TWO CINNAMIC ALLENIC ETHERS FROM THE FUNGUS CLITOCYBE EUCALYPTORUM*

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(Received 13 July 1992)

Key Word Index—*Clitocybe eucalyptorum*; Basidiomycetes; allenic ether; *cis-* and *trans-cinnamates*; eucalyptenes A and B.

Abstract—The structure of eucalyptene A, a fungal metabolite isolated from *Clitocybe eucalyptorum*, has been determined to be 4'-(2'',3'')-butadienyloxy) *trans*-cinnamic acid methyl ester on the basis of ¹H and ¹³C NMR evidence and chemical synthesis. Minor amounts of its *cis* isomer, eucalyptene B, were also isolated.

INTRODUCTION

During screening of the genus *Clitocybe* for secondary metabolites, we reported the isolation of a large number of protoilludane sesquiterpenes, viz. melledonals D and E from *C. elegans* [2], candicansol and 3-epi-illudol from *C. candicans* [3], and illudines A and B [4], illudalenol [4] and illudosin [5] from *C. illudens* (*O. olearius*). Further investigation of the *Clitocybe* genus led to the isolation from a strain of *C. eucalyptorum*, when grown in MPGA (malt, peptone, glucose and agar) medium, of a novel allenic ether of p-hydroxy-trans-cinnamic acid methyl ester, named eucalyptene A (3), together with some minor amounts of its *cis* isomer, eucalyptene B (4).

RESULTS AND DISCUSSION

Eucalyptene A (3) was isolated as a solid and its molecular formula C14H14O3 established by EI mass spectrometry, the peaks at m/z 215 [M – Me]⁺ and 178 $[M - C_4 H_4]^+$ indicating the loss of a methyl group and of a four-carbon fragment. Absorptions at 1965 and 1715 cm⁻¹ in the IR spectrum suggested the presence of allenic and conjugated ester groups. The ¹H NMR spectrum in chloroform-d (Table 1) showed the presence of a singlet at δ 3.79 due to a methoxyl group and AX and AA'XX' spin systems attributable to the protons of a trans-disubstituted double bond $({}^{3}J_{2,3} = 16.0 \text{ Hz})$ and of a paradisubstituted phenyl moiety. In addition, it displayed an A₂MX₂ spin system due to the C-1", C-2" and C-4" protons. The broad-band ¹H-decoupled and the fully ¹H-coupled ¹³CNMR spectra confirmed the presence of 14 carbon atoms ($1 \times Me$, $2 \times CH_2$, $7 \times CH$ and $4 \times C$). Four carbons were assigned to the -C(3)H=C(2)

H–C(1)O₂C(4)H₃ methyl acrylate moiety, six carbons, C-1' to C-6', to the aromatic ring, with the remaining four carbons, C-1" to C-4", being attributed to a 2,3-butadienyloxy group. In fact, the chemical-shift values of δ 86.7, 209.6 and 76.8 observed for C-2", C-3" and C-4" [6], together with the one-bond ¹H–¹³C couplings of 167 and 169 Hz exhibited by C-2" and C-4" [7], indicate that they are part of a –C(2")H=C(3")=C(4")H₂ moiety, while the presence of ¹H–¹H couplings of 6.8 and 2.5 Hz between the 1"-methylene protons, which were correlated with the oxygen-bearing carbon at δ 65.9, and H-2" and H"₂-4 defines the linkage between C-1" and C-2".

Definitive evidence for the proposed structure 3 followed from the synthesis of eucalyptene A shown in Scheme 1. This was accomplished by reacting 1-chloro-4-hydroxybut-2-yne [8] with p-hydroxy-trans-cinnamic acid methyl ester as the sodium salt in dry tetrahydro-furan. Chlorination with thionyl chloride of the resulting acetylenic ether 1 gave the chloro derivative 2 [9] which was reduced to eucalyptene A (3) with zinc-copper complex in methanol [10]. Finally, compound 3 on irradiation in isopropyl alcohol with 300 nm light yielded the *cis*-isomer 4, as evidenced by the presence in its ¹H NMR spectrum (Table 1) of a ¹H-¹H coupling of 12.2 Hz between the olefinic protons typical of a *cis*-geometry.

The spectroscopic properties of the synthesized compounds 3 and 4, i.e. ¹H NMR, IR and UV spectral data, were identical to those of the natural ones.

