, Piperazine-Azole-Fluoroquinolone Hybrids: Conventional and Microwave Irradiated Synthesis, Biological Activity Screening and Molecular Docking Studies, *Bioorganic Chemistry* (2019), doi: https://doi.org/10.1016/j.bioorg. 2019.01.009

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Piperazine-Azole-Fluoroquinolone Hybrids: Conventional and Microwave Irradiated Synthesis, Biological Activity Screening and Molecular Docking Studies

Arif Mermer^a, Ozlem Faiz^b, Ahmet Demirbas^a, Neslihan Demirbas^a*, Manikandan Alagumuthu^c, Sivakumar Arumugam^c

^{a)} Karadeniz Technical University, Department of Chemistry 61080 Trabzon TURKEY ^{b)} Recep Tayyip Erdogan University, Department of Chemistry, 53100 Rize TURKEY ^{c)} Dept. of Biotechnology, School of Bio-Sciences and Technology, VIT, Vellore-632014, India.

Abstract

A series of new 1,2,4-triazole and 1,3,4-oxadiazole derivatives was obtained via several steps sequential reactions of phenyl piperazine. Then, these compounds were converted to the corresponding fluoroquinolone hybrids via one pot three component Mannich reaction. All the reactions were examined under conventional and microwave mediated conditions, and optimum conditions were determined. The effect of different solvents and microwave power on microwave prompted reactions was investigated as well. All the newly synthesized compounds were characterized by FTIR, ¹H NMR, ¹³C NMR and EI MS spectral techniques. The antimicrobial activity, DNA gyrase and Topoisomerase IV inhibition potentials were performed. The results obtained showed that fluoroquinolone hybrids possess good antimicrobial activity. Moreover, Fluoroquinolone-azole-piperazine hybrids synthesized in the present study displayed excellent DNA gyrase inhibition. To unveil the interaction mode of compounds to receptor, a molecular docking study was performed. With an average least binding energy of -9.5 kcal/mol, all compounds were found to have remarkable inhibitory potentials against DNA gyrase (*E. coli*).

Keywords: Fluoroquinolone; 1,2,4-triazole; Microwave irradiation; Antimicrobial activity; DNA gyrase; Topoisomerase IV; Molecular docking.

1. Introduction

The increasing multidrug resistance among bacteria, viruses and fungi to currently used drugs has emerged an alarming and reemerging microbial threat and has become a major public health concern worldwide with nearly 15 million deaths every year. Several pathogens which appeared to be under control have once again been fatal since they have developed a variety of strategies to enhance their survival skills in the presence of antimicrobial agents.

Estimates suggested that unless significant progress has been made, drug resistant infections may result in ten million deaths annually by 2050. In order to manage drug resistance, the development of novel chemotypes which act upon novel molecular targets, has emerged as an urgent and crucial requirement. However, there have only been prepared a few molecules with new mode of action to medical use. More recently, the molecular hybridization concept based on the combination of structural units of two or more drug fragments through the fusion in one molecular framework with improved properties has been adopted as a new and attractive strategy [1-11]. The key advantage of hybrid molecules consisting of several pharmacophore groups each with different mode of action is to inhibit bacterial targets via unique binding sites or via novel modes of action or to exhibit dual mode of action [12-20]. Moreover, molecular hybrids can display better pharmacokinetic profile, therapeutic index and more importantly low tendency to resistance. Quinolones have been the centre of considerable scientific and clinical interest since their discovery in the early 1960s. This is because they potentially offer many of the attributes of an ideal antibiotic, combining high potency, a broad spectrum of activity, good bioavailability, oral and intravenous formulations, high serum levels, a large volume of distribution indicating concentration in tissues and a potentially low incidence of side-effects. Much research has attempted to make these potential attributes real. The first quinolone nalidixic acid was developed, but it took more than a decade before additional compounds, such as flumequin, norfloxacin and enoxacin became available for clinical use. The main use for all these agents was the treatment of urinary tract infection [21]. Since the discovery of norfloxacin (NFLX) by Koga et al. in the early 1980s, quinolones have become one of the most attractive agents in the chemotherapy of both community-acquired and serious hospital-acquired infections [21-23].

These drugs perform their activity by binding to two type II bacterial topoisomerase enzymes, DNA gyrase and topoisomerase IV. This causes to permanent double stranded DNA breaks, and results in cell death [24]. However, the rapidly increasing level of resistance towards quinolone class antibacterial drugs has limited their therapeutic efficacy and served the scientists to design either novel structural classes of inhibitors targeting gyrase and topoisomerase IV; or to discover new quinolone hybrids [25, 26]. During the past 30 years, hundreds of quinolone derivatives have been discovered, and a large body of structure– activity relationship (SAR) has been accumulated. The activity of quinolone class antibacterial agents arises from bicyclic heterocyclic ring and also the structure of peripheral substituents and their spatial relationship. These groups can provide additional affinity

towards bacterial enzymes, enhance the cell penetration or alter pharmacokinetic properties [27]. The *in silico* studies on quinolone class antibiotics have showed that the basic group at C-7 position is the most suitable site for chemical modifications. it constitutes an area with a great influence on potency, spectrum and safety and most of studies aiming the discovery of new quinolone hybrids have been focused on the functionalization at C-7 in quinolone skeleton [24, 28-30]. In general, 5- and 6-membered *N*-containing rings including piperazinyl, pyrrolidinyl and piperidinyl groups have been proven as optimal substituents [24, 31].

N-Containing heterocycles which have been regarded as important tools medicinally, have given a new aspect to new drug development studies. The therapeutic efficacy of 1,2,4triazole and 1,3,4-oxadiazole derivatives have been well studied in the several pathological conditions including inflammation, cancer, pain, tuberculosis or hypertension [32-35]. The compounds including these units have been reported to have several biological activities such as antimicrobial, anticancer, antioxidant, analgesic, antihypertensive etc [36-38]. Microwave irradiated syntheses have been proven as an effective technique in terms of environment, reaction time, high yields, ease of work-up and isolation of products [39]. Moreover, with the emergence of 'green chemistry concept', the scientists have shifted their focus to solvent-free methods which provide facile reactions conditions with high yields of pure products and eliminate or minimize the use of organic solvents which are often expensive, toxic and difficult to remove [40-42]. In light of these aforementioned statements, we have focused on the design, ecofriendly synthesis and antibacterial evaluation of new piperazine-azolefluoroquinolone conjugates (Fig. 1). Phenyl piperazine scaffold has been selected as the key prototype structural unit and the combination of phenyl piperazine skeleton with azole and fluoroquinolone units has been achieved by both the conventional and microwave irradiated techniques.



Figure 1. General representation of the target compounds.

2. Results and Discussion

2.1. Chemistry

The designed fluoroquinolone-triazole hybrid compounds **7a**, **b** and **8a-j** have been prepared according to the synthetic procedure presented in Scheme 1. In order to development more efficient synthetic procedures for these reactions, microwave (MW) irradiation has also been applied in addition to conventional methods. The compounds **2** and **3** have been synthesized following the procedure reported earlier [43, 44]. Initially, the carbo(thio)amide derivatives (**5a-e**) have been obtained from the nucleophilic attack of hydrazide-NH₂ to carbon atom of iso(thio)cyanate function. The evidence for the formation of compounds **5a-e** have been provided by FT IR, ¹H NMR, ¹³C NMR and EI-MS. For the synthesis of **6a-e**, two methods have been used including conventional heating under reflux conditions (Method 1) or microwave mediated conditions (Method 2). The latter method has provided more efficient procedure with much shorter reaction times varying between 15-20 min, whereas 9-15 hours have been needed for conventional method (Table 1). Moreover, MW irradiation has promoted the reaction yields from 77%-84% to 84%-90%. In method 2, higher microwave energy than 150 W has caused to a decrease in yields of compounds **6a-e**.

Compound	Micro	Microwave Irradiation			tional	Melting Point (°C)	nt
	Method			Method			
	Time	Power	Yield	Time	Yield		
	(min)	(W)	(%)	(h)	(%)		
4	15	150	92	10	84	201-203	
5a	15	150	89	10	81	231-232	
5b	15	150	86	13	77	207-208	
5c	15	150	85	11	80	203-204	
5d	15	150	90	9	84	223-224	
5e	20	150	84	15	78	202-203	
Entry		Time (min)	Power (W)	Yield (%)	S	Solvent	Cat (1(

Table 1. Time, power, yield data and melting point for compounds 4 and 5a-e

Entry	Time (min)	Power (W)	Yield (%)	Solvent	Catalyst (10%)
1	20	50	86	THF	-
2	25	70	84	THF	-
3	20	50	82	EtOH	-
4	25	70	80	EtOH	-
5	20	50	93	DMF	-
6	25	70	89	DMF	-
7	20	50	93	-	<i>p</i> -TSA
8	20	50	93	-	FeCl ₃
9	20	50	91	-	HCl



Our previous studies have showed that 1,2,4-triazoles containing a C=S (SH) group rather prefer the mercapto form, while the derivatives including C=O (OH) function are almost present in keto form [45-47]. The treatment of hydrazide (**3**) with carbon disulfide in the presence of KOH has yielded the corresponding 1,3,4-oxadiazole (**4**) under conventional heating and microwave mediated conditions. MW irradiation has decreased the reaction time from 10 h to 15 min and increased the reaction yield from 84% to 92%. The screening of the reaction condition has showed that the presence of catalysts has no significant impact on the yield of the desired compound. The best result has been obtained in DMF at 50 W maximum power without any catalyst (Table 2, Entry 5) at a yield of 93%. The remaining hybrid compounds, **7b**, **8b-j** have been obtained through applying the optimized conditions described above (Table 3). The structures of compounds **7a,b**; **8a-j** has been identified on the basis of FT IR, ¹H NMR, ¹³C NMR and mass spectrometric data.



Scheme 1. Synthetic pathway for the preparation of new compounds. i: BrCH₂COOEt, TEA, THF; ii: NH₂NH₂.H₂O, EtOH; iii: CS₂, KOH(aq), EtOH; iv: R₁NCX; v: NaOH(aq), EtOH; vi: HCHO, DMF, Ciprofloxacin or Norfloxacin

Commonwed	Microwave Irr	adiation Method	Conventional Method		
Compound -	Time (min)	Yield (%)	Time (h)	Yield (%)	
7a	20	91	24	86	
7b	20	93	24	86	
8 a	20	93	24	86	
8 b	20	89	24	82	
8c	20	91	24	85	
8d	20	94	24	88	
8e	20	96	24	86	
8f	20	90	24	81	
8g	20	95	24	89	
8h	20	92	24	82	
8i	20	89	24	79	
8j	20	88	24	81	

Table 3. The results of reaction yield and time for the synthesized compounds **7a,b** and **8a-j** in Microwave Irradiation and Conventional Method

2.2. Biological Activity

2.2.1. Antimicrobial activity

Antimicrobial activity of all compounds obtained were presented in Table 4 (Only positive results were presented). Among these, carbo(thio)amide and triazole derivatives did not show any activity against the test microorganism. The fluoroquinolone derivatives exhibited good, moderate and excellent activity almost all microorganism with the MIC values between 0.125-65 μ g/mL. Compare with the standard drug Ciprofloxacin, these compounds showed good-moderate activity except for **7b**, **8b** and **8h** which showed the same MIC values for *K*. *pneumoniae* and only **8h** showed 0.5 μ g/mL MIC value against *A. haemolyticus*. **7b** and **8b** showed excellent activity against *S. aureus* against ampicillin used as a standard drug for gram positive microorganisms. At the same time, **7b**, **8b**, **8h**, **8j** exhibited better activity against *E. faecalis* than the standard drug for Gram-negative microorganism. For *E. coli*, all compounds showed excellent activity against Gentamicin, also **7b**, **8b** and **8h** were found to have excellent activity for *K. pneumoniae* with the MIC value 0.25 μ g/mL. On the other hand, excellent activity for *K. pneumoniae* with the MIC value 0.25 μ g/mL. On the other hand, by the MIC value of *K. pneumoniae* with the MIC value 0.25 μ g/mL.

antimicrobial activity against test microorganisms when the results were evaluated collectively.

