Accepted Manuscript

Development of posaconazole-based analogues as hedgehog signaling pathway inhibitors

Kelly A. Teske, Radha Charan Dash, Shana R. Morel, Lianne Q. Chou, Robert J. Wechsler-Reya, M. Kyle Hadden

PII: S0223-5234(18)31015-8

DOI: https://doi.org/10.1016/j.ejmech.2018.11.056

Reference: EJMECH 10914

To appear in: European Journal of Medicinal Chemistry

Received Date: 24 September 2018

Revised Date: 13 November 2018

Accepted Date: 22 November 2018

Please cite this article as: K.A. Teske, R.C. Dash, S.R. Morel, L.Q. Chou, R.J. Wechsler-Reya, M.K. Hadden, Development of posaconazole-based analogues as hedgehog signaling pathway inhibitors, *European Journal of Medicinal Chemistry* (2018), doi: https://doi.org/10.1016/j.ejmech.2018.11.056.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Development of Posaconazole-based Analogues as Hedgehog Signaling Pathway Inhibitors

Kelly A. Teske^{1,#}, Radha Charan Dash¹, Shana R. Morel¹ Lianne Q. Chou², Robert J. Wechsler-Reya², and M. Kyle Hadden^{1,*}

¹Department of Pharmaceutical Sciences, University of Connecticut, 69 N Eagleville Rd, Unit 3092, Storrs, CT, USA 06269-3092

²Tumor Initiation and Maintenance Program, NCI-Designated Cancer Center, Sanford Burnham

Prebys Medical Discovery Institute, 10901 North Torrey Pines Road, La Jolla CA 92037, United

States

*Corresponding author: <u>kyle.hadden@uconn.edu</u>, Ph: 1-860-486-8446.

[#]Current address: Department of Chemistry, Western Michigan University, Email: Kelly.teske@wmich.edu.

ABSTRACT

Inhibition of the hedgehog (Hh) signaling pathway has been validated as a therapeutic strategy to treat basal cell carcinoma and holds potential for several other forms of human cancer. Itraconazole and posaconazole are clinically useful triazole anti-fungals that are being repurposed as anti-cancer agents based on their ability to inhibit the Hh pathway. We have previously demonstrated that removal of the triazole from itraconazole does not affect its ability to inhibit the Hh pathway while abolishing its primary side effect, potent inhibition of Cyp3A4. To develop structure-activity relationships for the related posaconazole scaffold, we synthesized and evaluated a series of *des*-triazole analogues designed through both ligand- and structure-based methods. These compounds demonstrated improved anti-Hh properties compared to posaconazole and enhanced stability without inhibiting Cyp3A4. In addition, we utilized a series of molecular dynamics and binding energy studies to probe specific interactions between the tetrahydrofuran region of the scaffold projects out of the binding site and that π - π interactions between the compound and Smoothened play a key role in stabilizing the bound analogues.

1. Introduction

The hedgehog (Hh) signaling pathway plays an essential role in embryonic development by regulating the proliferation and differentiation of cells in a time-dependent manner [1-3]. Hh signaling is significantly less active in adults with the exception of maintaining stem cell homeostasis, predominately in the skin and central nervous system [2]. Aberrant activation of the Hh signaling pathway has been implicated as a driving force in several cancers; most notably, basal cell carcinoma (BCC) and medulloblastoma (MB) [4-5]. Constitutive activation of the Hh pathway in these cancers is primarily caused by mutations in several key members of the Hh signal cascade, including Patched (Ptch), Smoothened (Smo), and Suppressor of fused (Sufu) [6]. Current FDA approved drugs that target the Hh pathway (vismodegib and sonidegib) function through direct antagonism of Smo; however, resistance due to point mutations in the binding pocket of these drugs on Smo has been identified in both BCC and MB patients who used these drugs for initial treatment [7-11]. The need to develop potent inhibitors that overcome the resistance observed with vismodegib and sonidegib has led to the identification of multiple small-molecule scaffolds that target different points along the Hh signaling pathway. [12-16]

Posaconazole (PSZ) and itraconazole (ITZ) are clinically approved azole antifungals that have been identified as Hh pathway inhibitors in vitro and in vivo [17-20]. Previous studies strongly suggest that both of these drugs inhibit the Hh pathway by directly binding Smo [17-18, 21-22]. PSZ and ITZ retain their anti-Hh activity in the presence of mutant forms of Smo that confer resistance to vismodegib and sonidegib, highlighting their potential as improved anticancer drugs targeting the Hh pathway. This activity is presumably a result of their ability to bind Smo in a manner distinct from the clinically approved drugs. We have undertaken a series of studies to more fully explore the anti-cancer potential of the PSZ scaffold through targeting the Hh pathway. Herein, we report the synthesis and evaluation of PSZ analogues designed to enhance both their Hh inhibitory activity and their drug-like properties. In addition, we have generated a homology model of Smo that was utilized in an extensive series of computational studies to probe potential intermolecular interactions between PSZ and Smo and to examine the conformational changes in Smo that drive its inactivation following antagonist binding.

2. Chemistry

2.1. PSZ Analogue Design

Multiple strategies were employed to rationally design improved analogues of PSZ for evaluation as anti-cancer agents that target the Hh signaling pathway. Our previous studies with the ITZ scaffold demonstrated that the triazole moiety, which is responsible for the potent inhibition of Cyp3A4 associated with the ITZ scaffold, is not required for inhibition of Hh signaling [17]. With this in mind, we designed an initial proof-of-concept *des*-triazole PSZ analogue, **1**, that removed the triazole while also incorporating a simplified alkyl side chain present in our most potent ITZ analogues (Figure 1, top). Based on our preliminary docking studies for PSZ in complex with Smo (*vide infra*), we incorporated modified side chains that contained either a primary alcohol or carboxylic acid to take advantage of enhance intermolecular interactions with the PSZ binding site on Smo (Figure 1, middle). Finally, we prepared several simplified PSZ analogues that replace the triazolone/side chain region with an aromatic moiety tethered to the central region of the scaffold through an amide bond (Figure 1, bottom). These compounds were prepared based on recent SAR results for the ITZ scaffold generated in our group [20].



Figure 1. Structure of PSZ and design strategies for PSZ analogues.

2.2. Compound Synthesis

PSZ analogue 1 was prepared through modification of a previously published synthesis of PSZ (Scheme 1) [23]. Friedel-Craft acylation of 1,3-difluorobenzene (2) with succinic anhydride (3), followed by conversion of the resulting ketone to the alkene through standard Wittig conditions provided carboxylic acid 5. Amidation of 5 with (4R)-(+)-4-benzyl-2-oxazolidinone (OXZ) affords OXZ-protected intermediate 6, which undergoes selective hydroxymethylation at the opposite face in the presence of titanium tetrachloride to form intermediate 7. Iodocyclization of 7 results in selective formation of the 2*S*,4*R* conformation around the intact tetrahydrofuran (THF) moiety (8). Reduction of 8 with sodium borohydride removes both the iodine and reduces the OXZ to the primary alcohol, which is subsequently tosylated to provide key *des*-triazole THF intermediate 10. Intermediate 11 was prepared as described previously [17] and coupled directly to 10 to afford desired analogue 1 with the *trans* conformation in 5% overall yield.



Scheme 1. Synthesis of THF Ring and final coupling to make *Des*-PSZ Analogue 1. *Reagents and conditions:* (a) AlCl₃, DCM, 45 $^{\circ}$ C, 3-4 h, 65%; (b) NaOtBu, MePPh₃Br, THF, 40 $^{\circ}$ C, 12 h, 100%; (C) i. DCM, TEA, pivaloyl chloride, ii. OXZ, DMAP, DMF, 45-50 $^{\circ}$ C, 12 h, 68%; (d) i. DCM, TiCl₄, DIPEA, 0 $^{\circ}$ C, 1 h, ii. *s*-trioxane, TiCl₄, DCM, 0 $^{\circ}$ C, 12 h, 50%; (e) I₂, pyridine, CH₃CN, R.T., 20 h, 61%; (f) NaBH₄, DMSO, 110 $^{\circ}$ C, 12 h, 62%; (g) Pyridine, TsCl, R.T., 12 h, 100%; (h) Cs₂CO₃, DMSO, 90 $^{\circ}$ C, 12 h, 63%.

Analogues incorporating either a carboxylic acid or primary alcohol in the side chain region of the PSZ scaffold were prepared by coupling either ethyl 3-bromopropanoate or ethyl bromoacetate to well-characterized intermediate **12** [17] under basic conditions, followed by demethylation with boron tribromide to afford esters **15** and **16**, respectively (Scheme 2). Subsequent coupling of the tosylated THF intermediate **10** to both esters resulted in **17** and **18**, which had been hydrolyzed to the carboxylic acid, presumably due to the basic conditions and elevated temperature required for the coupling. Alcohol **19** was prepared through reduction of **17** with lithium aluminum hydride. Surprisingly, attempts to reduce **18** to the corresponding alcohol were unsuccessful, resulting in multiple products that were difficult to characterize. Because **18**

was significantly less active than **17** against the Hh signaling pathway (Table 2 below), we did not optimize the conditions necessary to reduce **18** to the corresponding alcohol. A similar synthetic route was also utilized to synthesize an ITZ analogue incorporating the propanoic acid side chain (**21**) to explore whether incorporation of this moiety affected the activity of the *des*triazole ITZ scaffold (Scheme 3).



Scheme 2.Synthesis of Hydroxylated *Des*-triazole PSZ Analogues. *Reagents and conditions*: (a) ethyl 3-bromopropanoate or ethyl bromoacetate, Cs_2CO_3 , DMF, 60 °C – 100 °C, 12 h, quant; (b) BBr₃ (1M), DCM, 0 °C - R.T., 4 h, 29-35%; (c) Cs_2CO_3 , DMSO, 90 °C, 12 h, 36%; (d) LAH, THF, 0 °C to R.T., 63%.



Scheme 3. Synthesis of Hydroxylated Side Chain *Des*-triazole PSZ Analogues. *Reagents and conditions:* (a) Cs₂CO₃, DMSO, 90 °C, 12 h, 54%.

A series of 'right side' analogues based on previous ITZ SAR conducted in our lab were also synthesized (Scheme 4). They were prepared by coupling THF region **10** and the linker region precursor **22**, which provides the nitro-containing intermediate **23**. The nitro intermediate was reduced in the presence of 10% palladium on carbon and hydrazine monohydrate to produce aniline **24**. The carbamate **25** was generated through the addition of phenyl chloroformate to aniline **24** and subsequent treatment with hydrazide monohydrate provided carboxamide intermediate **26**. The final unsubstituted triazolone **27** was generated by heating carboxamide **26** with formamidine acetate. Finally, three analogues that replaced the triazolone side chain with an aromatic amide were prepared by directly coupling of aniline **24** to the requisite aromatic carboxylic acid (**28-30**, Scheme 5).



Scheme 4. Synthesis of 'Right-Side Analogues' – Linker Precursors. *Reagents and conditions*: (a) Cs_2CO_3 , DMSO, 90 °C, 12 h, 67%; (b) 10% Pd/C, EtOH, NH₂NH₂, reflux, 12 h, 90%; (c) Pyr, CICOOPh, 3 h, 40%; (d) NH₂NH₂-H₂O, reflux, 3h, 30%; (e) formamidine acetate, acetic acid, reflux, 3 h, 46%.



Scheme 5. Synthesis of 'Right-Side Analogues' – Amide Linkage. *Reagents and conditions:* (a) EDCI, DMAP, DCM, RT, 12 h, 27 - 63% or HATU, NMM, DMF, RT, 12h, 80%.

3. Initial Biological and Computational Studies for PSZ and Analogue 1.

3.1. *Hh inhibitory activity for PSZ and* 1.

To determine whether removal of the triazole moiety had any effect on the ability of the PSZ scaffold to inhibit Hh signaling, we evaluated whether **1** could down-regulate mRNA expression of the Hh-dependent target gene Gli1 in ASZ cells, a well-characterized Hh-dependent murine BCC cell line [24]. Interestingly, *des*-triazole analogue **1** was approximately 2.5-fold more potent than PSZ (IC₅₀ values 0.19 and 0.50 μ M, respectively; Table 1). A similar trend was observed with regards to the ability of **1** to inhibit the growth of a primary Hh-dependent MB derived from a conditional Patched knockout (Math1-Cre-ER:Ptch^{fl/fl}, termed Ptch-CKO) mouse [25].

Table 1. Ir	n Vitro and Pha	armacokinetic Ac	ctivity of PSZ	and initial Des	s-triazole PSZ	Analogue.

Cmpd	$IC_{50} \left(\mu M\right)^a$	Ptch-CKO $GI_{50} (\mu M)^b$	CYP3A4 IC ₅₀ (µM) ^c	P-gp Substrate ^c	T _{1/2} (min) ^c	Cl _{int} (µL/min/mg) ^c
PSZ	0.50 ± 0.1	1.5 ± 0.3	0.15	Yes	108	21.5
1	0.19 ± 0.03	0.61 ± 0.12	>20	No	61.8	37.4

 ${}^{a}IC_{50}$ values from ASZ cells represent the Mean \pm SEM of at least two separate experiments performed in triplicate following a 48 hr incubation.

^bValues represent the Mean ± SEM of at least two separate experiments performed in triplicate.

^cPK data generated by Pharmaron or Cyprotex as a CRO service via their standard protocols.

3.2. Computational Modeling of the Smoothened Receptor.

Several distinct lines of evidence strongly suggest that both PSZ and ITZ bind to the 7-TM domain of Smo in the same region where other Smo agonists and antagonists bind [17-18, 21-22]. Both analogues were active in *Ptch*^{-/-} knockout mouse embryonic fibroblasts (MEFs), but neither compound was inhibited Hh signaling in the presence of a constitutively active Smo mutant, indicating that inhibition of pathway signaling as at the level of Smo [17,21]. In addition, ITZ significantly displaced a tritiated Smo antagonist from HEK293 cells that stably express human Smo [22]. Both azoles retain in vitro and in vivo activity in the presence of several mutant forms of Smo resistant to other Smo antagonists, suggesting that the PSZ/ITZ scaffold may adopt a distinct binding conformation when in complex with Smo [17, 21]. With this in mind, we had three primary goals for our computational studies. First, we sought to predict the conformation these compounds adopt when binding to Smo to determine how they maintain potent anti-Hh activity in the presence of mutant forms of Smo. Second, we sought to correlate inhibition of the Hh pathway with binding energy calculations. Finally, we explored the conformational changes that occur in Smo following compound binding to validate the recent models predicting whether a Smo ligand is an agonist or antagonist [26, 27]. Taken together, this information could aid in the future design of PSZ/ITZ analogues as improved Hh pathway inhibitors. Our first step in this process was to generate a homology model of SMO based on previously published Smo structures in complex with known Smo antagonists (PDB IDs: 4JKV [26], 5V56 [28], and 5L7I [5]). We chose to generate a new model rather than use the existing structures because the complexes described above either removed or mutated multiple residues to allow for improved crystallization. We replaced these altered residues with the natural amino acids and refined the overall structure through homology modeling (Modeller 9.16).

The three-dimensional structure of the SMO receptor generated through our homology modeling process is analogous to the experimentally-derived structures previously disclosed. Smo contains an extracellular domain (ECD), which is composed of the well-characterized cysteine rich domain (CRD) and a linker domain (ELD) (Figure 2A). Other key regions of the Smo receptor include the seven transmembrane domain (7TM), extracellular loop 2 (ECL2) that connects TM4 and TM5, and the KTXXXW motif (K539-T553) located in helix VIII of the intracellular domain. ECL2 contains a β -hairpin that encompasses the primary binding pocket for SMO ligands. Orientation of the intracellular KTXXXW motif is important for both stabilizing

the entire Smo receptor in an inactive state and inducing a conformational change that results in Smo activation [28]. Stability of the KTXXXW motif in the inactive state is primarily governed by molecular interactions of T541, I544 and W545 in helix VIII with T251 and A254 in helix I. Most notably, the side chain hydroxyl of T541 forms a hydrogen bond with the backbone carbonyl of V536 in the intracellular end of helix VII and the indole hydrogen of W545 forms a hydrogen bond with the hydroxyl in the side chain of T251 in helix I (Supplemental Figure 1).

3.3. Binding modes for PSZ, 1, and 19

In order to explore the binding interaction of PSZ and 1 with Smo, a grid box was generated around the well-characterized Smo binding site. Both PSZ and 1 were docked into this grid on Smo utilizing the XP mode in Schrödinger Glide. The putative ligand binding site for these compounds is located at the extracellular surface of the β -hairpin in ECL2 and the α -helical extension of TM6 of the 7-TM domain. As shown in Figure 2, both PSZ and 1 adopt similar orientations and binding interactions inside the binding pocket. For both compounds, the triazole region is oriented towards the extracellular surface with the linker/side chain region penetrating deeply into the Smo binding pocket. The primary difference in the binding modes of the two compounds is in regards to the triazole ring of PSZ, which orients towards the solvent accessible surface of the binding pocket resulting in unfavorable polar binding interactions (Figure 2B and 2C). By contrast, removal of the triazole in analogue 1 results in reduced interactions with the water molecules present at the pocket surface, which may be a primary reason why analogue 1 demonstrates enhanced anti-Hh activity. Additional intermolecular interactions with Smo common to both compounds include a network of π - π and cation- π interactions between the phenyl ring adjacent to the ether and Y207, K395, R485, and/or F484 (Figure 3). The triazolom/side chain

ACCEPTED MANUSCRIPT

region of the scaffold forms two hydrogen bonds with polar residues (D384 and N219) while also exhibiting favorable non-polar interactions between the aliphatic side chain and several hydrophobic residues (Y394, E518 and P513).



Figure 2. Binding mode of PSZ and 1 in complex with Smo. (A) The binding orientations of PSZ (yellow) and 1 (blue) inside the binding pocket on Smo. The opening of the binding site is sandwiched between the ELD and ECL2. (B) The furan region of PSZ and 1 orient out of the binding pocket towards the ELD. (C) Compared to PSZ, 1 penetrates more deeply inside the binding pocket.

Based on these predicted binding modes for PSZ and 1, we sought to design an improved analogue that focused on exploring whether an analogue of 1 that replaced the *sec*-butyl with a simplified hydrophilic side chain would maintain potent anti-Hh activity. In addition, the

incorporation of the terminal alcohol could potentially improve the solubility of the scaffold. Prior to the synthesis of this analogue (**19**), the compound was docked into our Smo model. Overall, the orientation and binding mode of **19** was comparable to both PSZ and **1** (Figure 3). The difluorophenyl ring and tetrahydrofuran are oriented at the solvent accessible surface, while the triazolone/side chain penetrated deeply into the pocket. The primary alcohol on the side chain formed a hydrogen bond with D384, mimicking the interaction between this amino acid residue and the secondary alcohol in the side chain of PSZ (Figure 3). Finally, neither PSZ, **1**, nor **19** demonstrated specific intermolecular interactions with the primary residue mutations that have been linked to vismodegib (D477G and E522K) and sonidegib resistance (N223D, L225R, and G457S), providing evidence as to why these compounds maintain activity in the presence of mutated forms of Smo.



Figure 3. Two-dimensional Smo:ligand contact maps for PSZ (A), **1** (B), and **19** (C) in complex with our Smo structural model. Color code is as follows: red residue, negatively charged residue; purple, positively charged residue; green, hydrophobic residue; light blue, polar residue; gray, solvent exposed area; pink arrow, hydrogen bond; green line, pi-pi interactions.

4. Biological Activity of Second Generation PSZ Analogues.

4.1. Anti-Hh activity of PSZ analogues.

Preliminary docking studies with our Smo model encouraged the synthesis of **19** as well as two additional hydroxylated analogues, **17** and **18**. When evaluated in ASZ cells for Hh inhibition, **19**, which incorporates a propanol side chain, was most active amongst the hydroxylated analogues ($IC_{50} = 0.020 \mu M$). Overall, **17-19** had significantly increased anti-Hh activity when compared to PSZ and our initial proof-of-concept analogue **1**, highlighting the potential of introducing a hydrophilic side chain in addition to the removal of the triazole ring when improving this scaffold. Curious about the impact a hydroxylated side chain would have on the Hh activity of the ITZ scaffold, the non-stereochemically defined propanoic acid side chain analogue, **21**, was synthesized. Interestingly, **21** was more active than **19** by approximately 2-fold.

Based on our recent SAR for the ITZ scaffold [20], PSZ analogues 23-30 focused on incorporating modifications to the triazolone/side chain region or 'right-side' of the scaffold. Analogues 23-27 integrated the *des*-triazole THF region with the linker intermediates leading to the un-substituted triazolone ring while an amide linkage in analogues 28-30 introduced phenyl rings with hydrogen bonding potential. Evaluation in ASZ cells revealed the truncated nitro (23) and aniline (24) PSZ analogues had weakened Hh inhibition compared to 1 indicating an extended scaffold is required for optimal activity. The carbamate (25), carboxamide (26), and triazolone (27) PSZ analogues had similar Hh activity as their *trans*-ITZ counterparts previously published from our group [20]. When investigating the amide linkage analogues, we found that the addition of a *meta*-acetyl phenyl ring (28), increased anti-Hh activity ($IC_{50} = 0.024 \mu M$) that was

comparable to the hydroxylated analogues (**17-19**). A *meta*-hydroxylated phenyl ring (**29**) maintained Hh activity similar to **1** while the addition of a *meta*-pyridyl group (**30**) abolished Hh inhibition in ASZ cells.

Cmpd	$IC_{50} \left(\mu M\right)^{a, b}$
PSZ	0.50 ± 0.1
1	0.19 ± 0.03
17	0.032 ± 0.01
18	0.15 ± 0.02
19	0.020 ± 0.01
21	0.012 ± 0.005
23	0.74 ± 0.15
24	0.33 ± 0.12
25	>10
26	0.22 ± 0.08
27	0.19 ± 0.10
28	0.024 ± 0.009
29	0.19 ± 0.03
30	>10

^aIC₅₀ values represent the Mean \pm SEM of at least two separate experiments performed in triplicate. ^bAll analogues evaluated following 48 h incubation.

4.2. Preliminary pharmacokinetic profiles of PSZ and analogues.

Based on the promising and improved potency of the PSZ analogues, we evaluated several pharmacokinetic parameters of representative compounds (Tables 1 and 3). All of the compounds evaluated were minimally soluble in PBS at pH 7.4. As expected, removal of the triazole moiety completely abolished the ability of PSZ analogues (1, 21, and 28) to inhibit Cyp3A4. Interestingly, while PSZ was identified as a substrate for P-glycoprotein (Pgp), 1 was not a substrate, suggesting the triazole also plays a role in efflux pump recognition for the PSZ scaffold (Table 1). In addition, none of the compounds inhibited the hERG channel at concentrations up to 25 μ M. Overall, PSZ was more stable than ITZ; however, removal of the triazole from PSZ resulted in a significant decrease in metabolic stability for analogue 1 (T_{1/2} =

ACCEPTED MANUSCRIPT

108 and 61.8 min, respectively). By contrast, analogues **28-30** had improved metabolic stability compared to both PSZ and **1** as evidenced by $T_{1/2}$ values ranging from 115 to >180 min and internal clearance rates as low at 2.23 µl/min/mg.

Table 3. Preliminary Pharmacokinetic Activity of PSZ and Analogues.					
Cmpd	Solubility (µM) ^a	^с СҮРЗА4 IC ₅₀ (µМ)	hERG Inhibition (%) ^b	T _{1/2} (min) ^c	$\begin{array}{c} Cl_{int} \\ \left(\mu L/min/mg\right)^c \end{array}$
PSZ	0.781	0.15	14.7	108 ± 5.3	21.5
ITZ	0.8	0.04	< 1	27.0 ± 6.2	
1		>20		61.8 ± 2.1	37.4
19	1.56			C	
21	0.781	>20	3.6)
28	3.13	>20	4.2	115 ± 9.8	20.0
29	3.13			>180	2.23
30	1.56			> 180	7.15

^aKinetic solubility determined in PBS at pH 7.4.

^bPercent inhibition of the hERG channel at 25 μ M.

 $^{c}T_{1/2}$ and Cl_{int} were measured in human liver microsomes at 1 μ M.

--- indicates values were not determined.

4.3. Molecular dynamics studies for PSZ, 1, and 19.

4.3.1. Structural conformation studies. The molecular docking described above represents a single snapshot of the binding complex between Smo and our PSZ-based ligands. In order to more closely probe the binding interactions and conformational changes generated when PSZ or its analogues bind with Smo, we performed additional molecular dynamics (MD, 50 ns) studies on the Smo:PSZ, Smo:1, and Smo:19 docked structures. First, we utilized the root mean square deviations (RMSDs) of the heavy atoms in the Smo:ligand complexes to evaluate the dynamic structural changes that occurred during the MD simulations (Figure 4). The overall Smo:ligand complex was more stable for analogues 1 and 19 compared to PSZ, which suggests a tighter binding affinity (Table 3). A similar trend was observed for the RMSD values of the ligand, further highlighting that the two analogues are more stable in complex with Smo. The RMSD

values of the 7-TM domain and the KTXXXW motif suggest that these regions of Smo were in a stable conformation throughout the simulation. By contrast, the CRD is more flexible, most likely due to fluctuation of the ELD. The KTXXXW motif, which drives downstream activation of the pathway following a conformational change [27], demonstrated stable RMSD values for each complex providing strong evidence that Smo adopts its inactive conformation upon binding to the three small molecules (Table 3). To further probe the activation/inactivation state of Smo in complex with our analogues, we evaluated the conformational stability of helix VI/TM6 by RMSD analysis of the backbone atoms in this region. The α -helical extension of TM6 remained stable throughout the 50 ns simulation time for two primary reasons. First, a cation- π interaction between R451 of TM6 and W535 in TM7 was maintained throughout the simulation. This specific interaction is known to play a critical role in stabilizing Smo in an inactive conformation [27]. Second, a network of hydrogen bonds between several water molecules and amino residues 475-479 stabilize the non-proline kink of the α -helical extension and TM6.



Figure 4. RMSD for heavy atoms of the Smo:PSZ (A), Smo:1 (B), and SMO:19 complexes. 7-TM (black), ligand (blue), CRD (red), KTXXXW motif (green), TM6 (magenta), and R451-W535 (gray) with respect to the equilibrated structure are plotted as a function of MD simulation time.

 Table 3. RMSD comparisons for different domains of the Smo:PSZ, Smo:1, and Smo:19

 complexes.

	Smo:PSZ	Smo:1	Smo:19
Ligand	3.13 ± 0.5	1.95 ± 0.3	1.98 ± 0.4
Complex	8.85 ± 1.4	5.88 ± 1.3	4.48 ± 0.7
CRD	4.30 ± 0.5	5.41 ± 1.6	4.24 ± 0.7
7-TM	3.30 ± 0.6	2.96 ± 0.3	2.89 ± 0.4
KTXXXW	2.97 ± 0.2	2.53 ± 0.2	2.51 ± 0.4
TM6	1.43 ± 0.1	1.62 ± 0.2	1.59 ± 0.1
R451-W535	0.74 ± 0.2	0.70 ± 0.2	0.66 ± 0.2

4.3.2. Binding energy studies. The RMSD values for all the heavy atoms of the ligands and the 7-TM domain of the Smo:PSZ and Smo:**1** complex converged and equilibrated after 2 ns, which suggests that these binding conformations are appropriate to utilize for analysis of the binding

energies of the complexes. By contrast, convergence and equilibration of the Smo:19 complex was achieved after 10 ns; therefore, the stable trajectories of the last 45 ns of the simulation were employed to calculate the average binding energies for the three complexes using the MM/GBSA method. The predicted binding energies of PSZ, 1, and 19 with Smo are in general agreement with experimental inhibition of Hh inhibition described above, with the $\Delta G_{\text{binding}}$ for the Smo:1 and Smo:19 complexes (-39.41 and -36.07 kcal/mol, respectively) significantly lower than the Smo:PSZ complex (-27.90 kcal/mol) (Table 4). To gain a clearer understanding of which individual energy terms have the greatest impact on the calculated binding affinities, four individual energy components were calculated and compared. Overall, the van der Waals (ΔE_{vdw}) interactions served as the major contributor to the binding energies. The polar contributions (ΔG_{polar}) negatively affected compound binding, primarily through polar solvation effects that antagonized binding affinity.

	PSZ		19
$\Delta G_{binding}$	$\textbf{-27.90} \pm 4.9^a$	-39.41 ± 4.3	-36.07 ± 6.5
ΔG_{vdw}	-57.96 ± 5.2	-67.79 ± 3.3	$\textbf{-68.13} \pm \textbf{4.2}$
ΔG_{elect}	-11.07 ± 3.2	-8.08 ± 2.8	-18.64 ± 3.2
ΔG_{polar}	47.67 ± 7.1	43.95 ± 7.5	58.11 ± 8.7
$\Delta G_{nonpolar}$	-6.54 ± 0.4	-7.48 ± 0.2	-7.41 ± 0.3

Table 4. MM/PBSA binding energies (kcal/mol) for Smo:ligand complexes.

^aAll values are kcal/mol.

To evaluate the structural and energetic convergence of the complexes, averages of the MM/PBSA $\Delta G_{\text{binding}}$ values were plotted as a function of simulation time (Figure 5). These plots allowed us to determine the most stable structure across the entire simulation. For Smo:PSZ, the most energetically stable conformation occurred at 21 ns. In this complex, the difluorophenyl ring and the triazole moiety of PSZ are located between the ELD and ECL2 of Smo and have

minimal interactions with the residues present at the surface groove adjacent to the binding site (Figure 6A). The central piperazine adopts a pseudo chair conformation within the binding site that is stabilized by two strong π – π interactions between the phenyl ring adjacent to the furan and Y207 and the phenyl ring adjacent to the triazolone ring with F484. In addition, the piperazine demonstrates strong hydrophobic interactions with multiple residues in the binding site (M301, L303, Y394, K395, E481, and R485). The triazolone adopts a unique orientation such that the carbonyl forms a hydrogen bond (52% occupancy, interatomic distance 2.37Å) with the backbone of S385. The hydroxyl group on the propyl side chain forms a hydrogen bond with the side chain of D384 (40% occupancy rate, interatomic distance 2.63Å) while the aliphatic regions of the side chain are stabilized by multiple van der Waals contacts.



Figure 5. Binding energies for Smo:ligand complexes over time. The lowest energy conformation was utilized to explore intermolecular interactions between Smo and PSZ (blue), 1 (red), and **19** (green).

In a similar manner, the binding conformation of Smo:1 at 35 ns was used to explore the key interactions between the ligand and receptor (Figure 6B). When the triazole moiety is

ACCEPTED MANUSCRIPT

removed, the conformation of the THF region of the scaffold reorients such that the difluorophenyl ring of **1** interacts with the same region previously occupied by the triazole of PSZ. The van der Waals contacts between the difluorophenyl ring and multiple residues at the surface of the binding site (L303, R296, K204, S205 and I215) are stronger than those for the triazole in PSZ, which provides insight into the improved activity of the *des*-triazole analogues. The biaryl piperazine in **1** also adopts a chair conformation that is stabilized by π - π and π -cation interactions between the phenyl ring adjacent to the furan and F484 and K395, respectively. Interestingly, in this structure, N1 of the triazolone moiety is protonated, resulting in the formation of a salt bridge with the carboxylate side chain of D384 (49% occupancy, interatomic distance 1.81Å). Additional stabilization of the triazolone ring is provided by a π - π interaction with Y394 and a hydrogen bond with the side chain of N219. Finally, the highly flexible aliphatic side chain forms multiple hydrophobic contacts with several amino acids at the bottom of the binding pocket.

The most energetically stable conformation of Smo:19 (20 ns) demonstrated that analogue 19 penetrates deeper into the binding site than 1, allowing for a π -cation interaction between the difluorophenyl ring and K395 that is not present for the other two compounds (Figure 6C). The slightly deeper positioning of 19 also allows for the phenoxy ether to form a hydrogen bond with the backbone of E208 (36% occupancy, interatomic distance 1.81Å). The central diaryl piperazine of 19 is stabilized in a chair orientation by the same π - π interactions between the two flanking phenyl rings and Y207 and F484 seen in the Smo:PSZ structure. Finally, the primary hydroxyl of the propanol side chain forms a hydrogen bond with D384 (58% occupancy with interatomic distance 1.85Å).

5. CONCLUSION

We have broadened our understanding of the optimal structural motifs required for potent inhibition of the Hh signaling pathway for small molecule analogues based on the general PSZ and ITZ scaffolds. Our studies continue to demonstrate that the triazole moiety present in the parent scaffold is not required for potent anti-Hh activity and that its removal abrogates the primary off-target side effect associated with PSZ/ITZ administration. Interestingly, removal of the triazole in the PSZ scaffold results in a significant increase in Hh pathway inhibition and additional modifications to the 'right-side' of the scaffold have resulted in analogues with low nanomolar IC_{50} values in Hh-dependent cell lines. To more fully understand our experimental SAR, we performed extensive computational studies, the results of which provide important context for further analogue preparation. The THF region extends out of the binding pocket and removal of the triazole decreases unfavorable polar interactions with water molecules at the binding surface, which supports the improved activity of the *des*-triazole analogues. Finally, the incorporation of a hydrophilic moiety in the triazolone/side chain region of the scaffold can increase activity through improved hydrogen bond interactions with key binding site residues.



Figure 6. Energetically stabilized structures of Smo in complex with PSZ (A), **1** (B), and **19** (C). Intermolecular interactions between Smo and the ligand are represented as dotted lines. Yellow, hydrogen bond; Dark blue, π - π interaction; Red, π -cation interaction. **6.** Experimental section

6.1. General Information.

Starting materials and solvents were purchased from Sigma-Aldrich or Fisher Scientific. All reactions were performed under an argon atmosphere. Intermediates 11 and 20 were prepared as previously described [17]. NMR data was collected on a Bruker AVANCE 500 MHz spectrometer and analysis performed using MestReNova. HRMS data was obtained at the Mass Spectrometry Facility at the University of Connecticut. FT-IR analysis was performed on a Bruker Alpha Platinum ATR instrument using OPUS software (v 7.2). All PSZ analogues evaluated in the biological assays were greater than 95% purity. Purity was determined by injecting 25 µL of a 1 mg/mL solution each PSZ analogue dissolved in HPLC-grade MeCN into an Agilent Manual FL-Injection Valve (600 bar) on an Agilent 1100/1200 Series HPLC equipped with an Agilent Eclipse Plus C18 (4.6 x 100 mm) column and Agilent 1100 Series Photodiode Array Detector. The mobile phase consisted of 70% MeCN/30% H₂O. All analogues were run at a flow rate of 1.0 mL/min for 12 min and purity was assessed at 254 nm.

6.2. Chemical synthesis.

6.2.1. Synthesis of des-triazole intermediates.

6.2.1.1. ((3S,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-yl)methanol (9). To a solution of **8** (150 mg, 0.3 mmol) in anhydrous DMSO, NaBH₄ (43 mg, 1.1 mmol, 4 equiv.) was added. The reaction was stirred at 100 \Box C overnight. The reaction is cooled to 0 \Box C before being quenched by the dropwise addition of NH₄Cl (aqueous, saturated). Once bubbling ceased, the quenched reaction is extracted using EtOAc (4x). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated. The crude was purified by column chromatography (SiO₂, 10-30% EtOAc in Hexanes). The fractions containing product were collected and dried by rotary evaportation to obtain a clear, colorless oil (Yield = 40 mg, 62%, 40 mg). ¹HNMR (500 MHz, CDCl₃) δ 7.57 (ddd, *J* = 34.4, 15.6, 8.8 Hz, 1H), 6.91 – 6.78 (m, 2H), 4.23 – 4.18 (m, 1H), 4.04 (t, *J* = 8.1 Hz, 1H), 3.91 – 3.86 (m, 1H), 3.75 – 3.66 (m, 2H), 2.72 (dt, *J* = 14.6, 7.4 Hz, 1H), 2.58 – 2.51 (m, 1H), 2.48 – 2.35 (m, 1H), 1.87 (dd, *J* = 12.7, 8.4 Hz, 1H), 1.62 (s, 2H), 1.55

(s, 1H). ¹³CNMR (126 MHz, CDCl₃) δ 163.04, 162.94, 161.07, 159.96, 158.08, 157.99, 127.68, 127.63, 127.60, 127.56, 110.77, 110.58, 104.55, 104.34, 104.13, 82.80, 82.77, 77.30, 77.04, 76.79, 69.89, 64.76, 42.31, 42.03, 41.86, 41.83, 41.59, 28.29. DART-HRMS: *m*/*z* [M+H]⁺ calcd. for [C₁₂H₁₃F₂O₂]⁺, 227.0883; found 227.0899.

6.2.1.2. ((3S,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-yl)methyl 4-

methylbenzenesulfonate (10). Alcohol 9 (33 mg, 0.145 mmol, 1 equiv.), trimethylamine (0.29 mmol, 0.032 mL, 2 equiv.) and 4-DMAP (18 mg, 0.145 mmol, 1 equiv.) were dissolved in DCM at room temperature. Next, TsCl was added slowly. The reaction was stirred for 2 hours at room temperature. Upon completion, the reaction was diluted with EtOAc and washed with H₂O and brine. The organic layer was collected, dried over Na₂SO₄, filtered and concentrated. The crude was purified by column chromatography (SiO₂, 10-25% EtOAc in Hexanes). The fractions containing product were collected and dried by rotary evaportation to obtain a clear, colorless oil (Yield = 55 mg, ~100%). ¹HNMR (500 MHz, CDCl₃) δ 7.82 (d, J = 8.1 Hz, 1H), 7.74 (d, J = 8.1 Hz, 1H), 7.49 - 7.45 (m, 1H), 7.37 (d, J = 8.1 Hz, 1H), 7.36 (d, J = 8.1 Hz, 1H), 6.88 - 6.75 (m, 2H), 4.15 – 3.60 (m, 5H), 2.50 (m, 4H), 2.50 (m, 1H, hidden by CH₃) 2.41 – 2.37 (m, 1H), 1.91 – 1.77 (m, 1H), 1.55 (s, 2H), 1.50 (s, 1H). ¹³CNMR (126 MHz, CDCl₃) δ 163.07, 161.11, 159.91, 158.04, 145.02, 132.88, 129.96, 129.89, 129.60, 127.92, 127.87, 127.53, 127.45, 127.41, 127.07, 110.89, 110.72, 110.01, 109.98, 104.63, 104.42, 104.21, 82.74, 77.31, 77.05, 76.80, 71.29, 70.98, 69.75, 69.18, 42.02, 41.49, 41.46, 39.19, 39.00, 29.72, 28.26, 21.66. DART-HRMS: *m/z* [M+H]⁺ cald. for $[C_{19}H_{21}F_2O_4S]^+$, 383.1050; found 383.1117. Also found $[M-CH_3]^+ = 367.0803$ and $[M+NH_4]^+ = 400.1388.$

6.2.2. Synthesis of proof-of-concept analogue 1.

6.2.2.1. General synthetic procedure for coupling the des-triazole THF region to linker intermediate. Cs_2CO_3 (10 equiv.) and the requisite linker (1 equiv.) were diluted in anhydrous DMSO and stirred for 30 minutes at room temperature before addition of the des-triazole THF region in DMSO. The mixture was heated to 90 °C and stirred for 12 h. The reaction was quenched with water and extracted using EtOAc (2x) and DCM (2x). For **17** and **18**, the water layer was acidified to pH = 2 to ensure complete extraction of the carboxylic acid into the organic phase. The organic layers were combined, dried over Na₂SO₄, filtered and concentrated. The crude analogue was purified by column chromatography (SiO₂, 10-30% Acetone in Hexanes).

6.2.2.2. 2-((*R*)-sec-butyl)-4-(4-(4-(4-((((3*R*,5*R*)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-yl)methoxy)phenyl)-piperazin-1-yl)phenyl)-2,4-dihydro-3*H*-1,2,4-triazol-3-one (**1**). Analogue **1** was prepared by coupling THF region intermediate **10** and linker/side chain intermediate **11** as described above. Dark grey solid. (Yield = 63%). ¹HNMR (500 MHz, CDCl₃) δ 7.66 (s, 1H), 7.59 – 7.54 (dd, *J* = 16.2, 8.2 Hz, 1H), 7.47 (d, *J* = 8.5 Hz, 2H), 7.07 (d, *J* = 8.5 Hz, 2H), 6.98 (m, 1H), 6.89 (d, *J* = 7.6 Hz, 3H), 6.85 – 6.80 (m, 1H), 4.34 (m, 1H), 4.14 – 4.11 (t, *J* = 7.7 Hz, 1H), 3.97 – 3.96 (m, 3H), 3.85 – 3.74 (m, 1H), 3.41 (m, 4H), 3.28 (m, 4H), 2.64 – 2.62 (m, 2H), 2.10 – 1.87 (m, 2H), 1.79 – 1.75 (m, 1H), 1.65 (s, 3H), 1.43 (d, *J* = 6.7 Hz, 3H), 0.95 (t, *J* = 7.3 Hz, 3H). ¹³CNMR (126 MHz, CDCl₃) δ 152.08, 150.59, 145.72, 127.74, 127.66, 125.98, 123.59, 118.57, 116.70, 115.32, 104.50, 104.36, 104.15, 82.80, 70.16, 52.72, 50.75, 49.27, 42.07, 39.54, 29.72, 28.47, 28.39, 19.27, 10.81. DART-HRMS: m/z [M+H]⁺ cald. for [C₃₄H₄₀F₂N₅O₃]⁺, 604.3021; found 604.3085. IR (solid) vmax: 2932, 2842, 1702, 1513, 1230, 1041, 966, 823. Purity = 95.1%.

6.2.3. Synthesis of des-triazole analogues containing a modified side chain.

ACCEPTED MANUSCRIPT

125.94, 125.50, 123.67, 118.66, 116.61, 114.56, 110.01, 109.98, 77.34, 77.09, 76.83, 63.88, 61.83, 61.69, 55.60, 50.83, 49.16, 46.81, 14.16, 14.10. DART-HRMS: m/z [M+H]⁺ cald. for $[C_{23}H_{28}N_5O_4]^+$, 437.2063; found 438.2149.

6.2.3.3. ethyl 3-(4-(4-(4-(4-(4-hydroxyphenyl)piperazin-1-yl)phenyl)-5-oxo-4,5-dihydro-1H-1,2,4triazol-1-yl)propanoate (15). To a solution of 13 (1 equiv.) in anhydrous DCM as 0 °C was added 1M BBr₃ in DCM (4-5 equiv.) dropwise. The mixture was stirred for 30 min at 0 °C before warming to RT and stirred for an additional 3 h. The reaction was quenched by pouring the mixture over ice, followed by washing with DCM (2x) and EtOAc (2x). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated. Column chromatography (SiO₂, 1-5% MeOH in DCM) afforded 15 as a light tan solid (Yield = 28%). ¹HNMR (500 MHz, CDCl₃) δ 7.64 (s, 1H), 7.40 (d, *J* = 8.9 Hz, 2H), 7.02 (d, *J* = 9.0 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 6.75 (d, *J* = 8.8 Hz, 2H), 4.25 – 4.22 (m, 4H), 3.40 – 3.38 (m, 4H), 3.25 – 3.13 (m, 4H), 2.89 – 2.86 (t, *J* = 7.1 Hz, 2H), 1.32 – 1.30 (t, *J* = 7.1 Hz, 3H). ¹³CNMR (126 MHz, CDCl₃) δ 170.87, 152.24, 150.90, 150.69, 145.04, 134.66, 125.24, 124.16, 118.84, 116.53, 116.00, 110.01, 109.98, 77.31, 77.05, 76.80, 60.90, 51.01, 49.10, 41.46, 33.35, 14.18. DART-HRMS: *m*/z [M+H]⁺ cald. for [C₂₃H₂₈N₅O₄]⁺, 438.2063; found 438.2150.

6.2.3.4. *ethyl* 2-(4-(4-(4-(4-(4-hydroxyphenyl)piperazin-1-yl)phenyl)-5-oxo-4,5-dihydro-1H-1,2,4*triazol-1-yl*)*acetate* (16). Alcohol 16 was prepared using precursor 14 and BBr3 as described above. Tan solid. (Yield = 23%). ¹HNMR (500 MHz, CDCl₃) δ 8.41 (s, 1H), 7.71 (s, 1H), 7.42 (d, *J* = 8.3 Hz, 1H), 7.20 (d, *J* = 8.5 Hz, 1H), 7.04 – 6.99(m, 2H), 6.93 – 6.90 (t, *J* = 9.1 Hz, 2H), 6.82 – 6.77 (m, 2H), 4.69 (d, *J* = 8.7 Hz, 1H), 4.47 (s, 1H), 4.30 – 4.22 (m, 2H), 3.38 –3.24 (m, 8H), 1.35 – 1.31 (m, 3H). ¹³CNMR (126 MHz, CDCl₃) δ 168.51, 162.68, 150.85, 150.56, 150.10, 150.07, 145.56, 135.14, 132.85, 125.96, 123.79, 118.84, 116.70, 116.61, 115.96, 114.58, 61.86, 61.50, 55.61, 52.65, 50.89, 49.23, 49.17, 47.79, 46.83, 46.70, 40.99, 29.71, 14.14. DART-HRMS: m/z [M+H]⁺ cald. for [C₂₂H₂₆N₅O₄]⁺, 424.1907; found 424.1958.

yl)methoxy)phenyl)piperazin-1-yl)phenyl)-5-oxo-4,5-dihydro-1H-1,2,4-triazol-1-yl)propanoic

acid (17). Analogue 17 was prepared by coupling des-triazole intermediate 10 and linker region intermediate 15 as described above. White solid. (Yield = 42%) ¹HNMR (500 MHz, CDCl₃) δ 7.63 (s, 1H), 7.59 – 7.51 (m, 2H), 7.44 (d, *J* = 8.9 Hz, 1H), 7.07 (d, *J* = 9.0 Hz, 2H), 7.00 – 6.97 (m, 3H), 6.90 – 6.79 (m, 5H), 4.29 – 4.21 (m, 1H), 4.14 – 4.07 (m, 1H), 4.03 – 3.91 (m, 4H), 3.8 – 3.72 (m, 1H), 3.42 – 3.40 (m, 3H), 3.32 – 3.31 (m, 1H), 3.28 – 3.27 (m, 4H), 2.71 – 2.51 (m, 3H), 2.11 – 1.97 (m, 2H), 1.66 (s, 1H), 1.65 (s, 2H). ¹³CNMR (126 MHz, CDCl₃) δ 176.37, 162.98, 161.01, 153.50, 152.29, 150.77, 145.57, 134.42, 127.72, 125.49, 123.73, 118.69, 118.60, 118.04, 116.67, 116.50, 115.31, 110.79, 110.63, 110.01, 109.98, 104.57, 104.36, 104.16, 82.80, 77.32, 77.06, 76.81, 70.43, 70.16, 70.10, 50.77, 50.21, 49.12, 48.00, 42.99, 42.04, 39.51, 39.42, 36.79, 29.71, 28.36, 20.87. DART-HRMS: *m*/z [M+H]⁺ cald. for [C₃₃H₃₆F₂N₅O₅]⁺, 620.2060; found [M – 114.9653]⁺, 505.2407 *m*/z. IR (solid) *v*max: 2931, 1705, 1614, 1512, 1231.6, 1039, 966, 822. Purity = 95.1%.

6.2.3.6. $2-(4-(4-(4-(4-(((3R,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-yl)methoxy)phenyl)piperazin-1-yl)phenyl)-5-oxo-4,5-dihydro-1H-1,2,4-triazol-1-yl)acetic acid (18). Analogue 18 was prepared by coupling des-triazole intermediate 10 and linker region intermediate 16 as described above. Tan solid. (Yield = 27%) ¹HNMR (500 MHz, CDCl₃) <math>\delta$

7.63 – 7.51 (m, 1H), 7.00 – 6.94 (m, 5H), 6.90 – 6.88 (m, 3H), 6.81 – 6.79 (m, 2H), 4.29 – 4.26 (m, 1H), 4.14 – 4.11 (m, 1H), 3.99 – 3.94 (m, 3H), 3.87 – 3.72 (m, 2H), 3.40 (s, 1H), 3.33 – 3.26 (m, 8H), 2.65 – 2.62 (m, 2H), 1.98 (d, J = 4.3 Hz, 1H), 1.65 (s, 2H), 1.58 (s, 1H), 1.30 (s, 3H). ¹³CNMR (126 MHz, CDCl₃) δ 163.11, 161.01, 153.31, 148.19, 145.76, 129.69, 127.65, 118.49, 117.91, 116.63, 115.30, 110.79, 110.61, 104.57, 104.36, 104.15, 82.71, 70.12, 50.80, 50.08, 42.07, 39.54, 29.72, 28.36. DART-HRMS: m/z [M+H]⁺ cald. for [C₃₂H₃₄F₂N₅O₅]⁺, 606.2450; found [M – 101.0026]⁺, 505.2424 m/z. IR (solid) vmax: 2928, 2854, 2229, 1614, 1511, 1230, 966, 823; Purity = 97.2%.

6.2.3.7. 4-(4-(4-(4-((((3R,5R))-5-(2,4-difluorophenyl))-5-methyltetrahydrofuran-3yl)methoxy)phenyl)piperazin-1-yl)phenyl)-2-(3-hydroxypropyl)-2,4-dihydro-3H-1,2,4-triazol-3one (**19**). Carboxylic acid **17** (24 mg, 0.039 mmol) was dissolved in THF (5 mL) and cooled to 0 \Box C. LiAlH₄ (2M in THF, 0.062 mmol, 0.031 mL) was added dropwise. The mixture was warmed to RT and stirred for 4 h. The mixture was cooled to 0 \Box C and saturated NH₄Cl was added dropwise and the resulting solution was washed with EtOAc (3x). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated. Column chromatography (SiO₂, 1% MeOH in DCM) afforded alcohol **19** as a tan solid. (Yield = 17 mg, 87%). ¹HNMR (500 MHz, CDCl₃) δ 7.64 (s, 1H), 7.61 – 7.52 (m, 2H), 7.43 (d, *J* = 8.9 Hz, 1H), 7.06 (d, *J* = 9.1 Hz, 1H), 7.00 – 6.97 (m, 4H), 6.89 – 6.87 (m, 3H), 6.82 – 6.79 (m, 2H), 4.29 – 4.21 (m, 1H), 4.14 – 4.07 (m, 2H), 4.03 – 3.91 (m, 4H), 3.87 – 3.71 (m, 2H), 3.40 (m, 2H), 3.31 (m, 2H), 3.27 (s, 4H), 2.63 – 2.62 (m, 3H), 2.12 – 1.98 (m, 2H), 1.65 (s, 3H), 1.37 – 1.33 (dd, *J* = 13.6, 6.7 Hz, 2H). ¹³CNMR (126 MHz, CDCl₃) δ 163.04, 161.11, 160.07, 158.01, 153.39, 152.22, 150.77, 148.03, 145.76, 134.29, 127.73, 123.64, 118.58, 118.50, 117.92, 116.65, 116.61, 115.31, 111.50, 110.78, 110.63, 110.01, 109.98, 104.57, 104.36, 104.15, 82.74, 77.29, 77.03, 76.78, 70.46, 70.19, 70.12, 69.90, 50.81, 50.73, 50.11, 49.19, 47.98, 43.01, 42.05, 39.54, 39.45, 28.38, 22.35, 14.07. DART-HRMS: *m*/*z* [M+H]⁺ cald. for [C₃₃H₃₈F₂N₅O₄]⁺, 605.2814; found [M – 100.0.390]⁺, 505.2424. IR (solid) *v*max: 2932, 2828, 2232, 1689, 1614, 1511, 1230, 1112, 1039, 966, 821. Purity = 96.2%.

6.2.3.8. 3-(4-(4-(4-((2-(2,4-dichlorophenyl)-2-methyl-1,3-dioxolan-4-

yl)methoxy)phenyl)piperazin-1-yl)phenyl)-5-oxo-4,5-dihydro-1H-1,2,4-triazol-1-yl)propanoic acid (21). Analogue 21 was prepared by coupling des-triazole intermediate 20 and linker region intermediate 15 as described above. White solid. (Yield = 20 mg, 54%) ¹H NMR (500 MHz, CDCl₃) δ 7.77 (d, J = 8.4 Hz, 1H), 7.69 – 7.62 (m, 2H), 7.57 (s, 1H), 7.45 – 7.39 (m, 2H), 7.29 – 7.23 (m, 1H), 7.08 – 7.06 (m, 1H), 7.00 – 6.98 (s, 2H), 6.95 – 6.93 (m, 2H), 6.78 – 6.77 (m, 1H), 4.67 - 4.64 (m, 1H), 4.42 - 4.33 (m, 2H), 4.19 - 4.14 (m, 1H), 4.07 - 4.00 (m, 2H), 3.91 - 3.77 (m, 2H), 3.42 - 3.39 (m, 3H), 3.30 - 3.25 (m, 5H), 1.86 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 153.06, 153.06, 152.54, 152.54, 150.80, 150.75, 147.98, 145.94, 145.94, 139.35, 139.13, 138.14, 134.63, 134.58, 132.84, 132.77, 131.21, 131.16, 130.98, 129.00, 128.91, 128.59, 126.81, 126.75, 125.49, 123.70, 123.62, 118.50, 118.46, 118.42, 117.92, 116.66, 115.52, 115.27, 110.00, 109.17, 109.11, 77.30, 77.05, 76.79, 75.07, 75.07, 74.77, 74.77, 73.96, 73.96, 73.62, 73.62, 69.29, 69.29, 68.43, 68.43, 67.94, 67.37, 67.37, 67.31, 67.01, 67.01, 50.73, 50.64, 50.64, 50.14, 49.16, 49.16, 48.23, 48.23, 47.25, 31,61, 29.72, 25.90, 25.74, 25.74, 25.66, 22.67. DART-HRMS: *m/z* [M+H]⁺ cald. for $[C_{32}H_{34}Cl_2N_5O_6]^+$, 654.1808; found $[M - 115.0225]^+$, 539.1583 *m/z*. IR (solid) vmax: 2937, 2881, 2826, 1705, 1509, 1225, 1193, 1034, 819, 731, 500. Purity = 30.1 % : 69.0% (cis: trans), total = 99.1%.

ACCEPTED MANUSCRIPT

6.2.4. Synthesis of triazolone/side chain truncated analogues.

6.2.4.1. 1-(4-(((3R,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-yl)methoxy)phenyl)-4-(4-nitrophenyl)piperazine (23). Analogue 23 was prepared by coupling des-triazole intermediate 10 and linker region intermediate 22 as described above. Orange oil. (Yield = 40 mg, 67%). ¹HNMR (500 MHz, CDCl₃) δ 8.21 – 8.19 (m, 2H), 7.65 – 7.51 (m, 1H), 6.98 – 6.80 (m, 8H), 4.29 – 4.26 (m, 1H), 4.19 – 4.10 (m, 1H), 4.00 – 3.94 (m, 2H), 3.87 – 3.72 (m, 1H), 3.63 – 3.60 (m, 4H), 3.28 – 3.26 (m, 4H), 2.98 – 2.95 (m, 1H), 2.66 – 2.52 (m, 2H), 2.07 – 1.95 (m, 1H), 1.65 (s, 1H), 1.58 (s, 2H). ¹³CNMR (126 MHz, CDCl₃) δ 154.79, 145.38, 130.16, 127.72, 127.64, 126.99, 125.98, 118.68, 115.38, 115.33, 113.38, 112.92, 104.36, 104.15, 82.81, 70.19, 70.08, 69.90, 60.31, 50.47, 47.29, 42.03, 39.53, 28.37. DART-HRMS: m/z [M+H]⁺ cald. for [C₂₈H₃₀F₂N₃O₄]⁺, 510.2126; found 510.2205. IR (solid) vmax: 2932, 2850, 1596, 1509, 1322, 1229, 1115, 1038, 944, 826, 754. Purity = 95.6%.

6.2.4.2. $4-(4-(((3R,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-yl)methoxy)phenyl)piperazin-1-yl)aniline (24). Pd/C (10 wt. %, 0.05 equiv.) and ethanol were added to a round bottom flask purged with argon. Intermediate 23 (20 mg, 0.04 mmol) was added followed by dropwise addition of hydrazine monohydrate (13 mg, 0.40 mmol, 0.013 mL) and the solution was refluxed for 12 h. The mixture was cooled, filtered through celite and washed with ethanol. The filtrate was dried and the resulting crude gray solid was purified by column chromatography (SiO₂, 20-30% Acetone in Hexanes). White solid. (Yield = 17 mg, 90%). ¹HNMR (500 MHz, CDCl₃) <math>\delta$ 7.65 – 7.54 (m, 1H), 6.99 – 6.97 (m, 2H), 6.92 – 6.85 (m, 5H), 6.83 – 6.79 (m, 2H), 6.73 – 6.71 (m, 2H), 4.29 – 4.26 (m, 1H), 4.14 – 4.11 (m, 1H), 3.97 –

3.94 (m, 3H), 3.81 – 3.72 (m, 1H), 3.49 (br, s, 1H), 3.27 – 3.23 (m, 8H), 2.63 – 2.62 (m, 2H), 2.00 – 1.95 (m, 1H), 1.65 (s, 3H), 1.58 (s, 1H). ¹³CNMR (126 MHz, CDCl₃) δ 162.97, 158.04, 153.12, 145.98, 144.46, 140.40, 127.74, 127.67, 118.86, 118.34, 116.24, 115.27, 115.22, 110.80, 110.61, 104.56, 104.35, 104.15, 82.70, 70.20, 70.14, 69.92, 51.26, 50.92, 42.06, 39.77, 39.55, 29.72, 28.38. DART-HRMS: *m*/*z* [M+H]⁺ cald. for [C₂₈H₃₂F₂N₃O₂]⁺, 480.2384; found 480.2450. IR (solid) *v*max: 3459, 3364, 2974, 2821, 1614, 1510, 1496, 1221, 1038, 942, 819, 515. Purity = 98.1%.

(4-(4-(((3R,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-6.2.4.3. phenyl yl)methoxy)phenyl)piperazin-1-yl)phenyl)carbamate (25). Aniline 24 (100 mg, 0.42 mmol) was dissolved in CHCl₃ (15 mL) under argon followed by the addition of pyridine (6.72 mmol, 0.27 mL). The solution was cooled to 0 °C and phenylchloroformate (33 mg, 0.42 mmol, 0.025 mL) was added dropwise. The mixture was stirred at room temperature for 3 h before being diluted with water and washed with DCM (3x). The crude mixture was purified by column chromatography (SiO₂, 20-30% Acetone in Hexanes). White solid. (Yield = 50 mg, 40%). ¹HNMR (500 MHz, CDCl₃) δ 7.65 – 7.54 (m, 1H), 7.45 – 7.40 (m, 4H), 7.29 – 7.23 (m, 3H), 7.02 - 6.94 (m, 4H), 6.89 - 6.88 (m, 3H), 6.86 - 6.79 (m, 1H), 4.30 - 4.27 (dd, J = 8.5, 7.5 Hz, 1H), 4.14 – 4.11 (m, 1H), 4.00 – 3.94 (m, 2H), 3.87 – 3.72 (m, 1H), 3.35 – 3.32 (m, 4H), 3.28 – 3.25 (m, 4H), 2.99 - 2.94 (m, 1H), 2.67 - 2.61 (m, 1H), 2.56 - 2.52 (m, 1H), 2.07 - 1.98 (m, 1H), 1.65 (s, 2H), 1.58 (s, 1H). ¹³CNMR (126 MHz, CDCl₃) δ 159.99, 158.02, 153.26, 150.76, 145.82, 129.39, 127.69, 125.59, 121.67, 120.34, 118.46, 118.42, 117.16, 115.31, 115.31, 115.26, 110.81, 110.64, 104.36, 104.15, 82.71, 70.21, 70.13, 69.92, 50.81, 49.98, 42.06, 39.77, 39.56, 28.39. DART-HRMS: m/z [M+H]⁺ cald. for [C₃₅H₃₆F₂N₃O₄]⁺, 600.2596; found 600.2595. IR (solid) *v*max: 3334, 3295, 3044, 2966, 2831, 1707, 1534, 1488, 1197, 1030, 820, 687. Purity = 96.2%.

N-(4-(4-(4-((3R,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-6.2.4.4. yl)methoxy)phenyl)piperazin-1-yl)phenyl)hydrazinecarboxamide (26). Compound 25 (40 mg, 0.067 mmol) was dissolved in 1,4-dioxane (2 mL). Hydrazine monohydrate (12 mg, 0.36 mmol, 0.011 mL, 5.4 equiv.) was added and the solution was refluxed for 3 h. The solution was cooled to RT, diluted with water and washed with EtOAc (3x). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated and purified with column chromatography (SiO₂, 20-30%) Acetone in Hexanes). Tan solid. (Yield = 10.4 mg, 30%). ¹HNMR (500 MHz, CDCl₃) δ 8.08 (s, 1H), 7.61 (s, 1H), 7.57 - 7.54 (m, 1H), 7.46 (d, J = 8.0 Hz, 2H), 7.01 - 6.94 (m, 3H), 6.89 - 6.79 (m, 3H), 4.29 - 4.26 (t, J = 7.3 Hz, 1H), 4.14 - 4.11 (t, J = 7.0 Hz, 1H), 3.97 (br, s, 2H), 3.88 - 1003.72 (m, 1H), 3.40 (s, 1H), 3.32 - 3.27 (m, 7H), 2.98 - 2.95 (m, 1H), 2.63 - 2.62 (m, 1H), 2.56 -2.52 (m, 1H), 1.65 (s, 2H), 1.57 (s, 1H). ¹³CNMR (126 MHz, CDCl₃) δ 162.97, 153.71, 147.67, 147.11, 145.91, 131.06, 127.74, 121.17, 118.43, 117.25, 115.28, 115.24, 104.36, 104.15, 70.19, 70.13, 69.91, 50.84, 50.21, 42.05, 39.76, 39.54, 28.38, 25.26. DART-HRMS: *m/z* [M+H]⁺ cald. for [C₂₉H₃₄F₂N₅O₃]⁺, 538.2551; found. IR (solid) vmax: 3349, 3206, 2927, 2874, 2819, 1677, 1508, 1223, 1014, 817, 625, 516. Purity = 96.7%.

6.2.4.5. 4-(4-(4-(4-(((3R,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3yl)methoxy)phenyl)piperazin-1-yl)phenyl)-2,4-dihydro-3H-1,2,4-triazol-3-one (**27**). A solution of formamidine acetate (8.7 mg, 0.084 mmol) and **26** (10 mg, 0.019 mmol) in anhydrous DMF (5 mL) was stirred at 130 °C for 5 h. The mixture was cooled to RT, diluted with water, and washed with EtOAc (3x). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated. Purification by column chromatography (SiO₂, 15-30% Acetone in Hexanes) afforded triazolone **27**. The fractions containing product were collected and dried by rotary evaportation. Tan solid. (Yield = 4.7 mg, 46%). ¹HNMR (500 MHz, CDCl₃) δ 8.55 (d, *J* = 11.6 Hz, 1H), 8.38 (s, 1H), 7.63 – 7.48 (m, 2H), 7.24 (d, *J* = 11.5 Hz, 1H), 7.06 – 6.79 (m, 8H), 4.29 – 4.26 (m, 1H), 4.14 – 4.11 (m, 1H), 3.98 – 3.94 (m, 2H), 3.87 – 3.72 (m, 1H), 3.40 (s, 1H), 3.37 – 3.27 (m, 7H), 2.63 – 2.51 (m, 2H), 2.08 – 1.90 (m, 1H), 1.65 (s, 1H), 1.58 (s, 2H). ¹³CNMR (126 MHz, CDCl₃) δ 162.54, 158.58, 153.28, 148.76, 129.35, 127.68, 121.57, 121.41, 118.48, 117.31, 116.85, 115.31, 104.36, 70.20, 50.77, 49.79, 49.65, 42.08, 39.55, 29.72, 28.38. DART-HRMS: *m*/*z* [M+H]⁺ cald. for [C₃₀H₃₂F₂N₅O₃]⁺, 548.2395; found 548.2406. IR (solid) *v*max: 2923, 2852, 1690, 1509, 1226, 1076, 1018, 966, 944, 816, 730, 516. Purity = 96.7%.

6.2.5. Synthesis of analogues containing an amide.

6.2.5.1. 3-acetyl-N-(4-(4-(4-(((3R,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3yl)methoxy)phenyl)piperazin-1-yl)phenyl)benzamide (**28**). To a dry round bottom flask purged with Ar_(g) was added **24** (17 mg, 0.035 mmol), 3-acetyl benzoic acid (18 mg, 0.11 mmol), N-(3dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (EDCI) (21 mg, 0.11 mmol) and 4dimethylaminopyridine (13 mg, 0.11 mmol). The solids were dissolved with anhydrous DCM (8 mL) and the mixture was stirred for 12 h at RT before diluting with water and washing with DCM (3x). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated. The crude product was purified by column chromatography (SiO₂, 30% Acetone in Hexanes). White solid. (Yield = 5.2 mg, 27%). ¹HNMR (500 MHz, CDCl₃) δ 8.48 (s, 1H), 8.18 – 8.16 (m, 2H), 7.85 (s, 1H), 7.68 – 7.54 (m, 4H), 7.06 – 6.80 (m, 7H), 4.30 – 4.27 (dd, J = 8.7, 7.3 Hz, 1H), 4.14 - 4.11 (m, 1H), 4.00 - 3.94 (m, 2H), 3.87 - 3.73 (m, 1H), 3.40 - 3.37 (m, 4H), 3.30 - 3.28 (m, 4H), 2.73 (s, 3H), 2.65 - 2.53 (m, 2H), 2.07 - 1.95 (m, 1H), 1.65 (s, 3H). ¹³CNMR (126 MHz, CDCl₃) δ 197.40, 164.45, 162.98, 161.01, 160.08, 158.02, 153.28, 148.70, 145.85, 137.41, 135.64, 131.74, 131.38, 130.19, 129.28, 127.69, 126.29, 121.80, 118.49, 118.45, 116.87, 115.31, 115.26, 104.56, 104.36, 104.15, 82.71, 70.20, 70.13, 69.92, 50.80, 49.78, 42.05, 39.77, 39.55, 29.72, 28.38, 26.77. DART-HRMS: m/z [M+H]⁺ cald. for [C₃₇H₃₈F₂N₃O₄]⁺, 626.2752; found 626.2807. IR (solid) *v*max: 3305, 2944, 2830, 1642, 1591, 1511, 1497, 1227, 1023, 818, 706, 537, 515. Purity = 100%.

6.2.5.2. N-(4-(4-(4-(((3R,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-yl)methoxy)phenyl)piperazin-1-yl)phenyl)-3-hydroxybenzamide (**29**). To a dry round bottom flask purged with argo, 3-hydroxybenzoic acid (4 mg, 0.03 mmol) and HATU (15 mg, 0.04 mmol) were dissolved in DMF (5 mL). Next, NMM (12 mg, 0.12 mmol, 0.013 mL) was added and the mixture was stirred at RT for 30 min followed by the addition of**24** $(16 mg, 0.03 mmol). The mixture was stirred at RT for 12 h, poured over ice cold water (~5 mL) and washed with EtOAc (3x). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated. The crude product was purified by column chromatography (SiO₂, 30% Acetone in Hexanes). White solid. (Yield = 10 mg, 80%). ¹HNMR (500 MHz, CDCl₃) <math>\delta$ 8.18 – 8.10 (m, 1H), 7.83 – 7.54 (m, 5H), 7.44 – 7.39 (m, 2H), 7.18 (d, *J* = 7.3 Hz, 1H), 7.09 – 6.79 (m, 7H), 4.28 – 4.26 (m, 1H), 4.12 – 4.10 (t, *J* = 7.8 Hz, 1H), 3.97 (m, 2H), 1.57 (s, 1H). ¹³CNMR (126 MHz, CDCl₃) δ 156.31, 155.97, 153.28, 151.10, 148.63, 145.86, 136.88, 130.03, 127.79, 122.69, 121.73, 121.19, 118.61, 118.49, 116.92, 115.31, 104.36, 104.15, 82.71, 70.20, 70.13, 50.79, 49.81, 42.05, 39.55,

ACCEPTED MANUSCRIPT

29.72, 28.37. DART-HRMS: *m*/*z* [M+H]⁺ cald. for [C₃₅H₃₆F₂N₃O₄]⁺, 600.2596; found 600.2681. IR (solid) *v*max: 3269, 2925, 2825, 1676, 1642, 1511, 1225, 1020, 817, 731, 685, 590, 532, 516. Purity = 100%

6.2.5.3. N-(4-(4-(4-(((3R,5R)-5-(2,4-difluorophenyl)-5-methyltetrahydrofuran-3-yl)methoxy)phenyl)piperazin-1-yl)phenyl)nicotinamide (**30**). Analogue**30**was prepared utilizing the procedure described above for analogue**28** $. White solid. (Yield = 11.4 mg, 63%). ¹HNMR (500 MHz, CDCl₃) <math>\delta$ 9.13 (s, 1H), 8.81 (d, J = 3.3 Hz, 1H), 8.25 – 8.24 (d, J = 7.6 Hz, 1H), 7.87 (s, 1H), 7.60 – 7.47 (m, 4H), 7.05 – 6.79 (m, 7H), 4.29 – 4.26 (dd, J = 8.7, 7.3 Hz, 1H), 4.14 – 4.11 (dd, J = 8.6, 7.4 Hz, 1H), 3.99 – 3.94 (m, 2H), 3.85 – 3.74 (m, 1H), 3.39 – 3.36 (m, 4H), 3.29 – 3.27 (m, 4H), 2.99 – 2.94 (m, 1H), 2.67 – 2.52 (m, 2H), 2.07 – 1.95 (m, 1H), 1.65 (s, 2H), 1.58 (s, 1H). ¹³CNMR (126 MHz, CDCl₃) δ 163.63, 162.98, 161.17, 153.29, 152.43, 148.83, 147.81, 145.82, 135.26, 130.97, 129.92, 127.62, 123.68, 121.93, 118.49, 116.82, 115.30, 110.78, 110.65, 104.57, 104.36, 104.15, 82.74, 70.20, 70.13, 50.79, 49.72, 42.05, 39.55, 28.39. DART-HRMS: m/z [M+H]⁺ cald. for [C₃₄H₃₅F₂N₄O₃]⁺, 585.2599; found 585.2649. IR (solid) *v*max: 3312, 2932, 2874, 2826, 1737, 1646, 1614, 1592, 1242, 966, 945, 849, 748. Purity = 97.4%.

6.3. Cell Culture Studies.

Protocols for general cell culture, qPCR, and Hh inhibition in ASZ cells have been previously described [17, 20]. All PSZ analogues were tested for Hh inhibitory activity by measuring their inhibition of endogenous Gli1 mRNA expression in a murine BCC cell line (Ptch^{+/-}), ASZ. Cells at 80% confluence were plated in 154CF growth media (2% FBS, ThermoFisher) in a 96-well plate (10,000 cells/well). Cells were incubated for 24 hours (37°C, 5% CO₂) before media

without fetal bovine serum (FBS) was added. Cells were again incubated for 24 hours ($37^{\circ}C$, 5% CO₂) before being treated with DMSO (vehicle control), PSZ, or PSZ analoguea at varying concentrations ($10 - 0.001 \mu$ M). Compounds were incubated with the cells for 48 hours ($37^{\circ}C$, 5% CO₂). Cells were visually examined to ensure there was no non-specific cytotoxicity present for the concentrations evaluated. Cells were lysed, mRNA was extracted, and Act B (internal control) and Gli1 expression levels were measured by qRT-PCR using the TaqMan Fast Cells-to-C_T kit (ThermoFisher). Compound concentrations that significantly decreased actin B levels were excluded form our qRT-PCR calculations. Data was analyzed using GraphPad Prism 5 and IC₅₀ values computed as mean \pm SEM from at least three separate experiments performed in triplicate.

Acknowledgements

The authors gratefully acknowledge support of this work by the National Institutes of Health/National Cancer Institute (CA190617). ASZ001 cells were kindly provided by Dr. Ervin Epstein (Children's Hospital Oakland Research Institute).

Appendix A. Supporting Information. Synthetic protocols and characterization of previously disclosed intermediates. Protocols for the computational modeling and PK studies. ¹H, ¹³C and 2D NMR (HMBC and HSQC) for all new intermediates and final *des*-triazole PSZ analogues. Additional figures.

Abbreviations

ITZ: itraconazole; PSZ: posaconazole; