Accepted Manuscript

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PII: S0223-5234(15)00033-1

DOI: 10.1016/j.ejmech.2015.01.013

Reference: EJMECH 7629

To appear in: European Journal of Medicinal Chemistry

Received Date: 30 September 2014

Revised Date: 22 November 2014

Accepted Date: 8 January 2015

Please cite this article as: K.M. Naidu, H.N. Nagesh, M. Singh, D. Sriram, P. Yogeeswari, K.V. Gowri Chandra Sekhar Novel 6-(piperazin-1-yl)phenanthridine amide and sulphonamide derivatives as potent *Mycobacterium tuberculosis* H37Rv inhibitors, *European Journal of Medicinal Chemistry* (2015), doi: 10.1016/j.ejmech.2015.01.013.

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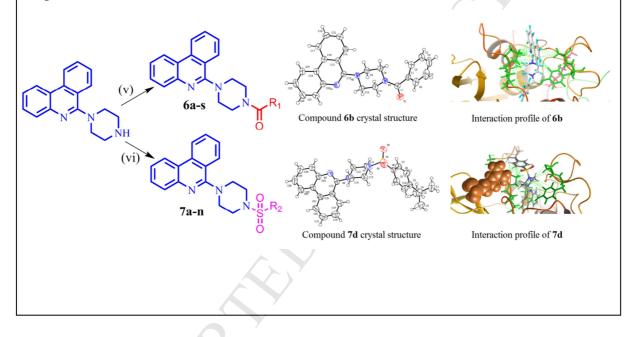
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33 novel compounds are synthesized and evaluated for their anti-TB activity. Amongst these, 3 compounds exhibited excellent anti-TB activity with MIC 1.56 μ g/mL and SI for these compounds was >46.



Novel 6-(piperazin-1-yl)phenanthridine amide and sulphonamide derivatives as potent *Mycobacterium tuberculosis* H37Rv inhibitors

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Abstract: A series of thirty three novel 6-(piperazin-1-yl)phenanthridine amide and sulphonamide analogues were synthesized, characterized and screened for their *in vitro* antimycobacterial activity against *Mycobacterium tuberculosis* (MTB) H37Rv strain. These compounds exhibited minimum inhibitory concentration (MIC) between $1.56 - \ge 50 \ \mu g/mL$. Out of these derivatives, few compounds **61**, **67**, **7b**, **7f**, **7g** and **7k** exhibited moderate activity (MIC = $6.25 \ \mu g/mL$) and compounds **6b**, **6e**, **6k**, **6n**, **7h**, **7i** and **7n** displayed good activity (MIC = $3.13 \ \mu g/mL$), whereas compounds **6m**, **6s** and **7d** exhibited excellent anti-tubercular activity (MIC = $1.56 \ \mu g/mL$). In addition, MTT assay was accomplished on the active analogues of the series against mouse macrophage (RAW 264.7) cells to evaluate the toxicity profile of the newly synthesized compounds and selectivity index of the compounds was determined. Additionally, compounds **6b** and **7d** were docked to the ATPase domain of *Mycobacterium tuberculosis* GyrB protein to know the interaction profile and structures of compounds **6b** and **7d** were further substantiated through single crystal XRD.

Keywords: Phenanthridine; antimycobacterial activity; piperazine sulphonamide; *Mycobacterium tuberculosis*; single crystal XRD.

1. Introduction

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Tuberculosis (TB) is a contagious disease caused by tubercular bacilli. In spite of several efforts by scientists across the world, this ancient scourge cannot be curtailed. It remains as one of the major public health concerns after the human immunodeficiency virus. Worldwide in 2013, 9 million people fell ill with TB, including 1.1 million cases among people living with HIV. 1.5 million died from TB, including 0.36 million who were HIV-positive. An estimated 0.48 million people developed multidrug-resistant TB (MDR-TB) and there were an estimated 0.21 million deaths from MDR-TB. Around 9.0 % of MDR-TB cases have extensively drug-resistant TB (XDR-TB). Around 100 countries have XDR-TB cases. More than 50 companies are involved in the development of new TB drugs. Since last four decades there are no new anti TB drugs except for Bedaquiline and Delamanid which became the first novel TB drugs approved. These two are used only for the treatment of pulmonary MDR-TB patients in serious or life-threatening conditions [1,2]. TB drug discovery and development has not progressed to the extent necessary to annihilate the illness completely. Hence, it is extremely essential to examine new compounds to cure against drug resistant forms of this mortal disease along with minimal side effects. Focus should also be on developing compounds which not only reduce the cost, but also the treatment period.

Phenanthridine derivatives are significant core moieties found in a variety of natural products, important class of alkaloids and other synthetic vital molecules with a wide range of biological activities and applications, including antibacterial [3,4], anticancer [5,6], antimalarial [7], anti-HIV [8], anti-HCV agents [9], antiplasmodial [10] and in particular as antitubercular agents [11-15]. Recently, Cappoen et al reported 1,2,3,4,8,9,10,11-Octahydrobenzo[*j*]phenanthridine-7,12-diones as new leads against *Mycobacterium tuberculosis* [11]. Our group also recently reported synthesis and evaluation of anti-tubercular activity of 6-(4-substitutedpiperazin-1-yl) phenanthridine analogues as anti-TB agents [12].

Lipophilicity of compounds plays an important role in the biological activities which they display. Attaching an alkyl/aryl amide and sulphonamides enhances the lipophilicity of compounds [16,17]. Consequently, alkyl/aryl amide and alkyl/aryl sulphonamide derivatives are known to exhibit wide range of pharmacological activities and physiochemical properties. Indeed, these alkyl/aryl amide and alkyl/aryl sulphonamide analogues display enhanced antibacterial and anti-TB activity [17-22].

In the quest for developing novel anti-TB agents we recently reported two compounds with sulphonamide moiety [13]. Interestingly out of the two compounds, one of them exhibited excellent anti-TB activity (MIC = $1.56 \mu g/mL$) and this prompted us to explore further structure activity relationship (SAR) of the compounds and hence prepared this series of

compounds based on sulphonamides. Since carboxamides are analogous to sulphonamides, we also decided to synthesise various amide derivatives and study their SAR. Hence, we designed new (substituted)(4-(phenanthridin-6-yl)piperazin-1-yl)methanone and 6-(4-((4-substituted)sulfonyl)piperazin-1-yl)phenanthridine derivatives expecting increased biological activity to combat this fatal disease. Design strategy to achieve title compounds [11-15, 17-22] is depicted in **figure 1**.

2. Chemistry

In this report, we chalked out for the synthesis of 6-(4-substitutedpiperazin-1-yl) phenanthridine derivatives starting from 9-fluorenone as outlined in Scheme 1. We synthesized compound 5 according to our reported protocol with slight modification [12]. 9fluorenone upon treatment with hydroxylamine hydrochloride and sodium acetate afforded Nhydroxy-9*H*-fluoren-9-imine (2). Further heating 2 with polyphosphoric acid and phosphorus pentoxide gave phenanthridin-6(5H)-one (3). 6-chlorophenanthridine (4) was synthesized by refluxing 3 with phosphorus oxychloride and N,N-dimethylaniline. Cardinal synthon 6-(piperazin-1-yl)phenanthridine (5) was prepared by irradiating a mixture of 6chlorophenanthridine, anhydrous triethylamine piperazine, (TEA) and N.Ndimethylformamide (DMF) under microwave conditions.

Initially, we tried monitoring the reaction conditions with various bases and solvents as summarised in **Table 1**. We set off our investigation for the synthesis of **6b**, under various reaction conditions. As a model reaction, compound (5) was treated with various acids and amide coupling reagents were employed to get desired compound (entry 1-11). Initially, we employed TEA as base, 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC.HCl) and 1hydroxybenzotriazole (HOBt) as amide coupling reagents in the presence of dichloromethane (DCM) as solvent at 0 °C to rt. Resultant mixture was stirred for 8h to yield 6b in about 58% (entry 1). Changing the solvent to DMF under similar reaction conditions yielded 6b in about 52%. Alternatively, changed amide coupling reagents such as 1-[Bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxid hexafluoro phosphate (HATU), (Benzotriazol-1-yloxy)tris(dimethylamino)phosphonium hexafluorophosphate (BOP) (Benzotriazol-1-yloxy)tripyrrolidinophosphonium and hexafluorophosphate (PyBOP) and solvent as N,N-diisopropylethylamine (DIPEA) under similar condition yields are 38-42% (entry 3-5). Next we hope to improve the yields, 1-Propanephosphonic anhydride (T_3P) was employed to alleviate the yield (entry 6-11). Initially, the reaction was carried out with various bases and solvents (entry 6-7) to give the desired compound in moderate yield. The reaction was further optimised with TEA and DCM

at various intervals of time (entry 8-11), to give **6b** in good yield. Found entries 9-10 potently improved the yield and actuate reaction period 6-8 h and reputable technique. Having the optimised reaction conditions handy, a series of **19** compounds **6a-s** was synthesized in good yield, thus validating the scope of the present protocol.

Further, compound **5** was reacted with various sulfonyl chlorides, using TEA as base and DCM as solvent at 0 °C to rt for 1-2 h to yield **7a-n** in good yields. All the title compounds were purified by column chromatography. Both analytical and spectral data (¹H NMR, ¹³C NMR, FTIR, Elemental analysis and MS) of all the synthesized compounds were confirmed prior to their evaluation of antimycobacterial activity.

In general, nucleophilic aromatic substitution at 6-chlorophenanthridine was confirmed by FTIR analysis of key synthon **5** which showed the disappearance of IR absorption peak at ~754 cm⁻¹ due to aromatic C-Cl stretching frequencies and further disappearance of weak band at ~3280 cm⁻¹ in the IR spectra of 6-(piperazin-1-yl)phenanthridine (**5**) confirms the formation of title compounds (**6a-s** and **7a-n**). In general, ¹H NMR spectrum of final compounds displayed 8 aromatic protons of phenanthridine in the range δ 7.32 - 8.74 and 8 aliphatic piperazine (-CH₂-) protons were observed in the range δ 3.42 - 4.28. The spectral data of all the title compounds are provided in experimental section.

3. Results and discussion

3.1. Antimycobacterial activity

All the synthesized compounds were tested for their ability to inhibit the growth of MTB H37Rv strain by Microplate Alamar Blue Assay (MABA) with compound concentration ranging from 50 to 0.78 µg/mL. Isoniazid, Rifampicin and Pyrazinamide were used as the positive drug standards. The *in vitro* test results for title compounds tabulated in **Table 2** indicate that MIC ranges from 1.56 to \geq 50 µg/mL. Compounds with MIC 1.56 – 12.5 µg/mL were further subjected to cytotoxicity studies. In these analogues, compounds **6m**, **6s** and **7d** are found to be excellent promising candidates by inhibiting 99% growth of MTB H37Rv (ATCC 27294) strain at 1.56 µg/mL concentration.

Among the synthesized compounds, electronic effects of substituent play an important role in displaying anti-TB activity. 6-(piperazin-1-yl)phenanthridine (**5**) was inhibiting 99% growth of MTB H37Rv strain at 25 μ g/mL [12]. Amongst, phenanthridine amides (**6a-s**) and phenanthridine sulphonamide (**7a-n**) derivatives MTB activity summary, SAR studies are described based on activity of compound **5**. Introduction of cyanoacetyl group (**6a**) at the 4th position of compound **5**, retained the anti-TB activity with MIC 25 μ g/mL. Presence of amine an electron donating group (EDG) at either ortho (**6g**) or para (**6h**) position on phenyl ring

decreased the activity by ≥ 2 fold. Introduction of nitro group, an electron withdrawing group (EWG), at meta (6e) position on phenyl ring greatly enhanced anti-TB activity by 8 fold with MIC 3.13 µg/mL. On the other hand nitro group at di-meta (6f) and para (6d) position on phenyl ring, decreased activity by 1-2 folds. Presence of chloro an EWG at para (6c) position on phenyl ring improved anti-TB activity by 2 fold with MIC 12.5 µg/mL. Benzoyl group (6b) at 4th position of compound 5 significantly increased anti-TB activity by 8 folds with MIC 3.13 µg/mL; cinnamoyl group (6j) at 4th position of 5 increased activity by 2 folds with MIC 12.5 µg/mL and 2-(naphthalen-1-yl)acetyl group (6i) at 4th position of compound 5 critically decreased the anti-TB activity by >2 folds with MIC >50 μ g/mL. To establish extensive SAR we also varied heterocyclic compounds such as five membered rings (furan and thiophene), six membered ring (pyridyls) and indole analogues. Furan-2-carbonyl group (60) and thiophene-2-carbonyl group (6p) at 4th position of 6-(piperazin-1-yl)phenanthridine (5) retained and decreased anti-TB activity by 1-2 folds with MIC 25 μ g/mL and 50 μ g/mL respectively. Introduction of six member heterocyclic rings such as nicotinoyl (61), picolinoyl (6k), 6-formicacid picolinoyl (6n) and isonicotinoyl groups (6m) at 4th position of 5 greatly enhanced the anti-TB activity by 4 – 16 folds with MIC 6.25 – 1.56 µg/mL. While, 2-(1Hindol-3-yl)acetyl (6q), 3-(1H-indol-3-yl)propanoyl (6r) and 4-(1H-indol-3-yl)butanoyl groups (6s) at 4th position of 5 witnessed either retention (6q) or greatly enhanced (6r and **6s**) the anti-TB activity by 1 - 16 folds. In conclusion, **6a-s** presence of EDGs on phenyl ring and five membered heterocyclic rings decreased anti-TB activity while EWGs, six membered and benzannulated heterocyclic rings increased the activity (exception 6d).

SAR profile of the fourteen phenanthridine sulphonamide (**7a-n**) analogues is as follows. In the presence of simple methanesulfonyl group (**7a**) at the 4th position of compound **5** the anti-TB activity is retained with MIC 25 μ g/mL. Benzenesulfonyl group (**7b**) at the 4th position of compound **5** increased the anti-TB activity by 4 fold with MIC 6.25 μ g/mL. Presence of EDGs in the phenyl ring increased the activity in general exception being (**7c**). 2,4,6-triethyl groups (**7e**) on phenyl ring improved anti-TB activity by 2 fold with MIC 12.5 μ g/mL. *Tert*butyl group (**7d**) at para position on phenyl ring remarkably enriched the anti-TB activity by 16 folds with MIC 1.56 μ g/mL. Methoxy group at para (**7k**) position on phenyl ring increased the anti-TB activity by 4 folds with MIC 6.25 μ g/mL. Presence of electron withdrawing halogens (F (**7g**), Cl (**7h**) and Br (**7i**)) at para position on phenyl ring impressively enhanced the anti-TB activity by 4-8 folds with MIC 6.25 – 3.13 μ g/mL, including compound **7f**. Electron withdrawing nitro (**7l**) and acetyl (**7m**) groups at para positions on phenyl ring either retained or decreased activity by 1–2 folds. 5-bromothiophene-2-sulfonyl group at 4th

position of 6-(piperazin-1-yl)phenanthridine (5) prominently augmented the anti-TB activity by 8 folds with MIC 3.13 μ g/mL. In general, EWGs on phenyl ring enhanced or retained the anti-TB activity exception being **71**. Wrapping up the results, we observe that in the series synthesized, sulphonamide analogues were more active than the amides.

3.2. Molecular modelling studies

To scrutinize the interaction profile, the compounds (6b and 7d) were docked to the ATPase domain of MTB GyrB protein found from protein data bank (PDB ID-3ZKB) [23] with additional precision mode (XP) of Glide module [24]. The interaction patterns of the analogues were studied by starting with analysis of co-crystallized ligand docking pattern. The crystal ligand phosphoaminophosphonic acid-adenylate ester (ANP) was involved in prominent H-bonding connections, one of the interactions between the amino group of the ANP moiety and the oxygen atom of Asp79 at bond length was 1.78Å as shown in Figure 2. The direction of this ligand in the protein active site disclosed the presence of a hydrophobic pocket where the ANP moiety was stabilized by the interactions with val 99, Ile 84, lys 108 and Tyr 114. The docking pose of compounds 6b and 7d were portrayed in figure 3 and figure 4 respectively. Compound 6b showed strong hydrogen bond interaction with Lys 108 by1.97Å distance. Both the compounds displayed common potent hydrophobic interactions with val 98, val 99, Ile 84, Pro 85, ala 113 and Tyr 114, whereas compound 7d showed extra hydrophobic interaction with Ala 87 and strong π - π stacking interaction (CPK model of protein in Figure 4) with Arg 141. These hydrophobic interactions lead towards the specificity of compounds with this protein, assigning to its activity.

3.3. Cytotoxicity assay

Among the series, most active compounds (MIC $\leq 12.5 \ \mu g/mL$) were subjected to Promega Cell Titer 96 non-radioactive cell proliferation assay to evaluate the *in vitro* cytotoxicity against mouse macrophage (RAW264.7) cell lines. The approximate IC₅₀ values and selectivity index (SI) values [25] are tabulated in **Table 3.** In these all active compounds (MIC $\leq 12.5 \ \mu g/mL$) displayed SI profile >7. while good anti-TB active compounds **6b**, **6e**, **6k**, **6n**, **7h**, **7i** and **7n** (MIC 3.13 $\mu g/mL$) exhibited SI profile >19. The most active anti-TB compounds **6m**, **6s**, and **7d** exhibited excellent SI profile >46. This outcome indicates that novel chemophores **6m**, **6s** and **7d** are not toxic and might be considered for further structural modification of phenanthridine piperazine amide/sulphonamide analogues as leads to accomplish the treatment of this deadly disease.

3.4. X-ray crystallographic study

The X-ray crystallographic analysis of the compounds **6b** and **7d** was carried out: Crystals were grown from the slow evaporation of a 1:3:6 ratio of methanol/dichloromethane/ethyl acetate solvent mixture at rt, to get white flake crystals. A desirable crystal under triclinic system with space groups $P\overline{1}$ of **6b**, crystal size/mm³ $0.6 \times 0.5 \times 0.4$ and crystal under monoclinic system with space groups $P2_1/n$ of **7d**, crystal size/mm³ $0.6 \times 0.5 \times 0.4$ are spectively. These crystals were kept at 293 K during data collection. Using Computer programs SHELXL97 (Sheldrick, 2008) the structure was solved with the unknown structure solution program using unknown and refined with the SHELXS-97 (Sheldrick, 2008) [26]. The crystal data, data collection and structure refinement details are summarized in **Table 4** and molecular structure is given as an ORTEP diagram pictured in **Figure 5**. Crystallographic Data Centre (CCDC). These compounds have been assigned the following deposition numbers CCDC 1012376 and CCDC 1012377 respectively. The X-ray structure parameters and complete details of compound **6b** and **7d** are provided in supplementary material.