Compound	Microorganisms and minimal inhibition concentration (µg/mL)					
	Gram-positive Gram-negative microorganism					nism
	microo	rganism				
	Sa	Ef	Ec	Ра	Кр	Ah
7a	16	32	0.5	8	4	16
7b	2	8	0.125	2	0.25	1
8 a	16	64	2	16	4	-
8b	2	8	0,125	2	0.25	1
8c	16	32	0.5	16	2	-
8d	8	16	0.125	4	0.5	4
8e	16	32	1	16	2	16
8f	8	16	0.125	4	0.5	2
8g	16	32	0.5	8	1	8
8h	4	8	0.125	2	0.25	0.5
8i	16	32	0.5	16	2	16
8j	8	8	0.125	4	2	4
Cip.	1	0.5	0.03	0.25	0.25	0.25
Amp.	1.56	12.5		-	-	-
Gen.	-	-	0.78	1.56	0.39	0.78

Compound Microorganisms and minimal inhibition concentration (µg/mL)	Table 4. Scre	eening for the activity of newly synthesized compounds 7a,b and 8a-j.
	Compound	Microorganisms and minimal inhibition concentration ($\mu g/mL$)

Sa:Staphylococcus aureus ATCC 25923, Ef:Enterococcus faecalis ATCC 29212, Ec:Escherichia coli ATCC 25922, Pa: Pseudomonas aeruginosa ATCC 27853, Kp:Klebsiella pneumoniae ATCC 13883, Ah:Acinetobacter haemolyticus ATCC 19002. Cip.: Ciprofloxacin, Amp.: Amphicilin, Gen.: Gentamicin

2.2.2. Results of DNA gyrase and Topo-IV inhibition studies

DNA gyrase and Topoisomerase IV are type-II topoisomerases enzymes, involving in the negative supercoiling which is a vital part of bacterial survival. Thus, we have assessed the inhibition potentials of present study compounds against these two enzymes. The results of DNA gyrase and Topo-IV inhibition study is depicted in Table 6. As per our observations, the activity has been totally a dose dependent (Fig. 2a). Compounds **5a** to **6e** have been found totally inactive for the activity. On the other hand, fluoroquinolone-azole-piperazine hybrids have showed excellent activity with lowest IC₅₀ values. Remarkably, compounds **8c**, **8f**, **8i**, **8j**, **7a** and **7b** were the excellent group compounds for DNA gyrase inhibition (Fig. 2b). At this point we have recognized the consistence between the molecular docking and the enzyme inhibition study in which the result has favored DNA gyrase. Meanwhile, the low selective TOPO-IV inhibition activity was also recognized from the obtained IC₅₀ values. Among all compounds, **8j** was exhibited most potent activity with a calculated relative % activity of 85 % and IC₅₀ of 0.75 μ M. A maximum activity was found at the 10⁻⁶ concertation, which indicates the necessity of low amount of target candidate drug to have a maximum activity.

The activity variation and druggability suggestions against the present study compound was assessed by comparing the DNA gyrase and Topo-IV inhibition study results of the compounds with the standard drug, Novobiocin's (Novo) results. Novo exhibited a significant DNA gyrase as well as TOPO-IV activities. A relative % activity of 82.52 % and an IC₅₀ value of 1.0 μ M was showed by Novo against DNA gyrase inhibition. In figure 2a&b, the results of Novo and potentially identified compound **8j** were compared. In the dose response curve, even though the curve elevation started equally and slopped at same concentrations, a slight variation could be seen between **8j** and Novo. This activity equality or slightly elevated activity of **8j** indicate its druggability potentials as a future DNA gyrase inhibitor.

Comp. No.	IC50 (µM) DNA Gyrase	IC ₅₀ (µМ) DNA Торо-IV
4	10.12 ± 0.14	> 25.00
5a	> 25.00	> 25.00
5b	> 25.00	> 25.00
5c	> 25.00	> 25.00
5d	> 25.00	> 25.00
5e	> 25.00	> 25.00
6a	> 25.00	> 25.00
6b	> 25.00	> 25.00
6c	> 25.00	> 25.00
6d	> 25.00	> 25.00
6e	> 25.00	> 25.00
7a	2.25 ± 0.28	15.24 ± 1.05
7b	3.25 ± 0.34	12.48 ± 1.25
8a	5.24 ± 0.88	> 25.00
8b	4.88 ± 0.76	> 25.00
8c	1.25 ± 0.22	14.36 ± 1.45
8d	2.05 ± 0.38	15.78 ± 1.57
8 e	1.84 ± 0.15	> 25.00
8 f	1.00 ± 0.18	> 25.00
8g	0.88 ± 0.18	16.46 ± 1.74
8h	1.50 ± 0.12	18.67 ± 1.68
8 i	0.95 ± 0.08	6.78 ± 1.08
8 j	0.75 ± 0.07	8.24 ± 1.64

Table 5. Results of DNA gyrase and DNA Topo-IV inhibition studies.



Figure 2a. Dose response curve of 8j against DNA gyrase inhibition. Figure 2b. DNA gyrase and TOPO-IV inhibition potential of compounds 7a, 7b and 8a-j. *Novobiocin.

2.2.3. Results of molecular docking studies

From the molecular docking study results, the attained free energy value reveals that the compounds **8a-j** having a fortunate selectivity towards DNA gyrase over TOPO-IV (ParC, PDB ID: 1ZVU) (Fig. 3). Based on the molecular mechanistic values, we have categorized the compounds as most active, intermediates and inactive groups. Mostly, the first category was used to evaluate *in vitro* experiments. The most significant binding affinity of the docked compounds was recognized through the obtained inhibitory constant (*ki*). Compounds **7a,b** and **8a-j** fortunately have established with a significant binding affinity (*ki* values < $1 \pm 0.5 \mu$ M). The average and sum of all docking molecular mechanics values have given a clear-cut idea about the activity selectivity of compounds towards the DNA gyrase inhibition (average least binding energy value of -11.214 kcal/mol). Meanwhile, TOPO-IV activity average was also reliable (-8.24 kcal/mol). While discussing about the ligand efficiency, with a significant average value of -0.32, fluoroquinolone-1,2,4-triazole hybrids were found to have efficiency towards 2XCT (DNA gyrase). The average inhibitory constant (*ki*) indicates the possible bioactivity potentials of the compounds in low molecular concentrations towards both DNA gyrase (0.0035 μ M) as well as the TOPO-IV (0.5 μ M).



Figure 3. Results of molecular docking studies. 3A. Ligand efficiency values; 3B. Binding energy values; 3C. Obtained inhibitory constant (*ki*) value for PDB ID: 2XCT; 3D. Obtained inhibitory constant (*ki*) value for PDB ID: 1ZVU

2.2.4. Molecular assessments of Receptor-Ligand interactions of DNA gyrase and TOPO-IV and compounds 7a, 7b and 8a-j

To unveil the mechanism of action and molecular interaction by means of hydrogen bonding and non-covalent bonding (π - π interaction and π -cation interactions), the established best docked poses have been analyzed. Various drug efficacy factors have been contributing in these types' communications/interactions between drug and receptor. These factors are mainly controlling the strength, duration, and different types of the drug-receptor interaction. In fact, these factors are decisively dictating the strength of the interaction established by a drug (by constituting the complex) against its receptor. This has been known as the affinity. Therefore, the binding affinity is considered as one of the chief in silico measurement to adjudge on new chemical entity's drug efficacy.

Various intermolecular forces are involved in the interaction establishment between drug and binding sites such as Van der Waals forces (weak bonds and transient reversible effects), Hydrogen bonds (intermediate bonds and transient reversible effects), Covalent bonds (strong bonds and long-lasting or irreversible effects) and non-covalent bonding (electromagnetic interactions among molecules). Fig. 4a-b has illustrated the interaction of top scored

compounds to DNA gyrase (PDB ID: 2XCT) and TOPO-IV (PDB ID: 1ZVU). Ser1173 was the main amino acid residue to establish hydrogen bond and Arg1033to establish the non-covalent cation- π interaction DNA gyrase (PDB ID: 2XCT). Docking results were also suggested to evaluate further possible *in vitro* and *in vivo* analysis to unveil the drug ability of the present study compounds.



Figure 4A. Interaction of compound **8a** to DNA Gyrase (PDB: 2XCT). **Figure 4B**. Interaction of compound **8a** to DNA Gyrase (PDB: 2XCT) binding pocket.

3. Experimental

3.1. General

All the chemicals were purchased from Fluka Chemie AG Buchs and Sigma Aldrich and used without further purification. Melting points of the synthesized compounds were determined in open capillaries on a Büchi B-540 melting point apparatus and are uncorrected. Reactions were monitored by thin-layer chromatography (TLC) on silica gel 60 F254 aluminium sheets. The mobile phase was ethyl acetate: diethyl ether (1:1), and detection was made using UV light. FT-IR spectra were recorded using a Perkin Elmer 1600 series FTIR spectrometer. ¹H NMR and ¹³C NMR spectra were registered in DMSO- d_6 on a BRUKER AVENE II 400 MHz NMR spectrometer (400.13 MHz for ¹H and 100.62 MHz for ¹³C). The chemical shifts are given in ppm relative to Me₄Si as an internal reference, *J* values are given in Hz. The mass spectra were obtained on a Quattro LC-MS (70 eV) Instrument. Microwave irradiated reactions were performed in a CEM Discovery monomod synthesis reactor.

3.1.1. 5-[(4-Phenylpiperazin-1-yl)methyl]-1,3,4-oxadiazole-2-thiol (4)

Method 1. Compound **3** (10 mmol) and CS_2 (15 mmol) were added to a solution of KOH (10 mmol) in 10 mL H₂O and 10 mL ethanol and the reaction mixture was refluxed for 10 h.

Then, the reaction content was acidified with conc. HCl to pH 6. The precipitate formed was filtered off, washed with H_2O and recrystallized from ethanol to afford the desired compound. Yield: 84%.

Method 2. The solution of compound **3** (10 mmol), CS_2 (15 mmol) and KOH (10 mmol) in H_2O +ethanol was irradiated in monomod microwave reactor in closed vessel with the pressure control at 100 °C, 150 W maximum power for 15 min (hold time). After the completion of the reaction, (monitored by TLC), the mixture was acidified with conc. HCl to pH 6. The precipitate formed was filtered off, washed with H_2O and recrystallized from ethanol to afford the desired compound. Yield: 92 %.

FT-IR (υ_{max} , cm⁻¹): 3011 (ar-CH), 2855 (-SH), 1602 (C=N). ¹H NMR (DMSO- d_6 , δ ppm): 2.65 (d, 4H, *J*=4.0 Hz, 2CH₂), 3.15 (s, 4H, 2CH₂), 3.74 (s, 2H, CH₂), 6.79 (t, 1H, *J*=12.0 Hz, arH), 6.93 (d, 2H, *J*=8.0 Hz, arH), 7.21 (t, 2H, *J*=12.0 Hz, arH), 14.42 (s, 1H, SH). ¹³C NMR (DMSO- d_6 , δ ppm): 48.52 (2CH₂), 51.56 (CH₂), 52.42 (2CH₂), arC: [116.00 (2CH), 119.45 (CH), 129.37 (2CH), 151.30 (C)], 161.09 (oxadiazole C-5), 178.49 (oxadiazole C-2). EI MS m/z (%): 106.04 (44), 163.08 (33), 175.16 (49), 277.23 ([M+1]⁺, 100), 315.22 ([M+K]⁺, 40).

3.1.2. General method for the synthesis of compounds 5a-e

A mixture of compound **3** (10 mmol) and the corresponding iso(thio)cyanate (15 mmol) in dichloromethane was stirred at room temperature for 24 h (monitored by TLC). After evaporating the solvent under reduced pressure, a solid obtained. The crude product was purified by crystallization from appropriate solvents.

3.1.2.1. *N*-Phenyl-2-[(4-phenylpiperazin-1-yl)acetyl]hydrazincarbothioamide (5a)

Recrystallized from acetone. Yield: 95 %, Mp. 184-185 °C. FT-IR (υ_{max} , cm⁻¹): 3306, 3273 ve 3210 (3NH), 3112 (ar-CH), 1665 (C=O), 1235 (C=S). ¹H NMR (DMSO-*d*₆, δ ppm): 2.69 (s, 4H, 2CH₂), 3.18 (s, 6H, 3CH₂), 6.78 (t, 1H, *J*=16.0 Hz, arH), 7.17-7.24 (m, 3H, arH), 7.35 (t, 2H, *J*=16.0 Hz, arH), 7.45 (d, 2H, *J*=5.0 Hz, arH), 9.59 (bs, 2H, 2NH), 9.92 (bs, 1H, NH). ¹³C NMR (DMSO-*d*₆, δ ppm): 46.71 (2CH₂), 52.71 (2CH₂), 59.95 (CH₂), arC: [113.11 (2CH), 115.15 (CH), 124.19 (2CH), 126.19 (3CH), 129.83 (2CH), 135.90 (C), 137.31 (C)], 155.15 (C=O), 181.69 (C=S). EI MS *m*/*z* (%): 173.27 (96), 208.31 (52), 220.26 (49), 294.40 (49), 369.44 ([M]⁺, 100).

