A One-Pot, Three-Step Synthesis of α-Aminophosphonic Acids

Sergey V. Ryabukhin,*^{a,b} Dmitriy M. Panov,^b Alexander N. Shivanyuk,^a Andrey S. Plaskon,^b Evgeniy N. Zarudnitskiy,^c Oleg Lukin^d

^a The Institute of High Technologies, Kyiv National Taras Shevchenko University, 4 Glushkov str., Building 5, Kyiv 03187, Ukraine Fax +38(050)6424763; E-mail: s.v.ryabukhin@gmail.com

- ^b Department of Chemistry, Kyiv National Taras Shevchenko University, 64 Volodymyrska str., Kyiv 01601, Ukraine
- ^c Institute of Organic Chemistry, National Academy of Sciences of Ukraine, 5 Murmanska str., Kyiv 02094, Ukraine

^d Curplyx DRS, JSC Macrochem 7th Floor, KOMOD Business Center, 4 Lunacharski str., Kyiv 02002, Ukraine

Received: 05.02.2014; Accepted after revision: 06.03.2014

Abstract: A total of 20 α -aminophosphonic acids are synthesized in 73–89% yields via a one-pot, three-step procedure involving chlorotrimethylsilane-promoted condensation of carbonyl compounds and primary amines yielding azomethine intermediates, subsequent reaction with tris(trimethylsilyl) phosphite to give the corresponding trimethylsilylphosphonic esters and finally, mild cleavage with methanol–water. The wide scope and simple set-up and work-up procedures of the reported method allow the synthesis of a diverse set of α -aminophosphonic acids possessing two different functional groups.

Key words: α-aminophosphonic acids, aldehydes, amines, chlorotrimethylsilane, tris(trimethylsilyl) phosphite, one-pot synthesis

 α -Aminophosphonic acids 1 (Scheme 1) are analogues of α -amino acids in which the carboxylic group is replaced with a dihydroxyphosphoryl functionality.¹ α -Aminophosphonic acids possess a wide spectrum of biological activity.² For example, oligopeptides incorporating an α aminophosphonic acid group are excellent inhibitors of proteolytic enzymes.³ Additionally, many derivatives of α -aminophosphonic acids have been shown to possess antibacterial,⁴ antiviral⁵ and antifungal⁶ activities. Some α aminophosphonic acids, such as glyphosate, glufosinate and bilanafos are environmentally friendly commercial herbicides.⁷ In connection with the important applications of the title structures, a range of synthetic pathways to α aminophosphonic acids has been developed. Previously reported synthetic routes to α -aminophosphonic acids are summarized in Scheme 1.



Scheme 1 A basic retrosynthetic analysis of α-aminophosphonic acids

SYNTHESIS 2014, 46, 2079–2084 Advanced online publication: 11.04.2014 DOI: 10.1055/s-0033-1341064; Art ID: SS-2014-Z0091-OP © Georg Thieme Verlag Stuttgart · New York A few synthetic methodologies have been developed to vary the nature of the R^1 and R^2 groups in order to influence the biological activity of the α -aminophosphonic acids. For example, N-substituted α -aminomethanephosphonic acids were synthesized in one step through a Mannich-type reaction of a primary amine (R¹NH₂), formaldehyde (CH₂O) and phosphonic acid (H_3PO_3) .⁸ Both the R¹ and R² groups can be varied via a two-step approach comprising the synthesis and hydrolysis of α-aminophosphonates A (Scheme 1). Compounds A were prepared through either a one-pot Kabachnik-Fields reaction⁹ of carbonyl compounds, primary amines, and a dialkyl phosphonate [(AlkO)₂P(O)H] (Scheme 1), or via hydrophosphonylation of azomethines with a dialkyl phosphonate.¹⁰ A mild Lewis acid catalyzed hydrophosphonylation of azomethines afforded α-aminophosphonates in high yields.¹¹ Dialkyl α-aminophosphonates of type A were prepared in 85–98% yields through a one-pot chlorotrimethylsilane-promoted reaction of aldehydes with primary amines, followed by a lithium perchlorate (LiClO₄) activated reaction with a trialkyl phosphite $[P(OAlk)_3]^{12}$ Compounds A were also converted into α aminophosphonic acids through an acid-catalyzed hydrolysis¹³ or, in the case of dibenzylphosphonates, by catalytic hydrogenation.¹⁴ The reaction of α -aminophosphonates with bromotrimethylsilane (Me₃SiBr)¹⁵ or iodotrimethylsilane (Me₃SiI)¹⁶ resulted in the corresponding trimethylsilyl esters, which were then hydrolyzed with methanol-water. To avoid the transesterification step, trimethylsilylphosphonates were obtained directly through an Abramov-type reaction of tris(trimethylsilyl) phosphite [(Me₃SiO)₃P] with various electrophilic reagents. For example, the reaction of tris(trimethylsilyl) phosphite with carbonyl compounds followed by mild hydrolysis afforded α -hydroxyphosphonic acids,¹⁷ whereas treatment of acid chlorides or anhydrides with two molar equivalents of tris(trimethylsilyl) phosphite gave the corresponding hydroxymethylene bisphosphonic acids.¹⁸ Trimethylsilyl esters of α -aminophosphonic acids were also synthesized through the reactions of tris(trimethylsilyl) phosphite with aminoacetals¹⁹ and azomethines.²⁰

However, the procedures mentioned above cannot be used for the parallel synthesis of α -aminophosphonic acids on account of either a narrow scope, or technical complications with the two-step procedures, such as the isolation and purification of intermediates. In this contribution, we describe a one-pot, three-step synthesis of α -aminophosphonic acids. The process involves: (1) the chlorotrimethylsilane-promoted generation of azomethines from various aldehydes and amines, (2) reaction with tris(trimethylsilyl) phosphite to give trimethylsilylphosphonate intermediates, and (3) mild hydrolysis to afford the target α -aminophosphonic acids.