4. Conclusion:

In this communication, phenanthridine derivatives were synthesized with moderate to excellent yields. The preliminary *in vitro* anti-TB screening results, toxicity studies and SI profile results of compounds **6b**, **6e**, **6k**, **6n**, **7h**, **7i** and **7n** (MIC 3.13 μ g/mL) displayed good anti-TB activity and SI profile >19. Compounds **6m**, **6s**, and **7d** (MIC 1.56 μ g/mL) exhibited excellent anti-TB activity and SI profile >46. Detailed *in vivo* studies and the exact mechanistic studies of **6m**, **6s**, and **7d** analogues are scope of future study in our lab. The 6-(4-substitutedpiperazin-1-yl)phenanthridine skeleton is established to be an important motif for further development of anti-tubercular agents.

5. Experimental Section:

5.1. Materials and methods:

Chemicals and solvents were procured from commercial sources and are analytically pure. Thin-layer chromatography (TLC) was carried out on aluminium-supported silica gel plates (Merck 60 F254) with visualization of components by UV light (254 nm). Column chromatography was carried out on silica gel (Merck 230-400 mesh). ¹H NMR spectra and ¹³C NMR spectra were recorded at 300 MHz using a Bruker AV 300 spectrometer or 400 MHz using a Bruker AV 400 spectrometer (Bruker CO., Switzerland) in CDCl₃ or DMSO-D₆ solution with tetramethylsilane as the internal standard, and chemical shift values (δ) were given in ppm. Microwave reactions were performed in closed vessel using Biotage Initiator

microwave synthesizer (Uppsala, Sweden). IR spectra were recorded on a FT-IR spectrometer (Schimadzu) and peaks are reported in cm⁻¹. Melting points were determined on an electro thermal melting point apparatus (Stuart-SMP30) in open capillary tubes and are uncorrected. Elemental analyses were analysed by Elementar Analysensysteme GmbH vario MICRO cube CHNS/O Analyzer. Mass spectra (ESI-MS) were recorded on Schimadzu MS/ESI mass spectrometer.

5.2 Synthesis of title compounds (6a-s and 7a-n)

For **6a-s**: 6-(piperazin-1-yl)phenanthridine (1.14 mmol) was dissolved in DCM (4 mL) in an oven dry 25mL two necked RBF. TEA (3.41 mmol) and amide coupling reagent T_3P (1.36 mmol) followed by anhydrous substituted alkyl / aryl acids (1.14 mmol) were added at 0 °C and stirred at rt for 6h.

For **7a-n**: 6-(piperazin-1-yl)phenanthridine (1.14 mmol) was dissolved in DCM (4 mL) in an oven dry 25mL two necked RBF. TEA (3.41 mmol) followed by anhydrous substituted aryl sulfonyl chloride (1.36 mmol) were added at 0 °C and stirred for 2h at rt.

Completion of the reaction was monitored by TLC using 0 - 6% MeOH in DCM as mobile phase. After the reaction was complete, reaction mixture was quenched in ice cold water and extracted with DCM (3 x 10 mL), combined organic layers were washed with saturated brine solution and dried on anhydrous Na_2SO_4 then evaporated *in vacuo*. Obtained residue was purified by column chromatography using gradient 0 - 6% MeOH in DCM, to give the title compounds (**6a-s** and **7a-n**).

5.2.1. 3-oxo-3-(4-(phenanthridin-6-yl)piperazin-1-yl)propanenitrile (6a)

Appearance: off white solid; yield = 78%; mp = 139-140 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.56 (d, J = 8.4 Hz, 1H), 8.42 (d, J = 7.2 Hz, 1H), 8.02 (d, J = 8.4 Hz, 1H), 7.90 (d, J = 8.0 Hz, 1H), 7.82-7.48 (m, 4H), 3.82 – 3.54 (t, 8H), 3.46 (s, 2H). ¹³C NMR (100.61 MHz, CDCl₃) δ 170.14, 161.68, 142.40, 136.22, 134.45, 131.41, 130.37, 128.83, 127.19, 125.14, 124.17, 122.13, 121.58, 120.26, 119.85, 51.60, 47.12, 31.28. FT-IR: v 2248 (CN); ESI-MS: (m/z) calcd. for C₂₀H₁₈N₄O, 330.14, found: 331.22 (M + H)⁺. Anal. Calcd for C₂₀H₁₈N₄O; (%) C 72.71, H 5.49, N 16.96; found: C 72.82, H 5.54, N 17.04.

5.2.2. (4-(phenanthridin-6-yl)piperazin-1-yl)(phenyl)methanone (6b)

Appearance: white solid; yield = 85%; mp = 108-109 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.59 (d, J = 8.4 Hz, 1H), 8.46 (d, J = 7.2 Hz, 1H), 8.22 (d, J = 7.6 Hz, 1H), 7.91 (d, J = 8.0 Hz, 1H), 7.82 (t, J = 7.6 Hz, 1H), 7.78 (m, 2H), 7.66-7.45 (m, 6H), 4.12 – 3.51 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 170.74, 159.62, 143.50, 135.74, 135.02, 133.41, 130.37, 130.12,

129.83, 128.83, 128.59, 127.15, 126.95, 126.14, 125.17, 122.83, 121.91, 121.26, 51.63, 47.82. ESI-MS: (m/z) calcd. for $C_{24}H_{21}N_3O$, 367.16, found: 368.23 (M + H)⁺. Anal. Calcd for $C_{24}H_{21}N_3O$; (%) C 78.45, H 5.76, N 11.44; found: C 78.61, H 5.81, N 11.53.

5.2.3. (4-chlorophenyl)(4-(phenanthridin-6-yl)piperazin-1-yl)methanone (6c)

Appearance: off white solid; yield = 81%; mp = 132-133 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.59 (d, *J* = 8.4 Hz, 1H), 8.46 (d, *J* = 8.8 Hz, 2H), 8.25 (d, *J* = 8.2 Hz, 1H), 7.96 (d, *J* = 10.8 Hz, 1H), 7.91 (d, *J* = 8.0 Hz, 1H), 7.84 (d, *J* = 8.8 Hz, 2H), 7.81 – 7.48 (m, 4H), 3.67 – 3.44 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 170.52, 160.14, 144.22, 136.18, 135.81, 132.87, 131.08, 130.64, 129.54, 128.63, 128.32, 127.11, 126.84, 126.33, 125.46, 122.57, 121.87, 121.18, 52.28, 47.54. ESI-MS: (m/z) calcd. for C₂₄H₂₀ClN₃O, 401.12, found: 402.25 (M + H)⁺. Anal. Calcd for C₂₄H₂₀ClN₃O; (%) C 71.73, H 5.02, N 10.46; found: C 71.81, H 5.09, N 10.53.

5.2.4. (4-nitrophenyl)(4-(phenanthridin-6-yl)piperazin-1-yl)methanone (6d)

Appearance: yellow solid; yield = 68%; mp = 148-149 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 8.62 (d, J = 8.4 Hz, 1H), 8.48 (d, J = 8.0, 1H), 8.25 (d, J = 8.4 Hz, 1H), 8.20 (d, J = 9.6 Hz, 2H), 7.97 (d, J = 7.6 Hz, 1H), 7.92 (d, J = 8.0 Hz, 2H), 7.82 – 7.54 (m, 4H), 3.72 – 3.68 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 172.12, 164.51, 146.86, 142.02, 139.53, 137.84, 132.07, 130.84, 129.74, 128.44, 128.01, 127.69, 126.80, 126.11, 125.47, 123.78, 122.35, 120.88, 52.23, 48.59. ESI-MS: (m/z) calcd. for C₂₄H₂₀N₄O₃, 412.15, found: 413.22 (M + H)⁺. Anal. Calcd for C₂₄H₂₀N₄O₃; (%) C 69.89, H 4.89, N 13.58; found: C 69.98, H 4.95, N 13.70.

5.2.5. (3-nitrophenyl)(4-(phenanthridin-6-yl)piperazin-1-yl)methanone (6e)

Appearance: pale yellow solid; yield = 63%; mp = 230-231 °C; ¹H NMR (400 MHz, DMSOd₆) δ 8.69 (s, 1H), 8.63 (d, *J* = 8.4 Hz, 1H), 8.42 (d, *J* = 8.0, 1H), 8.26 (d, *J* = 8.4 Hz, 1H), 8.21 (d, *J* = 9.2 Hz, 1H), 7.97 (d, *J* = 7.6 Hz, 1H), 7.93 (m, 1H), 7.92 (d, *J* = 8.0 Hz, 1H), 7.82 – 7.54 (m, 4H), 3.69 – 3.57 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 171.08, 162.31, 147.36, 143.47, 141.11, 139.62, 135.24, 132.04, 131.84, 130.02, 129.21, 128.68, 128.32, 127.34, 126.01, 125.82, 125.21, 123.52, 121.76, 120.88, 53.62, 47.84. ESI-MS: (m/z) calcd. for C₂₄H₂₀N₄O₃, 412.15, found: 413.20 (M + H)⁺. Anal. Calcd for C₂₄H₂₀N₄O₃; (%) C 69.89, H 4.89, N 13.58; found: C 69.96, H 4.96, N 13.66.

5.2.6. (3,5-dinitrophenyl)(4-(phenanthridin-6-yl)piperazin-1-yl)methanone (6f)

Appearance: yellow solid; yield = 64%; mp = 196-197 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 9.08 - 8.83 (s, 3H), 8.46 (d, J = 8.0, 1H), 8.22 (d, J = 8.4 Hz, 1H), 8.18 (d, J = 7.6 Hz, 1H), 7.93 (d, J = 8.0 Hz, 1H), 7.85 – 7.48 (m, 4H), 3.71 – 3.62 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 172.32, 159.22, 154.08, 144.75, 142.47, 139.81, 138.02, 133.83, 132.38, 130.39, 129.21, 128.52, 127.01, 125.44, 124.23, 122.89, 121.22, 119.65, 52.63, 47.82. ESI-MS: (m/z) calcd. for C₂₄H₁₉N₅O₅, 457.13, found: 458.21 (M + H)⁺. Anal. Calcd for C₂₄H₁₉N₅O₅; (%) C 63.02, H 4.19, N 15.31; found: C 63.11, H 4.24, N 15.39.

5.2.7. (2-aminophenyl)(4-(phenanthridin-6-yl)piperazin-1-yl)methanone (6g)

Appearance: white solid; yield = 60%; mp = 164-165 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 8.38 (d, J = 8.0 Hz, 1H), 8.22 (d, J = 7.6 Hz, 1H), 8.07 (d, J = 8.0 Hz, 1H), 7.91 (d, J = 8.4 Hz, 1H), 7.80 – 7.18 (m, 6H), 7.12 (d, J = 8.4 Hz, 1H), 7.09 (d, J = 8.4 Hz, 1H), 6.02 (br, 2H), 3.68 – 3.52 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 169.78, 158.13, 146.87, 142.42, 141.44, 138.23, 137.54, 131.12, 130.66, 130.08, 129.23, 128.84, 126.45, 124.87, 124.12, 123.88, 121.86, 120.22, 119.82, 118.52, 52.42, 46.57. ESI-MS: (m/z) calcd. for C₂₄H₂₂N₄O, 382.17, found: 383.26 (M + H)⁺. Anal. Calcd for C₂₄H₂₂N₄O; (%) C 75.37, H 5.80, N 14.65; found: C 75.51, H 5.85, N 14.54.

5.2.8. (4-aminophenyl)(4-(phenanthridin-6-yl)piperazin-1-yl)methanone (6h)

Appearance: off white semi solid; yield = 62%; mp = Not determined (ND); ¹H NMR (400 MHz, DMSO-d₆) δ 8.57 (d, *J* = 8.4 Hz, 1H), 8.43 (d, *J* = 8.0, 1H), 8.22 (d, *J* = 7.8 Hz, 1H), 7.98 (d, *J* = 8.4 Hz, 1H), 7.84 (m, 1H), 7.74 (d, *J* = 8.4 Hz, 2H), 7.71 – 7.52 (m, 3H), 7.18 (d, *J* = 8.2 Hz, 2H), 5.84 (br, 2H), 3.78 – 3.54 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 170.84, 163.58, 147.18, 146.09, 140.25, 134.28, 131.12, 130.82, 129.98, 128.22, 127.84, 127.02, 126.45, 125.34, 124.72, 123.21, 122.87, 119.24, 52.64, 47.82. ESI-MS: (m/z) calcd. for C₂₄H₂₂N₄O, 382.17, found: 383.22 (M + H)⁺. Anal. Calcd for C₂₄H₂₂N₄O; (%) C 75.37, H 5.80, N 14.65; found: C 75.54, H 5.86, N 14.52.

5.2.9. 2-(naphthalen-1-yl)-1-(4-(phenanthridin-6-yl)piperazin-1-yl)ethanone (6i)

Appearance: pale yellow solid; yield = 75%; mp = 181-182 °C; ¹H NMR (400 MHz, DMSOd₆) δ 8.80 (d, J = 8.4 Hz, 1H), 8.65 (d, J = 8.0 Hz, 1H), 8.28 (d, J = 8.8 Hz, 1H), 8.14 (d, J = 8.0 Hz, 1H), 7.96 (d, J = 7.8 Hz, 1H), 7.91 (d, J = 8.0 Hz, 1H), 7.82 (d, J = 7.6 Hz, 1H), 7.78 – 7.42 (m, 7H), 7.31 (d, J = 8.0 Hz, 1H), 4.28 (s, 2H), 3.93 – 3.40 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 172.09, 163.24, 150.86, 148.13, 141.18, 140.45, 138.16, 134.22, 133.72, 132.07, 131.28, 130.88, 129.12, 128.33, 126.57, 125.78, 124.02, 123.31, 123.04, 122.89, 122.23, 121.64, 121.02, 120.58, 50.82, 48.54. ESI-MS: (m/z) calcd. for C₂₉H₂₅N₃O, 431.19, found: 432.33 (M + H)⁺. Anal. Calcd for C₂₉H₂₅N₃O; (%) C 80.72, H 5.84, N 9.74; found: C 80.79, H 5.87, N 9.81. 5.2.10. (E)-1-(4-(phenanthridin-6-yl)piperazin-1-yl)-3-phenylprop-2-en-1-one (6j)

Appearance: off white greenish solid; yield = 72%; mp = 154-156 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 8.58 (d, *J* = 8.4 Hz, 1H), 8.24 (d, *J* = 8.0 Hz, 1H), 8.08 (d, *J* = 7.8 Hz, 1H), 7.88 (d, *J* = 8.0 Hz, 1H), 7.79 (d, *J* = 8.0 Hz, 2H), 7.75 – 7.48 (m, 7H), 7.38 (d, *J* = 15.2 Hz, 1H), 7.29 (d, *J* = 15.8 Hz, 1H), 3.68 – 3.54 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 172.08, 161.22, 154.16, 148.12, 140.82, 138.45, 133.16, 132.84, 131.90, 128.37, 127.78, 125.23, 124.88, 123.65, 122.44, 122.21, 121.84, 121.22, 120.36, 118.41, 51.59, 46.38. ESI-MS: (m/z) calcd. for C₂₆H₂₃N₃O, 393.18, found: 394.27 (M + H)⁺. Anal. Calcd for C₂₆H₂₃N₃O; (%) C 79.36, H 5.89, N 10.68; found: C 79.48, H 5.93, N 10.75.

5.2.11. (4-(phenanthridin-6-yl)piperazin-1-yl)(pyridin-2-yl)methanone (6k)

Appearance: off white semi solid; yield = 78%; mp = ND; ¹H NMR (400 MHz, CDCl₃) δ 8.59 (d, J = 8.8 Hz, 1H), 8.46 (d, J = 8.4 Hz, 1H), 8.25 (d, J = 8.8 Hz, 1H), 8.14 (m, 1H), 7.96 (d, J = 7.8 Hz, 1H), 7.91 (d, J = 8.0 Hz, 1H), 7.82 (d, J = 7.6 Hz, 1H), 7.78 – 7.48 (m, 5H), 3.92 – 3.52 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 171.23, 160.89, 154.04, 151.12, 149.34, 142.81, 140.86, 133.17, 132.34, 130.14, 129.13, 128.47, 126.05, 124.60, 123.37, 122.88, 122.14, 121.22, 120.80, 51.74, 46.58. ESI-MS: (m/z) calcd. for C₂₃H₂₀N₄O, 368.16, found: 369.24 (M + H)⁺. Anal. Calcd for C₂₃H₂₀N₄O; (%) C 74.98, H 5.47, N 15.21; found: C 75.07, H 5.41, N 15.14.

5.2.12. (4-(phenanthridin-6-yl)piperazin-1-yl)(pyridin-3-yl)methanone (61)

Appearance: off white solid; yield = 74%; mp = 153-154 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 8.81 (s, 1H), 8.72 (d, *J* = 8.4 Hz, 2H), 8.64 (d, *J* = 8.0 Hz, 1H), 8.28 (d, *J* = 8.8 Hz, 1H), 7.94 (m, 2H), 7.83 (d, *J* = 8.0 Hz, 1H), 7.78 – 7.51 (m, 4H), 3.98 – 3.44 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 170.84, 162.45, 153.76, 153.23, 151.88, 141.54, 139.72, 132.65, 132.11, 130.81, 129.33, 128.28, 126.32, 124.86, 123.42, 122.46, 122.02, 121.85, 120.43, 52.68, 47.54. ESI-MS: (m/z) calcd. for C₂₃H₂₀N₄O, 368.16, found: 369.28 (M + H)⁺. Anal. Calcd for C₂₃H₂₀N₄O; (%) C 74.98, H 5.47, N 15.21; found: C 75.05, H 5.45, N 15.12.