3.1.2.2. N-Benzyl-2-[(4-phenylpiperazin-1-yl)acetyl]hydrazincarbothioamide (5b)

Recrystallized from acetone. Yield: 90 %, Mp. 193-194 °C. FT-IR (υ_{max} , cm⁻¹): 3204 (3NH), 3052 (ar-CH), 1671 (C=O), 1233 (C=S). ¹H NMR (DMSO- d_6 , δ ppm): 2.64 (s, 4H, 2CH₂), 3.15 (s, 2H, CH₂), 3.38 (s, 2H, CH₂ + su), 3.70 (s, 4H, 2CH₂), 7.02 (d, 2H, *J*= 8.0 Hz, arH), 7.06-7.43 (m, 5H, arH), 8.05 (d, 2H, *J*=8.0 Hz, arH), 9.59 (bs, 2H, 2NH), 9.94 (bs, 1H, NH). ¹³C NMR (DMSO- d_6 , δ ppm): 46.69 (CH₂), 51.82 (2CH₂), 54.10 (2CH₂), 59.93 (CH₂), arC: [113.13 (d, *J*=7.0 Hz, 2CH), 126.16 (d, *J*=5.0 Hz, 2CH), 127.47 (CH), 129.13 (CH), 129.57 (d, *J*=5.0 Hz, 2CH), 130.55 (2CH), 133.46 (C), 155.08 (C)], 169.52 (C=O), 192.95 (C=S). EI MS *m*/*z* (%): 106 (100), 135 (42), 320 (53), 331 (48), 384 ([M+1]⁺, 48).

3.1.2.3. *N*-(4-Fluorophenyl)-2-[(4-phenylpiperazin-1-yl)acetyl]hydrazin carbothioamide (5c)

Recrystallized from acetone. Yield: 92 %, Mp. 188-190 °C. FT-IR (v_{max} , cm⁻¹): 3184 (3NH), 3032 (ar-CH), 1669 (C=O), 1226 (C=S). ¹H NMR (DMSO- d_6 , δ ppm): 2.66 (s, 4H, 2CH₂), 3.16 (d, 4H, *J*=16.0 Hz, 2CH₂), 3.36 (s, H₂O+CH₂), 6.78 (t, 1H, *J*=12.0 Hz, arH), 6.94 (d, 2H, *J*=8.0 Hz, arH), 7.16-7.23 (m, 4H, arH), 7.41 (s, 2H, arH), 9.63 (bs, 2H, 2NH), 9.90 (bs, 1H, NH). ¹³C NMR (DMSO- d_6 , δ ppm): 48.48 (2CH₂), 53.24 (2CH₂), 60.28 (CH₂), arC: [115.24 (d, *J*=22.0 Hz, 2CH), 115.83 (2CH), 119.23 (2CH), 123.99 (CH), 129.39 (2CH), 135.95 (C), 151.45 (C), 152.02-154.50 (d_{C-F}, *J*=248.0 Hz, C)], 169.23 (C=O), 181.60 (C=S). EI MS *m/z* (%): 106 (93), 388 ([M+1]⁺, 100), 389 ([M+2]⁺, 86).

3.1.2.4. N-(4-Ethyl)-2-[(4-phenylpiperazin-1-yl)acetyl]hydrazincarbothioamide (5d)

Recrystallized from acetone. Yield: 90 %, Mp. 175 °C. FT-IR (υ_{max} , cm⁻¹): 3232 (3NH), 3063 (ar-CH), 1670 (C=O), 1234 (C=S). ¹H NMR (DMSO- d_6 , δ ppm): 1.08 (t, 3H, *J*=16.0 Hz, CH₃), 2.67 (s, 4H, 2CH₂), 3.13 (s, 2H, 2CH₂), 3.18 (s, 4H, 2CH₂), 3.48 (s, 2H, CH₂), 6.78 (t, 1H, *J*=16.0 Hz, arH), 6.94 (d, 2H, *J*=8.0 Hz, arH), 7.22 (t, 2H, *J*=16.0 Hz, arH), 7.82 (bs, 1H, NH), 9.12 (bs, 1H, NH), 9.65 (bs, 1H, NH). ¹³C NMR (DMSO- d_6 , δ ppm): 14.92 (CH₃), 38.91 (CH₂), 48.37 (2CH₂), 53.19 (2CH₂), 59.85 (CH₂), arC: [115.84 (2CH), 119.26 (CH), 129.39 (2CH), 151.40 (C)], 176.89 (C=O), 190.63 (C=S). EI MS *m/z* (%): 322 ([M+1]⁺, 100), 323 ([M+2]⁺, 85).

3.1.2.5. N-Benzyl-2-[(4-phenylpiperazin-1-yl)acetyl]hydrazincarboxamide (5e)

Recrystallized from acetone. Yield: 86 %, Mp. 211-212 °C. FT-IR (v_{max} , cm⁻¹): 3380 ve 3304 (3NH), 3037 (ar-CH), 1713 (C=O), 1663 (C=O). ¹H NMR (DMSO-*d*₆, δ ppm): 2.62 (s, 4H,

2CH₂), 3.09 (s, 4H, 2CH₂), 3.48 (bs, H₂O+2CH₂), 4.25 (s, 2H, CH₂), 7.01 (s, 3H, arH), 7.28 (s, 7H, arH), 7.91 (s, 1H, NH), 8.05 (s, 1H, NH), 9.53 (bs, 1H, NH). ¹³C NMR (DMSO- d_6 , δ ppm): 43.14 (CH₂), 46.68 (2CH₂), 52.64 (2CH₂), 59.89 (CH₂), arC: [113.05 (2CH), 126.18 (CH), 126.94 (CH), 127.02 (2CH), 127.41 (CH), 127.48 (CH), 128.53 (CH), 128.59 (CH), 137.28 (C), 155.12 (C)], 158.65 (C=O), 169.53 (C=O). EI MS *m*/*z* (%): 113.02 (60), 368.36 ([M+1]⁺, 100).

3.1.3. General method for the synthesis of compounds 6a-e

Method 1. A solution of corresponding carbo(thio)amide **5a-e** (10 mmol) in ethanol/water (1:1) was refluxed in the presence of 2 N NaOH for 10 h (for **6a**), 13 h (for **6b**), 11 h (for **6c**), 9h (for **6d**) and 15 h (for 6e) (the progress of the reaction was monitored by TLC). Then, the resulting solution was cooled to room temperature and acidified to pH 5 with 37% HCl. The precipitate formed was filtered off, washed with water, and recrystallized from an appropriate solvent to give the target compound.

Method 2. The mixture of the corresponding compound **5** (10 mmol) and 2 N NaOH in ethanol (10 mL) was irradiated in monomod microwave reactor in closed vessel (physical parameters, reaction time and yields were given in Table 4). Then the resulting solution was cooled to room temperature and acidified to pH 5 with 37% HCl. The precipitate formed was filtered off, wash with water, and recrystallized from an appropriate solvent to give the target compound.

3.1.3.1. 4-Phenyl-5-[(4-phenylpiperazin-1-yl)methyl]-4H-1,2,4-triazol-3-thiol (6a)

Recrystallized from ethanol. FT-IR (v_{max} , cm⁻¹): 3095 (ar-CH), 2759 (-SH), 1598 (C=N). ¹H NMR (DMSO- d_6 , δ ppm): 2.50 (t, 4H, J=5.0 Hz, 2CH₂), 3.19 (s, 4H, 2CH₂), 3.40 (s, 2H, CH₂), 7.04 (d, 2H, J=8.0 Hz, arH), 7.43 (t, 4H, J=12.0 Hz, arH), 7.66-7.70 (m, 2H, arH), 8.06 (d, 2H, J=8.0 Hz, arH), 13.42 (s, 1H, SH). ¹³C NMR (DMSO- d_6 , δ ppm): 45.54 (CH₂), 48.59 (2CH₂), 51.45 (d, J=22.0 Hz, 2CH₂), arC: [116.76 (2CH), 126.53 (CH), 128.39 (2CH), 129.61 (CH), 130.25 (2CH), 130.87 (CH), 134.08 (CH), 148.40 (C), 151.57 (C)], 150.20 (triazol C-5), 169.61 (triazol C-3). EI MS *m*/*z* (%): 205.16 (100), 265.35 (42), 307.30 (35), 352.33 ([M+1]⁺, 92), 374.28 ([M+Na]⁺, 30).

3.1.3.2 4-Benzyl-5-[(4-phenylpiperazin-1-yl)methyl]-4H-1,2,4-triazol-3-thiol (6b)

Recrystallized from ethanol. FT-IR (υ_{max} , cm⁻¹): 3090 (ar-CH), 2751 (-SH), 1597 (C=N). ¹H NMR (DMSO- d_6 , δ ppm): 2.38 (s, 4H, 2CH₂), 3.27 (s, 4H, 2CH₂), 3.54 (s, 2H, CH₂), 4.89 (s, 2H, CH₂), 6.96 (s, 1H, arH), 7.53 (s, 2H, arH), 7.58 (s, 4H, arH), 8.02 (s, 3H, arH), 13.89 (s, 1H, SH). ¹³C NMR (DMSO- d_6 , δ ppm): 46.67 (CH₂), 51.42 (CH₂), 51.88 (CH₂), 51.97 (2CH₂), arC: [117.72 (2CH), 126.13 (2CH), 127.61 (CH), 129.39 (2CH), 130.02 (CH), 130.78 (2CH), 139.14 (C), 148.40 (C), 150.79 (C)], 153.05 (triazol C-5), 174.54 (triazol C-3). EI MS m/z (%): 134.98 (44), 208.18 (50), 288.46 (45), 388.32 ([M+Na]⁺, 100).

3.1.3.3. 4-(4-Fluorophenyl)-5-[(4-phenylpiperazin-1-yl)methyl]-4*H*-1,2,4-triazol-3-thiol (6c)

Recrystallized from ethylacetate. FT-IR (ν_{max} , cm⁻¹): 3087 (ar-CH), 2830 (-SH), 1589 (C=N). ¹H NMR (DMSO- d_6 , δ ppm): 2.37 (t, 4H, *J*=12.0 Hz, 2CH₂), 3.28 (t, 4H, *J*=12.0 Hz, 2CH₂), 3.42 (s, 2H, CH₂), 6.97 (d, 2H, *J*=8.0 Hz, arH), 7.36 (s, 1H, arH), 7.40 (d, 2H, *J*=8.0 Hz, arH), 7.55 (q, 2H, *J*=16.0 Hz, arH), 8.03 (d, 2H, *J*=8.0 Hz, arH), 13.90 (s, 1H, SH). ¹³C NMR (DMSO- d_6 , δ ppm): 46.66 (CH₂), 50.58 (2CH₂), 51.89 (2CH₂), arC: [113.13 (2CH), 116.28 (d, *J*=23.0 Hz, 2CH), 121.22 (CH), 126.13 (2CH), 131.17 (d, *J*=9.0 Hz, 2CH), 137.41 (C), 149.59 (C), 161.29-163.74 (d_{C-F}, *J*=245.0 Hz, C)], 155.05 (triazol C-5), 168.95 (triazol C-3). EI MS *m*/*z* (%): 175 (100), 285 (64), 301 (63), 315 (62), 369 ([M]⁺, 46).