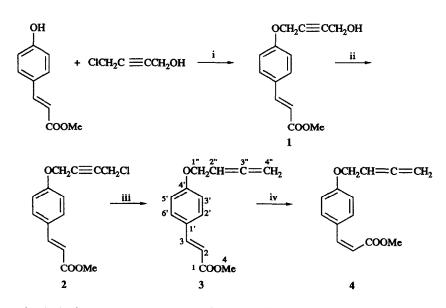
Allenic functions in basidiomycetous fungi are formed by isomerization of acetylenes by isomerases; in particular, the genus *Clitocybe* produces a series of polyacetylenes active as inhibitors of fungi and of seed germination [11, 12]. Cinnamic acids are widely present in higher plants and fungi, and are active as growth inhibitors of plants and spore germination [13]. A similar allenic ether, chestersiene, has been isolated from the fungus Hypoxylon chestersii and it is the only other report of the

^{*}Part 40 in the series 'Secondary Mould Metabolites'. For Part 39 see ref. [1].

Atom	3			4	
	δ_{c}	δ_{H}	J (H, H) (Hz)	$\delta_{ m H}$	J (H, H) (Hz)
1	167.8				
2	115.4 (162)*	6.31 d	16.0	5.83 d	12.2
3	144.5 (155)	7.65 dtt	16.0, 0.6, 0.4	6.85 br d	12.2
4	51.6 (147)	3.79 s		3.73 s	
1′	129.7				
2′	130.3 (158)	7.47 m		7.69 m	
3′	115.2 (161)	6.91 m		6.90 m	
4′	160.2				
5'	115.2 (161)	6.91 m		6.90 m	
6'	130.3 (158)	7.47 m		7.69 m	
1″	65.9 (148)	4.60 dt	6.8, 2.5	4.61 dt	6.8, 2.5
2″	86.7 (167)	5.38 tt	6.8, 6.6	5.39 tt	6.8, 6.6
3″	209.6				
4″	76.8 (169)	4.89 dt	6.6, 2.5	4.88 dt	6.6, 2.5

Table 1. ¹³C and ¹H NMR spectral data of compounds 3 and 4 (in CDCl₃)

*Values in parentheses are one-bond ¹H-¹³C coupling constants in Hz.



Scheme 1. Synthesis of eucalyptene A (3). Reagents: (i) NaH-THF; (ii) SOCl₂-THF; (iii) Cu, Zn-MeOH; (iv) hv (300 nm)-2-propanol.

natural occurrence of an aromatic allenic ether [14]. Eucalyptenes A (3) and B (4) exhibit antifungal activity, revealed by means of bioautography on *Cladosporium cladosporioides* for amounts as low as 50 μ g.

EXPERIMENTAL

Mps: uncorr. Flash CC was performed on silica gel (0.04–0.06 mm), TLC on HF₂₅₄ silica gel. MS were recorded at 60 eV. ¹H NMR were recorded at 300 MHz, ¹³C NMR at 63 MHz. Chemical shifts are in ppm (δ) from TMS as int. standard. The apparatus used for irradiation of **3** was a Rayonet RPR-100 (Southern New England UV Co.) equipped with 16 fluorescent lamps irradiating at 300 nm.

Cultivation of fungus and isolation of eucalyptene. The CBS 433.72 strain of *C. eucalyptorum* was inoculated in 40 Roux flasks containing MPGA (100 ml) (malt ext.-peptone-glucose-agar, $20:5:20:15 \text{ g} \text{ l}^{-1}$) with a mycelium suspension. After 3 weeks, flasks were extracted with EtOAc containing 1% MeOH, the combined extracts dried (Na₂SO₄) and evapd to give a crude extract (1.1 g). This mixt. was chromatographed on a silica gel column using hexane-EtOAc (4:1) and purified further by prep. TLC (PLC) with hexane-EtOAc (9:1) to yield eucalyptenes A (3) (150 mg) and B (4) (20 mg).

4'-(2",3"-butadienyloxy)trans-Cinnamic acid methyl ester (3) (eucalyptene A). Solid, mp 68° (from hexane). (Found: C, 72.88; H, 6.10. $C_{14}H_{14}O_3$ requires: C, 73.02; H, 6.13%.) EIMS (rel. int.) m/z: 230 [M]⁺ (45), 215 (15), 192 (72), 178 (100) and 149 (69); CIMS (isobutane) m/z: 231 [MH]⁺. UV $\lambda_{max}^{95\% EtOH}$ nm: 204, 220 and 302 (ε 12050, 10950 and 19060). ¹H and ¹³C NMR in Table 1.