5.2.13. (4-(phenanthridin-6-yl)piperazin-1-yl)(pyridin-4-yl)methanone (6m)

Appearance: off white semi solid; yield = 81%; mp = ND; ¹H NMR (400 MHz, CDCl₃) δ 8.74 (d, J = 8.8 Hz, 2H), 8.45 (d, J = 8.0 Hz, 1H), 8.18 (d, J = 8.4 Hz, 1H), 7.96 (d, J = 7.8 Hz, 1H), 7.92 (d, J = 8.4 Hz, 2H), 7.82 (d, J = 7.6 Hz, 1H), 7.75 – 7.44 (m, 4H), 3.88 – 3.54 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 172.06, 159.15, 152.74, 150.89, 147.21, 144.39, 139.82, 133.45, 130.23, 128.06, 127.14, 126.23, 124.18, 123.42, 122.01, 121.79, 120.12, 52.36, 47.52. ESI-MS: (m/z) calcd. for $C_{23}H_{20}N_4O$, 368.16, found: 369.23 (M + H)⁺. Anal. Calcd for $C_{23}H_{20}N_4O$; (%) C 74.98, H 5.47, N 15.21; found: C 75.12, H 5.40, N 15.16.

5.2.14. 6-(4-(phenanthridin-6-yl)piperazine-1-carbonyl)picolinic acid (6n)

Appearance: off white semi solid; yield = 65%; mp = ND; ¹H NMR (400 MHz, DMSO-d₆) δ 11.58 (s, 1H), 8.88 (d, J = 8.4 Hz, 1H), 8.82 (d, J = 9.2 Hz, 1H), 8.61 (m, 1H), 8.57 (d, J = 8.4 Hz, 1H), 8.26 (d, J = 8.0 Hz, 1H), 8.04 (d, J = 8.4 Hz, 1H), 7.87 (d, J = 7.8 Hz, 1H), 7.79 – 7.46 (m, 4H), 3.82 – 3.52 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 172.36, 171.81, 160.12, 152.45, 150.86, 148.32, 143.75, 140.48, 135.08, 134.53, 131.24, 129.04, 128.14, 126.87, 124.12, 123.44, 122.81, 122.63, 121.07, 120.24, 52.72, 47.50. ESI-MS: (m/z) calcd. for C₂₄H₂₀N₄O₃, 412.15, found: 413.28 (M + H)⁺. Anal. Calcd for C₂₄H₂₀N₄O₃; (%) C 69.89, H 4.89, N 13.58; found: C 69.98, H 4.95, N 13.64.

5.2.15. furan-2-yl(4-(phenanthridin-6-yl)piperazin-1-yl)methanone (60)

Appearance: off white solid; yield = 76%; mp = 118-119 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.59 (d, J = 8.8 Hz, 1H), 8.44 (d, J = 8.0 Hz, 1H), 8.18 (d, J = 8.4 Hz, 1H), 8.02 (d, J = 8.8 Hz, 1H), 7.92 (d, J = 8.0 Hz, 1H), 7.86 – 7.54 (m, 4H), 7.43 (d, J = 7.8 Hz, 1H), 7.22 (m, 1H), 4.94 – 3.68 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 169.74, 157.68, 148.15, 142.44, 135.22, 133.45, 130.87, 130.12, 129.18, 128.84, 128.04, 127.21, 126.90, 126.12, 125.18, 122.18, 121.98, 118.24, 51.68, 48.84. ESI-MS: (m/z) calcd. for C₂₂H₁₉N₃O₂, 357.14, found: 358.25 (M + H)⁺. Anal. Calcd for C₂₂H₁₉N₃O₂; (%) C 73.93, H 5.36, N 11.76; found: C 73.99, H 5.41, N 11.83.

5.2.16. (4-(phenanthridin-6-yl)piperazin-1-yl)(thiophen-2-yl)methanone (6p)

Appearance: white solid; yield = 81%; mp = 143-145 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 8.72 (d, J = 8.8 Hz, 1H), 8.58 (d, J = 8.0 Hz, 1H), 8.42 (d, J = 8.0 Hz, 1H), 8.23 (d, J = 8.0 Hz, 1H), 8.08 (d, J = 8.4 Hz, 1H), 7.90 (d, J = 8.0 Hz, 1H), 7.88 – 7.37 (m, 5H), 4.89 – 3.53 (t, 8H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 170.02, 158.88, 149.22, 141.89, 136.72, 131.93, 130.12, 129.94, 129.34, 128.84, 128.11, 127.26, 126.95, 125.87, 125.03, 122.48, 121.68, 120.87, 52.46, 47.52. ESI-MS: (m/z) calcd. for C₂₂H₁₉N₃OS, 373.12, found: 374.27 (M + H)⁺. Anal. Calcd for C₂₂H₁₉N₃OS; (%) C 70.75, H 5.13, N 11.25; found: C 70.83, H 5.19, N 11.15.

5.2.17. 2-(1H-indol-3-yl)-1-(4-(phenanthridin-6-yl)piperazin-1-yl)ethanone (6q)

Appearance: off white solid; yield = 80%; mp = 162-163 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 8.24 (d, J = 8.0 Hz, 1H), 8.08 (d, J = 7.6 Hz, 1H), 7.91 (d, J = 8.0 Hz, 1H), 7.88 (d, J = 8.0 Hz, 1H), 7.84 (m, 1H), 7.79 (d, J = 7.2 Hz, 1H), 7.58 – 7.48 (m, 3H), 7.42 (d, J = 8.0 Hz,

1H), 7.34 (s, 1H), 7.20 (m, 2H), 6.08 (br, 1H), 3.88 (t, 4H), 3.80 (s, 2H), 3.52 (t, 4H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 168.54, 159.62, 144.83, 143.23, 140.23, 138.64, 135.49, 135.05, 133.63, 132.84, 129.47, 128.43, 127.19, 126.82, 126.11, 124.46, 123.23, 121.52, 119.88, 119.04, 117.74, 108.04, 52.64, 48.48, 40.68. ESI-MS: (m/z) calcd. for C₂₇H₂₄N₄O 420.19, found: 421.15 (M + H)⁺. Anal. Calcd for C₂₇H₂₄N₄O: (%) C 77.12, H 5.75, N 13.32; Found: C 77.24, H 5.88, N 13.41.

5.2.18. 3-(1H-indol-3-yl)-1-(4-(phenanthridin-6-yl)piperazin-1-yl)propan-1-one (6r)

Appearance: off white solid; yield = 83%; mp = 120-121 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 8.19 (d, J = 8.0 Hz, 1H), 8.02 (d, J = 7.8 Hz, 1H), 7.94 (d, J = 7.6 Hz, 1H), 7.86 (d, J = 7.2 Hz, 1H), 7.80 (m, 1H), 7.76 (d, J = 7.8 Hz, 1H), 7.61 – 7.44 (m, 3H), 7.41 (d, J = 8.0 Hz, 1H), 7.38 (s, 1H), 7.22 (m, 2H), 6.84 (br, 1H), 3.92 – 2.70 (t, 12H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 170.22, 162.48, 145.16, 144.06, 141.18, 139.21, 136.42, 135.84, 133.82, 133.08, 128.92, 128.11, 127.64, 126.74, 126.32, 124.22, 123.68, 120.94, 119.24, 118.48, 117.38, 110.28 52.12, 47.39, 39.94, 32.98. ESI-MS: (m/z) calcd. for C₂₈H₂₆N₄O 434.21, found: 435.28 (M + H)⁺. Anal. Calcd for C₂₈H₂₆N₄O: (%) C 77.39, H 6.03, N 12.89; Found: C 77.46, H 6.12, N 12.94.

5.2.19. 4-(1H-indol-3-yl)-1-(4-(phenanthridin-6-yl)piperazin-1-yl)butan-1-one (6s)

Appearance: off white solid; yield = 75%; mp = 148-149 °C; ¹H NMR (400 MHz, DMSO-d₆) δ 8.22 (d, *J* = 8.0 Hz, 1H), 8.07 (d, *J* = 7.6 Hz, 1H), 7.98 (d, *J* = 7.8 Hz, 1H), 7.89 (d, *J* = 7.4 Hz, 1H), 7.84 (m, 1H), 7.72 (d, *J* = 7.4 Hz, 1H), 7.62 – 7.39 (m, 3H), 7.35 (d, *J* = 8.0 Hz, 1H), 7.30 (s, 1H), 7.18 (m, 2H), 6.42 (br, 1H), 3.89 – 2.60 (t, 12H), 1.68 (m, 2H). ¹³C NMR (100.61 MHz, DMSO-d₆) δ 170.84, 163.04, 144.87, 143.21, 141.85, 139.42, 135.08, 134.91, 133.09, 132.44, 129.48, 128.74, 127.04, 126.23, 123.11, 122.41, 122.26, 121.39, 120.12, 119.94, 118.87, 111.46, 52.88, 46.24, 40.37, 30.42, 39.87. ESI-MS: (m/z) calcd. for C₂₉H₂₈N₄O 448.22, found: 449.24 (M + H)⁺. Anal. Calcd for C₂₉H₂₈N₄O: (%) C 77.65, H 6.29, N 12.49; Found: C 77.72, H 6.38, N 12.38.

5.2.20. 6-(4-(methylsulfonyl)piperazin-1-yl)Phenanthridine (7a):

Appearance: off white semi solid; yield = 78%; mp = ND; ¹H NMR (400 MHz, CDCl₃) δ 8.26 (d, J = 8.0 Hz, 1H), 8.13 (d, J = 7.8 Hz, 1H), 7.97 (d, J = 8.0 Hz, 1H), 7.81 (d, J = 7.4 Hz, 1H), 7.74 – 7.41 (m, 4H), 3.87 – 3.53 (t, 8H), 3.09 (s, 3H). ¹³C NMR (100.61 MHz, CDCl₃) δ 164.24, 148.76, 140.08, 133.13, 131.67, 130.43, 129.88, 123.24, 122.41, 121.82, 120.34, 119.92, 114.34, 53.65, 48.21, 38.98. ESI-MS: (m/z) calcd. for C₁₈H₁₉N₃O₂S 341.12, found: 342.14 $(M + H)^+$. Anal. Calcd for $C_{18}H_{19}N_3O_2S$: (%) C 63.32, H 5.61, N 12.31; found: C 63.64, H 5.74, N 12.28.

5.2.21. 6-(4-(phenylsulfonyl)piperazin-1-yl)phenanthridine (7b)

Appearance: off white solid; yield = 88%; mp = 238-239 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.57 (d, *J* = 8.0 Hz, 1H), 8.41 (d, *J* = 7.6 Hz, 1H), 8.02 (d, *J* = 7.8 Hz, 2H), 7.90 (dd, *J* = 9.0 Hz, *J* = 0.8 Hz, 1H), 7.82 (d, *J* = 7.4 Hz, 1H), 7.77 – 7.39 (m, 7H), 3.68 – 3.44 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 162.86, 152.08, 141.12, 138.47, 132.24, 131.82, 130.12, 129.64, 124.28, 123.81, 122.89, 121.24, 121.08, 120.42, 119.92, 118.42, 112.47, 50.48, 48.07. ESI-MS: (m/z) calcd. for C₂₃H₂₁N₃O₂S 403.13, found: 403.22 (M + H)⁺. Anal. Calcd for C₂₃H₂₁N₃O₂S: (%) C 68.46, H 5.25, N 10.41; found: C 68.55, H 5.32, N 10.60.

5.2.22. 6-(4-tosylpiperazin-1-yl)phenanthridine (7c)

Appearance: pale yellow solid; yield = 93%; mp = 188-189 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.48 (d, J = 8.2 Hz, 1H), 8.01 (d, J = 7.6 Hz, 1H), 7.91 (dd, J = 8.4 Hz, J = 0.8 Hz, 1H), 7.88 (d, J = 8.4 Hz, 1H), 7.80 (d, J = 7.8 Hz, 2H), 7.79 (d, J = 8.0 Hz, 2H), 7.74 – 7.46 (m, 4H), 3.64 – 3.38 (t, 8H), 2.94 (s, 3H). ¹³C NMR (100.61 MHz, CDCl₃) δ 162.42, 153.14, 144.46, 139.08, 138.81, 132.11, 131.81, 129.64, 128.43, 124.82, 123.08, 122.28, 121.63, 120.42, 119.92, 118.42, 113.22, 52.18, 47.11, 24.08. ESI-MS: (m/z) calcd. for C₂₄H₂₃N₃O₂S 417.15, found: 417.24 (M + H)⁺. Anal. Calcd for C₂₄H₂₃N₃O₂S: (%) C 69.04, H 5.55, N 10.06; found: C 69.11, H 5.64, N 10.12.

5.2.23. 6-(4-(4-tert-butylphenylsulfonyl)piperazin-1-yl)phenanthridine (7d)

Appearance: white solid; yield = 88%; mp = 154-155 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.58 (d, *J* = 8.0 Hz, 1H), 8.45 (d, *J* = 8.2 Hz, 1H), 8.04 (d, *J* = 8.0 Hz, 1H), 7.91 (dd, *J* = 8.4 Hz, *J* = 0.8 Hz, 1H), 7.78 (dd, *J* = 6.4 Hz, *J* = 1.6 Hz, 1H), 7.74 (dd, *J* = 8.8 Hz, *J* = 1.6 Hz, 1H), 7.76 (m, 1H), 7.74 (d, *J* = 7.2 Hz, 1H), 7.64 – 7.55 (m, 4H), 7.51 (d, *J* = 8.0 Hz, 1H), 3.60 – 3.35 (t, 8H), 1.37 (s, 9H). ¹³C NMR (100.61 MHz, CDCl₃) δ 159.24, 156.71, 143.45, 134.98, 132.83, 130.30, 128.82, 128.62, 127.79, 126.81, 126.17, 125.95, 125.14, 122.85, 122.65, 121.86, 121.03, 50.48, 46.03, 35.23, 31.13. ESI-MS: (m/z) calcd. for C₂₇H₂₉N₃O₂S 459.19, found: 460.25 (M + H)⁺. Anal. Calcd for C₂₇H₂₉N₃O₂S: (%) C 70.56, H 6.36, N 9.14; found: C 70.64, H 6.28, N 9.24.

5.2.24. 6-(4-(mesitylsulfonyl)piperazin-1-yl)phenanthridine (7e)

Appearance: white solid; yield = 81%; mp = 118-119 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.48 (d, *J* = 8.0 Hz, 1H), 8.24 (d, *J* = 8.0 Hz, 1H), 7.98 (dd, *J* = 8.8 Hz, *J* = 1.6 Hz, 1H), 7.82 (dd, *J* = 7.2 Hz, *J* = 1.2 Hz, 1H), 7.72 – 7.58 (m, 4H), 7.24 (s, 2H), 3.62 – 3.42 (t, 8H), 2.88 – 2.62

(s, 9H). ¹³C NMR (100.61 MHz, CDCl₃) δ 162.38, 157.24, 148.12, 145.31, 139.45, 138.08, 131.78, 131.41, 130.21, 127.09, 125.22, 124.62, 123.26, 122.08, 121.96, 121.14, 120.42, 51.04, 47.21, 31.08, 30.88. ESI-MS: (m/z) calcd. for C₂₆H₂₇N₃O₂S, 445.18, found: 446.23 (M + H)⁺. Anal. Calcd for C₂₆H₂₇N₃O₂S; (%) C 70.08, H 6.11, N 9.43; found: C 70.14, H 6.23, N 9.59.

5.2.25. 6-(4-(4-chloro-2,5-dimethylphenylsulfonyl)piperazin-1-yl)phenanthridine (7f)

Appearance: white solid; yield = 87%; mp = 163-164 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.58 (d, *J* = 8.4 Hz, 1H), 8.46 (d, *J* = 8.0 Hz, 1H), 8.12 (d, *J* = 8.8 Hz, 1H), 7.91 (d, *J* = 7.8 Hz, 1H), 7.85 (s, 1H), 7.78 – 7.53 (m, 4H), 7.34 (s, 1H), 3.56 – 3.46 (t, 8H), 2.62 – 2.42 (s, 6H). ¹³C NMR (100.61 MHz, CDCl₃) δ 164.21, 158.73, 149.08, 142.74, 141.03, 139.22, 137.63, 137.24, 134.09, 131.11, 130.87, 129.32, 128.08, 126.43, 123.28, 122.92, 121.03, 120.25, 119.46, 51.82, 47.04, 36.42, 30.12. ESI-MS: (m/z) calcd. for C₂₅H₂₄ClN₃O₂S, 465.12, found: 466.15 (M + H)⁺. Anal. Calcd for C₂₅H₂₄ClN₃O₂S; (%) C 64.44, H 5.19, N 9.02; found: C 64.58, H 5.30, N 9.14.

5.2.26. 6-(4-(4-fluorophenylsulfonyl)piperazin-1-yl)phenanthridine (7g)

Appearance: pale yellow solid; yield = 74%; mp = 188-189 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.57 (d, *J* = 8.2 Hz, 1H), 8.42 (d, *J* = 8.0 Hz, 1H), 8.03 (d, *J* = 8.2 Hz, 1H), 7.90 (d, *J* = 8.2 Hz, 1H), 7.48 – 7.29 (m, 8H), 3.64 – 3.32 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 161.86, 160.08, 151.11, 149.23, 138.14, 137.67, 135.62, 132.16, 131.23, 128.32, 126.45, 124.82, 123.88, 122.44, 121.21, 120.08, 114.05, 52.24, 46.42. ESI-MS: (m/z) calcd. for C₂₃H₂₀FN₃O₂S, 421.12, found: 422.25 (M + H)⁺. Anal. Calcd for C₂₃H₂₀FN₃O₂S; (%) C 65.54, H 4.78, N 9.97; found: C 65.62, H 4.88, N 10.08.

5.2.27. 6-(4-(4-chlorophenylsulfonyl)piperazin-1-yl)phenanthridine (7h)

Appearance: white solid; yield = 82%; mp = 196-197 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.59 (d, J = 10.8 Hz, 1H), 8.46 (dd, J = 10.8 Hz, J = 1.6 Hz 1H), 8.25 (dd, J = 11.2 Hz, J = 1.2 Hz 1H), 7.96 (dd, J = 10.8 Hz, J = 1.2 Hz 1H), 7.82 – 7.48 (m, 4H), 7.28 (d, J = 6.8 Hz, 1H), 7.24 (d, J = 7.2 Hz, 1H), 3.67 – 3.44 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 163.14, 159.28, 150.88, 142.23, 139.86, 134.07, 132.47, 131.52, 129.71, 128.91, 127.20, 124.88, 124.25, 123.62, 122.89, 121.03, 120.21, 51.72, 46.05. ESI-MS: (m/z) calcd. for C₂₃H₂₀ClN₃O₂S; 437.09, found: 438.15 (M + H)⁺. Anal. Calcd for C₂₃H₂₀ClN₃O₂S; (%) C 63.08, H4.60, N 9.59; found: 63.19, H4.64, N 9.65.