We previously reported a chlorotrimethylsilane-promoted combinatorial synthesis of azomethines from various amines and carbonyl compounds.²¹ In this connection, our attention was drawn by the aforementioned high-yielding, one-pot preparation of dialkyl a-aminophosphonates via the chlorotrimethylsilane/lithium perchlorate mediated reaction of aldehydes, primary amines, and a trialkyl phosphite.¹² The large excess of lithium perchlorate used in the reported procedure to facilitate the addition of the trialkyl phosphite to the azomethine intermediate complicated the isolation and purification steps. This fact makes the procedure, to a large degree, impractical from the combinatorial synthesis standpoint. In an attempt to optimize the one-pot preparation of α -aminophosphonates, we carried out a series of model experiments in which, as shown in Scheme 2, aniline (2a) was reacted with an equimolar amount of benzaldehyde (3a) in the presence of different quantities of chlorotrimethylsilane and lithium perchlorate in acetonitrile. After sonication of the solution containing 2a, 3a, chlorotrimethylsilane and lithium perchlorate at room temperature for two hours, triethyl phosphite $[P(OEt)_3]$ was added and the mixture was sonicated for another two hours. The formation of product 4a was monitored by ³¹P NMR spectroscopy of the reaction mixture.



Scheme 2 Preparation of diethyl α -aminophosphonate **4a**. *Reagents and conditions*: (i) Me₃SiCl, LiClO₄, r.t.; (ii) P(OEt)₃, r.t. See text for details.

The first experiment in which 2a, 3a, chlorotrimethylsilane and lithium perchlorate were mixed in a molar ratio of 1:1:1:10 showed a moderate conversion of 38%. Interestingly, the exclusion of lithium perchlorate in a second experiment, while leaving all the other reagents and conditions unchanged, led to the formation of nearly the same amount of 4a (42%). This observation seemed to indicate that chlorotrimethylsilane not only promoted the formation of the azomethine intermediate, but also the addition of triethyl phosphite to the latter. Therefore, it was envisaged that addition of a larger amount of chlorotrimethylsilane would lead to an increase in the conversion. Indeed, the use of two and three equivalents of chlorotrimethylsilane (with respect to 2a or 3a) afforded compound 4a in high (81%) and nearly quantitative (98%) conversions, respectively. Consequently, the optimization studies revealed that a 1:1:3 ratio of reactants ($2a:3a:Me_3SiCl$) in acetonitrile at room temperature were the best conditions for the synthesis of compound 4a. Our subsequent attempts at cleavage of the ethyl groups of 4a in the same reaction pot, by the action of bromotrimethylsilane and varying the reaction temperature were unsuccessful. On the basis of the obtained results and those reported in the literature (see the introductory section), we reasoned that the use of tris(trimethylsilyl) phosphite instead of triethyl phosphite should not influence the yield of the corresponding trimethylsilyl analogue of 4a (Scheme 2). At the same time, trimethylsilylphosphonates are known to hydrolyze smoothly, thereby simplifying the preparation of the α -aminophosphonic acids in the same reaction pot.



Scheme 3 Preparation of aminophosphonic acid **1aa**. *Reagents and conditions*: (i) Me₃SiCl, r.t.; (ii) P(OSiMe₃)₃, r.t.; (iii) MeOH–H₂O.

The one-pot, three-step synthesis of α -aminophosphonic acids was optimized using the reaction of aniline (2a) and benzaldehyde (3a) (Scheme 3) as a model system, by varying the solvent and the quantity of chlorotrimethylsilane. In a series of experiments similar to those carried out with the use of triethyl phosphite (see Scheme 2), it was found that addition of tris(trimethylsilyl) phosphite to the azomethine intermediate proceeded smoothly in the presence of three equivalents of chlorotrimethylsilane (with respect to 2a or 3a) in acetonitrile, 1,4-dioxane or THF to give α -aminophosphonate 5 (Scheme 3). Compound 5 was hydrolyzed in situ by simply triturating with methanol-water. It was found that the use of a 1:1:3:1 ratio of reactants [2a:3a:Me₃SiCl:P(OSiMe₃)₃] in anhydrous acetonitrile gave the highest yield of 1aa (optimized conditions). It is noteworthy that the simultaneous addition of 2a, 3a, chlorotrimethylsilane and tris(trimethylsilyl) phosphite under the optimized conditions, followed by triturating the reaction mixture with methanol-water gave an inseparable mixture of 1aa (70%) and the corresponding α -hydroxyphosphonic acid (30%). It was proved in a separate experiment [by the reaction of 3a with $P(OSiMe_3)_3$ in the presence of Me_3SiCl)] that the latter compound stemmed from a chlorotrimethylsilane-mediated addition of tris(trimethylsilyl) phosphite to aldehyde **3a**. Therefore, it is important to ensure complete conversion of 2a and 3a into the azomethine intermediate prior to the addition of tris(trimethylsilyl) phosphite. The optimization studies showed that a reaction time of two hours was sufficient for complete generation of the azomethine. As shown in Scheme 4, the optimum reaction conditions were applied to primary amines 2a-m (Figure 1) and carbonyl compounds 3a-j (Figure 2) to afford 20 α -aminophosphonic acids 1 in 73–89% yields. Compounds 1 normally precipitated from the reaction mixtures at room temperature. They were purified by filtering and washing with methanol. When precipitates did not form, the reaction mixtures were reduced in volume through partial evaporation of the solvent and crude compounds 1 were triturated with methanol, filtered, and washed with cold methanol. These simple work-up procedures afforded compounds 1 in >95% purity according to the NMR and LC–MS analyses.



Scheme 4 Preparation of aminophosphonic acids **1** using the optimized conditions: (i) Me₃SiCl, r.t., sonication, 2 h; (ii) P(OSiMe₃)₃, r.t., sonication, 2 h; (iii) MeOH–H₂O work-up.



Figure 1 The selection of amines used for the synthesis of α -aminophosphonic acids 1

The structures and compositions of compounds 1 were confirmed by ¹H, ¹³C and ³¹P NMR spectroscopy, LC–MS and elemental analysis. The ¹H NMR spectra of α -aminophosphonic acids 1 derived from aldehydes contained signals for the R¹ and R² groups and a pronounced doublet (²*J*_{H,P} = 15–25 Hz) at 4.1–5.3 ppm for the methine proton at the α -position relative to the phosphonate group. The ¹H NMR (DMSO-*d*₆) signals of the protons due to the NH and OH groups of compounds 1 were difficult to observe, presumably on account of their mutual exchange. Interest-



Figure 2 The selection of carbonyl compounds used for the synthesis of α -aminophosphonic acids 1

ingly, seven out of the 20 phosphonic acids reported here have been prepared before by other methods, and the reported melting regions for these compounds were often quite different to those obtained here (see the Supporting Information for details). The differences found in the reported and observed melting regions might be due to the different isolation procedures.