3.1.3.4. 4-Ethyl-5-[(4-phenylpiperazin-1-yl)methyl]-4H-1,2,4-triazol-3-thiol (6d)

Recrystallized from ethanol. FT-IR (ν_{max} , cm⁻¹): 3089 (ar-CH), 2762 (-SH), 1597 (C=N). ¹H NMR (DMSO-*d*₆, δ ppm): 1.30 (t, 3H, *J*=16.0 Hz, CH₃), 2.58 (s, 4H, 2CH₂), 3.12 (t, 4H, *J*=12.0 Hz, 2CH₂), 3.64 (s, 2H, CH₂), 4.04 (q, 2H, *J*=8.0 Hz, CH₂), 6.78 (t, 1H, *J*=16.0 Hz, arH), 6.93 (d, 2H, *J*=8.0 Hz, arH), 7.21 (q, 2H, *J*=8.0 Hz, arH), 13.63 (s, 1H, SH). ¹³C NMR (DMSO-*d*₆, δ ppm): 13.72 (CH₃), 39.26 (CH₂), 48.69 (2CH₂), 52.33 (CH₂), 52.85 (2CH₂), arC: [115.97 (2CH), 119.44 (CH), 129.38 (2CH), 149.47 (C)], 151.36 (triazol C-5), 167.36 (triazol C-3). EI MS *m/z* (%): 106 (100), 213 (41), 304 ([M+1]⁺, 24).

3.1.3.5. 4-Benzyl-5-[(4-phenylpiperazin-1-yl)methyl]-1*H*-1,2,4-triazol-5-one (6e)

Recrystallized from acetone. FT-IR (υ_{max} , cm⁻¹): 3083 (ar-CH), 1578 (C=N). ¹H NMR (DMSO- d_6 , δ ppm): 2.50 (t, 4H, J=4.0 Hz, 2CH₂), 3.32 (s, 2H, CH₂), 3.36 (s, H₂O+CH₂), 4.89 (s, 2H, CH₂), 6.97 (d, 2H, J=8.0 Hz, arH), 7.26 (d, 3H, J=8.0 Hz, arH), 7.31 (s, 1H, arH), 7.34 (d, 2H, J=8.0 Hz, arH), 8.04 (d, 2H, J=12.0 Hz, arH), 11.75 (s, 1H, NH). ¹³C NMR (DMSO-

*d*₆, δ ppm): 44.25 (CH₂), 46.41 (2CH₂), 52.13 (2CH₂), 53.28 (CH₂), arC: [113.00 (2CH), 121.38 (CH), 126.13 (2CH), 127.48 (CH), 128.50 (2CH), 128.87 (2CH), 137.40 (C), 144.65 (C)], 154.96 (triazol C-3), 156.06 (triazol C-5). EI MS *m/z* (%): 168.93 (68), 205.22 (42), 372.33 ([M+Na]⁺, 100).

3.1.4. General method for the synthesis of compounds 7a,b and 8a-j

Method 1. The solution of ciprofloxacin (for 7b, 8b, 8d, 8f, 8h, 8j) or norfloxacin (for 7a, 8a, 8c, 8e, 8g, 8i), (10 mmol) in dimethyl formamide was stirred at room temperature in the presence of formaldehyde (37%, 30 mmol) for 15 min. Then, the corresponding compound 4 or 6 (10 mmol) was added into it and stirred for additional 24 h. The reaction mixture was poured to ice-water and a solid obtained. This crude product was recrystallized from an appropriate solvent to give the desired compound.

Method 2. A mixture of ciprofloxacin or norfloxacin (10 mmol), the corresponding compound **7** or **8** (10 mmol) and formaldehyde (37%, 30 mmol) in DMF was irradiated in a monomod microwave reactor in a closed vessel under pressure control at 50 W for 20 min. The solid obtained was purified by recrystallization from an appropriate solvent.

3.1.4.1. 1-Ethyl-6-fluoro-4-oxo-7-(4-{[5-[(4-phenylpiperazin-1-yl)methyl]-2-tioxo-1,3,4oxadiazole-3(2*H*)-yl]methyl}piperazin-1-yl)-1,4-dihydroquinolin-3-carboxylic acid (7a)

Recrystallized from ethyl acetate:ethanol (3:1). Mp. 153-154 °C. FT-IR (ν_{max} , cm⁻¹): 3065 (ar-CH), 1721 (C=O), 1520 (C=N), 1238 (C=S). ¹H NMR (DMSO-*d*₆, δ ppm): 1.41 (t, 3H, *J*=12.0 Hz, CH₃), 2.66 (s, 4H, 2CH₂), 2.74 (s, 2H, CH₂), 2.90 (s, 4H, 2CH₂), 2.94 (s, 4H, 2CH₂), 3.13 (s, 4H, 2CH₂), 3.78 (s, 2H, CH₂), 4.58 (d, 2H, *J*=5.0 Hz, CH₂), 5.07 (s, 2H, CH₂), 6.67 (t, 1H, *J*=16.0 Hz, arH), 6.91 (d, 2H, *J*=8.0 Hz, arH), 7.19 (t, 3H, *J*=12.0 Hz, arH), 7.96 (s, 1H, arH), 8.95 (s, 1H, CH), 15.34 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 14.81 (CH₃), 48.80 (2CH₂), 45.92 (2CH₂), 49.81 (2CH₂), 51.66 (2CH₂), 52.46 (2CH₂), 69.93 (CH₂), arC: [106.47 (CH), 107.55 (C), 111.66 (d, *J*=23.0 Hz, CH), 115.98 (2CH), 119.44 (CH), 119.71 (C), 129.36 (2CH), 137.63 (C), 145.76 (d, *J*=11.0 Hz, C), 151.33 (C), 152.05-154.53 (d_{C-F}, *J*=248.0 Hz, C)], 148.98 (CH), 159.47 (oxadiazole C-5), 166.56 (C=O), 176.63 (C=O), 178.48 (oxadiazole C-2). EI MS *m*/*z* (%): 205.21 (100), 361.27 (50), 517.26 (45), 630.28 ([M+Na]⁺, 40).

3.1.4.2. 1-Cyclopropyl-6-fluoro-4-oxo-7-(4-{[5-[(4-phenylpiperazin-1-yl)methyl]-2-tioxo-1,3,4-oxadiazole-3(2*H*)-yl]methyl}piperazin-1-yl)-1,4-dihydroquinolin-3-carboxylic acid (7b)

Recrystallized from ethyl acetate:ethanol (3:1). Mp. 176 °C. FT-IR (ν_{max} , cm⁻¹): 3014 (ar-CH), 1724 (C=O), 1598 (C=N), 1261 (C=S). ¹H NMR (DMSO-*d*₆, δ ppm): 1.17 (s, 2H, CH₂), 1.32 (d, 2H, *J*=8.0 Hz, CH₂), 2.66 (s, 4H, 2CH₂), 2.74 (s, 2H, CH₂), 2.90 (s, 2H, CH₂), 2.96 (s, 4H, 2CH₂), 3.13 (s, 4H, 2CH₂), 3.79 (s, 2H, CH₂), 3.96 (s, 1H, CH), 5.07 (s, 2H, CH₂), 6.77 (t, 1H, *J*=16.0 Hz, arH), 6.90 (d, 2H, *J*=8.0 Hz, arH), 7.19 (t, 2H, *J*=16.0 Hz, arH), 7.55 (d, 1H, *J*=8.0 Hz, arH), 7.87 (d, 1H, *J*=12.0 Hz, arH), 8.66 (s, 1H, CH), 15.18 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 8.04 (2CH₂), 36.30 (CH), 48.60 (2CH₂), 49.74 (CH₂), 49.82 (2CH₂), 51.66 (2CH₂), 52.46 (2CH₂), 69.91 (CH₂), arC: [107.02 (CH), 107.20 (C), 111.42 (d, *J*=23.0 Hz, CH), 115.97 (2CH), 119.04 (C), 119.28 (d, *J*=32.0 Hz, CH), 129.37 (2CH), 134.69 (C), 139.58 (C), 145.44 (d, *J*=10.0 Hz, C), 151.32 (C), 152.18-154.66 (d_{C-F}, *J*=248.0 Hz, C)], 148.43 (CH), 159.46 (oxadiazole C-5), 166.38 (C=O), 176.80 (C=O), 178.49 (oxadiazole C-2). EI MS *m/z* (%): 619.02 ([M] ⁺, 100).

3.1.4.3. 1-Ethyl-6-fluoro-4-oxo-7-[4-({4-phenyl-3-[(4-phenylpiperazin-1-yl)methyl]-5-tioxo-4,5-dihydro-1*H*-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1,4-dihydroquinolin-3-carboxylic acid (8a)

Recrystallized from ethyl acetate:ethanol (3:1). Mp. 203-205 °C. FT-IR (v_{max} , cm⁻¹): 3021 (ar-CH), 1721 (C=O), 1548 (C=N), 1261 (C=S). ¹H NMR (DMSO- d_6 , δ ppm): 1.44 (s, 3H, CH₃), 2.40 (s, 4H, 2CH₂), 2.92 (s, 4H, 2CH₂), 3.02 (s, 4H, 2CH₂), 3.39 (s, 4H, 2CH₂), 3.48 (s, 2H, CH₂), 4.58 (s, 2H, CH₂), 5.22 (s, 2H, CH₂), 6.75 (s, 1H, arH), 6.82 (s, 1H, arH), 7.17 (s, 3H, arH), 7.52 (d, 5H, *J*=16.0 Hz, arH), 7.91 (s, 1H, arH), 8.91 (s, 1H, CH), 15.26 (bs, 1H, OH). ¹³C NMR (DMSO- d_6 , δ ppm): 14.80 (CH₃), 46.63 (2CH₂), 49.56 (2CH₂), 49.97 (CH₂), 50.10 (2CH₂), 51.67 (CH₂), 51.89 (2CH₂), 69.02 (CH₂), arC: [106.41 (CH), 107.54 (C), 113.12 (2CH), 116.35 (d, *J*=23.0 Hz, 2CH), 119.76 (C), 122.45 (2CH), 124.50 (2CH), 126.11 (2CH), 131.28 (CH), 137.54 (d, *J*=23.0 Hz, 2C), 139.50 (C), 142.33 (C), 148.40 (C), 152.30-155.02 (d_{C-F}, *J*=272.0 Hz, C)], 148.94 (CH), 158.65 (triazol C-5), 166.60 (C=O), 170.01 (triazol C-3), 176.63 (C=O). EI MS *m*/*z* (%): 205.18 (100), 460.27 (26), 682.14 ([M]⁺, 34).

3.1.4.4.1-Cyclopropyl-6-fluoro-4-oxo-7-[4-({4-phenyl-3-[(4-phenylpiperazin-1-yl]methyl]-5-tioxo-4,5-dihydro-1H-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1,4-dihydroquinolin-3-carboxylic acid (8b)

Recrystallized from ethyl acetate:ethanol (3:1). Mp. 243-244 °C. FT-IR (ν_{max} , cm⁻¹): 3018 (ar-CH), 1728 (C=O), 1599 (C=N), 1255 (C=S). ¹H NMR (DMSO-*d*₆, δ ppm): 1.18 (s, 2H, CH₂), 1.33 (s, 2H, CH₂), 2.40 (s, 4H, 2CH₂), 2.92 (s, 4H, 2CH₂), 3.03 (s, 4H, 2CH₂), 3.27 (s, 4H, 2CH₂), 3.48 (s, 2H, CH₂), 3.82 (s, 1H, CH), 5.22 (s, 2H, CH₂), 6.75 (d, 1H, *J*=5.0 Hz, arH), 6.82 (d, 2H, *J*=8.0 Hz, arH), 7.17 (d, 2H, *J*=5.0 Hz, arH), 7.50 (d, 2H, *J*=8.0 Hz, arH), 7.55 (d, 4H, *J*=8.0 Hz, arH), 7.89 (t, 1H, *J*=12.0 Hz, arH), 8.65 (s, 1H, CH), 15.13 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 8.07 (2CH₂), 36.31 (CH), 48.46 (2CH₂), 49.90 (CH₂), 50.09 (2CH₂), 51.90 (2CH₂), 52.42 (2CH₂), 68.95 (CH₂), arC: [106.93 (CH), 107.21 (C), 111.40 (d, *J*=24.0 Hz, CH), 115.80 (2CH), 118.50 (CH), 119.03 (d, *J*=8.0 Hz, C), 119.34 (2CH), 128.73 (CH), 129.33 (CH), 129.41 (2CH), 129.78 (CH), 134.97 (C), 139.59 (C), 145.58 (C), 148.49 (C), 152.19-154.67 (d_{C-F}, *J*=248.0 Hz, C)], 148.49 (CH), 151.23 (triazol C-5), 166.37 (C=O), 169.80 (triazol C-3), 176.79 (C=O). EI MS *m/z* (%): 205.15 (100), 696.11 ([M+2]⁺, 83).