5.2.28. 6-(4-(4-bromophenylsulfonyl)piperazin-1-yl)phenanthridine (7i)

Appearance: pale yellow solid; yield = 78%; mp = 224-225 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.64 (d, J = 8.4 Hz, 1H), 8.61 (d, J = 7.6 Hz, 1H), 8.08 (d, J = 8.4 Hz, 1H), 7.88 (m, 1H), 7.84 (d, J = 7.2 Hz, 1H), 7.82 – 7.51 (m, 5H), 7.22 (d, J = 7.2 Hz, 2H), 3.51 – 3.20 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 163.80, 159.12, 148.46, 144.02, 139.14, 138.07, 134.61, 133.72, 131.98, 131.34, 127.45, 126.82, 123.74, 122.46, 121.84, 121.03, 120.42, 51.68, 47.04. ESI-MS: (m/z) calcd. for C₂₃H₂₀BrN₃O₂S, 481.04, found: 482.13 (M + H)⁺. Anal. Calcd for C₂₃H₂₀BrN₃O₂S; (%) C 57.27, H 4.18, N 8.71; found: C 57.34, H 4.23, N 8.85.

5.2.29. 6-(4-(4-(trifluoromethyl)phenylsulfonyl)piperazin-1-yl)phenanthridine (7j)

Appearance: pale yellow solid; yield = 74%; mp = 163-164 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.57 (d, *J* = 8.2 Hz, 1H), 8.44 (d, *J* = 7.6 Hz, 1H), 8.02 (d, *J* = 8.4 Hz, 1H), 7.98 – 7.49 (m, 9H), 3.63 – 3.38 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 161.24, 156.68, 149.12, 142.81, 138.37, 134.12, 131.28, 130.98, 130.12, 129.35, 128.84, 128.02, 126.87, 124.07, 123.89, 121.25, 120.81, 119.48, 52.38, 48.65. ESI-MS: (m/z) calcd. for C₂₄H₂₀F₃N₃O₂S; 471.12, found: 472.18 (M + H)⁺. Anal. Calcd for C₂₄H₂₀F₃N₃O₂S; (%) C 61.14, H 4.28, N 8.91; found: C 61.18, H 4.31, N 8.86.

5.2.30. 6-(4-(4-methoxyphenylsulfonyl)piperazin-1-yl)phenanthridine (7k)

Appearance: pale yellow solid; yield = 78%; mp = 208-209 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.76 (d, J = 8.4 Hz, 1H), 8.63 (d, J = 7.6 Hz, 1H), 8.09 (d, J = 8.4 Hz, 1H), 7.88 (m, 1H), 7.82 (dd, J = 8.4 Hz, J = 1.2 Hz, 1H), 7.78 – 7.74 (m, 2H), 7.69 (dd, J = 8.0 Hz, J = 0.8 Hz, 1H), 7.65 (dd, J = 8.0 Hz, J = 0.8 Hz, 1H), 7.55 (m, 1H), 7.22 (d, J = 7.2 Hz, 2H), 3.88 (s, 3H), 3.47 – 3.32 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 164.08, 160.28, 151.42, 140.23, 133.86, 132.46, 131.27, 130.52, 128.74, 127.91, 126.24, 124.88, 124.21, 123.63, 122.87, 120.03, 118.77, 51.78, 46.53. ESI-MS: (m/z) calcd. for C₂₄H₂₃N₃O₃S; 433.14, found: 434.23 (M + H)⁺. Anal. Calcd for C₂₄H₂₃N₃O₃S; (%) C 66.49, H 5.35, N 9.69; found: C 66.54, H 5.42, N 9.77.

5.2.31. 6-(4-(4-nitrophenylsulfonyl)piperazin-1-yl)phenanthridine (71)

Appearance: pale yellow solid; yield = 81%; mp = 198-199 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.57 (d, *J* = 8.4 Hz, 1H), 8.45 (d, *J* = 7.8 Hz, 3H), 8.06 (d, *J* = 8.4 Hz, 1H), 8.02 (d, *J* = 8.4 Hz, 2H), 7.91 (dd, *J* = 8.0 Hz, *J* = 0.8 Hz, 1H), 7.79 – 7.48 (m, 4H), 3.64 – 3.38 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 161.25, 154.71, 146.36, 137.11, 132.87, 132.23, 131.08, 128.52, 127.79, 126.82, 126.18, 125.94, 125.14, 123.65, 121.86, 121.02, 120.48, 50.79, 47.04. ESI-MS: (m/z) calcd. for C₂₃H₂₀N₄O₄S; 448.12, found: 449.22 (M + H)⁺. Anal. Calcd for C₂₃H₂₀N₄O₄S; (%) C 61.59, H 4.49, N 12.49; found: C 61.64, H 4.61, N 12.54.

5.2.32. 1-(4-(4-(phenanthridin-6-yl)piperazin-1-ylsulfonyl)phenyl)ethanone (7m)

Appearance: off white solid; yield = 76%; mp = 204-205 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.60 (d, J = 8.8 Hz, 1H), 8.46 (d, J = 8.0 Hz, 2H), 8.40 (d, J = 8.2 Hz, 1H), 8.12 (d, J = 8.4 Hz, 1H), 8.04 (d, J = 8.0 Hz, 2H), 7.88 (dd, J = 8.0 Hz, J = 1.2 Hz, 1H), 7.81 – 7.44 (m, 4H), 3.64 – 3.38 (t, 8H), 2.72 (s, 3H). ¹³C NMR (100.61 MHz, CDCl₃) δ 171.47, 160.68, 152.23, 145.22, 138.22, 132.46, 132.67, 130.97, 129.42, 128.84, 127.46, 126.17, 124.24, 124.08, 123.68, 121.22, 120.88, 120.56, 50.64, 46.03, 29.44. ESI-MS: (m/z) calcd. for C₂₅H₂₃N₃O₃S; 445.14, found: 446.25 (M + H)⁺. Anal. Calcd for C₂₅H₂₃N₃O₃S; (%) C 67.40, H 5.20, N 9.43; found: C 67.49, H 5.24, N 9.47.

5.2.33. 6-(4-(5-bromothiophen-2-ylsulfonyl)piperazin-1-yl)phenanthridine (7n)

Appearance: pale yellow solid; yield = 80%; mp = 162-163 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.58 (d, *J* = 8.4 Hz, 1H), 8.45 (d, *J* = 7.8 Hz, 1H), 8.08 (d, *J* = 8.2 Hz, 1H), 8.93 (d, *J* = 8.0 Hz, 1H), 7.81 – 7.50 (m, 4H), 7.38 (d, *J* = 7.2 Hz, 1H), 7.16 (d, *J* = 7.4 Hz, 1H), 3.66 – 3.40 (t, 8H). ¹³C NMR (100.61 MHz, CDCl₃) δ 162.24, 151.47, 144.78, 139.41, 134.89, 133.01, 131.22, 128.41, 126.84, 126.12, 125.08, 124.86, 123.25, 123.11, 122.47, 120.78, 119.23, 50.48, 46.14. ESI-MS: (m/z) calcd. For C₂₁H₁₈BrN₃O₂S₂; 487.00, found: 487.13 (M + H)⁺. Anal. Calcd for C₂₁H₁₈BrN₃O₂S₂; (%) C 51.64, H 3.71, N 8.60; found: C 51.78, H 3.75, N 8.69.

5.3. Microplate Alamar Blue Assay (MABA)

The anti-mycobacterial activities of title compounds **6a-s** and **7a-n** were evaluated against MTB H37Rv (ATCC 27294) strain using MABA [27,28]. 200 μ L of sterile deionized water was added to all outer-perimeter wells of sterile 96-well plates to minimize evaporation of the medium in the test wells during incubation. The wells in rows B to G in columns 3 to 11 received 100 μ l of 7H9GC broth. 100 μ L of 2× drug solutions were added to the wells in rows B to G in columns 2 and 3. By using a multichannel pipette, 100 μ l was transferred from column 3 to column 4, and the contents of the wells were mixed well. Identical serial 1:2 dilutions were continued through column 10, and 100 μ l of excess medium was discarded from the wells in column 10.

 100μ L of MTB inoculum was added to the wells in rows B to G in columns 2 to 10. Add 100μ L of medium to B11 and C11 (media control), 100μ L of MTB inoculum to D11 and E11 and 100μ L of MTB inoculum with 3-5% DMSO to F11 and G11 (solvent control).

The plates were sealed with parafilm and were incubated at $37^{\circ}C$ for 5 days. $50\mu L$ of a freshly prepared 1:1 mixture of $10\times$ Alamar Blue (Accumed International, Westlake, Ohio)

reagent and 10% Tween 80 was added to well D11. The plates were reincubated at 37°C for 24 h. If well D11 turned pink, the reagent mixture was added to all wells in the microplate (if the well remained blue, the reagent mixture would be added to another control well and the result would be read on the following day). The microplates were resealed with parafilm and were incubated for an additional 24 h at 37°C, and the colors of all wells were recorded. A blue color in the well was interpreted as no growth, and a pink color was scored as growth. A few wells appeared violet after 24 h of incubation, but they invariably changed to pink after another day of incubation and thus were scored as growth (while the adjacent blue wells remained blue). The MIC was defined as the lowest drug concentration which prevented a color change from blue to pink.

5.4. Molecular modelling studies:

The 2D structures of synthesized compounds (6a-s and 7a-n) were sketched and converted to 3D using Ligprep, module of Schrodinger suite package 2013. ATPase domain of Mycobacterium tuberculosis GyrB protein was selected and prepared for docking by protein preparation wizard which was a part of the Maestro software [29]. Bond orders and formal charges were added for hetero groups, and hydrogens were added to all atoms in the system. Water molecules within 5 Å distance were removed. For each structure, a brief relaxation was performed using an all-atom constrained minimization carried out with the Impact Refinement module (Impref). (Impact v5.8, Schro"dinger, LLC, New York, NY) using the OPLS-2005 force field to alleviate steric clashes that may exist in the original PDB structure. The minimization was terminated when the energy converged or the RMSD reached a maximum cut off of 0.30 Å. The binding site was defined by a rectangular box surrounding the ANP and coordinates of grid box were X = -21.987185, Y = 26.604429, Z = -50.838890. Prepared ligands were docked into the ANP binding pocket of MTB GyrB protein with Glide 5.0 (Glide v6.0 Schrodinger, LLC, New York, NY) [30]. Glide energy grids were generated for each of the prepared complexes. Glide XP (Extra Precision) docking with ligand flexibility was performed. A total of ten ligand conformations were allowed and finally top score conformation was selected as active conformation. Molecules were analyzed based on docking score, interacting amino acids and hydrogen bonds.

5.5. Cytotoxicity assay

Most active anti-TB compounds (MIC $\leq 12.5 \ \mu g/mL$) were further examined for toxicity in RAW 264.7 cell lines at the concentration of 50 $\mu g/mL$. After 72 h of exposure, viability was

assessed on the basis of cellular conversion of MTT into a formazan product using the Promega Cell Titer 96 non-radioactive cell proliferation assay [31].

Acknowledgements

The financial assistance provided by Department of Biotechnology, (No.BT/PR4801/MED/29/370/2012), New Delhi, India is gratefully acknowledged.

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Figure 1: Design strategy to achieve title compounds

Figure 2: Interaction profile of ANP with ATPase domain of *Mycobacterium tuberculosis* GyrB protein. The hydrophobic pocked is highlighted with the residues interacting with the crystal ligand

Figure 3: Interaction profile of compound **6b** with ATPase domain of *Mycobacterium tuberculosis* GyrB protein. The various hydrophobic interactions of compound **6b** are noticeable.

Figure 4: Interaction profile of compound **7d** with ATPase domain of *Mycobacterium tuberculosis* GyrB protein. The different polar contacts, π - π interaction and the hydrophobic interactions with compound **7d** are noticeable

Figure 5: ORTEP plot for the compound **6b** and **7d**. All the non-hydrogen atoms are presented by their 50% probability thermal ellipsoids

Scheme 1: General synthetic procedure for title compounds

Table 1: Optimisation of reaction conditions of **6b**^{*a*}

Table 2: Antimycobacterial activities of compound 6a-s and 7a-n against MTB H37Rv

Table 3: IC_{50} (µg/mL) and SI values of active compounds

Table 4: Crystal data and structure refinement details for 6b and 7d

Entry	Solvent	Base	Reagent	Temperature (°C)	Time	Yield $(\%)^b$
1	DCM	TEA	EDC.HCl, HOBt	rt	8 h	58
2	DMF	TEA	EDC.HCl, HOBt	rt	8 h	52
3	DMF	DIPEA	HATU	rt	8 h	41
4	DMF	DIPEA	BOP	rt	8 h	42
5	DMF	DIPEA	PyBOP	rt	8 h	38
6	DCM	Pyridine	T ₃ P	rt	4 h	54
7	DCM	TEA	T ₃ P	rt	2 h	56
8	DCM	TEA	T ₃ P	rt	4 h	68
9	DCM	TEA	T_3P	rt	6 h	84
10	DCM	TEA	T ₃ P	rt	8 h	85
11	DCM	TEA	T ₃ P	rt	12 h	84

Table 1: Optimisation of reaction conditions of $\mathbf{6b}^a$

) /

^{*a*}All the reactions were carried out with **5** (1.0 equiv.), base (2.0 equiv.) and reagent (1.0-1.6 equiv.) in solvent, ^{*b*}Isolated yield after column chromatography.

S.No.	Entry	$\mathbf{R}_1/\mathbf{R}_2$	MIC(µg/mL) against MTB H ₃₇ Rv strain
1	ба	N	25
2	6b		3.13
3	6с	CI	12.5
4	6d	0 ₂ N-	50
5	6e		3.13
6	6f		25
7	6g	NH ₂	>50
8	6h	H ₂ N-	50
9	6i		>50
10	6ј		12.5
11	6k		3.13
12	61		6.25
13	6m		1.56
14	6n		3.13

Table 2: Antimycobacterial activities of	f compounds 6a-s and 7a-n	against MTB H37Rv

15	60		25
16	6р		50
17	6q	HN	25
18	6r	HN	6.25
19	6s	HN	1.56
20	7a	H ₃ C	25
21	7b		6.25
22	7c		>50
23	7d		1.56
24	7e		12.5
25	7f	ci	6.25
26	7g	F	6.25
27	7h	CI	3.13
28	7i	Br	3.13
29	7j	F ₃ C	25

30	7k		6.25	
31	71	0 ₂ N-	50	
32	7m		25	
33	7n	Br	3.13	
- - -	Isoniazid Rifampicin Pyrazinamide		0.1 0. 2 6.25	
		Ar		
	A			
	R			
R.				
F				

ATB H37Rv 3.13 12.5 3.13 12.5 3.13 6.25 1.56 3.13 6.25	at 50 µg/mL 22.16 26.50 34.64 22.12 42.16 19.16 27.90 26.52	approximation 112.81 94.33 72.17 113.01 59.29 130.48 89.60	values 36.15 7.54 23.13 9.04 19.00 20.87 57.43
12.5 3.13 12.5 3.13 6.25 1.56 3.13	26.50 34.64 22.12 42.16 19.16 27.90	94.33 72.17 113.01 59.29 130.48 89.60	7.54 23.13 9.04 19.00 20.87
3.13 12.5 3.13 6.25 1.56 3.13	34.64 22.12 42.16 19.16 27.90	72.17 113.01 59.29 130.48 89.60	23.13 9.04 19.00 20.87
12.5 3.13 6.25 1.56 3.13	22.12 42.16 19.16 27.90	113.01 59.29 130.48 89.60	9.04 19.00 20.87
3.13 6.25 1.56 3.13	42.16 19.16 27.90	59.29 130.48 89.60	19.00 20.87
6.25 1.56 3.13	19.16 27.90	130.48 89.60	20.87
1.56 3.13	27.90	89.60	
3.13			57.43
	26.52		
6.25		94.26	30.21
	34.64	72.17	11.54
1.56	34.12	73.27	46.96
6.25	30.72	81.38	13.02
1.56	28.92	86.44	55.41
12.5	26.52	94.26	7.54
6.25	34.64	72.17	11.54
6.25	32.76	76.31	12.21
3.13	20.92	119.50	38.30
3.13	28.16	88.77	28.45
6.25	26.52	94.26	15.08
3.13	42.16	59.29	38.01
	1.56 12.5 6.25 6.25 3.13 3.13 6.25	1.56 28.92 12.5 26.52 6.25 34.64 6.25 32.76 3.13 20.92 3.13 28.16 6.25 26.52 3.13 42.16	1.56 28.92 86.44 12.5 26.52 94.26 6.25 34.64 72.17 6.25 32.76 76.31 3.13 20.92 119.50 3.13 28.16 88.77 6.25 26.52 94.26 3.13 42.16 59.29

Compound	6b	7d
Formula	C ₂₄ H ₂₁ N ₃ O	$C_{27}H_{29}N_3O_2S$
Sum formula weight	367.44	459.60
Temperature/K	293	293
Crystal system	Triclinic	Monoclinic
Space group	Pī	$P2_{1}/n$
a (Å)	9.4717 (13)	12.5058(13)
b (Å)	9.6936 (14)	14.6774 (17)
c (Å)	11.8432 (16)	13.3524 (14)
Angle α , β , γ (°)	73.767 (12), 74.207 (12), 65.560 (14)	90, 89.451 (9), 90
Volume (Å ³)	935.1 (2)	2450.8(5)
Z	2	4
F(000)	388.0	976.0
$D_{\text{calc }(g/\text{mm}^3)}$	1.305	1.246
$\mu (mm^{-1})$	0.081	0.16
Absorption correction:	Multi-scan	Multi-scan
Radiation wavelength (Å)	0.71073	0.71073
Radiation type	MoK_{α}	MoK_{α}
Radiation monochromator	Graphite	Graphite
$(h, k, l)_{max}$, $(h, k, l)_{min}$	(12, 13, 14), (-12, -13, -14)	(17, 20, 15), (-9, -16, -18)
2Θ range for data collection	$6.6 - 58.2^{\circ}$	$6.4 - 58.4^{\circ}$
<i>T</i> min, <i>T</i> max	0.246, 1.000	0.922, 1.000
No. of measured, independent	6305, 5019, 2213	7784, 6663, 3305
and observed $[I > 2\sigma(I)]$		
reflections	0.038	0.018
$ \begin{array}{c} \mathbf{R}_{\text{int}} \\ (\sin \theta / \lambda)_{\text{max}} (\text{\AA}^{-1}) \end{array} $	0.684	0.687
$(\sin \theta/\lambda)_{max}$ (A) Crystal size/mm ³	0.084 $0.6 \times 0.5 \times 0.4$	$0.6 \times 0.5 \times 0.4$
$R[F^2 > 2\sigma(F^2)], wR(F^2), S$	0.0757 (2213), 0.2206	0.050 (3305), 0.121
$\mathbf{K}[\mathbf{I}^{*} \geq 2\mathbf{U}(\mathbf{I}^{*})], \ \mathbf{W}\mathbf{K}(\mathbf{\Gamma}^{*}), \ \mathbf{S}$		
No. of parameters	(4075), 0.95 253	(5149), 1.02 298
$\Delta \rho_{\text{max}}, \Delta \rho_{\text{min}} (e \text{ Å}^{-3})$	0.27, -0.28	0.14, -0.25
Structure refinement	SHELXL97 (Sheldrick, 1	,
Suucluie ieimememem	STIELAL97 (SHEIUIICK,	2000)