In conclusion, the described one-pot, three-step synthetic procedure was validated by preparing 20 structurally diverse α -aminophosphonic acids. The main benefits of the described method are easily available reagents, ambient conditions for all three reaction steps, and simple reaction work-up and product purification. These advantages indicate that the method should be easily adaptable for the parallel synthesis of sizable collections of α -aminophosphonic acids.

Solvents were purified according to standard procedures. Starting materials were purchased from Acros, Merck or Fluka. Analytical TLC was performed using Polychrom SI F254 plates. Column chromatography was performed using Kieselgel Merck 60 (230–400 mesh) silica gel as the stationary phase. A VWR USC 500 T ultrasonic bath was used for sonication. Melting points were measured on a Buchi melting point apparatus and are uncorrected. NMR spectra were recorded on a Bruker 170 Avance 500 spectrometer (500 MHz for ¹H and NOE, 125 MHz for ¹³C and 202 MHz for ³¹P), and a Varian Unity Plus 400 spectrometer (400.4 MHz for ¹H), using TMS as the internal standard. Mass spectra were recorded on an Agilent 1100 LCMSD SL instrument (CI) and Agilent 5890 Series II 5972 GC–MS instrument (EI). Elemental analyses were carried out on a Vario MICRO Cube (Elementar) Elemental Micro-Analyzer.

a-Aminophosphonic Acids 1; General Procedure

Me₃SiCl (0.43^2 g, 4 mmol) was added dropwise to a solution of amine **2** (1 mmol) and carbonyl compound **3** (1 mmol) in anhydrous MeCN (1.5-2 mL), and the resulting mixture was sonicated for 2 h at r.t. P(OSiMe₃)₃ (0.328 g, 1.1 mmol) was added dropwise and the mixture was sonicated for another 2 h at r.t. The mixture was diluted with MeOH (5 mL) and H₂O (0.1 mL, 5 mmol). Dilution normally caused precipitation of the product. The precipitated product was filtered and washed with MeOH (1 mL). In cases where no precipitation occurred, the mixture was further sonicated for 2 h and then left to stand for 24 h at 0 °C. Alternatively, the solvents were removed under reduced pressure and the residue was triturated with cold MeOH (2 mL) and filtered.

[Phenyl(phenylamino)methyl]phosphonic Acid (1aa) Yield: 0.23 g (87%); colorless crystals; mp 115–117 °C.

¹H NMR (500 MHz, DMSO- d_6): $\delta = 4.67$ (d, ² $J_{P,H} = 24.2$ Hz, 1 H), 6.52 (t, ³ $J_{H,H} = 7.3$ Hz, 1 H), 6.68 (d, ³ $J_{H,H} = 7.8$ Hz, 2 H), 7.02 (t, ³ $J_{H,H} = 7.8$ Hz, 2 H), 7.20 (t, ³ $J_{H,H} = 7.3$ Hz, 1 H), 7.28 (t, ³ $J_{H,H} = 7.3$ Hz, 2 H), 7.47 (d, ³ $J_{H,H} = 7.8$ Hz, 2 H).

¹³C NMR (125 MHz, DMSO-*d*₆): δ = 56.7 (d, ¹*J*_{C,P} = 142 Hz), 114.1, 117.3, 127.2, 128.2, 128.7 (d, ³*J*_{C,P} = 5 Hz), 129.1, 138.7, 147.6 (d, ²*J*_{C,P} = 14 Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 18.6$.

MS (APSI): $m/z = 262 [M - 1]^+$.

Anal. Calcd for $C_{13}H_{14}NO_3P$: C, 59.32; H, 5.36; N, 5.32. Found: C, 59.47; H, 5.27; N, 5.42.

[Phenyl(p-tolylamino)methyl]phosphonic Acid (1ba) Yield: 0.22 g (85%); colorless crystals; mp 139–141 °C.

¹H NMR (500 MHz, DMSO-*d*₆): δ = 2.10 (s, 3 H), 4.63 (d, ²*J*_{P,H} = 24.5 Hz, 1 H), 6.58 (d, ³*J*_{H,H} = 8.1 Hz, 2 H), 6.82 (d, ³*J*_{H,H} = 8.1 Hz, 2 H), 7.19 (t, ³*J*_{H,H} = 7.1 Hz, 1 H), 7.27 (t, ³*J*_{H,H} = 7.1 Hz, 2 H), 7.45 (d, ³*J*_{H,H} = 7.1 Hz, 2 H).

¹³C NMR (125 MHz, DMSO-*d*₆): δ = 20.5, 56.9 (d, ¹*J*_{C,P} = 145 Hz), 114.2, 125.8, 127.1, 128.2, 128.6 (d, ³*J*_{C,P} = 5 Hz), 129.6, 138.8, 145.3 (d, ²*J*_{C,P} = 14 Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 18.9$.

MS (APSI): $m/z = 276 [M - 1]^+$.

Anal. Calcd for $C_{14}H_{16}NO_3P$: C, 60.65; H, 5.82; N, 5.05. Found: C, 60.52; H, 5.69; N, 5.12.

[(Benzylamino)(Phenyl)methyl]phosphonic Acid (1ca) Yield: 0.21 g (78%); colorless crystals; mp 238–240 °C.

¹H NMR (500 MHz, DMSO-*d*₆): δ = 4.09 (d, ²*J*_{H,H} = 13.4 Hz, 1 H), 4.15 (d, ²*J*_{H,H} = 13.4 Hz, 1 H), 4.27 (d, ²*J*_{P,H} = 17.0 Hz, 1 H), 7.30–7.35 (m, 8 H), 7.45 (d, ³*J*_{H,H} = 6.3 Hz, 2 H).