3.1.4.5. 1-Ethyl-6-fluoro-4-oxo-7-[4-({4-benzyl-3-[(4-phenylpiperazin-1-yl)methyl]-5tioxo-4,5-dihydro-1*H*-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1,4-dihydroquinolin-3carboxylic acid (8c)

Recrystallized from ethyl acetate:acetone (1:1). Mp. 248-249 °C. FT-IR (ν_{max} , cm⁻¹): 3063 (ar-CH), 1719 (C=O), 1546 (C=N), 1259 (C=S). ¹H NMR (DMSO-*d*₆, δ ppm): 1.41 (s, 3H, CH₃), 2.44 (s, 4H, 2CH₂), 2.87 (s, 4H, 2CH₂), 2.94 (s, 4H, 2CH₂), 3.33 (s, 4H, 2CH₂), 3.52 (s, 2H, CH₂), 4.58 (s, 2H, CH₂), 5.17 (s, 2H, CH₂), 5.43 (s, 2H, CH₂), 6.75 (s, 1H, arH), 6.82 (s, 2H, arH), 7.17 (s, 3H, arH), 7.27 (s, 2H, arH), 7.30 (s, 2H, arH), 7.90 (d, 1H, *J*=8.0 Hz, arH), 8.94 (s, 1H, CH), 15.33 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 14.81 (CH₃), 46.74 (CH₂), 47.87 (CH₂), 48.20 (2CH₂), 49.52 (CH₂), 49.91 (CH₂), 50.16 (CH₂), 52.33 (2CH₂), 52.63 (2CH₂), 69.02 (CH₂), arC: [106.38 (CH), 107.54 (C), 115.84 (2CH), 119.33 (2CH), 119.64 (C), 127.29 (2CH), 127.43 (CH), 128.87 (2CH), 129.36 (2CH), 136.44 (C), 137.63 (C), 145.72 (C), 148.61 (C), 152.03-154.50 (d_{C-F}, *J*=247.0 Hz, C)], 148.97 (CH), 151.24 (triazol C-5), 166.57 (C=O), 169.85 (triazol C-3), 176.61 (C=O). EI MS *m/z* (%): 694.29 (92), 696.23 ([M]⁺, 100).

3.1.4.6.1-Cyclopropyl-6-fluoro-4-oxo-7-[4-({4-benzyl-3-[(4-phenylpiperazin-1-yl]methyl]-5-tioxo-4,5-dihydro-1H-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1,4-dihydroquinolin-3-carboxylic acid (8d)

Recrystallized from ethyl acetate:petroleum ether (3:1). Mp. 176-178 °C. FT-IR (v_{max} , cm⁻¹): 3028 (ar-CH), 1728 (C=O), 1542 (C=N), 1256 (C=S). ¹H NMR (DMSO- d_6 , δ ppm): 1.13 (s, 2H, CH₂), 1.33 (s, 2H, CH₂), 2.45 (s, 4H, 2CH₂), 2.92 (s, 6H, 3CH₂), 2.96 (s, 4H, 2CH₂), 3.30 (s, 4H, 2CH₂), 3.53 (s, 2H, CH₂), 3.81 (s, 1H, CH), 5.21 (s, 2H, CH₂), 5.41 (s, 2H, CH₂), 6.76 (s, 1H, arH), 6.83 (s, 2H, arH), 7.18 (d, 2H, *J*=6.0 Hz, arH), 7.28 (s, 3H, arH), 7.30 (d, 2H, *J*=5.0 Hz, arH), 7.57 (s, 1H, arH), 7.90 (d, 1H, *J*=12.0 Hz, arH), 8.66 (s, 1H, CH), 15.17 (s, 1H, OH). ¹³C NMR (DMSO- d_6 , δ ppm): 8.06 (2CH₂), 36.30 (CH), 47.72 (2CH₂), 47.88 (CH₂), 48.24 (2CH₂), 49.83 (CH₂), 50.14 (2CH₂), 52.34 (2CH₂), 52.65 (2CH₂), 69.03 (CH₂), arC: [106.96 (CH), 107.21 (C), 111.43 (d, *J*=23.0 Hz, CH), 115.87 (d, *J*=4.0 Hz, 2CH), 119.09 (C), 119.34 (CH), 127.30 (CH), 127.43 (CH), 127.86 (CH), 128.86 (CH), 128.88 (CH), 129.36 (2CH), 136.43 (C), 139.59 (C), 148.67 (d, *J*=10.0 Hz, C), 151.24 (C), 152.18-154.66 (d_{C-F}, *J*=248.0 Hz, C)], 148.43 (CH), 156.07 (triazol C-5), 166.39 (C=O), 169.86 (triazol C-3), 176.81 (C=O). EI MS *m/z* (%): 205.21 (100), 644.26 (32), 710.23 ([M+2]⁺, 73).

3.1.4.7. 1-Ethyl-6-fluoro-4-oxo-7-[4-({4-fluorophenyl-3-[(4-phenylpiperazin-1-yl)methyl]-5-tioxo-4,5-dihydro-1*H*-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1,4-dihydroquinolin-3carboxylic acid (8e)

Recrystallized from ethyl acetate:ethyl acetate (1:1). Mp. 164-165 °C. FT-IR (v_{max} , cm⁻¹): 3063 (ar-CH), 1724 (C=O), 1599 (C=N), 1258 (C=S). ¹H NMR (DMSO-*d*₆, δ ppm): 1.42 (t, 3H, *J*=12.0 Hz, CH₃), 2.39 (s, 8H, 4CH₂), 2.95 (s, 4H, 2CH₂), 3.00 (s, 4H, 2CH₂), 3.47 (s, 2H, CH₂), 4.59 (d, 2H, *J*=8.0 Hz, CH₂), 5.21 (s, 2H, CH₂), 6.76 (s, 1H, arH), 6.85 (q, 2H, *J*=8.0 Hz, arH), 7.18 (d, 3H, *J*=8.0 Hz, arH), 7.40 (d, 3H, *J*=8.0 Hz, arH), 7.55 (d, 2H, *J*=5.0 Hz, arH), 8.95 (s, 1H, CH), 15.35 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 14.83 (CH₃), 48.49 (2CH₂), 49.53 (CH₂), 49.92 (CH₂), 50.11 (2CH₂), 51.94 (2CH₂), 52.43 (2CH₂), 70.97 (CH₂), arC: [106.40 (CH), 107.55 (C), 111.62 (d, *J*=23.0 Hz, CH), 115.70 (d, *J*=6.0 Hz, CH), 115.83 (d, *J*=22.0 Hz, 2CH), 119.35 (2CH), 119.66 (C), 129.35 (2CH), 131.12 (d, *J*=9.0 Hz, 2CH), 137.64 (C), 145.79 (C), 148.63 (d, *J*=17.0 Hz, C), 151.29 (C), 152.06-154.54 (d_{C-F}, *J*=248.0 Hz, C), 163.79-166.58 (d_{C-F}, *J*=279.0 Hz, C)], 148.97 (CH), 161.35 (triazol C-5), 168.62 (C=O), 169.95 (triazol C-3), 176.61 (C=O). EI MS *m*/*z* (%): 205.21 (100), 524.29 (27), 70.88 ([M]⁺, 70).

3.1.4.8. 1-Cyclopropyl-6-fluoro-4-oxo-7-[4-({4-fluorophenyl-3-[(4-phenylpiperazin-1-yl)methyl]-5-tioxo-4,5-dihydro-1*H*-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1,4dihydroquinolin-3-carboxylic acid (8f)

Recrystallized from ethyl acetate: ethyl acetate (1:1). Mp. 145-146 °C. FT-IR (v_{max} , cm⁻¹): 3070 (ar-CH), 1723 (C=O), 1599 (C=N), 1257 (C=S). ¹H NMR (DMSO-*d*₆, δ ppm): 1.18 (s, 2H, CH₂), 1.33 (d, 2H, *J*=8.0 Hz, CH₂), 2.40 (s, 8H, 4CH₂), 2.48 (s, 2H, CH₂), 2.94 (s, 8H, 4CH₂), 3.82 (s, 1H, CH), 5.21 (s, 2H, CH₂), 6.76 (t, 1H, *J*=12.0 Hz, arH), 6.86 (q, 2H, *J*=6.0 Hz, arH), 7.18 (q, 2H, *J*=8.0 Hz, arH), 7.40 (t, 2H, *J*=20.0 Hz, arH), 7.53-7.59 (m, 3H, arH), 7.91 (d, 1H, *J*=12.0 Hz, arH), 8.66 (s, 1H, CH), 15.20 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 8.07 (2CH₂), 36.32 (CH), 48.49 (2CH₂), 49.91 (CH₂), 50.06 (2CH₂), 51.94 (2CH₂), 52.43 (2CH₂), 70.97 (CH₂), arC: [106.99 (CH), 107.22 (C), 111.43 (d, *J*=22.0 Hz, CH), 115.84 (d, *J*=5.0 Hz, 2CH), 116.21 (d, *J*=6.0 Hz, CH), 119.04 (C), 119.35 (CH), 129.35 (2CH), 131.12 (d, *J*=9.0 Hz, 2CH), 139.62 (C), 145.56 (d, *J*=9.0 Hz, C), 148.62 (d, *J*=16.0 Hz, C), 151.27 (d, *J*=4.0 Hz, C), 152.21-154.69 (d_{C-F}, *J*=248.0 Hz, C), 166.39-168.62 (d_{C-F}, *J*=223.0 Hz, C], 148.45 (CH), 161.35 (triazol C-5), 163.79 (C=O), 169.97 (triazol C-3), 176.80 (C=O). EI MS *m*/*z* (%): 205.15 (48), 380.35 (67), 522.29 (44), 609.17 (49), 611.17 (50), 708.16 (82), 710.17 (85),712.12 ([M]⁺, 100), 704.12 ([M+2]⁺, 96).

3.1.4.9. 1-Ethyl-6-fluoro-4-oxo-7-[4-({4-ethyl-3-[(4-phenylpiperazin-1-yl)methyl]-5-tioxo-4,5-dihydro-1*H*-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1,4-dihydroquinolin-3carboxylic acid (8g)

Recrystallized from ethyl acetate:ethanol (3:1). Mp. 193-194 °C. FT-IR (ν_{max} , cm⁻¹): 3095 (ar-CH), 1718 (C=O), 1600 (C=N), 1243 (C=S). ¹H NMR (DMSO-*d*₆, δ ppm): 1.16 (s, 3H, CH₃), 1.33 (d, 3H, *J*=8.0 Hz, CH₃), 2.60 (s, 4H, 2CH₂), 2.74 (s, 4H, 2CH₂), 2.92 (s, 4H, 2CH₂), 3.27 (s, 4H, 2CH₂), 3.70 (s, 2H, CH₂), 3.81 (s, 2H, CH₂), 4.11 (d, 2H, *J*=5.0 Hz, CH₂), 5.13 (s, 2H, CH₂), 6.77 (t, 1H, *J*=12.0 Hz, arH), 6.89 (d, 2H, *J*=8.0 Hz, arH), 7.19 (t, 2H, *J*=12.0 Hz, arH), 7.54 (d, 1H, *J*=5.0 Hz, arH), 7.87 (d, 1H, *J*=16.0 Hz, arH), 8.64 (s, 1H, CH), 15.13 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 13.50 (CH₃), 14.74 (CH₃), 48.65 (2CH₂), 49.51 (2CH₂), 49.87 (CH₂), 50.11 (2CH₂), 52.15 (2CH₂), 52.88 (2CH₂), 68.53 (CH₂), arC: [106.38 (CH), 107.54 (C), 111.66 (d, *J*=23.0 Hz, CH), 115.93 (2CH), 119.54 (d, *J*=15.0 Hz, CH), 119.68 (C), 129.43 (2CH), 137.62 (C), 145.75 (d, *J*=10.0 Hz, C), 148.36 (C), 152.02-154.50 (d_{C-F}, *J*=248.0 Hz, C)], 148.96 (CH), 151.28 (triazol C-5), 166.55 (C=O), 168.46 (triazol C-3), 176.59 (C=O). EI MS *m*/*z* (%): 205.21 (100), 517.26 (35), 658.54 ([M+1+Na]⁺, 38).