 Table 4: Crystal data and structure refinement details for 6b and 7d

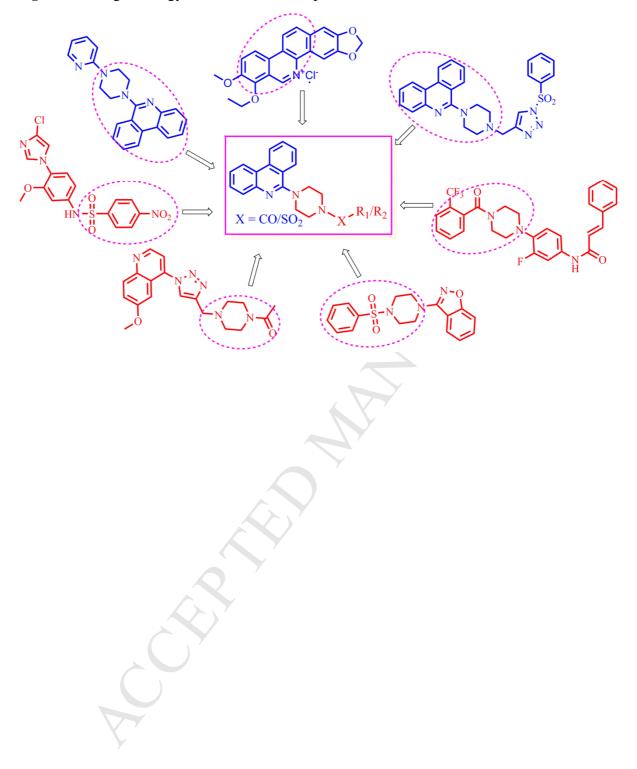
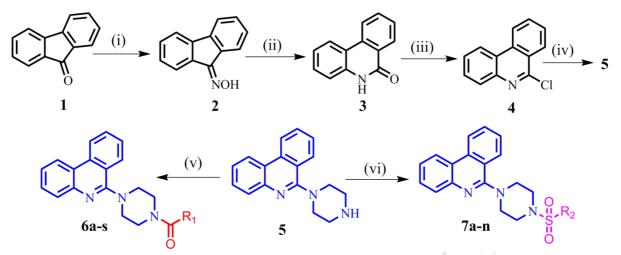


Figure 1: Design strategy to achieve title compounds

Scheme 1: General synthetic procedure to attain title compounds



Reagents and conditions: (i) NH₂OH.HCl (1.2 equiv), NaOAc (1.2 equiv), EtOH:H₂O (2:1), rt 2 h (ii) PPA (8.0 equiv), P₂O₅ (0.5 equiv), heating at 150 °C, 1 h (iii) POCl₃ (6.0 equiv), *N*,*N*-dimethylaniline (0.5 equiv), reflux 4h (iv) piperazine (8.0 equiv), Et₃N (2.0 equiv), DMF, MW, 700 Watt, 15 Min (v) substituted alkyl/aryl acids (1.05 equiv), T₃P (1.2 equiv), Et₃N (2.0 equiv), CH₂Cl₂, rt, 6–8 h (vi) substituted alkyl/aryl sulfonyl chlorides (1.1 equiv), Et₃N (1.2 equiv), CH₂Cl₂, rt, 1–2 h.

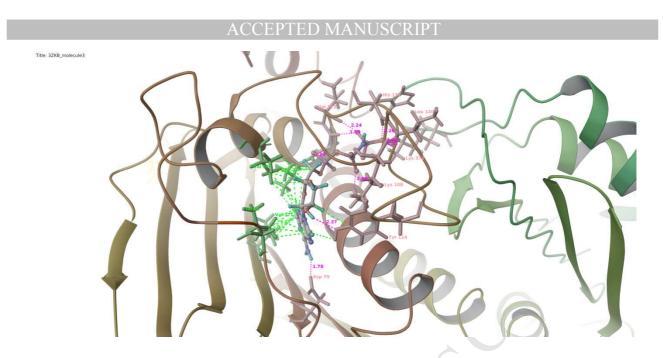


Figure 2: Interaction profile of ANP with ATPase domain of *Mycobacterium tuberculosis* GyrB protein. The hydrophobic pocked is highlighted with the residues interacting with the crystal ligand.

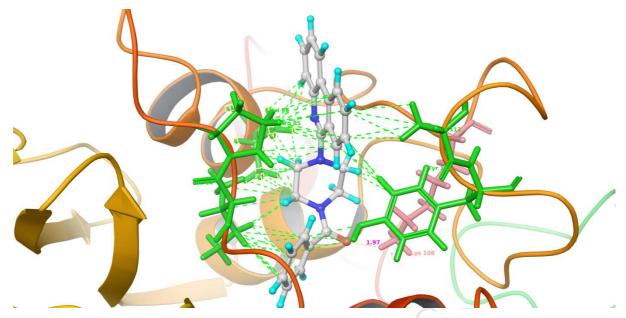


Figure 3: Interaction profile of compound **6b** with ATPase domain of *Mycobacterium tuberculosis* GyrB protein. The various hydrophobic interactions of compound **6b** are noticeable.

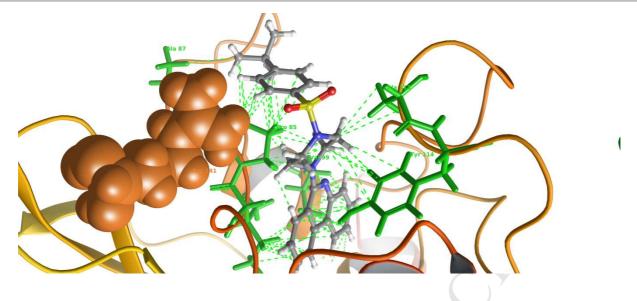
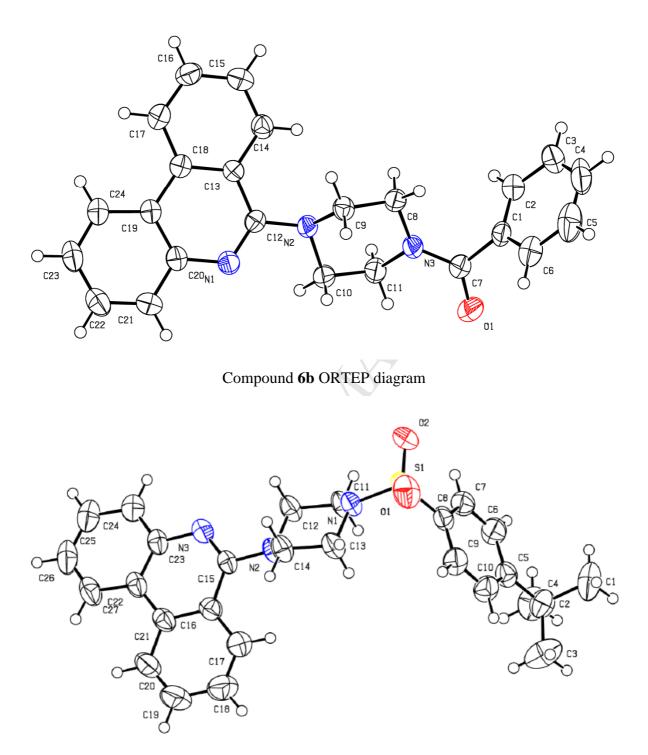


Figure 4: Interaction profile of compound **7d** with ATPase domain of *Mycobacterium tuberculosis* GyrB protein. The different polar contacts, π - π interaction and the hydrophobic interactions with compound **7d** are noticeable.

Figure 5: ORTEP plot for the compound **6b** and **7d**. All the non-hydrogen atoms are presented by their 50% probability thermal ellipsoids.



Compound 7d ORTEP diagram

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- \Rightarrow 33 novel phenanthridines were synthesized and evaluated for anti-TB activity.
- \Rightarrow 9 analogues (MIC \leq 3.13) exhibited excellent anti-TB activity against MTB H₃₇R_V.
- \Rightarrow In vitro toxicity studies were accomplished for most active compounds.
- \Rightarrow 6b and 7d compounds were further substantiated by single crystal X-ray studies.
- ⇒ Compounds **6b** and **7d** were docked to ATPase domain of MTB GyrB protein.

Supplementary Data

Novel 6-(piperazin-1-yl)phenanthridine amide and sulphonamide derivatives as potent *Mycobacterium tuberculosis* H37Rv inhibitors

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Perumal Yogeeswari,^c Kondapalli Venkata Gowri Chandra Sekhar^{a*}

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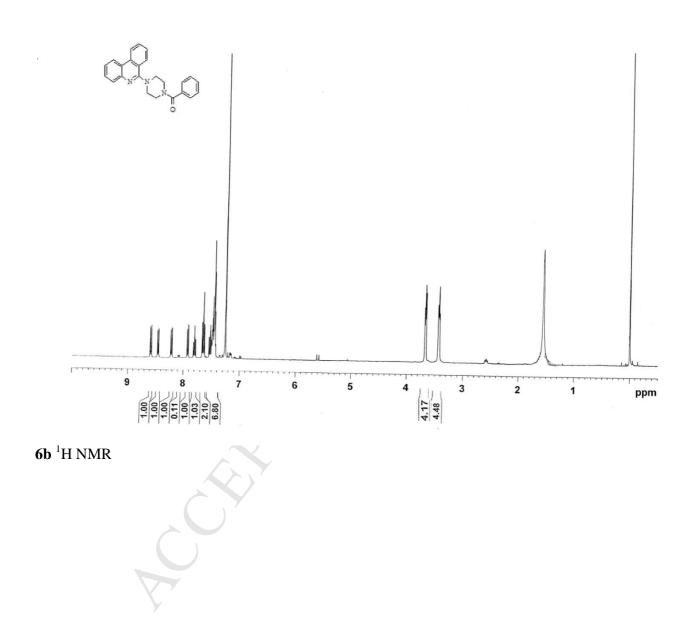
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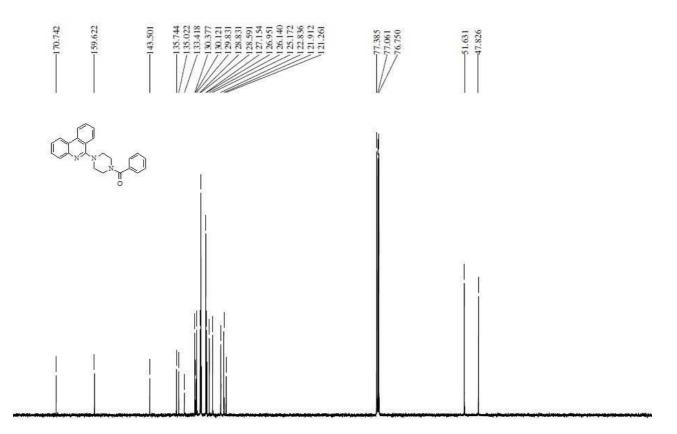
- 1. SpectrasS3-S152. Crystal refinement details and tables 1S-10S: Full list of atomic coordinates thermal
 - parameters, bond distances and angles of **6b** and **7d** crystal structure S15-S26

AND MARKER

1. Spectras

1.1.¹H and ¹³C NMR spectra's of represented final compounds



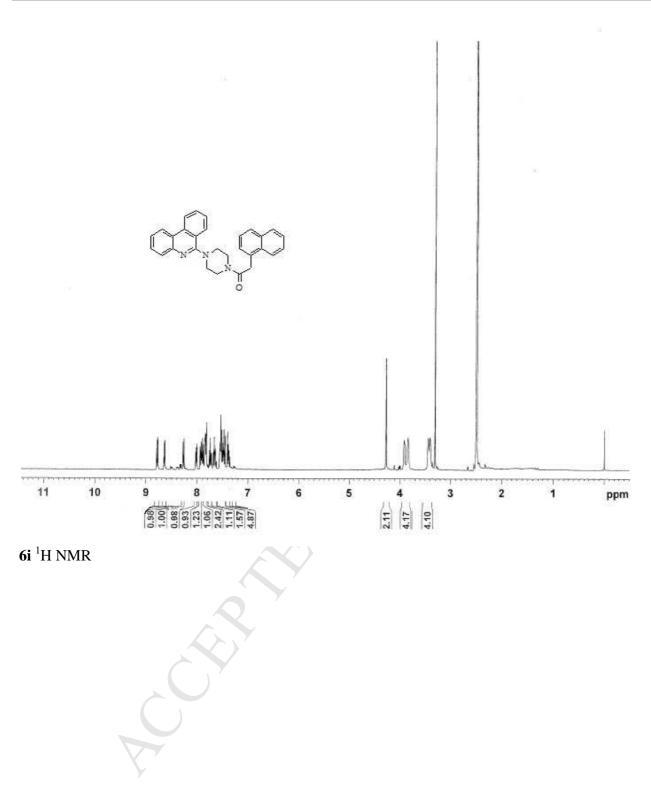


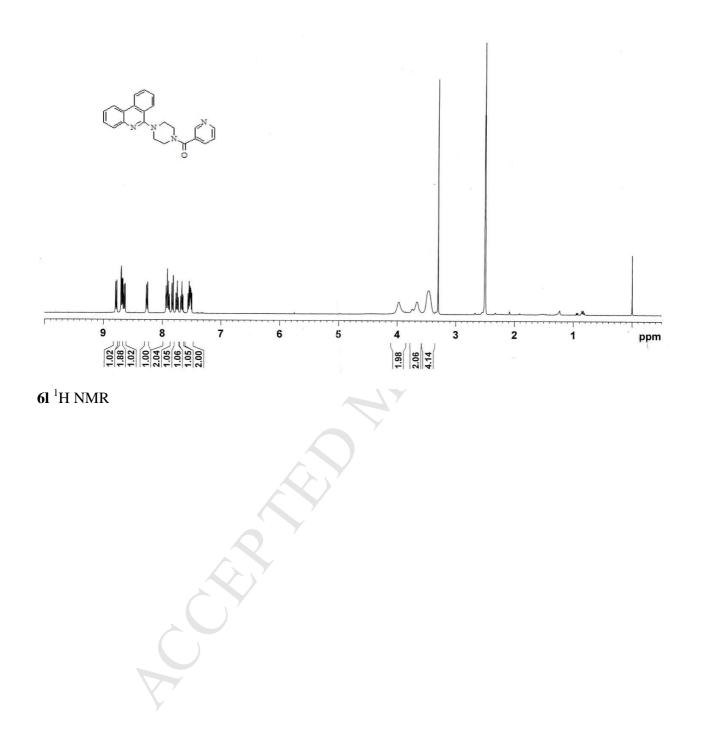


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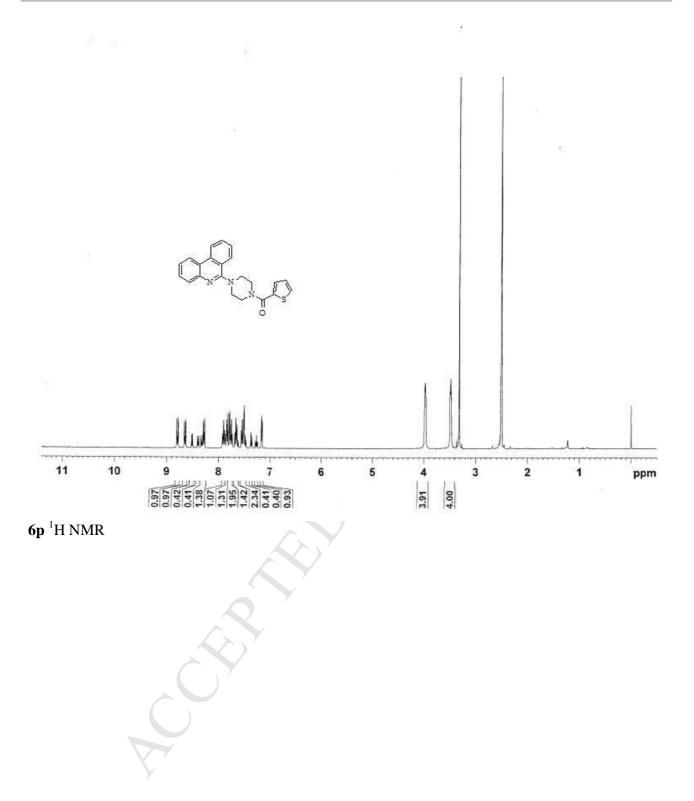
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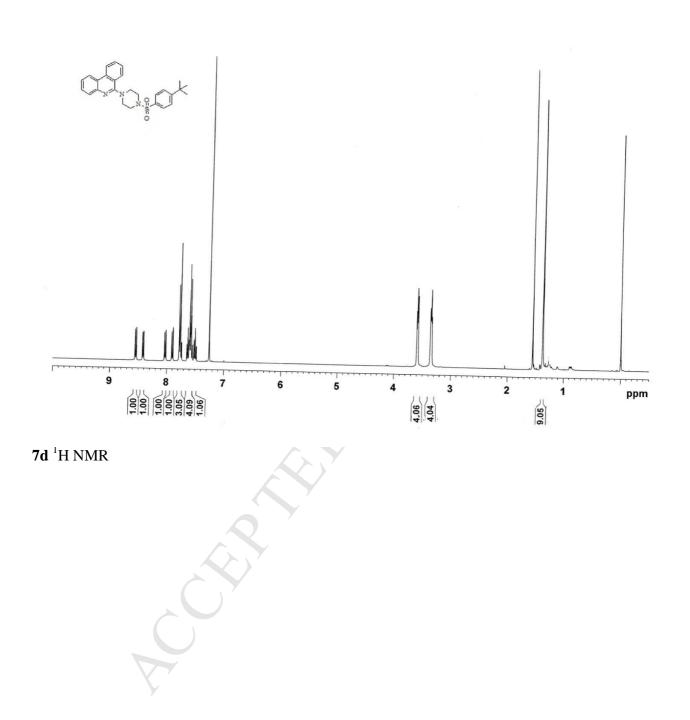