¹³C NMR (125 MHz, DMSO-*d*₆): δ = 50.3 (d, ³*J*_{C,P} = 5 Hz), 57.7 (d, ¹*J*_{C,P} = 147 Hz), 128.6, 128.7, 129.1 (d, ³*J*_{C,P} = 2 Hz), 129.3, 129.8, 129.9, 130.8, 131.1 (d, ²*J*_{C,P} = 5 Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 11.5$.

MS (APSI): $m/z = 276 [M - 1]^+$.

Anal. Calcd for $C_{14}H_{16}NO_3P$: C, 60.65; H, 5.82; N, 5.05. Found: C, 60.55; H, 5.70; N, 5.16.

[(Phenethylamino)(phenyl)methyl]phosphonic Acid (1da) Yield: 0.22 g (75%); colorless crystals; mp 263–265 °C.

¹H NMR (500 MHz, DMSO-*d*₆): δ = 2.89–3.00 (m, 2 H), 3.00–3.10 (m, 1 H), 3.14–3.22 (m, 1 H), 4.51 (d, ²*J*_{P,H} = 17.0 Hz, 1 H), 7.05 (d, ³*J*_{H,H} = 7.3 Hz, 2 H), 7.12 (t, ³*J*_{H,H} = 7.3 Hz, 1 H), 7.15–7.21 (m, 2 H), 7.31–7.38 (m, 3 H), 7.52 (d, ³*J*_{H,H} = 6.8 Hz, 2 H).

¹³C NMR (125 MHz, DMSO-*d*₆): δ = 31.5, 48.4, 59.2 (d, ${}^{1}J_{C,P}$ = 144 Hz), 126.9, 128.6, 128.7, 128.8, 129.1, 129.7 (d, ${}^{2}J_{C,P}$ = 6 Hz), 131.3 (d, ${}^{3}J_{C,P}$ = 5 Hz), 137.2.

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 11.9$.

MS (APSI): $m/z = 290 [M - 1]^+$.

Anal. Calcd for $C_{15}H_{18}NO_3P$: C, 61.85; H, 6.23; N, 4.81. Found: C, 61.64; H, 6.10; N, 4.84.

[(Isopropylamino)(phenyl)methyl]phosphonic Acid (1ea) Yield: 0.18 g (79%); colorless crystals; mp 241–243 °C.

¹H NMR (500 MHz, CF₃COOD): δ = 1.56 (t, ³*J*_{H,H} = 7.6 Hz, 6 H), 3.50–3.74 (m, 1 H), 5.02 (d, ²*J*_{P,H} = 14.7 Hz, 1 H), 7.63 (s, 5 H). ¹³C NMR (125 MHz, CF₃COOD): δ = 17.6, 19.6, 48.8 (d, ³*J*_{C,P} = 4 Hz), 59.2 (d, ¹*J*_{C,P} = 145 Hz), 126.8, 128.4, 129.7, 130.6. ³¹P NMR (202 MHz, CF₃COOD): δ = 14.4.

MS (APSI): $m/z = 228 [M - 1]^+$.

Anal. Calcd for $C_{10}H_{16}NO_3P$: C, 52.40; H, 7.04; N, 6.11. Found: C, 52.32; H, 6.89; N, 6.27.

[(Butylamino)(phenyl)methyl]phosphonic Acid (1fa) Yield: 0.17 g (73%); colorless crystals; mp 231–233 °C

¹H NMR (500 MHz, DMSO-*d*₆): δ = 0.76 (t, ${}^{3}J_{\rm H,H}$ = 7.3 Hz, 3 H), 1.10–1.24 (m, 2 H), 1.53–1.67 (m, 2 H), 2.74–2.79 (m, 1 H), 2.87–2.95 (m, 1 H), 4.46 (d, ${}^{2}J_{\rm P,H}$ = 16.9 Hz, 1 H), 7.34–7.38 (m, 3 H), 7.52 (d, ${}^{3}J_{\rm H,H}$ = 7.2 Hz, 2 H).

¹³C NMR (125 MHz, DMSO-*d*₆): δ = 12.9, 19.4, 27.1, 47.0 (d, ${}^{3}J_{C,P}$ = 5 Hz), 59.6 (d, ${}^{1}J_{C,P}$ = 144 Hz), 128.7, 129.0, 129.7 (d, ${}^{2}J_{C,P}$ = 5 Hz), 131.5 (d, ${}^{3}J_{C,P}$ = 4 Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 12.0$.

MS (APSI): $m/z = 242 [M - 1]^+$.

Anal. Calcd for $C_{11}H_{18}NO_3P$: C, 54.32; H, 7.46; N, 5.76. Found: C, 54.18; H, 7.55; N, 5.88.

[(Cyclohexylamino)(phenyl)methyl]phosphonic Acid (1ga) Yield: 0.20 g (78%); colorless crystals; mp 192–194 °C.

¹H NMR (500 MHz, DMSO-*d*₆): δ = 0.90–1.11 (m, 3 H), 1.21–1.39 (m, 2 H), 1.48–1.58 (m, 1 H), 1.63–1.69 (m, 2 H), 1.95–2.08 (m, 1 H), 2.89–2.97 (m, 1 H), 4.51 (d, ${}^{2}J_{\rm P,H}$ = 17.1 Hz, 2 H), 7.40 (s, 3 H), 7.55 (s, 2 H).

¹³C NMR (125 MHz, DMSO- d_6): δ = 23.6, 23.7, 27.4, 30.1 (d, ³ $J_{C,P}$ = 4 Hz), 57.4 (d, ¹ $J_{C,P}$ = 145 Hz), 126.6, 128.5 (d, ² $J_{C,P}$ = 6 Hz), 129.7, 130.7.

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 12.1$.

MS (APSI): $m/z = 268 [M - 1]^+$.

Anal. Calcd for $C_{13}H_{20}NO_3P$: C, 57.99; H, 7.49; N, 5.20. Found: C, 57.82; H, 7.35; N, 5.28.

({[2-(Dimethylamino)ethyl]amino}[phenyl]methyl)phosphonic Acid (1ha)

Yield: 0.21 g (82%); colorless crystals; mp 162–163 °C.