3.1.4.10.1-Cyclopropyl-6-fluoro-4-oxo-7-[4-({4-ethyl-3-[(4-phenylpiperazin-1-yl]methyl]-5-tioxo-4,5-dihydro-1H-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1,4-dihydroquinolin-3-carboxylic acid (8h)

Recrystallized from ethyl acetate: ethanol (3:1). Mp. 229-230 °C. FT-IR (ν_{max} , cm⁻¹): 3073 (ar-CH), 1726 (C=O), 1543 (C=N), 1252 (C=S). ¹H NMR (DMSO-*d*₆, δ ppm): 1.33 (t, 4H, *J*=12.0 Hz, 2CH₂), 1.41 (t, 3H, *J*=12.0 Hz, CH₂), 2.58 (s, 4H, 2CH₂), 2.74 (s, 4H, 2CH₂), 2.90 (s, 4H, 2CH₂), 3.34 (s, H₂O+2CH₂), 3.69 (s, 1H, CH), 4.10 (d, 2H, *J*=8.0 Hz, CH₂), 4.58 (d, 2H, *J*=8.0 Hz, CH₂), 5.12 (s, 2H, CH₂), 6.78 (d, 1H, *J*=8.0 Hz, arH), 6.88 (d, 2H, *J*=8.0 Hz, arH), 7.15-7.21 (m, 3H, arH), 7.89 (d, 1H, *J*=16.0 Hz arH), 8.94 (s, 1H, CH), 15.33 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 8.04 (2CH₂), 13.51 (CH₃), 36.28 (CH), 44.07 (CH₂), 48.65 (2CH₂), 49.80 (2CH₂), 50.07 (2CH₂), 52.16 (2CH₂), 52.89 (2CH₂), 68.53 (CH₂), arC: [106.92 (CH), 107.20 (C), 111.39 (d, *J*=23.0 Hz, CH), 115.93 (2CH), 119.01 (d, *J*=7.0 Hz, C), 119.46 (CH), 129.37 (2CH), 139.57 (C), 145.47 (d, *J*=10.0 Hz, C), 148.38 (C), 152.16-154.64 (d_{C-F}, *J*=248.0 Hz, C)], 148.40 (CH), 151.28 (triazol C-5), 166.35 (C=O), 168.47 (triazol C-3), 176.76 (C=O). EI MS *m*/*z* (%): 205.15 (49), 646.26 ([M] ⁺, 90), 648.27 ([M+2]⁺, 100).

3.1.4.11. 7-[4-({4-Benzyl-5-oxo-3-[(4-phenylpiperazin-1-yl)methyl]-4,5-dihydro-1*H*-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1-ethyl-6-fluoro-4-oxo-1,4-dihydroquinolin-3-carboxylic acid (8i)

Recrystallized from ethyl acetate:ethanol (3:1). Mp. 253-255 °C. FT-IR (ν_{max} , cm⁻¹): 3059 (ar-CH), 1703 (C=O), 1677 (C=O), 1600 (C=N). ¹H NMR (DMSO-*d*₆, δ ppm): 1.41 (t, 3H, *J*=12.0 Hz, CH₃), 2.44 (s, 4H, 2CH₂), 2.82 (s, 4H, 2CH₂), 3.23 (s, 4H, 2CH₂), 3.39 (s, 6H, 3CH₂), 4.58 (d, 2H, *J*=5.0 Hz, CH₂), 4.96 (s, 2H, CH₂), 6.94 (d, 2H, *J*=8.0 Hz, arH), 7.17 (d, 1H, *J*=8.0 Hz, arH), 7.25-7.33 (m, 6H, arH), 7.91 (d, 1H, *J*=16.0 Hz, arH), 8.02 (d, 2H, *J*=8.0 Hz, arH), 8.94 (s, 1H, CH), 15.33 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 14.78 (CH₃), 44.87 (CH₂), 46.38 (2CH₂), 49.53 (2CH₂), 49.78 (2CH₂), 52.12 (2CH₂), 52.97 (2CH₂), 66.05 (CH₂), arC: [106.35 (CH), 107.52 (C), 112.92 (2CH), 116.04 (2CH), 119.80 (C), 126.13 (2CH), 127.41 (2CH), 127.80 (CH), 128.92 (2CH), 137.38 (C), 137.58 (d, *J*=9.0 Hz, C), 143.57 (C), 145.82 (d, *J*=10.0 Hz, C), 152.04-154.96 (d_{C-F}, *J*=292.0 Hz, C)], 148.91 (CH), 155.23 (triazol C-3), 161.87 (triazol C-5), 166.59 (C=O), 176.58 (C=O). EI MS *m*/*z* (%): 488.32 (47), 575.20 (88), 577.15 (53), 680.16 ([M]⁺, 100).

3.1.4.12. 7-[4-({4-Benzyl-5-oxo-3-[(4-phenylpiperazin-1-yl)methyl]-4,5-dihydro-1*H*-1,2,4-triazol-1-yl}methyl)piperazin-1-yl]-1-Cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinolin-3-carboxylic acid (8j)

Recrystallized from ethyl acetate: ethanol (3:1). Mp. 199-200 °C. FT-IR (ν_{max} , cm⁻¹): 3059 (ar-CH), 1710 (C=O), 1670 (C=O), 1600 (C=N). ¹H NMR (DMSO-*d*₆, δ ppm): 1.20 (s, 2H, CH₂), 1.33 (s, 2H, CH₂), 2.44 (s, 4H, 2CH₂), 2.73 (s, 2H, CH₂), 3.25 (s, 4H, 2CH₂), 3.34 (s, H₂O+4CH₂), 3.39 (s, 2H, CH₂), 3.89 (s, 1H, CH), 5.03 (s, 2H, CH₂), 6.97 (d, 2H, *J*=8.0 Hz, arH), 7.28-7.35 (m, 6H, arH), 7.58 (t, 1H, *J*=16.0 Hz, arH), 7.91 (d, 1H, *J*=12.0 Hz, arH), 8.04 (d, 2H, *J*=8.0 Hz, arH), 8.66 (s, 1H, CH), 15.22 (s, 1H, OH). ¹³C NMR (DMSO-*d*₆, δ ppm): 8.04 (2CH₂), 36.30 (CH), 44.81 (CH₂), 46.42 (CH₂), 48.39 (CH₂), 50.36 (2CH₂), 51.10 (CH₂), 52.16 (2CH₂), 53.04 (2CH₂), 67.53 (CH₂), arC: [106.92 (CH), 107.23 (C), 111.42 (d, *J*=26.0 Hz, CH), 113.10 (2CH), 119.02 (C), 126.15 (2CH), 127.44 (CH), 127.53 (CH), 127.80 (2CH), 128.91 (2CH), 137.43 (d, *J*=6.0 Hz, C), 139.57 (C), 143.58 (C), 152.12-154.08 (d_{C-F}, *J*=194.0 Hz, C)], 145.63 (triazol C-3), 148.44 (CH), 155.02 (triazol C-5), 166.39 (C=O), 176.83 (C=O). EI MS *m/z* (%): 581.10 (92), 583.11 (100), 694.16 ([M]⁺, 36).

3.2. Antimicrobial activity

The test microorganisms were obtained from the Hifzissihha Institute of Refik Saydam (Ankara, Turkey) and were as follows: *Staphylococcus aureus* ATCC 25923, *Enterococcus faecalis* ATCC 29212, *Escherichia coli* ATCC 25922, *Pseudomonas aeruginosa* ATCC 27853, *Klebsiella pneumoniae* ATCC 13883, *Acinetobacter haemolyticus* ATCC 19002. All the newly synthesized compounds were weighed and dissolved in DMSO to prepare extract stock solution of 10.000 μ g/mL. The antimicrobial effects of the substances were tested quantitatively in respective broth media by using double micro-dilution and the minimal inhibition concentration (MIC) values (μ g/mL) were determined. The antibacterial were performed in Mueller-Hinton broth (MH) (Difco, Detroit, MI) at pH.7.3 and buffered Yeast Nitrogen Base (Difco, Detroit, MI) at pH 7.0, respectively. The micro dilution test plates were incubated for 18–24 h at 35 °C. Brain Heart Infusion broth (BHI) (Difco, Detroit, MI) was used for *M. smegmatis*, and incubated for 48–72 h at 35 °C [48]. Ampicillin (10 μ g), gentamicin (10 μ g) and ciprofloxacin (10 μ g) were used as standard antibacterial respectively. Dimethylsulphoxide with dilution of 1:10 was used as solvent control. All results were presented in Table 3.

3.3. In vitro inhibitory activity screening and determination of IC_{50} values on E. coli gyrase

The assay for determining IC₅₀ values (Inspiralis) was performed on black streptavidincoated 96-well microtiter plates (Thermo Scientific Pierce). The plate was first rehydrated with the wash buffer supplied (20mMTris-HCl (pH 7.6),137mM NaCl, 0.01% (w/v) BSA, 0.05% (v/v) Tween 20). Biotinylated oligonucelotide was immobilized onto the wells. The excess of oligonucleotide was then washed off and the enzyme assay carried out in the wells. The final reaction volume of 30 mL in buffer (35 mM Tris HCl pH 7.5; 24 mM KCl; 4 mM MgCl₂; 2 mM DTT; 1.8 mM spermidine; 1 mM ATP; 6.5% (w/v) glycerol; 0.1 mg/mL albumin) contained 1.5 U of DNA gyrase from E. coli 0.75 mg of relaxed pNO1 plasmid, and 3 mL of inhibitors solution in 10% DMSO and 0.008% Tween® 20. Reactions were incubated for 30 min at 37 °C and, after addition of the TF buffer (50 mM NaOAc pH 5.0, 50 mM NaCl and 50 mM MgCl2), which terminated the enzymatic reaction, for another 30 min at room temperature to allow triplex formation (biotin-oligonucleotide-plasmid). The unbound plasmid was then washed off using TF buffer, and a solution of SybrGOLD stain in T10 buffer (10 mM Tris-HCl pH 8.0 and 1 mM EDTA) was added. After mixing, the fluorescence (excitation, 485 nm; emission, 535 nm) was read using a BioTek's Synergy H4 microplate reader. Preliminary screening was performed at inhibitor concentrations of 100 mM and 10 mM. For the most potent compounds IC₅₀ was determined with 7 concentrations of the inhibitors. IC₅₀ values were calculated using GraphPad Prism software and represent the concentration of inhibitor where the residual activity of the enzyme is 50 % in three independent measurements; the final result is given as their average value. Novobiocin (IC₅₀) ¹/₄ 0.17 mM (lit. 0.08 mM) for E. coli DNA gyrase was used as a positive control [49].

3.4. *In vitro* inhibitory activity screening and determination of IC₅₀ values on E. coli topoisomerase IV

The assay for the determination of IC_{50} values (Inspiralis) was performed on the black streptavidin-coated 96-well microtiter plates (Thermo Scientific Pierce). The plate was first rehydrated with the supplied wash buffer (20 mM Tris-HCl (pH 7.6), 137 mM NaCl, 0.01% (w/v) BSA, 0.05% (v/v) Tween® 20) and biotinylated oligonucelotide was immobilized onto the wells. The excess of oligonucleotide was then washed off, and the enzyme assay carried out in the wells. The final reaction volume of 30 mL in buffer (40 mM HEPES KOH (pH 7.6), 100 mM potassium glutamate, 10 mM magnesium acetate, 10 mM DTT, 1 mM ATP, 0.05

mg/mL albumin) contained 1.5 U of topoisomerase IV from E. coli or S. aureus, 0.75 mg of supercoiled pNO1 plasmid, and 3 mL of inhibitors solution in 10% DMSO and 0.008% Tween® 20. Reactions were incubated for 30 min at 37 °C, and after addition of the TF buffer (50 mM NaOAc pH 5.0, 50 mM NaCl and 50 mM MgCl2), which terminated the enzymatic reaction, for another 30 min at room temperature to allow triplex formation (biotin-oligonucleotide-plasmid). The unbound plasmid was then washed off using TF buffer and the solution of SybrGOLD stain in T10 buffer (10 mM Tris-HCl pH 8.0 and 1mM EDTA) was added. After mixing, fluorescence (excitation, 485 nm; emission, 535 nm) was read using a BioTek's Synergy H4 microplate reader. Preliminary screening was performed at inhibitor concentrations of 100 mM and 10 mM. For most potent compounds IC_{50} was determined with 7 concentrations of the inhibitors. IC_{50} values were calculated using GraphPad Prism software and represent the concentration of inhibitor where the residual activity of the enzyme is 50% in three independent measurements; the final result is given as their average value. Novobiocin (IC_{50} ¹/₄ 11 mM (lit. 10 mM) for E. coli topoisomerase IV was used as a positive control.