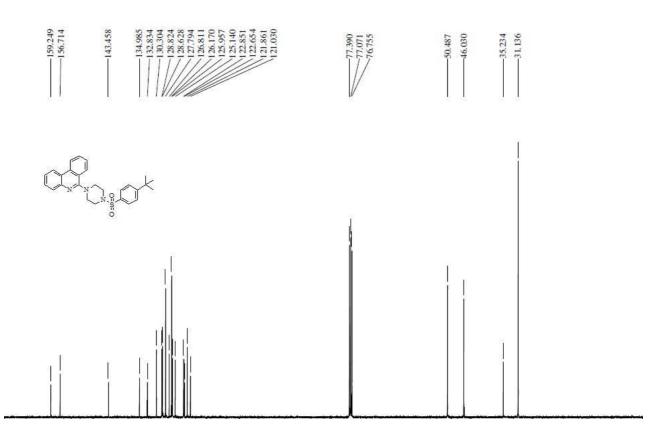


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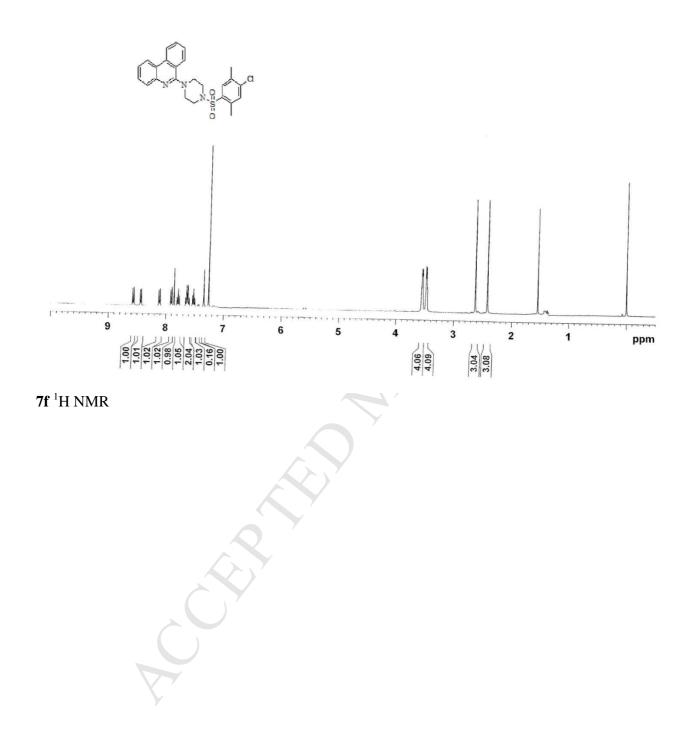


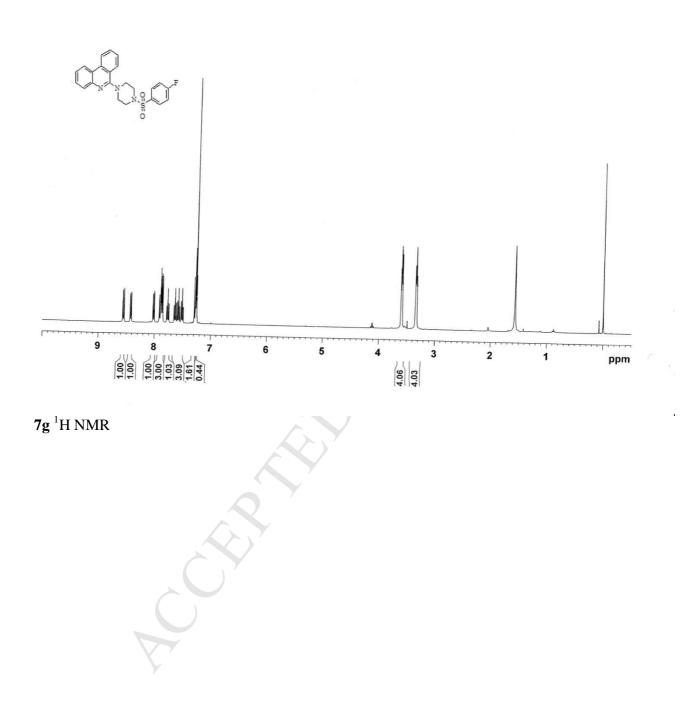
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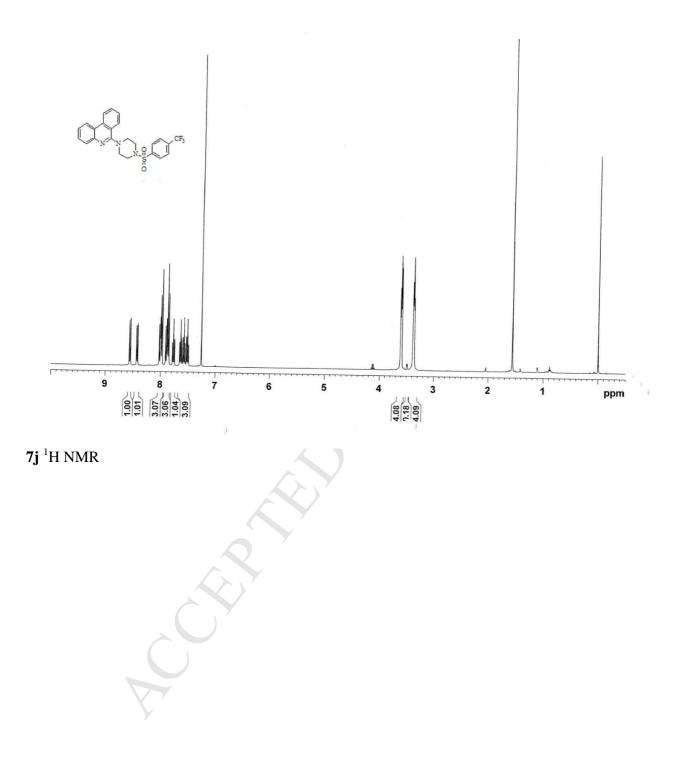


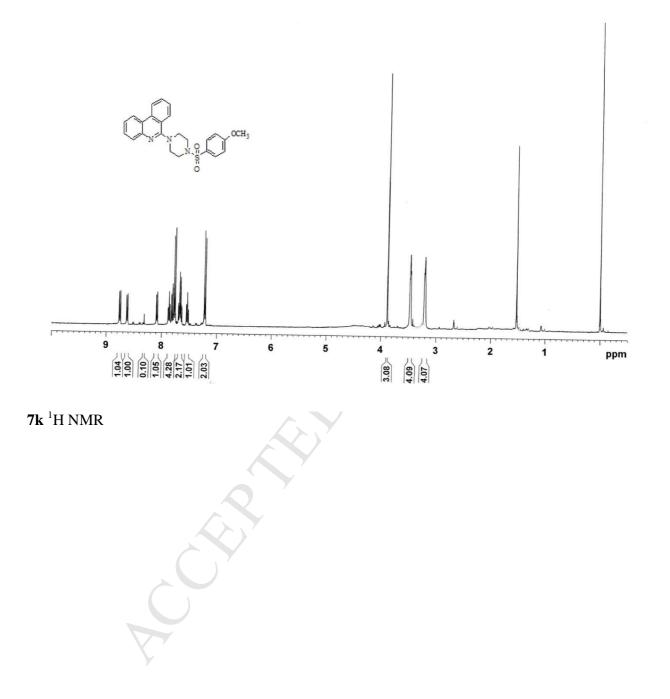


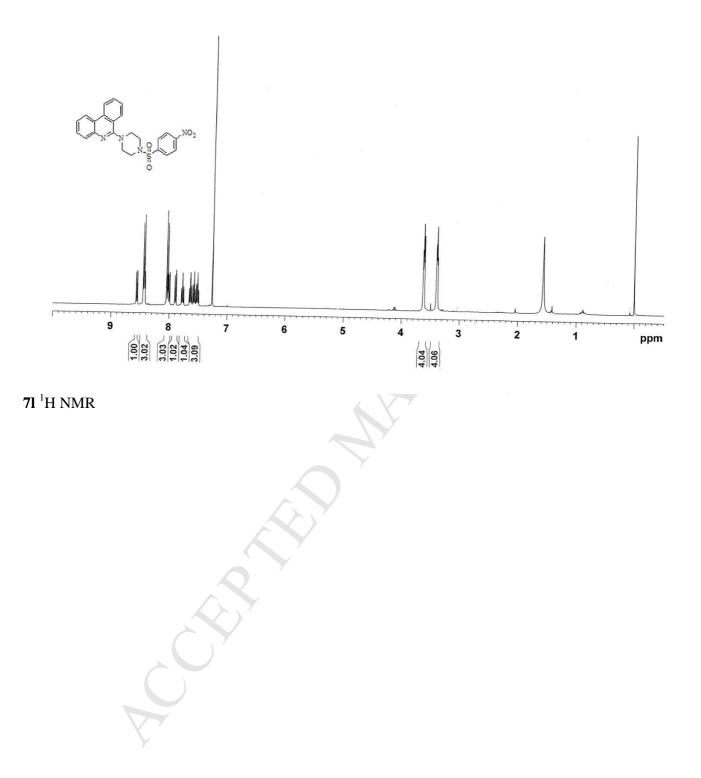
7d ¹³C NMR

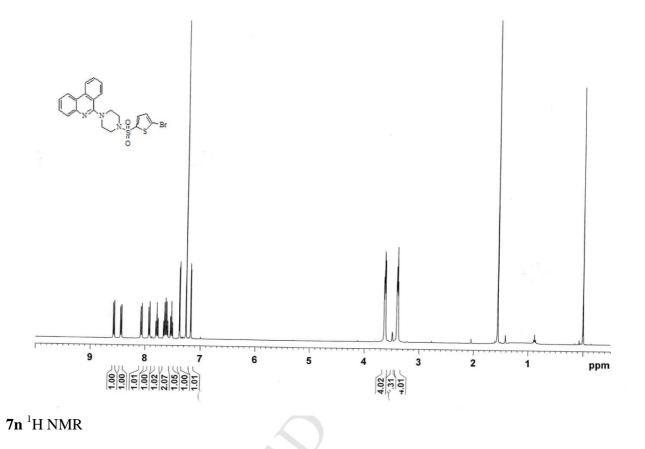












2. Crystal refinement details and tables 1S-10S: Full list of atomic coordinates thermal parameters, bond distances and angles of **6a** and **7d** crystal structures

Computing details

Program(s) used to refine structure: SHELXL97 (Sheldrick, 2008).

Crystal data for **6b**

CHNO $M_r = 367.44$ Triclinic, $P\overline{1}$ a = 9.4717 (13) Å b = 9.6936 (14) Å c = 11.8432 (16) Å $a = 73.767 (12)^{\circ}$ $\beta = 74.207 (12)^{\circ}$ $\gamma = 65.560 (14)^{\circ}$ $V = 935.1 (2) \text{ Å}^{3}$ Z = 2 F(000) = 388 $D_x = 1.305 \text{ Mg m}^{-3}$ Mo K α radiation, $\lambda =$ 0.71073 Å Cell parameters from 571 reflections $\theta = 3.3-29.3^{\circ}$ $\mu = 0.08 \text{ mm}^{-1}$ T = 293 K, white $0.6 \times 0.5 \times 0.4 \text{ mm}$

Data collection

Radiation source: fine-focus sealed tube Graphite monochromator Absorption correction: multi-scan ? $T_{min} = 0.246, T_{max} = 1.000$ 6305 measured reflections 5019 independent reflections

Refinement

Refinement on F^2 Least-squares matrix: full $R[F^2 > 2\sigma(F^2)] = 0.076$ $wR(F^2) = 0.221$ S = 0.954075 reflections 253 parameters 0 restraints Primary atom site location: structureinvariant direct methods 2213 reflections with $I > 2\sigma(I)$ $R_{int} = 0.038$ $\theta_{max} = 29.1^{\circ}, \theta_{min} = 3.3^{\circ}$ $h = -12 \rightarrow 12$ $k = -13 \rightarrow 13$ $l = -14 \rightarrow 14$

Secondary atom site location: difference Fourier map Hydrogen site location: inferred from neighbouring sites H atoms treated by a mixture of independent and constrained refinement $w = 1/[\sigma^2(F_o^2) + (0.1054P)^2]$ where $P = (F_o^2 + 2F_c^2)/3$ $(\Delta/\sigma)_{max} = 0.047$ $\Delta\rho_{max} = 0.27$ e Å⁻³ $\Delta\rho_{min} = 0.28$ e Å⁻³

Special details

Geometry. All s.u.'s (except the s.u. in the dihedral angle between two l.s. planes) are estimated using the full covariance matrix. The cell s.u.'s are taken into account individually in the estimation of s.u.'s in distances, angles and torsion angles; correlations between s.u.'s in cell parameters are only used when they are defined by crystal symmetry. An approximate (isotropic) treatment of cell s.u.'s is used for estimating s.u.'s involving l.s. planes.

Refinement. Refinement of F^2 against ALL reflections. The weighted R-factor wR and goodness of fit S are based on F^2 , conventional R-factors R are based on F, with F set to zero for negative F^2 . The threshold expression of $F^2 > 2\sigma(F^2)$ is used only for calculating R-factors(gt) etc. and is not relevant to the choice of reflections for refinement. R-factors based on F^2 are statistically about twice as large as those based on F, and R- factors based on ALL data will be even larger.

Atom	x	у	z	$U_{ m iso}*/U_{ m eq}$
C2	0.0324 (3)	1.0737 (3)	0.1190 (2)	0.0545 (8)
H2	-0.0145	1.0010	0.1364	0.065*
C3	-0.0592 (4)	1.2241 (4)	0.1328 (3)	0.0705 (11)
H3	-0.1674	1.2527	0.1591	0.085*
N1	0.5774 (2)	0.2605 (2)	0.39124 (18)	0.0385 (5)
C13	0.3312 (3)	0.3338 (3)	0.5330(2)	0.0311 (6)
C18	0.3902 (3)	0.2004 (3)	0.6187 (2)	0.0326 (6)
C19	0.5486 (3)	0.0918 (3)	0.5881 (2)	0.0327 (6)
C12	0.4338 (3)	0.3564 (3)	0.4190 (2)	0.0320 (6)
N2	0.3716(2)	0.4846 (2)	0.33013 (17)	0.0356 (5)
C20	0.6364 (3)	0.1286 (3)	0.4749 (2)	0.0356 (6)
C17	0.2917 (3)	0.1777 (3)	0.7303 (2)	0.0432 (7)
H17	0.3289	0.0900	0.7874	0.052*
C14	0.1745 (3)	0.4354 (3)	0.5595 (2)	0.0392 (6)
H14	0.1326	0.5204	0.5019	0.047*
C24	0.6195 (3)	-0.0452 (3)	0.6650(2)	0.0443 (7)
H24	0.5624	-0.0723	0.7393	0.053*
C15	0.0826 (3)	0.4109 (3)	0.6691 (2)	0.0467 (7)
H15	-0.0200	0.4803	0.6856	0.056*
N3	0.2904 (2)	0.7503 (2)	0.14794 (18)	0.0405 (6)
C16	0.1418 (3)	0.2834 (3)	0.7555 (2)	0.0473 (7)
H16	0.0802	0.2693	0.8305	0.057*
C8	0.2291 (3)	0.7624 (3)	0.2738 (2)	0.0392 (6)
H8A	0.1270	0.7521	0.2974	0.047*
H8B	0.2155	0.8631	0.2844	0.047*
C9	0.3407 (3)	0.6385 (3)	0.3514 (2)	0.0363 (6)
H9A	0.4391	0.6554	0.3337	0.044*
H9B	0.2954	0.6433	0.4348	0.044*
O1	0.3656 (2)	0.8506 (2)	-0.04484 (17)	0.0617 (7)
C21	0.7934 (3)	0.0290 (3)	0.4435 (2)	0.0469 (7)
H21	0.8522	0.0525	0.3690	0.056*
C7	0.2915 (3)	0.8695 (3)	0.0563 (2)	0.0399 (6)
C23	0.7720 (4)	-0.1397 (3)	0.6319 (3)	0.0524 (8)
H23	0.8175	-0.2303	0.6839	0.063*
C10	0.4540 (3)	0.4759 (3)	0.2071 (2)	0.0474 (8)
H10A	0.4826	0.3734	0.1922	0.057*

Table 1S: Fractional Atomic Coordinates (×10⁴) and Equivalent Isotropic Displacement Parameters ($\mathring{A}^2 \times 10^3$) for 6b. U_{eq} is defined as 1/3 of of the trace of the orthogonalised U_{IJ} tensor.