¹H NMR (500 MHz, DMSO-*d*₆): δ = 2.48 (s, 6 H), 2.67–2.81 (m, 1 H), 2.84–2.96 (m, 1 H), 3.06–3.16 (m, 2 H), 4.08 (d, ${}^{2}J_{P,H}$ = 18.1 Hz, 1 H), 7.25–7.37 (m, 1 H), 7.53–7.62 (m, 2 H).

¹³C NMR (125 MHz, DMSO-*d*₆): δ = 42.9, 44.5, 55.5 (d, ${}^{3}J_{C,P}$ = 4 Hz), 61.5 (d, ${}^{1}J_{C,P}$ = 137 Hz), 127.4, 128.3, 129.5, 137.5 (d, ${}^{2}J_{C,P}$ = 5 Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 11.7$.

MS (APSI): $m/z = 257 [M - 1]^+$.

Anal. Calcd for $C_{11}H_{19}N_2O_3P$: C, 51.16; H, 7.42; N, 10.85. Found: C, 51.03; H, 7.38; N, 11.01.

({[3-(Dimethylamino)propyl]amino}[phenyl]methyl)phosphonic Acid (1ia)

Yield: 0.23 g (82%); colorless crystals; mp 156–158 °C.

¹H NMR (500 MHz, DMSO-*d*₆): δ = 2.12–2.22 (m, 2 H), 2.68 (s, 6 H), 2.83–2.90 (m, 1 H), 3.01–3.21 (m, 3 H), 4.51 (d, ${}^{2}J_{P,H}$ = 16.5 Hz, 1 H), 7.37–7.47 (m, 3 H), 7.58–7.67 (m, 2 H).

¹³C NMR (125 MHz, DMSO- d_6): δ = 20.8, 42.3, 44.4, 53.9, 60.2, (d, ${}^{1}J_{C,P}$ = 138 Hz), 128.8, 128.9, 130.0 (d, ${}^{3}J_{C,P}$ = 5 Hz), 132.8 (d, ${}^{2}J_{C,P}$ = 4 Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 10.8$.

MS (APSI): $m/z = 271 [M - 1]^+$.

Anal. Calcd for $C_{12}H_{21}N_2O_3P{:}\ C,\ 52.93;\ H,\ 7.77;\ N,\ 10.29.$ Found: C, 52.80; H, 7.72; N, 10.34.

[({2-[Methyl(phenyl)amino]ethyl}amino)(phenyl)methyl]phosphonic Acid (1ja)

Ŷield: 0.28 g (89%); colorless crystals; mp 222–224 °C.

¹H NMR (500 MHz, CF₃COOD): δ = 3.48 (s, 3 H), 3.47–3.57 (m, 2 H), 4.26–4.38 (m, 2 H), 4.86 (d, ${}^{2}J_{P,H}$ = 17.2 Hz, 1 H), 7.45–7.62 (m, 7 H), 7.62–7.67 (m, 3 H).

¹³C NMR (125 MHz, CF₃COOD): δ = 42.5, 46.8, 54.8, 62.2 (d, ${}^{1}J_{C,P} = 143$ Hz), 119.9, 125.9, 128.4 (d, ${}^{3}J_{C,P} = 6$ Hz), 129.8, 131.1, 131.3, 131.9, 138.1 (d, ${}^{2}J_{C,P} = 9$ Hz).

³¹P NMR (202 MHz, CF₃COOD): δ = 12.5, 12.6.

MS (APSI): $m/z = 319 [M - 1]^+$.

Anal. Calcd for C₁₆H₂₁N₂O₃P: C, 59.99; H, 6.61; N, 8.75. Found: C, 59.87; H, 6.54; N, 8.80.

[({2-[(4-Chlorophenyl)thio]ethyl}amino)(phenyl)methyl]phosphonic Acid (1ka) Yield: 0.30 g (82%); colorless crystals; mp 258–260 °C.

¹H NMR (500 MHz, CF₃COOD): δ = 3.23–3.32 (m, 3 H), 3.41–3.51 (m, 1 H), 4.86 (d, ${}^{2}J_{P,H} = 17.8$ Hz, 1 H), 7.18–7.30 (m, 4 H), 7.38– 7.48 (m, 4 H), 7.48–7.62 (m, 1 H).

¹³C NMR (125 MHz, CF₃COOD): δ = 29.8, 46.2, 61.1 (d, ¹J_{CP} = 144 Hz), 126.3, 128.3 (d, ${}^{3}J_{C,P} = 5$ Hz), 129.4, 129.5, 129.7, 130.7, 132.1, 134.8.

³¹P NMR (202 MHz, CF₃COOD): δ = 13.5.

MS (APSI): $m/z = 356 [M - 1]^+$.

Anal. Calcd for C₁₅H₁₇ClNO₃PS: C, 50.35; H, 4.79; N, 3.91. Found: C, 50.22; H, 4.71; N, 4.04.

[(4-Fluorophenyl)(phenylamino)methyl]phosphonic Acid (1ab) Yield: 0.24 g (84%); colorless crystals; mp 187–189 °C.

¹H NMR (500 MHz, DMSO- d_6): $\delta = 4.70$ (d, ² $J_{P,H} = 24.4$ Hz, 1 H), 6.53 (t, ${}^{3}J_{H,H} = 7.1$ Hz, 1 H), 6.67 (d, ${}^{3}J_{H,H} = 8.0$ Hz, 2 H), 7.01 (t, ${}^{3}J_{\text{H,H,F}} = 8.0 \text{ Hz}, 2 \text{ H}$, 7.11 (t, ${}^{3}J_{\text{H,H,F}} = 8.0 \text{ Hz}, 2 \text{ H}$), 7.45–7.55 (m, 2 H).

¹³C NMR (125 MHz, DMSO- d_6): $\delta = 55.7$ (d, ¹ $J_{CP} = 146$ Hz), 113.8, 115.0 (d, ${}^{2}J_{C,F}$ = 22 Hz), 117.1, 129.2, 130.5 (m), 135.1, 147.8 (d, ${}^{2}J_{C,P} = 14$ Hz), 161.2 (d, ${}^{1}J_{C,F} = 243$ Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 16.8$.

MS (APSI): $m/z = 280 [M - 1]^+$.