4'-(2",3"-butadienyloxy)cis-Cinnamic acid methyl ester (4) (eucalyptene B). Oil. EIMS (rel. int.) m/z: 230 [M]⁺ (50), 215 (10), 192 (70), 178 (100) and 149 (70). IR $v_{max}^{Liquid film}$ cm⁻¹: 1965 (allene) and 1715 (ester CO).

4'-(2"-butynyloxy-4"-hydroxy)trans-Cinnamic acid methyl ester (1). To a stirred suspension of NaH (80%, 440 mg, 14.6 mmol) in dry THF (10 ml), a soln of phydroxycinnamic acid Me ester (1.3 g, 7.3 mmol) in 5 ml of dry THF was added dropwise. The suspension was then refluxed for 1 hr. After cooling, 1-chloro-4hydroxybut-2-yne (760 mg, 7.3 mmol) dissolved in dry THF was added dropwise. The mixt. was stirred under reflux for 48 hr. After cooling, H₂O was added and extraction was performed (Et₂O, 2×10 ml). The combined Et_2O extracts were dried (Na₂SO₄), the solvent evapd and the resulting solid was chromatographed on a flash silica gel column using hexane-EtOAc (2:1). Compound 1 was obtained as a solid (1.1 g, 60%), mp 98° (from EtOH). EIMS m/z: 246 [M]⁺, 228, 203 and 178. ¹H NMR (CDCl₃): δ 1.90 (1H, br s, OH), 3.80 (3H, s, OMe), 4.30 (2H, br s, H₂-4"), 4.73 (2H, br s, H₂-1"), 6.27 (1H, d, J = 15.8 Hz, H-2), 6.93 (2H, m, H-3' and H-5'), 7.45(2H, m, H-2' and H-6') and 7.60 (1H, d, J = 15.8 Hz, H-3).

4'-(2"-butynyloxy-4"-chloro)trans-Cinnamic acid methyl ester (2). Compound 1 (0.5 g, 2 mmol) was dissolved in dry THF (10 ml) and dry pyridine (0.2 ml) added under stirring together with thionyl chloride (0.12 g, 1 mmol) dissolved in dry THF (2 ml). The mixt. was refluxed for 12 hr, ice-H₂O added and extraction was performed (Et₂O, 2 × 10 ml). The combined Et₂O extracts were dried (Na₂SO₄) and the solvent evapd. The crude compound was purified by chromatography (hexane–EtOAc, 2: 1) to give **2** as a solid (0.5 g, 95%), mp 58°. EIMS m/z: 264 [M]⁺, 229, 197, 187, 178 and 147. ¹H NMR (CDCl₃): 3.79 (3H, s, OMe), 4.17 (2H, t, J = 2.1 Hz, H₂-4''), 4.78 (2H, t, J= 2.1 Hz, H₂-1''), 6.34 (1H, d, J = 15.9 Hz, H-2), 6.96 (2H, m, H-3' and H-5'), 7.50 (2H, m, H-2' and H-6') and 7.65 (1H, d, J = 15.9 Hz, H-3).

Eucalyptene A (3). The chloro derivative 2(0.2 g) was dissolved in dry MeOH (3 ml) and added under stirring to a suspension of Cu (0.1 g) and Zn (0.1 g) powders and the mixt. refluxed for 24 hr. The soln was then cooled, centrifuged and the solvent evapd under vacuum. The residue was recrystallized from hexane yielding a compound identical to the natural one.

Eucalyptene B (4). Eucalyptene A (3) (30 mg) on irradiation in isopropyl alcohol (20 ml) with 300 nm light yielded the *cis*-isomer, eucalyptene B (4), as an oil in a 60% yield.

Antifungal tests. These expts were performed by means of bioautography. Pure metabolites were loaded in the amounts of 50, 100 and 200 μ g on a TLC plate, which was, after development, sprayed with a conidial suspension of *C. cladosporioides* in Czapek Dox Broth. The plate was then incubated in a moist chamber and antifungal activity was evidenced by the appearance of a white spot, corresponding to the position of the active metabolite, surrounded by a grey-black fungal growth all over the plate.

Acknowledgement—This work was supported by Consiglio Nazionale delle Ricerche (CNR) Rome, Progetto finalizzato "Chimica fine II".

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