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H10B	0.5497	0.4966	0.1936	0.057*
C1	0.1948 (3)	1.0307 (3)	0.0790 (2)	0.0406 (7)
C6	0.2621 (4)	1.1407 (3)	0.0510(2)	0.0537 (8)
H6	0.3697	1.1140	0.0216	0.064*
C22	0.8589 (3)	-0.1015 (3)	0.5220 (3)	0.0536 (8)
H22	0.9631	-0.1655	0.5014	0.064*
C11	0.3458 (3)	0.5941 (3)	0.1243 (2)	0.0471 (8)
H11A	0.4011	0.5930	0.0423	0.057*
H11B	0.2555	0.5662	0.1332	0.057*
C4	0.0112 (5)	1.3314 (4)	0.1072 (3)	0.0765 (12)
H4	-0.0496	1.4323	0.1174	0.092*
C5	0.1710 (5)	1.2896 (4)	0.0666 (3)	0.0730 (10)
H5	0.2177	1.3624	0.0495	0.088*

Table 2S: Anisotropic Displacement Parameters ($\mathring{A}^2 \times 10^3$) for 6a. The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^{*2}U_{11}+...+2hka \times b \times U_{12}]$

Atom	$U^{ m 11}$	U^{22}	$U^{ m 33}$	U^{12}	U^{13}	U^{23}
C2	0.0522 (17)	0.0420 (16)	0.0530 (18)	–0.0105 (15)	-0.0063 (14)	0.0020 (13)
C3	0.069 (2)	0.0491 (19)	0.057 (2)	0.0063 (18)	-0.0096 (16)	-0.0003 (16)
N1	0.0366 (11)	0.0350 (11)	0.0412 (13)	-0.0118 (10)	-0.0078 (9)	-0.0046 (9)
C13	0.0303 (12)	0.0322 (13)	0.0328 (13)	-0.0114 (11)	-0.0075 (10)	-0.0073 (10)
C18	0.0360 (13)	0.0328 (13)	0.0364 (14)	-0.0179 (11)	-0.0092 (10)	-0.0068 (10)
C19	0.0382 (13)	0.0300 (12)	0.0348 (14)	-0.0130 (11)	-0.0123 (10)	-0.0073 (10)
C12	0.0368 (13)	0.0311 (12)	0.0326 (13)	-0.0166 (11)	-0.0069 (10)	-0.0056 (10)
N2	0.0426 (12)	0.0290 (11)	0.0309 (11)	-0.0098 (9)	-0.0050 (9)	-0.0056 (8)
C20	0.0379 (13)	0.0318 (13)	0.0410 (15)	-0.0143 (11)	-0.0122 (11)	-0.0057 (11)
C17	0.0479 (16)	0.0424 (15)	0.0391 (15)	-0.0194 (14)	-0.0104 (12)	–0.0005 (12)
C14	0.0356 (13)	0.0373 (14)	0.0434 (15)	–0.0125 (12)	-0.0109 (11)	-0.0035 (12)
C24	0.0496 (16)	0.0355 (14)	0.0444 (16)	–0.0127 (13)	-0.0149 (12)	-0.0007 (12)
C15	0.0367 (14)	0.0489 (16)	0.0503 (17)	–0.0114 (13)	-0.0018 (12)	–0.0155 (13)
N3	0.0492 (13)	0.0299 (11)	0.0327 (12)	-0.0061 (10)	-0.0055 (9)	-0.0060 (9)
C16	0.0444 (15)	0.0550 (17)	0.0400 (15)	-0.0221 (14)	0.0005 (12)	-0.0071 (13)
C8	0.0481 (15)	0.0326 (13)	0.0324 (14)	–0.0121 (12)	-0.0034 (11)	–0.0070 (11)
C9	0.0424 (14)	0.0305 (13)	0.0383 (14)	–0.0137 (12)	–0.0090 (11)	–0.0076 (11)
O1	0.0743 (15)	0.0511 (12)	0.0396 (12)	–0.0167 (11)	0.0076 (10)	-0.0052 (9)
C21	0.0376 (14)	0.0486 (16)	0.0481 (17)	–0.0108 (14)	-0.0008 (12)	–0.0137 (13)
C7	0.0428 (14)	0.0383 (14)	0.0350 (14)	-0.0134 (12)	-0.0080 (11)	-0.0028 (11)
C23	0.0630 (19)	0.0341 (14)	0.0536 (18)	-0.0050 (14)	-0.0262 (14)	-0.0031 (13)
C10	0.0562 (17)	0.0364 (14)	0.0330 (15)	–0.0061 (13)	-0.0008 (12)	–0.0057 (12)
C1	0.0504 (15)	0.0323 (13)	0.0317 (14)	–0.0110 (13)	–0.0106 (11)	0.0017 (11)
C6	0.0674 (19)	0.0451 (17)	0.0511 (18)	-0.0244 (15)	-0.0182 (14)	0.0005 (14)
C22	0.0466 (16)	0.0442 (16)	0.061 (2)	-0.0023 (14)	–0.0171 (14)	–0.0117 (15)
C11	0.0652 (18)	0.0333 (14)	0.0328 (14)	-0.0092 (13)	-0.0063 (12)	–0.0075 (11)
C4	0.120 (3)	0.0337 (17)	0.053 (2)	0.000 (2)	-0.030 (2)	-0.0027 (15)
C5	0.116 (3)	0.0432 (19)	0.068 (2)	-0.032 (2)	-0.038 (2)	0.0020 (16)
	× 1					

Table 3S: Bond Lengths for 6b

Atom—Atom	Length/Å	Atom—Atom	Length/Å
C2—C3	1.383 (4)	N3—C8	1.464 (3)
C2—C1	1.395 (4)	N3—C11	1.465 (3)
C2—H2	0.9300	C16—H16	0.9300
C3—C4	1.381 (5)	C8—C9	1.505 (3)

C3—H3	0.9300	C8—H8A	0.9700
N1—C12	1.305 (3)	C8—H8B	0.9700
N1—C20	1.389 (3)	C9—H9A	0.9700
C13—C14	1.407 (3)	C9—H9B	0.9700
C13—C18	1.412 (3)	O1—C7	1.233 (3)
C13—C12	1.454 (3)	C21—C22	1.363 (4)
C18—C17	1.414 (3)	C21—H21	0.9300
C18—C19	1.450 (3)	C7—C1	1.506 (4)
C19—C24	1.401 (3)	C23—C22	1.382 (4)
C19—C20	1.410 (3)	C23—H23	0.9300
C12—N2	1.414 (3)	C10—C11	1.508 (3)
N2—C10	1.461 (3)	C10—H10A	0.9700
N2—C9	1.477 (3)	C10—H10B	0.9700
C20—C21	1.412 (3)	C1—C6	1.380 (4)
C17—C16	1.375 (3)	C6—C5	1.376 (4)
C17—H17	0.9300	C6—H6	0.9300
C14—C15	1.372 (3)	C22—H22	0.9300
C14—H14	0.9300	C11—H11A	0.9700
C24—C23	1.370 (4)	C11—H11B	0.9700
C24—H24	0.9300	C4—C5	1.375 (5)
C15—C16	1.386 (3)	C4—H4	0.9300
C15—H15	0.9300	C5—H5	0.9300
N3—C7	1.347 (3)		

Table 4S: Bond Angles for 6b

8			
Atom—Atom—Atom	Angle/°	Atom—Atom—Atom	Angle/°
C3—C2—C1	120.3 (3)	N3—C8—H8B	109.6
C3—C2—H2	119.8	C9—C8—H8B	109.6
C1—C2—H2	119.8	H8A—C8—H8B	108.1
C4—C3—C2	119.6 (3)	N2-C9-C8	110.4 (2)
C4—C3—H3	120.2	N2—C9—H9A	109.6
C2—C3—H3	120.2	C8—C9—H9A	109.6
C12—N1—C20	118.5 (2)	N2—C9—H9B	109.6
C14—C13—C18	118.8 (2)	С8—С9—Н9В	109.6
C14—C13—C12	123.0 (2)	H9A—C9—H9B	108.1
C18—C13—C12	118.1 (2)	C22—C21—C20	120.2 (2)
C13—C18—C17	118.6 (2)	C22—C21—H21	119.9
C13—C18—C19	118.5 (2)	C20—C21—H21	119.9
C17—C18—C19	122.8 (2)	O1—C7—N3	122.3 (2)
C24—C19—C20	118.6 (2)	O1—C7—C1	119.5 (2)
C24—C19—C18	123.8 (2)	N3—C7—C1	118.2 (2)
C20—C19—C18	117.6 (2)	C24—C23—C22	120.6 (2)
N1—C12—N2	117.88 (19)	C24—C23—H23	119.7
N1-C12-C13	123.8 (2)	C22—C23—H23	119.7
N2—C12—C13	118.2 (2)	N2-C10-C11	108.34 (19)
C12—N2—C10	116.41 (18)	N2—C10—H10A	110.0
C12—N2—C9	116.7 (2)	C11—C10—H10A	110.0
C10—N2—C9	108.16 (19)	N2—C10—H10B	110.0
N1—C20—C19	123.5 (2)	C11—C10—H10B	110.0
N1—C20—C21	117.2 (2)	H10A—C10—H10B	108.4
C19—C20—C21	119.3 (2)	C6—C1—C2	119.2 (3)
C16—C17—C18	121.0 (2)	C6—C1—C7	120.8 (2)
C16—C17—H17	119.5	C2—C1—C7	119.8 (3)
C18—C17—H17	119.5	C5—C6—C1	120.3 (3)

C15—C14—C13	121.0 (2)	C5—C6—H6	119.8
C15—C14—H14	119.5	C1—C6—H6	119.8
C13—C14—H14	119.5	C21—C22—C23	120.6 (3)
C23—C24—C19	120.7 (2)	C21—C22—H22	119.7
C23—C24—H24	119.6	C23—C22—H22	119.7
C19—C24—H24	119.7	N3-C11-C10	112.4 (2)
C14—C15—C16	120.4 (2)	N3-C11-H11A	109.1
C14—C15—H15	119.8 ໌	C10—C11—H11A	109.1
C16—C15—H15	119.8	N3—C11—H11B	109.1
C7—N3—C8	125.4 (2)	C10—C11—H11B	109.1
C7—N3—C11	119.9 (2)	H11A—C11—H11B	107.8
C8—N3—C11	114.63 (19)	C3—C4—C5	120.2 (3)
C17—C16—C15	120.0 (2)	C3—C4—H4	119.9
C17—C16—H16	120.0	C5—C4—H4	119.9
C15—C16—H16	120.0	C6—C5—C4	120.4 (3)
N3—C8—C9	110.40 (19)	C6—C5—H5	119.8
N3—C8—H8A	109.6	C4—C5—H5	119.8
C9—C8—H8A	109.6		
		(
Table 5S: Dihedral angl	le for 6b		
0			

Table 5S: Dihedral angle for 6b

Two planes	Angle/°	Two planes	Angle/°
C1—C2—C3—C4	-0.2 (5)	C18—C17—C16—C15	2.9 (5)
C14—C13—C18—C17	-2.9 (4)	C14—C15—C16—C17	–2.1 (5)
C12—C13—C18—C17	179.6 (2)	C7—N3—C8—C9	-135.9 (3)
C14—C13—C18—C19	176.5 (2)	C11—N3—C8—C9	47.5 (3)
C12—C13—C18—C19	–1.0 (4)	C12-N2-C9-C8	–161.6 (2)
C13—C18—C19—C24	–179.4 (2)	C10-N2-C9-C8	64.9 (3)
C17—C18—C19—C24	0.0 (4)	N3-C8-C9-N2	-55.3 (3)
C13—C18—C19—C20	1.7 (4)	N1-C20-C21-C22	179.3 (3)
C17—C18—C19—C20	-178.9 (2)	C19—C20—C21—C22	0.2 (4)
C20-N1-C12-N2	–175.7 (2)	C8-N3-C7-O1	168.8 (3)
C20-N1-C12-C13	0.2 (4)	C11—N3—C7—O1	–14.9 (4)
C14—C13—C12—N1	–177.4 (3)	C8—N3—C7—C1	–12.9 (4)
C18—C13—C12—N1	0.0 (4)	C11—N3—C7—C1	163.5 (3)
C14—C13—C12—N2	–1.5 (4)	C19—C24—C23—C22	0.1 (5)
C18—C13—C12—N2	175.9 (2)	C12-N2-C10-C11	162.6 (2)
N1-C12-N2-C10	16.8 (4)	C9—N2—C10—C11	-63.7 (3)
C13—C12—N2—C10	-159.3 (2)	C3—C2—C1—C6	-1.4 (4)
N1—C12—N2—C9	-112.9 (3)	C3—C2—C1—C7	–175.8 (3)
C13—C12—N2—C9	70.9 (3)	O1—C7—C1—C6	–55.1 (4)
C12—N1—C20—C19	0.6 (4)	N3—C7—C1—C6	126.5 (3)
C12-N1-C20-C21	-178.5 (3)	O1—C7—C1—C2	119.3 (3)
C24—C19—C20—N1	179.4 (2)	N3—C7—C1—C2	–59.1 (4)
C18—C19—C20—N1	-1.6 (4)	C2—C1—C6—C5	2.3 (4)
C24—C19—C20—C21	–1.5 (4)	C7—C1—C6—C5	176.7 (3)
C18—C19—C20—C21	177.4 (2)	C20—C21—C22—C23	1.3 (5)
C13—C18—C17—C16	-0.3 (4)	C24—C23—C22—C21	–1.5 (5)
C19—C18—C17—C16	–179.7 (3)	C7—N3—C11—C10	134.6 (3)
C18—C13—C14—C15	3.7 (4)	C8—N3—C11—C10	-48.7 (3)
C12—C13—C14—C15	–178.9 (3)	N2-C10-C11-N3	56.0 (3)
C20-C19-C24-C23	1.4 (4)	C2—C3—C4—C5	0.9 (5)
C18—C19—C24—C23	–177.5 (3)	C1—C6—C5—C4	–1.6 (5)
<u>C13—C14—C15—C16</u>	–1.2 (4)	C3—C4—C5—C6	0.0 (5)

Crystal data 7d

CHNOS $M_r = 459.60$

Monoclinic, $P2_1/n$

$$a = 12.5058 (13) \text{ Å}$$

$$b = 14.6774 (17) \text{ Å}$$

$$c = 13.3524 (14) \text{ Å}$$

$$\beta = 89.451 (9)^{\circ}$$

$$V = 2450.8 (5) \text{ Å}^{\circ}$$

$$Z = 4$$

Data collection Radiation source: fine-focus sealed tube Graphite monochromator Absorption correction: multi-scan ? $T_{min} = 0.922, T_{max} = 1.000$ 7784 measured reflections 6663 independent reflections

Refinement

Refinement on F^2 Least-squares matrix: full $R[F^2 > 2\sigma(F^2)] = 0.050$ $wR(F^2) = 0.121$ S = 1.025149 reflections 298 parameters 0 restraints Primary atom site location: structureinvariant direct methods F(000) = 976.0 $D_x = 1.246 \text{ Mg m}^{-3}$ Mo K α radiation, $\lambda =$ 0.71073 Å Cell parameters from 1760 reflections $\theta = 3.2-29.2^{\circ}$ $\mu = 0.16 \text{ mm}^{-1}$ T = 293 K, white $0.6 \times 0.5 \times 0.4 \text{ mm}$

3305 reflections with $I > 2\sigma(I)$ $R_{int} = 0.018$ $\theta_{max} = 29.2^{\circ}, \theta_{min} = 3.2^{\circ}$ $h = -9 \rightarrow 17$ $k = -16 \rightarrow 20$ $l = -18 \rightarrow 15$

Secondary atom site location: difference Fourier map Hydrogen site location: inferred from neighbouring sites H atoms treated by a mixture of independent and constrained refinement $w = 1/[\sigma^2(F_o^2) + (0.0438P)^2 + 0.1582P]$ where $P = (F_o^2 + 2F_c^2)/3$ $(\Delta/\sigma)_{max} = 0.001$ $\Delta\rho_{max} = 0.14$ e Å⁻³ $\Delta\rho_{min} = -0.25$ e Å⁻³

Atom	x	у	Z	Uiso*/ U eq
S1	0.87179 (4)	0.19249 (4)	0.62715 (4)	0.05313 (17)
N2	0.90451 (12)	0.15341 (10)	0.29824 (11)	0.0437 (4)
01	0.85352 (12)	0.28653 (10)	0.64877 (12)	0.0704 (5)
02	0.95600 (10)	0.14456 (12)	0.67589 (11)	0.0700 (5)
N3	1.02859 (12)	0.10726 (10)	0.17912 (11)	0.0440 (4)
N1	0.89739 (12)	0.18566 (10)	0.50721 (11)	0.0444 (4)
C16	0.86887 (14)	0.18206 (13)	0.11995 (15)	0.0445 (5)
C8	0.75352 (14)	0.13109 (14)	0.64929 (14)	0.0465 (5)
C22	1.00718 (15)	0.13476 (13)	-0.00057 (14)	0.0462 (5)
C15	0.93860 (14)	0.14755 (12)	0.19725 (13)	0.0392 (4)
C21	0.90525 (15)	0.17769 (13)	0.01987 (15)	0.0482 (5)
C14	0.89796 (16)	0.24619 (13)	0.33966 (15)	0.0508 (5)
H14A	0.8606	0.2855	0.2933	0.061*
H14B	0.9694	0.2703	0.3486	0.061*
C23	1.06275 (14)	0.09828 (13)	0.08049 (14)	0.0441 (5)
C12	0.96299 (15)	0.09523 (14)	0.36699 (14)	0.0508 (5)
H12A	1.0351	0.1184	0.3749	0.061*
H12B	0.9677	0.0340	0.3399	0.061*
C13	0.83968 (16)	0.24495 (13)	0.43846 (15)	0.0504 (5)
H13A	0.8356	0.3062	0.4657	0.060*
H13B	0.7674	0.2225	0.4295	0.060*
C27	1.05135 (18)	0.12372 (16)	-0.09680 (16)	0.0622 (6)
H27	1.0162	0.1480	-0.1517	0.075*
C11	0.90694 (16)	0.09288 (13)	0.46703 (14)	0.0513 (5)
H11A	0.8364	0.0664	0.4597	0.062*
H11B	0.9470	0.0553	0.5131	0.062*
C5	0.56493 (15)	0.03232 (14)	0.67638 (15)	0.0490 (5)
C24	1.15876 (15)	0.05188 (14)	0.06368 (17)	0.0558 (5)
H24	1.1959	0.0279	0.1176	0.067*
C9	0.65609 (16)	0.17097 (15)	0.62998 (16)	0.0556 (6)
H9	0.6528	0.2311	0.6082	0.067*
C25	1.19867 (18)	0.04137 (16)	-0.03106 (18)	0.0656 (6)
H25	1.2621	0.0095		0.079*
н25 С6			-0.0415	
	0.66410 (16) 0.6676	-0.00622 (15)	0.69497 (17) 0.7171	0.0596 (6) 0.072*
H6		-0.0662		
C20	0.83837 (19)	0.20989 (17)	-0.05630 (18)	0.0695 (7)
H20	0.8620	0.2082	-0.1225	0.083*
C2	0.46293 (16)	-0.02121 (15)	0.69611 (18)	0.0589 (6)
C10	0.56375 (16)	0.12128 (14)	0.64320 (16)	0.0562 (6)
H10	0.4984	0.1486	0.6293	0.067*
C7	0.75710 (16)	0.04200 (15)	0.68149 (16)	0.0577 (6)
H7	0.8226	0.0145	0.6941	0.069*
C3	0.36522 (16)	0.02238 (18)	0.6496 (2)	0.0959 (10)
H3A	0.3746	0.0253	0.5782	0.144*
H3B	0.3562	0.0828	0.6758	0.144*
H3C	0.3030	-0.0134	0.6654	0.144*
C17	0.76540 (16)	0.21474 (15)	0.13892 (18)	0.0602 (6)
H17	0.7392	0.2157	0.2043	0.072*

Table 6S: Fractional Atomic Coordinates (×10⁴) and Equivalent Isotropic Displacement Parameters ($Å^2 \times 10^3$) for 6b. U_{eq} is defined as 1/3 of of the trace of the orthogonalised U_{IJ} tensor.

C26	1.1454 (2)	0.07774 (17)	-0.11104 (18)	0.0702 (7)
H26	1.1734	0.0711	-0.1754	0.084*
C4	0.47333 (19)	-0.11869 (16)	0.6557 (2)	0.0801 (8)
H4A	0.5347	-0.1474	0.6845	0.120*
H4B	0.4815	-0.1169	0.5841	0.120*
H4C	0.4103	-0.1527	0.6730	0.120*
C18	0.70249 (19)	0.24507 (19)	0.0633 (2)	0.0832 (8)
H18	0.6342	0.2670	0.0773	0.100*
C1	0.4469 (2)	-0.02644 (18)	0.80942 (19)	0.0864 (8)
H1A	0.5084	-0.0543	0.8392	0.130*
H1B	0.3845	-0.0622	0.8244	0.130*
H1C	0.4377	0.0339	0.8359	0.130*
C19	0.7404 (2)	0.2432 (2)	-0.0348 (2)	0.0903 (9)
H19	0.6977	0.2652	-0.0861	0.108*

Table 7S: Anisotropic Displacement Parameters ($\mathring{A}^2 \times 10^3$) for 6a. The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^{*2}U_{11}+...+2hka\times b\times U_{12}]$

Atom	$U^{\scriptscriptstyle 11}$	U^{22}	$U^{ m 33}$	U^{12}	$U^{ m 13}$	U^{23}
S1	0.0594(3)	0.0670 (4)	0.0330 (3)	-0.0039(3)	-0.001(2)	-0.0050(2)
N2	0.0618(9)	0.0366 (9)	0.0327 (9)	0.0049(7)	0.0026(7)	0.0039(7)
01	0.0942(11)	0.0632 (11)	0.0538 (10)	-0.0137(8)	0.0063(9)	-0.0220(8)
O2	0.0588(8)	0.1098 (14)	0.0418 (9)	-0.0016(8)	0.0125(7)	0.0081(8)
N3	0.0528(9)	0.0446 (10)	0.0345 (9)	-0.004(7)	0.0011(7)	0.0013(7)
N1	0.0562(9)	0.0451 (10)	0.0319 (9)	0.0048(7)	0.0012(7)	0.0021(7)
C16	0.0489(10)	0.0431 (12)	0.0416 (12)	-0.0099(9)	-0.0041(9)	0.0053(9)
C8	0.0546(11)	0.0520 (13)	0.0329 (11)	0.0074(9)	0.0047(9)	0.0016(9)
C22	0.0582(11)	0.0482 (12)	0.0320 (11)	-0.0205(9)	0.0004(9)	-0.0002(9)
C15	0.0500(10)	0.0361 (10)	0.0316 (10)	-0.0055(8)	0.0016(8)	0.0026(8)
C21	0.0580(11)	0.0487 (12)	0.0381 (12)	-0.0171(9)	-0.0081(9)	0.0070(7)
C14	0.0724(12)	0.0403 (12)	0.0395 (12)	0.0046(10)	0.0041(10)	0.0063(7)
C23	0.0550(11)	0.0421 (11)	0.0351 (11)	-0.0130(9)	0.0042(9)	-0.0030(9)
C12	0.0695(12)	0.0477 (12)	0.0352 (11)	0.0157(10)	0.0039(9)	0.0069(9)
C13	0.0675(12)	0.0423 (12)	0.0414 (12)	0.0107(10)	0.0020(10)	0.0013(9)
C27	0.0780(15)	0.0738 (16)	0.0346 (12)	-0.0287(12)	-0.0012(11)	-0.0012(11)
C11	0.0722(12)	0.0458 (12)	0.0358 (11)	0.0139(10)	0.0047(10)	0.0089(9)
C5	0.0563(11)	0.0497 (13)	0.0408 (12)	0.0075(10)	0.0065(9)	-0.0009(9)
C24	0.0585(12)	0.0577 (14)	0.0511 (14)	–0.0055(10)	0.0072(10)	-0.0018(11)
C9	0.0614(12)	0.0464 (13)	0.0590 (14)	0.0131(10)	0.0098(11)	0.0040(10)
C25	0.0682(13)	0.0660 (16)	0.0621 (16)	-0.0139(12)	0.0223(12)	-0.0115(13)
C6	0.0591 (12)	0.0527 (13)	0.0669 (16)	0.0101(11)	0.0069(11)	0.0179(11)
C20	0.0728(15)	0.0898 (19)	0.0462 (14)	-0.0140(13)	-0.0148(12)	0.0196(12)
C2	0.0579(12)	0.0540 (14)	0.0647 (16)	0.0018(10)	0.0112(11)	-0.0081(11)
C10	0.0523(11)	0.0527 (13)	0.0635 (15)	0.0154(10)	0.0085(10)	0.0024(11)
C7	0.0545(12)	0.0650 (15)	0.0536 (14)	0.0154(11)	0.0028(10)	0.0164(11)
C3	0.0566(14)	0.084 (2)	0.147 (3)	-0.0009(13)	–0.0066(16)	0.0004(18)
C17	0.0557 (12)	0.0663 (16)	0.0586 (15)	-0.0017(10)	-0.0005(11)	0.0094(11)
C26	0.0816(16)	0.0805 (18)	0.0479 (15)	-0.0313(14)	0.0240(13)	-0.0143(13)
C4	0.0890(16)	0.0647 (17)	0.086 (2)	-0.0047(13)	0.0095(15)	-0.0176(14)
C18	0.0584(14)	0.100(2)	0.091 (2)	0.0077(13)	-0.0093 (14)	0.0243(17)
C1	0.0956(17)	0.08(2)	0.0754 (19)	-0.0198(15)	0.0381(15)	-0.0135(15)
C19	0.0772(17)	0.120(3)	0.074 (2)	–0.0002 (16)	–0.0238 (15)	0.0337(18)

Atom—Atom	Length/Å	Atom—Atom	Length/Å
S1—O2	1.4286 (15)	C5—C6	1.388 (3)
S1—N1	1.6332 (16)	C5—C2	1.519 (3)
S1—C8	1.7546 (19)	C24—C25	1.364 (3)
N2—C15	1.413 (2)	C24—H24	0.9300
N2-C12	1.456 (2)	C9—C10	1.376 (3)
N2-C14	1.472 (2)	C9—H9	0.9300
N3—C15	1.292 (2)	C25—C26	1.373 (3)
N3—C23	1.387 (2)	C25—H25	0.9300
N1—C13	1.461 (2)	C6—C7	1.372 (3)
N1—C11	1.468 (2)	C6—H6	0.9300
C16—C17	1.401 (3)	C20—C19	1.348 (3)
C16—C21	1.409 (3)	C20—H20	0.9300
C16—C15	1.449 (3)	C2—C3	1.517 (3)
C8—C7	1.377 (3)	C2—C1	1.526 (3)
C8—C9	1.378 (3)	C2—C4	1.534 (3)
C22—C23	1.398 (3)	C10—H10	0.9300
C22—C27	1.403 (3)	C7—H7	0.9300
C22—C21	1.446 (3)	C3—H3A	0.9600
C21—C20	1.405 (3)	C3—H3B	0.9600
C14—C13	1.501 (2)	C3—H3C	0.9600
C14—H14A	0.9700	C17—C18	1.361 (3)
C14—H14B	0.9700	C17—H17	0.9300
C23—C24	1.397 (3)	C26—H26	0.9300
C12—C11	1.503 (2)	C4—H4A	0.9600
C12—H12A	0.9700	C4—H4B	0.9600
C12—H12B	0.9700	C4—H4C	0.9600
C13—H13A	0.9700	C18—C19	1.389 (4)
C13—H13B	0.9700	C18—H18	0.9300
C27—C26	1.367 (3)	C1—H1A	0.9600
C27—H27	0.9300	C1—H1B	0.9600
C11—H11A	0.9700	C1—H1C	0.9600
C11—H11B	0.9700	C19—H19	0.9300

Table 8S: Bond Lengths for 7d

Table 9S: Bond Angles for 7d

Atom—Atom—Atom	Angle/°	Atom—Atom—Atom	Angle/°
01—S1—02	120.07 (10)	C10—C5—C2	122.27 (18)
O1—S1—N1	106.69 (9)	C6—C5—C2	120.57 (19)
O2-S1-N1	106.14 (8)	C25—C24—C23	120.7 (2)
O1—S1—C8	109.23 (9)	C25—C24—H24	119.7
O2—S1—C8	107.11 (9)	C23—C24—H24	119.7
N1—S1—C8	106.90 (9)	C10—C9—C8	119.6 (2)
C15—N2—C12	114.66 (14)	C10—C9—H9	120.2
C15—N2—C14	115.48 (14)	C8—C9—H9	120.2
C12—N2—C14	109.40 (15)	C24—C25—C26	120.1 (2)
C15—N3—C23	118.76 (17)	C24—C25—H25	120.0
C13—N1—C11	111.22 (15)	C26—C25—H25	120.0
C13—N1—S1	119.09 (12)	C7—C6—C5	121.6 (2)
C11-N1-S1	115.46 (12)	C7—C6—H6	119.2
C17—C16—C21	118.45 (19)	C5—C6—H6	119.2

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$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C7—C8—C9	119.54 (18)	C21—C20—H20	119.5
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C7—C8—S1	120.69 (15)	C3—C2—C5	112.90 (19)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C9—C8—S1	119.71 (16)	C3—C2—C1	109.17 (19)
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H14A-C14-H14B108.2H3A-C3-H3C109.5N3-C23-C24117.09 (18)H3B-C3-H3C109.5N3-C23-C22123.21 (17)C18-C17-C16121.2 (2)C24-C23-C22119.68 (18)C18-C17-H17119.4N2-C12-C11109.94 (14)C16-C17-H17119.4N2-C12-H12A109.7C27-C26-C25120.5 (2)C11-C12-H12A109.7C25-C26-H26119.8N2-C12-H12B109.7C2-C4-H4A109.5N1-C13-C14108.7C2-C4-H4B109.5N1-C13-C14108.71 (15)H4A-C4-H4B109.5N1-C13-H13A109.9C2-C4-H4C109.5N1-C13-H13B109.9H4A-C4-H4C109.5C14-C13-H13B109.9C17-C18-C19119.9 (2)H13A-C13-H13B108.3C17-C18-H18120.0C26-C27-C22121.0 (2)C19-C18-H18120.0C26-C27-H27119.5C2-C1-H1A109.5N1-C11-H11A109.7C2-C1-H1C109.5N1-C11-H11A109.7C2-C1-H1C109.5N1-C11-H11B109.7H1A-C1-H1C109.5N1-C11-H11B109.7H1A-C1-H1C109.5N1-C11-H11B109.7C2-C1-H1C109.5N1-C11-H11B109.7H1B-C1-H1C109.5N1-C11-H11B109.7H1B-C1-H1C109.5N1-C11-H11B109.7C2-C1-H1C109.5N1-C11-H11B109.7H1B-C1-H1C109.5N1-C11-H11B109.7C2-C19-C18120.6 (2)H1				
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N3-C23-C22123.21 (17)C18-C17-C16121.2 (2)C24-C23-C22119.68 (18)C18-C17-H17119.4N2-C12-C11109.94 (14)C16-C17-H17119.4N2-C12-H12A109.7C27-C26-C25120.5 (2)C11-C12-H12A109.7C25-C26-H26119.8N2-C12-H12B109.7C25-C26-H26119.8C11-C12-H12B109.7C2-C4-H4A109.5H12A-C12-H12B108.2C2-C4-H4B109.5N1-C13-C14108.71 (15)H4A-C4-H4B109.5N1-C13-H13A109.9C2-C4-H4C109.5C14-C13-H13B109.9H4B-C4-H4C109.5C14-C13-H13B109.9C17-C18-C19119.9 (2)H13A-C13-H13B109.9C17-C18-H18120.0C26-C27-L22121.0 (2)C19-C18-H18120.0C26-C27-H27119.5C2-C1-H1A109.5N1-C11-H11A109.7C2-C1-H1B109.5N1-C11-H11A109.7C2-C1-H1B109.5N1-C11-H11B109.7C2-C1-H1C109.5N1-C11-H11B109.7C2-C1-H1C109.5N1-C11-H11B109.7H1A-C1-H1C109.5N1-C11-H11B109.7H1A-C1-H1C109.5N1-C11-H11B109.7H1A-C1-H1C109.5N1-C11-H11B109.7H1A-C1-H1C109.5N1-C11-H11B109.7C2-C10-C18120.6 (2)H11A-C11-H11B108.2C20-C19-C18120.6 (2)	H14A—C14—H14B	108.2	НЗА—СЗ—НЗС	
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N2C12H12A109.7C27C26C25120.5 (2)C11C12H12A109.7C27C26H26119.8N2C12H12B109.7C25C26H26119.8C11C12H12B109.7C2C4H4A109.5H12AC12H12B108.2C2C4H4B109.5N1C13C14108.71 (15)H4AC4H4B109.5N1C13H13A109.9C2C4H4C109.5C14C13H13A109.9H4AC4H4C109.5C14C13H13B109.9H4BC4H4C109.5C14C13H13B109.9C17C18C19119.9 (2)H13AC13H13B108.3C17C18H18120.0C26C27C22121.0 (2)C19C18H18120.0C26C27H27119.5C2C1H1A109.5N1C11C12109.83 (15)H1AC1H1B109.5N1C11H11A109.7C2C1H1C109.5N1C11H11B109.7H1BC1H1C109.5N1C11H11B109.7H1AC1H1C109.5N1C11H11B109.7C20C19C18120.6 (2)H11AC1H11B109.7C20C19C18120.6 (2)H11AC11H11B108.2C20C19H19119.7	C24—C23—C22	119.68 (18)	C18—C17—H17	119.4
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$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C11—C12—H12A	109.7	C27—C26—H26	
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C12—C11—H11B109.7C20—C19—C18120.6 (2)H11A—C11—H11B108.2C20—C19—H19119.7				
H11A—C11—H11B 108.2 C20—C19—H19 119.7				
C10—C5—C6 117.11 (18) C18—C19—H19 119.7				
	<u>C10—C5—C6</u>	117.11 (18)	C18—C19—H19	119.7

Table 10S: Dihedral angle for 7d

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Two planes	Angle/°	Two planes	Angle/°
O1-S1-N1-C13	-37.12 (16)	C15—N2—C12—C11	-168.92 (16)
O2—S1—N1—C13	-166.26 (15)	C14—N2—C12—C11	59.5 (2)
C8—S1—N1—C13	79.65(16)	C11—N1—C13—C14	-58.4 (2)
O1—S1—N1—C11	-173.48 (14)	S1—N1—C13—C14	163.59 (13)

O2-S1-N1-C11	57.38(16)	N2—C14—C13—N1	59.5 (2)
C8—S1—N1—C11	-56.70 (15)	C23—C22—C27—C26	0.7 (3)
O1—S1—C8—C7	-148.45 (17)	C21—C22—C27—C26	–176.19 (19)
O2—S1—C8—C7	–17.0 (2)	C13—N1—C11—C12	57.9 (2)
N1—S1—C8—C7	96.47 (18)	S1—N1—C11—C12	-162.41 (13)
O1—S1—C8—C9	34.50 (19)	N2-C12-C11-N1	-58.0 (2)
O2—S1—C8—C9	166.00 (16)	N3—C23—C24—C25	-178.98 (18)
N1—S1—C8—C9	-80.58 (18)	C22—C23—C24—C25	-0.3 (3)
C23—N3—C15—N2	178.28 (15)	C7—C8—C9—C10	0.0 (3)
C23—N3—C15—C16	1.8 (3)	S1—C8—C9—C10	177.13 (16)
C12—N2—C15—N3	-13.4 (2)	C23—C24—C25—C26	1.0 (3)
C14—N2—C15—N3	115.17 (18)	C10—C5—C6—C7	0.3 (3)
C12—N2—C15—C16	163.22 (16)	C2—C5—C6—C7	-177.4 (2)
C14—N2—C15—C16	-68.2 (2)	C16—C21—C20—C19	-1.0 (3)
C17-C16-C15-N3	171.27 (18)	C22-C21-C20-C19	174.9 (2)
C21-C16-C15-N3	-5.1 (3)	C10—C5—C2—C3	15.8 (3)
C17-C16-C15-N2	-5.1 (3)	C6—C5—C2—C3	-166.7 (2)
C21-C16-C15-N2	178.48 (16)	C10—C5—C2—C1	–104.7 (2)
C17—C16—C21—C20	2.8 (3)	C6—C5—C2—C1	72.9 (2)
C15—C16—C21—C20	179.32 (18)	C10—C5—C2—C4	137.4 (2)
C17—C16—C21—C22	-173.31 (17)	C6—C5—C2—C4	-45.0 (3)
C15—C16—C21—C22	3.3 (3)	C8—C9—C10—C5	0.7 (3)
C23—C22—C21—C20	-174.58 (18)	C6—C5—C10—C9	-0.8 (3)
C27—C22—C21—C20	2.3 (3)	C2C5C10C9	176.8 (2)
C23—C22—C21—C16	1.3 (3)	C5—C6—C7—C8	0.4 (4)
C27—C22—C21—C16	178.24 (18)	C9—C8—C7—C6	-0.6 (3)
C15—N2—C14—C13	168.12 (16)	S1-C8-C7-C6	–177.61 (17)
C12—N2—C14—C13	-60.7 (2)	C21—C16—C17—C18	-2.6 (3)
C15—N3—C23—C24	-178.08 (17)	C15—C16—C17—C18	–178.9 (2)
C15—N3—C23—C22	3.3 (3)	C22—C27—C26—C25	–0.1 (3)
C27—C22—C23—N3	178.04 (17)	C24—C25—C26—C27	-0.8 (3)
C21—C22—C23—N3	-4.8 (3)	C16—C17—C18—C19	0.5 (4)
C27—C22—C23—C24	-0.5 (3)	C21—C20—C19—C18	–1.1 (4)
C21—C22—C23—C24	176.59 (17)	C17—C18—C19—C20	1.4 (4)

S26