Anal. Calcd for C₁₃H₁₃FNO₃P: C, 55.52; H, 4.66; N, 4.98. Found: C, 55.43; H, 4.50; N, 5.08.

[(4-Methoxyphenyl)(phenylamino)methyl]phosphonic Acid (1ac)

Yield: 0.22 g (77%); colorless crystals; mp 162–163 °C.

¹H NMR (500 MHz, DMSO- d_6): $\delta = 3.71$ (s, 3 H), 4.58 (d, ² $J_{P,H} =$ 17.4 Hz, 1 H), 6.51 (t, ${}^{3}J_{H,H} = 7.8$ Hz, 1 H), 6.66 (d, ${}^{3}J_{H,H} = 7.8$ Hz, 2 H), 6.85 (d, ${}^{3}J_{H,H} = 7.8$ Hz, 2 H), 7.00 (t, ${}^{3}J_{H,H} = 7.8$ Hz, 2 H), 7.37 (d, ${}^{3}J_{\rm H,H} = 7.8$ Hz, 2 H).

¹³C NMR (125 MHz, DMSO- d_6): $\delta = 55.5$ (d, ¹ $J_{C,P} = 147$ Hz), 55.6, 113.7, 116.9, 129.1, 129.7 (d, ${}^{3}J_{C,P} = 5$ Hz), 130.7 (d, ${}^{3}J_{C,P} = 3$ Hz), 148.1 (d, ${}^{2}J_{C,P} = 15$ Hz), 159.2.

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 19.3$.

MS (APSI): $m/z = 292 [M - 1]^+$.

Anal. Calcd for C₁₄H₁₆NO₄P: C, 57.34; H, 5.50; N, 4.78. Found: C, 57.17; H, 5.43; N, 4.88.

[(Phenylamino)(*m*-tolyl)methyl]phosphonic Acid (1ad) Yield: 0.23 g (82%); colorless crystals; mp 130–132 °C.

¹H NMR (500 MHz, DMSO- d_6): $\delta = 2.27$ (s, 3 H), 4.60 (d, ² $J_{H,P} =$ 15.2 Hz, 1 H), 6.52 (t, ${}^{3}J_{H,H} = 7.1$ Hz, 1 H), 6.57 (d, ${}^{3}J_{H,H} = 8.1$ Hz, 2 H), 6.98–7.06 (m, 3 H), 7.18 (t, ${}^{3}J_{H,H} = 7.5$ Hz, 1 H), 7.25 (d, ${}^{3}J_{\rm H,H} = 7.5$ Hz, 1 H), 7.29 (s, 1 H).

¹³C NMR (125 MHz, DMSO- d_6): $\delta = 21.6$, 56.5 (d, ¹ $J_{C,P} = 149$ Hz), 113.7, 116.9, 125.8 (d, ${}^{3}J_{C,P} = 5$ Hz), 127.9 (d, ${}^{3}J_{C,P} = 3$ Hz), 128.1, 129.1, 129.2, 137.1, 138.9 (d, ${}^{3}J_{C,P} = 2$ Hz), 148.1 (d, ${}^{2}J_{C,P} = 14$ Hz). ³¹P NMR (202 MHz, DMSO- d_6): $\delta = 19.0$.

MS (APSI): $m/z = 276 [M - 1]^+$.

Anal. Calcd for $C_{14}H_{16}NO_3P$: C, 60.65; H, 5.82; N, 5.05. Found: C, 60.57; H, 5.70; N, 5.18.

[(Phenylamino)(thien-3-yl)methyl]phosphonic Acid (1ae) Yield: 0.24 g (88%); colorless crystals; mp 132–133 °C

¹H NMR (500 MHz, DMSO- d_6): $\delta = 4.82$ (d, ² $J_{P,H} = 23.5$ Hz, 1 H), 6.56 (t, ${}^{3}J_{H,H}$ = 7.2 Hz, 1 H), 6.74 (d, ${}^{3}J_{H,H}$ = 7.6 Hz, 2 H), 7.02 (t, ${}^{3}J_{\rm H,H} = 7.6$ Hz, 2 H), 7.16–7.22 (m, 1 H), 7.38–7.42 (m, 2 H).

¹³C NMR (125 MHz, DMSO- d_6): $\delta = 52.7$ (d, ¹ $J_{C,P} = 147$ Hz), 114.0, 117.5, 123.1 (d, ³ $J_{C,P} = 8$ Hz), 125.5, 128.7 (d, ³ $J_{C,P} = 4$ Hz), 129.2, 139.6, 147.6 (d, ${}^{2}J_{C,P} = 13$ Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 18.4$.

MS (APSI): $m/z = 268 [M - 1]^+$.

Anal. Calcd for C₁₁H₁₂NO₃PS: C, 49.07; H, 4.49; N, 5.20. Found: C, 48.95; H, 4.44; N, 5.31.

[(2-Hydroxyphenyl)(phenylamino)methyl]phosphonic Acid (1af)

Yield: 0.20 g (75%); colorless crystals; mp 131–134 °C.

¹H NMR (500 MHz, DMSO- d_6): $\delta = 5.01$ (d, ² $J_{P,H} = 24.3$ Hz, 1 H), 6.51 (t, ³ $J_{H,H} = 7.3$ Hz, 1 H), 6.68 (d, ³ $J_{H,H} = 8.2$ Hz, 2 H), 6.72 (t, ³ $J_{H,H} = 7.3$ Hz, 1 H), 6.89 (d, ³ $J_{H,H} = 7.3$ Hz, 1 H), 6.96–7.05 (m, 3 H), 7.36 (d, ${}^{3}J_{H,H} = 7.3$ Hz, 1 H).

¹³C NMR (125 MHz, DMSO- d_6): $\delta = 49.2$ (d, ¹ $J_{C,P} = 150$ Hz), 113.4, 115.8, 116.9, 119.4, 125.5, 128.1 (d, ${}^{3}J_{C,P} = 6$ Hz), 128.9 (d, ${}^{3}J_{C,P} =$ 4 Hz), 129.1, 148.8 (d, ${}^{2}J_{C,P} = 15$ Hz), 155.7 (d, ${}^{3}J_{C,P} = 6$ Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 20.4$.

MS (APSI): $m/z = 278 [M - 1]^+$.

Anal. Calcd for C₁₃H₁₄NO₄P: C, 55.92; H, 5.05; N, 5.02. Found: C, 55.87; H, 4.94; N, 5.17.

[(4-Acetamidophenyl)(benzylamino)methyl]phosphonic Acid (1cg)

Yield: 0.26 g (77%); colorless solid; mp 272-274 °C.

¹H NMR (500 MHz, DMSO- d_6): $\delta = 2.05$ (s, 3 H), 3.72 (d, ² $J_{H,H} =$ 11.3 Hz, 1 H), 3.80 (d, ${}^{2}J_{H,P}$ = 15.9 Hz, 1 H), 4.13 (d, ${}^{2}J_{H,H}$ = 11.3 Hz, 1 H), 7.30–7.45 (m, 7 H), 7.51–7.61 (m, 2 H), 10.18 (br s, 1 H).

¹³C NMR (125 MHz, DMSO- d_6): $\delta = 24.2, 49.3, 59.3$ (d, ¹ $J_{C,P} = 134$ Hz), 119.1, 129.0, 129.1 (d, ${}^{3}J_{C,P} = 5$ Hz), 130.3 (d, ${}^{2}J_{C,P} = 8$ Hz), 132.7, 134.6, 139.4, 162.8, 168.9.

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 9.1$.

MS (APSI): $m/z = 333 [M - 1]^+$.

Anal. Calcd for C₁₆H₁₉N₂O₄P: C, 57.48; H, 5.73; N, 8.38. Found: C, 57.57; H, 5.80; N, 8.26.

{[(4-Fluorobenzyl)amino](4-nitrophenyl)methyl}phosphonic Acid (11h)

Yield: 0.27 g (79%); colorless crystals; mp 244-246 °C.

¹H NMR (500 MHz, DMSO- d_6): $\delta = 4.14-4.21$ (m, 2 H), 4.51 (d, ${}^{2}J_{P,H} = 17.1$ Hz, 1 H), 7.13 (t, ${}^{3}J_{H,H,F} = 8.5$ Hz, 2 H), 7.50 (t, ${}^{3}J_{H,H,F} =$ 6.0 Hz, 2 H), 7.79 (d, ${}^{3}J_{H,H} = 8.2$ Hz, 2 H), 8.20 (d, ${}^{3}J_{H,H} = 8.2$ Hz, 2 H).

¹³C NMR (125 MHz, DMSO- d_6): $\delta = 49.7$, 58.2 (d, ¹ $J_{C,P} = 134$ Hz), 115.7 (d, ${}^{2}J_{C,F}$ = 22 Hz), 123.7, 128.1 (d, ${}^{3}J_{C,F}$ = 3 Hz), 131.1 (d, ${}^{3}J_{C,P} = 4$ Hz), 133.4 (d, ${}^{2}J_{C,P} = 11$ Hz), 140.9, 147.7, 162.4 (d, ${}^{1}J_{C,F} = 55$ Hz).

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 10.2$.

MS (APSI): $m/z = 339 [M - 1]^+$.

Anal. Calcd for $C_{14}H_{14}FN_2O_5P$: C, 49.42; H, 4.15; N, 8.23. Found: C, 49.49; H, 4.26; N, 8.11.

({[(1-Phenyl-1*H*-pyrazol-4-yl)methyl]amino}(thien-2-yl)methyl)phosphonic Acid (1mi)

Yield: 0.29 g (82%); colorless crystals; mp 246-247 °C.

¹H NMR (500 MHz, DMSO-*d*₆): δ = 4.43 (d, ²*J*_{H,H} = 13.7 Hz, 1 H), 4.57 (d, ²*J*_{H,H} = 13.7 Hz, 1 H), 5.32 (d, ²*J*_{P,H} = 17.6 Hz, 1 H), 7.14–7.22 (m, 1 H), 7.43 (br s, 1 H), 7.50–7.74 (m, 6 H), 8.56 (d, ³*J*_{H,H} = 8.5 Hz, 2 H).

¹³C NMR (125 MHz, DMSO-*d*₆): δ = 40.1, 57.1 (d, ${}^{1}J_{C,P}$ = 147 Hz), 114.1, 122.5, 126.2 (d, ${}^{3}J_{C,P}$ = 5 Hz), 125.1, 129.9, 130.5, 132.2 (d, ${}^{2}J_{C,P}$ = 7 Hz), 132.4, 133.5, 136.8, 137.4.

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 11.6$.

MS (APSI): $m/z = 348 [M - 1]^+$.

Anal. Calcd for $C_{15}H_{16}N_3O_3PS$: C, 51.57; H, 4.62; N, 12.03. Found: C, 51.73; H, 4.73; N, 11.87.

[1-(Phenylamino)cyclohexyl]phosphonic Acid (1aj) Yield: 0.19 g (74%); colorless crystals; mp 161–164 °C.

¹H NMR (500 MHz, DMSO-*d*₆): δ = 1.16–1.24 (m, 1 H), 1.37–1.52 (m, 5 H), 1.68–1.76 (m, 2 H), 2.12–2.18 (m, 2 H), 6.74 (t, ${}^{3}J_{\rm H,H}$ = 7.2 Hz, 1 H), 7.10 (t, ${}^{3}J_{\rm H,H}$ = 7.2 Hz, 2 H), 7.16 (d, ${}^{3}J_{\rm H,H}$ = 7.2 Hz, 2 H). ¹³C NMR (125 MHz, DMSO-*d*₆): δ = 20.4 (d, ${}^{3}J_{\rm C,P}$ = 4 Hz), 25.4 (d, ${}^{2}J_{\rm C,P}$ = 8 Hz), 29.6, 57.2 (d, ${}^{1}J_{\rm C,P}$ = 158 Hz), 119.2, 119.5, 128.6, 144.4.

³¹P NMR (202 MHz, DMSO- d_6): $\delta = 24.9$.

MS (APSI): $m/z = 254 [M - 1]^+$.

Anal. Calcd for $C_{12}H_{18}NO_3P$: C, 56.47; H, 7.11; N, 5.49. Found: C, 56.38; H, 7.00; N, 5.62.

Supporting Information for this article is available online at http://www.thieme-connect.com/ejournals/toc/synthesis.

References

- (a) Aminophosphonic and Aminophosphinic Acids: Chemistry and Biological Activity; Kukhar, V. P.; Hudson, H. R., Eds.; John Wiley & Sons: Chichester, 2000.
 (b) Kudzin, Z. H.; Kudzin, M. H.; Drabowicz, J.; Stevens, C. V. Curr. Org. Chem. 2011, 15, 2015. (c) Turcheniuk, K. V.; Kukhar, V. P.; Röschenthaler, G.-V.; Aceña, J. L.; Soloshonok, V. A.; Sorochinsky, A. E. RSC Adv. 2013, 3, 6693. (d) Ali, T. E. ARKIVOC 2014, (i), 21.
- (2) (a) Mucha, A.; Kafarski, P.; Berlicki, L. J. Med. Chem.
 2011, 54, 5955. (b) Romanenko, V. D.; Kukhar, V. P. Chem. Rev. 2006, 106, 3868. (c) Naydenova, E. D.; Todorov, P. T.; Troev, K. D. Amino Acids 2010, 38, 23.
- (3) Hiratake, J.; Oda, J. *Biosci. Biotechnol. Biochem.* **1997**, *61*, 211.

- (4) (a) Allen, J. G.; Atherton, F. R.; Hall, M. J.; Hassall, C. H.; Holmes, S. W.; Lambert, R. W.; Nisbet, L. J.; Ringrose, P. S. *Nature* 1978, 272, 56. (b) Pratt, R. F. *Science* 1989, 246, 917.
- (5) Chen, M.-H.; Chen, Z.; Song, B.-A.; Bhadury, P. S.; Yang, S.; Cai, X.-J.; Hu, D.-Y.; Xue, W.; Zeng, S. J. Agric. Food Chem. 2009, 57, 1383.
- (6) Maier, L.; Diel, P. J. Phosphorus, Sulfur Silicon Relat. Elem. 1991, 57, 57.
- (7) Duke, S. O.; Powles, S. B. Pest Manag. Sci. 2008, 64, 319.
- (8) (a) Moedritzer, K.; Irani, R. R. J. Org. Chem. 1966, 31, 1603. (b) Lazar, I.; Sherry, A. D. Synthesis 1995, 453. (c) Bazakas, K.; Lukes, I. J. Chem. Soc., Dalton Trans. 1995, 1133. (d) Convey, P. V.; Davey, R. J.; Griffin, J. L.; Whiting, A. Chem. Commun. 1998, 1467.
- (9) (a) Kabachnik, M. I.; Medved, T. Ya.; Dyatlova, N. M.; Arkhipova, O. G.; Rudomino, M. V. Russ. Chem. Rev. (Engl. Transl.) 1968, 38, 503. (b) Gallardo-Macias, R.; Nakayama, K. Synthesis 2010, 57.
- (10) (a) Alfonsov, V. A. *Phosphorus, Sulfur Silicon Relat. Elem.* **2008**, *183*, 2637. (b) Issleib, K.; Ralszuweit, A.; Richter, H.-J.; Tonk, W. Z. Chem. **1983**, *23*, 434.
- (11) Firouzabadi, H.; Iranpoor, N.; Sobhani, S. Synthesis 2004, 2692.
- (12) Saidi, M. R.; Azizi, N. Synlett 2002, 1347.
- (13) (a) Kudzin, Z. H.; Kotynski, A. *Synthesis* 1980, 1028.
 (b) Huber, R.; Knierzinger, A.; Obrecht, J.-P.; Vasella, A. *Helv. Chim. Acta* 1985, *68*, 1730. (c) Liavielle, G.; Hautefaye, P.; Schaeffer, C.; Boutin, J. A.; Cudennec, C. A.; Pierre, A. J. Med. Chem. 1991, *34*, 1998.
- (14) (a) Bird, J.; Mello, R. C. D.; Harper, G. P.; Hunter, D. J.; Karran, E. H. J. Med. Chem. 1994, 37, 158. (b) Jacobsen, E. N.; Joly, G. D. J. Am. Chem. Soc. 2004, 126, 4102.
- (15) (a) Maier, L.; Diel, P. J. *Phosphorus, Sulfur Silicon Relat. Elem.* **1991**, *57*, 57. (b) Cherenok, S.; Kalchenko, V.; Solovyov, A.; Tsymbal, I.; Failla, S.; Consiglio, G. A.; Finocchiaro, P. *Heteroat. Chem.* **2001**, *12*, 58.
- (16) Ntai, I.; Bachmann, B. O. Bioorg. Med. Chem. Lett. 2008, 18, 3068.
- (17) (a) Bouchemal, N.; Lecouvey, M.; Mallard, I.; Migianu, E. *Tetrahedron Lett.* 2004, 45, 4511. (b) Guliaiko, I. V.; Kolodiazhnyi, O. I. *Phosphorus, Sulfur Silicon Relat. Elem.* 2008, 183, 677.
- (18) (a) Migianu-Griffoni, E.; Mbemba, C.; Burgada, R.; Lecouvey, M.; Lecercle, D.; Taran, F. *Tetrahedron* 2009, 65, 1517. (b) Zhang, Y.; Leon, A.; Song, Y.; Studer, D.; Haase, C.; Koscielski, L. A.; Oldfield, E. *J. Med. Chem.* 2006, 49, 5804.
- (19) Nifant'ev, E. E.; Popkova, T. N.; Kukhareva, T. S.; Bekker, A. R. J. Gen. Chem. USSR (Engl. Transl.) 1988, 58, 1550.
- (20) (a) Boduszek, B.; Lipinski, M.; Kowalska, M. W. *Phosphorus, Sulfur Silicon Relat. Elem.* 1998, 143, 179.
 (b) Mlynarz, P.; Rydzewska, A.; Sliwinska, S.; Szymczyk, M. Phosphorus, Sulfur Silicon Relat. Elem. 2009, 184, 1496.
- (21) Ryabukhin, S. V.; Panov, D. M.; Plaskon, A. S.; Chuprina, A.; Tolmachev, A. A.; Shivanyuk, A. N. *Mol. Diversity* 2012, *16*, 625.