3.5. Molecular docking studies

Docking studies of most active (determined through enzyme inhibition studies) new fluoroquinolone-1,2,4-triazole hybrids (only **7a**, **7b** and **8a-j**) into the binding pockets of DNA gyrase (PDB ID: 1SUU) and TOPO-IV (PDB ID: 1S16) was carried out using Autodock4.2.6, Autodock Tools 1.5.6 and the Arguslab version 4.0.1. 3D crystal structures were retrieved from the protein data bank (PDB) (Source: www.rcsb.org/pdb/). All molecular docking tactics and parameters were referred from our previous reports [50-52]. Statistical mechanical analysis for the ligands and the lowest binding energy, ligand efficiency (it is used in drug discovery investigations to support in narrowing focus to candidate drugs with optimum amalgamations of physicochemical and pharmacological potentials [53]) and the inhibitory constant (*ki*) values were extracted. Molecular interaction (hydrogen bonding, π - π interaction, and π -cation interaction) results were validated.

4. Discussion

In our initial effort, to synthesis medicinally important small molecules, compound 8a, has been selected as a model product to determine the optimum reaction conditions. In this trial, the model reaction has been carried out in polar solvent including ethanol, tetrahydrofurane and DMF in the presence of different Lewis and Bronsted acid catalysts, such as *p*-TSA,

FeCl₃, InCl₃ and HCl under the microwave irradiated conditions. In all cases, progress of reaction has been monitored by TLC analysis. The condensation of the azoles, **4** and **6a-e** with fluoroquinolone class antibacterial drugs namely norfloxacin and ciprofloxacin has been carried out under the Mannich reaction conditions. A Mannich reaction which leads to the formation of aminoalkylated products has been descript as a one pot condensation between three components consisting of a substrate containing at least one active hydrogen, an aldehyde component and an amine reagent [54, 55]. *N*-Mannich bases constitute important tools to development of new drugs containing amine or amide function [56]. It has been believed that the group linked to parent amine through a Mannich reaction cause to increase the absorption of molecules through bio-membrans as a result of their diminished protonation causing to increase the lipophilicity of molecules at physiological pH values [57]. Furthermore, the transformation into iminium salts render the Mannich bases to soluble in aqueous solvents [58].

Bringing the achieved compounds to medicinal applications, these compounds were initially tested for their antimicrobial effects and DNA gyrase and TOPO-IV inhibitions. DNA gyrase and TOPO-IV inhibition result of all compounds was compared to the standard drug, Novobiocin. The obtained potentially good DNA gyrase and TOPO-IV inhibition by the compounds was further validated through binding affinity calculations and molecular interaction validations. DNA gyrase and TOPO-IV are the enzymes that are highly involved in the synthesis of bacterial genetic material, especially in the DNA supercoiling processes [59, 60]. Due to this reason these two enzymes are being the antibacterial drug targets. In the present study, compounds **7a**, **7b** and **8a-j** were showed remarkable antibacterial as well as effective DNA gyrase inhibition activities. Compound **8j** can be a future antibiotic.

4. Conclusion

We have designed the Microwave irradiated and conventional synthesis, antibacterial activity, DNA gyrase and topoisomerase IV inhibition of new piperazine functionalized fluoroquinolone hybrids. Compared to conventional method, by the microwave irradiation method, reaction yield and time was significantly decreased. At the same time, as a result of optimization studies for the synthesis of fluoroquinolone derivatives as target molecules, it was determined that Lewis acids and solvent were not affected. The antibacterial activity screening studies have revealed that new hybrid molecules consisting of fluoroquinolone-azole-piperazine conjugation (**7a,b** and **8a-j**) have good-excellent antibacterial activity against

the test microorganisms. Furthermore, these hybrids have exhibited good DNA gyrase inhibition with the IC₅₀ values varying between 0.134-1.84 μ g/mL. To unveil the interaction mode of compounds to receptor, a molecular docking study has been performed. With an average least binding energy of -9.5 kcal/mol, all compounds have been found to possess remarkable inhibitory potentials against DNA gyrase (*E. coli*). Conclusively, considering their bio-activity potentials, perhaps highly substitute (**8a-j**) piperazine-azole functionalized fluoroquinolone hybrids (**7a,b** and **8a-j**) could be the future antibiotics.

References

- [1] K. Lal, P. Yadav, A. Kumar, A. Kumar, A.K. Paul, Design, synthesis, characterization, antimicrobial evaluation and molecular modeling studies of some dehydroacetic acidchalcone-1,2,3-triazole hybrids, Bioorganic Chemistry 77 (2018) 236-244.
- [2] S.B. Ozdemir, N. Demirbas, A. Demirbas, N. Colak, F.A. Ayaz, Design, microwaveassisted and conventional synthesis of new hybrid compounds derived from 1-(4fluorophenyl) piperazine and screening for their biological activities, Chemistry Select 3 (2018) 2144-2151.
- [3] D.D. Subhedar, M.H. Shaikh, B.B. Shingate, L. Nawale, D. Sarkar, V.M. Khedkar, F.A. K. Khan, J.N. Sangshetti, Quinolidene-rhodanine conjugates: Facile synthesis and biological evaluation, Eur. J. Med. Chem. 125 (2017) 385-399.
- [4] T. Tomasic, S. Katsamakas, Z. Hodnik, J. Ilas, M. Brvar, T. Solmajer, S. Montalvio, P. Tammela, M. Banjanac, G. Ergovic, M. Anderluh, L.P. Masic, D. Kikelj, Discovery of 4,5,6,7-Tetrahydrobenzo[1,2-d]thiazoles as novel DNA gyrase inhibitors targeting the ATP-binding site, J. Med. Chem. 58 (2015) 5501-5521.
- [5] K. Hoshino, A. Kitamura, I. Morrissey, K. Sato, J. Kato, H. Ikeda, Comparison of Inhibition of Escherichia Coli Topoisomerase IV by Quinolones with DNA Gyrase Inhibition, Antimicrobial Agents and Chemotherapy, 38 (1994) 2623-2627.
- [6] A.M. Vijesh, A.M. Isloor, P. Shetty, S. Sundershan, H.K. Fun, New pyrazole derivatives containing 1, 2, 4-triazoles and benzoxazoles as potent antimicrobial and analgesic agents, Eur. J. Med. Chem. 62 (2013) 410-415.
- [7] M. Mentese, N. Demirbas, A. Mermer, S. Demirci, A. Demirbas, F.A. Ayaz, Novel azole-functionalited flouroquinolone hybrids: Design, conventional and microwave irradiated synthesis, evaluation as antibacterial and antioxidant agents, Letters in Drug Design and Discovery, 15 (2018) 46-64.
- [8] S. Demirci, A. Demirbas, S. Ulker, S. Alpay-Karaoglu, N. Demirbas, Synthesis of some heterofunctionalized penicillanic acid derivatives and investigation of their biological activities, Arch Pharm (Weinheim) 347 (2014) 200-20
- [9] A. Siwek, P. Staczek, J. Stefanska, Synthesis and structure activity relationship studies of 4-arylthiosemicarbazides as topoisomerase IV inhibitors with Gram-positive antibacterial activity. Search for molecular basis of antibacterial activity of thiosemicarbazides Eur. J. Med. Chem. 46 (2011) 5717-5726.
- [10] M.J. Mitton-Fry, S.J. Brickner, J.C. Hamel, R. Barham, L. Brennan, J.M. Casavant, X. Ding, S. Finegan, J. Hardink, T. Hoang, M.D. Huband, M. Maloney, A. Marfat, S.P. McCurdy, D. McLeod, C. Subramanyam, M. Plotkin, U. Reilly, J. Schafer, G.G. Stone,

D.P. Uccello, T. Wisialowski, K. Yoon, R. Zaniewski, C. Zook, Novel 3-fluoro-6methoxyquinoline derivatives as inhibitors of bacterial DNA gyrase and topoisomerase IV, Bioorg. Med. Chem. Lett. 27 (2017) 3353-3358.

- [11]I.A. Yule, L.G. Czaplewski, S. Pommier, D.T. Davies, S.K. Narramore, C.W.G. Fishwick, Pyridine-3-carboxamide-6-yl-ureas as novel inhibitors of bacterial DNA gyrase: Structure based design, synthesis, SAR and antimicrobial activity, Eur. J. Med. Chem. 86 (2014) 31-38.
- [12] S.S. Panda, S. Liaqat, A.S. Girgis, A. Samir, C.D. Hall, A.R. Katritzky, Novel antibacterial active quinolone–fluoroquinolone conjugates and 2D-QSAR studies, Bioorg. Med. Chem. Lett. 25 (2015) 3816-3821.
- [13] X. Wang W. Wei, P. Wang, Y.T. Tang, R.C. Deng, B. Li, S.S. Zhou, J.W. Zhang, L. Zhang, Z.P. Xiao, H. Ouyang, H.L. Zhu, Novel 3-arylfuran-2 (5H)-one-fluoroquinolone hybrid: design, synthesis and evaluation as antibacterial agent, Bioorg. Med. Chem. 22 (2014) 3620-3628.
- [14] X. Li, Y.K. Zhang, J.J. Plattner, W. Mao, M.R.K. Alley, Y. Xia, V. Hernandez, Y. Zhou, Z.C. Ding, J. Li, Z. Shao, H. Zhang, M. Xu, Synthesis and antibacterial evaluation of a novel tricyclic oxaborole-fused fluoroquinolone, Bioorg. Med. Chem. Lett. 23 (2013) 963-966.
- [15]S. Ceylan, H. Bektas, H. Bayrak, N. Demirbas, S. Alpay-Karaoglu, S. Ulker, Syntheses and biological activities of new hybrid molecules containing different heterocyclic moieties, Arc. Pharm. 346 (2013) 743-756.
- [16] E. Chugunova, N. Akylbekov, A. Bulatova, N. Gavrilov, A. Voloshina, N. Kulik, V. Zobov, A. Dobrynin, V. Syakaev, A. Burilov, Synthesis and biological evaluation of novel structural hybrids of benzofuroxan derivatives and fluoroquinolones, Eur. J. Med. Chem. 116 (2016) 165-172.
- [17]S.B. Ozdemir, N. Demirbas, Y.U. Cebeci, H. Bayrak, A. Mermer, S. Ceylan, A. Demirbas, Synthesis and Antimicrobial Activities of Hybrid Heterocyclic Molecules Based on 1-(4-Fluorophenyl) piperazine Skeleton, Letters in Drug Design and Discovery, 14 (2017) 1014-1034.
- [18]S. Basoglu, S. Ulker, S. Alpay-Karaoglu, N. Demirbas, Microwave-assisted synthesis of some hybrid molecules containing penicillanic acid or cephalosporanic acid moieties and investigation of their biological activities, Med. Chem. Res. 23 (2014) 3128-3143.
- [19] N. Zidar, T. Tomasic, H. Macut, A. Sirc, M. Brvar, S. Montalvao, P. Tammela, J. Ilas, D. Kikelj, New N-phenyl-4,5-dibromopyrrolamides and N-Phenylindolamides as ATPase inhibitors of DNA gyrase, Eur. J. Med. Chem. 117 (2016) 197-211.
- [20] A. Siwek, P. Staczek, J. Stefanska, Synthesis and structure activity relationship studies of 4-arylthiosemicarbazides as topoisomerase IV inhibitors with Gram-positive antibacterial activity. Search for molecular basis of antibacterial activity of thiosemicarbazides, Eur. J. Med. Chem. 46 (2011) 5717-5726.
- [21] M.I. Andersson, A.P. MacGowan, Development of the quinolones, Journal of Antimicrobial Chemotherapy, 51 (2003) 1–11.
- [22] M. Mentese, S. Demirci, S.B. Ozdemir, A. Demirbas, S. Ulker, N. Demirbas, Microwave assisted synthesis and antimicrobial activity evaluation of new heterofunctionalized norfloxacine derivatives, Letters in Drug Design and Discovery, 13 (2016) 1076-1090.
- [23] K. Itoh, Y. Kuramoto, H. Amano, D. Kazamori, A. Yazaki, Discovery of WQ-3810: Design, synthesis, and evaluation of 7-(3-alkylaminoazetidin-1-yl)fluoro-quinolones as orally active antibacterial agents, Eur. J. Med. Chem. 103 (2015) 354-360.

- [24] Y. Chai, Z.L. Wan, B. Wang, H.Y. Guo, M.L. Liu, Synthesis and in vitro antibacterial activity of 7-(4-alkoxyimino-3-amino-3-methylpiperidin-1-yl) fluoroquinolone derivatives, Eur. J. Med. Chem. 44 (2009) 4063-4069.
- [25] T. Tomasic, M. Mirt, M. Barancokova, J. Ilas, N. Zidar, P. Tammela, D. Kikelj, Design, synthesis and biological evaluation of 4,5-dibromo-N-(thiazol-2-yl)-1H-pyrrole-2carboxamide derivatives as novel DNA gyrase inhibitors, Bioorg. Med. Chem. 25 (2017) 338-349.
- [26] M. Alagumuthu, S. Arumugam, Molecular docking, discovery, synthesis, and pharmacological properties of new 6-substituted-2-(3-phenoxyphenyl)-4-phenyl quinoline derivatives; an approach to developing potentDNA gyrase inhibitors/antibacterial agents, Bioorg. Med. Chem. 25 (2017) 1448-1455.
- [27] K.C. Fang, Y.L. Chen, J.Y. Sheu, T.C. Wang, C.C. Tzeng, Synthesis, antibacterial, and cytotoxic evaluation of certain 7-substituted norfloxacin derivatives, J. Med. Chem. 43 (2000) 3809-3812.
- [28]G. El-Din, A.A. Abuo-Rahma, H.A. Sarhan, G.F.M. Gad, Design, synthesis, antibacterial activity and physicochemical parameters of novel N-4-piperazinyl derivatives of norfloxacin, Bioorg. Med. Chem. 17 (2009) 3879-3886.
- [29] P.C. Sharma, A. Jain, M.S. Yar, R. Pahwa, J. Singh, S. Goel, Synthesis and antibacterial evaluation of novel analogs of fluoroquinolones annulated with 6-substituted-2-aminobenzothiazoles, Arabian J. Chem. 8 (2015) 671-677.
- [30] M.Y. Mentese, H. Bayrak, Y. Uygun, A. Mermer, S. Ulker, S. Alpay Karaoglu, N. Demirbas, Microwave assisted synthesis of some hybrid molecules derived from norfloxacin and investigation of their biological activities, Eur. J. Med. Chem. 67 (2013) 230-242
- [31]Z. Dang, Y.S. Yang, R.Y. Ji, S.H. Zhang, Synthesis and antibacterial activity of novel fluoroquinolones containing substituted piperidines, Bioorg. Med. Chem. Lett. 17 (2007) 4523-4526.
- [32] R.G. Fiorot, J.F.A. Filho, T.M.C. Pereira, V.R. Lacerda, B. Santos, W. Romao, S.J.A. Greco, A. simple and convenient method for synthesis of new amino naphthoquinones derived from lawsone by catalytic multicomponent Mannich reaction, Tetrahedron Lett. 55 (2014) 4373-4377.
- [33]S.S. Mansoor, K. Aswin, K. Logaiya, S.P.N. Sudhan, An efficient synthesis of β-amino ketone compounds throughone-pot three-component Mannich-type reactions using bismuth nitrate as catalyst, J. Saudi Chemical Society, 19 (2015) 379-386.
- [34]H.A.M. El-Sherief, B.G.M. Youssif, S.N.A. Bukhari, M. Abdel-Aziz, H.M. Abdel-Rahman, Novel 1,2,4-triazole derivatives as potential anticancer agents: Design, synthesis, molecular docking and mechanistic studies, Bioorganic Chemistry 76 (2018) 314-325.
- [35]S.S. Thakkar, P. Thakor, H. Doshi, A. Ray, 1,2,4-Triazole and 1,3,4-oxadiazole analogues: Synthesis, MO studies, *in silico* molecular docking studies, antimalarial as DHFR inhibitor and antimicrobial activities, Bioorganic & Medicinal Chemistry 25 (2017) 4064-4075.
- [36] Y.U. Cebeci, S.B. Ozdemir, S. Ceylan, H. Bayrak, A. Demirbas, S. Alpay-Karaoglu, N. Demirbas, Microwave-assisted synthesis of some hybrid molecules derived from morpholine and investigation of their antimicrobial activities, Chemistry Select 2 (2017) 11402-11407.

- [37] S. Ceylan, H. Bayrak, A. Demirbas, S. Ulker, S. Alpay Karaoglu, N. Demirbas, Synthesis of some new hybride molecules containing several azole moieties and investigation of their biological activities, Russian J. Bioorg. Chem. 40 (2014) 314–329.
- [38] A. Mermer, N. Demirbas, Y. Sirin, H. Uslu, Z. Ozdemir, A. Demirbas, Conventional and microwave prompted synthesis, antioxidant, anticholinesterase activity screening and molecular docking studies of new quinolone-triazole hybrids, Bioorganic Chemistry 78 (2018) 236-248.
- [39] K.K. Sivakumar, A. Rajasekaran, P. Senthilkumar, P.P. Wattamwar, Conventional and microwave assisted synthesis of pyrazolone Mannich basespossessing anti-inflammatory, analgesic, ulcerogenic effect and antibacterial properties, Bioorg. Med. Chem. Lett. 24 (2014) 2940-2944.
- [40] A.S. Medvedeva, M.M. Demina, T.V. Konkova, T.L.H. Nguyen, A.V. Afonin, I.A. Ushakov, Microwave assisted solvent-and catalyst-free three-component synthesis of NH-1, 2, 3-triazoloimines, Tetrahedron 73 (2017) 3979-3985.
- [41] S. Abas, U. Moens, C. Escolano, Facile microwave-assisted synthesis of thioformamides from isocyanides and carbon disulfide, Tetrahedron Lett. 58 (2017) 2768–2770.
- [42] P. Sharma, T.S. Reddy, N.P. Kumar, K.R. Senwar, S.K. Bhargava, N. Shankaraiah. Conventional and microwave-assisted synthesis of new 1H-benzimidazole-thiazolidinedione derivatives: A potential anticancer scaffold, Eur. J. Med. Chem. 138 (2017) 234-245.
- [43] W.X. Sun, Y.J. Ji, Y. Wan, H.W. Han, H.Y. Lin, G.H. Lu, J.L. Qi, X.M. Wang, Y.H. Yang, Design and synthesis of piperazine acetate podophyllotoxin ester derivatives targeting tubulin depolymerization as new anticancer agents, Bioorg. Med. Chem. Lett., 27 (2017) 4066-4074.
- [44] P.J. Hergenrother, Preparation of N'-(2-hydroxybenzylidene)-2-(piperazin-1-yl) asetohidrazids as procaspase-activating compounds, (2013) US 20130096133.
- [45]H. Bayrak, A. Demirbas, N. Demirbas, S.A. Karaoglu, Synthesis of some new 1 2, 4triazoles starting from isonicotinic acid hydrazide and evaluation of their antimicrobial activities, Eur. J. Med. Chem. 44 (2009) 4362-4366.
- [45] S. Basoglu, A. Demirbas, S. Ulker, S. Alpay-Karaoglu, N. Demirbas, Design, synthesis and biological activities of some 7-aminocephalosporanic acid derivatives, Eur. J. Med. Chem. 69 (2013) 622-631.
- [46]M. Yolal, S. Basoglu, H. Bektas, et al., Synthesis of eperezolid-like molecules and evaluation of their antimicrobial activities, Russ. J. Bioorg. Chem. 38 (2012) 539-549.
- [47] S. Demirci, F. Aksakal, N. Colak, S. Ulker, A. Demirbas, N. Demirbas, Structure-Based hybridization, conventional and microwave irradiated synthesis, biological evaluation and molecular docking studies of new compounds derived from thiomorpholin, Letters Drug Design and Discovery, 14 (2017) 444-463.
- [47] R.A. Rane, S.D. Gutte, N.U. Sahu, Synthesis and evaluation of novel 1,3,4-oxadiazole derivatives of marine bromopyrrole alkaloids as antimicrobial agent, Bioorg. Med. Chem. Lett. 22 (2012) 6429-6432.
- [48] Clinical and Laboratory Standards Institute. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Approved Standard-Ninth Edition. CLSI document M07-A9, 2012, Wayne, PA.
- [49] Z. Jakopin, J. Ilas, M. Barancokova, M. Brvar, P. Tammela, M.S. Dolenc, T. Tomasic, D. Kikelj, Discovery of substituted oxadiazoles as a novel scaffold for DNA gyrase inhibitors, Eur. J. Med. Chem. 130 (2017) 171-184

- [50] Manikandan A, Pearl Moharil, Sathiskumar M, C Muñoz-Garay and Sivakumar A. Therapeutic investigations of novel Indoxyl-based indolines: A drug target validation and Structure-Activity Relationship of angiotensin-converting enzyme inhibitors with cardiovascular regulation and thrombolytic potential. *Eur. J. Med. Chem.* 141 (2017) 417-426.
- [51] Manikandan A, Ravichandran S, Sathiyanarayanan K.I, Sivakumar A. Efficacy and rationale of 2-(4-phenylquinolin-2-yl)phenols as COX-2 inhibitors; an approach to emergent the small molecules as the anti-inflammatory and analgesic therapeutics. *Inflammopharmacology*, 25 (2017) 621–631.
- [52] Manikandan A, Sivakumar A. Molecular explorations of substituted 2-(4phenylquinolin-2-yl) phenols as phosphoinositide 3-kinase inhibitors and anticancer agents. *Cancer Chemother Pharmacol* 79 (2017) 389-397.
- [53] Kuntz I. D, Chen K, Sharp K. A, Kollman P. A. The maximal affinity of ligands. *Proc Natl Acad Sci U S A*. 96 (1999) 9997–10002.
- [54]S.B. Dongare, H.V. Chavan, P.S.Bhale, Y.B. Mule, A.S. Kotmale, P. Babasaheb, P. Bandgar, A catalyst- and solvent-free multicomponent synthesis of 7-azagramine analogues via a Mannich type reaction, Chinese Chem. Lett. 27 (2016) 99-103.
- [55]G.K. Oloyede, I.E. Willie, O.O. Adeeko, Synthesis of Mannich bases: 2-(3-Phenylaminopropionyloxy)-benzoic acid and 3-Phenylamino-1-(2,4,6-trimethoxyphenyl)-propan-1-one, their toxicity, ionization constant, antimicrobial and antioxidant activities, Food Chemistry, 165 (2014) 515-521.
- [56] Y.J. Zhao, W. Wei, Z.G. Su, G.H. Ma, Poly (ethylene glycol) prodrug for anthracyclines via N-Mannich base linker: Design, synthesis and biological evaluation, International Journal of Pharmaceutics, 379 (2009) 90-99.
- [57] Clinical and Laboratory Standards Institute. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Approved Standard-Ninth Edition. CLSI document M07-A9 (2012) Wayne, PA.
- [58]Z. Jakopin, J. Ilas, M. Barancokova, M. Brvar, P. Tammela, M.S. Dolenc, T. Tomasic, D. Kikelj, Discovery of substituted oxadiazoles as a novel scaffold for DNA gyrase inhibitors, Eur. J. Med. Chem. 130 (2017) 171-184.
- [59] Dilek I, Xiamei Z, Christine S.L, and David C.H. Dual Targeting of DNA Gyrase and Topoisomerase IV: Target Interactions of Garenoxacin (BMS-284756, T-3811ME), a New Desfluoroquinolone. Antimicrob. Agents Chemother. 46 (2002) 3370-3380.
- [60] Chris U, and Nicholas R.C. Contrasting Enzymatic Activities of Topoisomerase IV and DNA Gyrase from *Escherichia coli**. The Journal of Biological Chemistry 271 (1996) 31549-31555.

Highlights

- *E. Coli* DNA gyrase and Topoisomerase IV inhibition of newly synthesized fluoroquinolone derivatives
- Screening antimicrobial activity of novel drug-like compounds.
- Docking studies were performed for the most active compounds and interaction modes with enzyme active sites were determined.
- Conventional and microwave prompted synthesis of new fluoroquinolone-triazole •

Graphical abstract

Piperazine-Azole-Fluoroquinolone Hybrids: Conventional and Microwave Irradiated Synthesis, Biological Activity Screening and Molecular Docking Studies



Arif Mermer^a, Ozlem Faiz^b, Ahmet Demirbas^a, Neslihan Demirbas^a*, Manikandan Alagumuthu^c, Sivakumar Arumugam^c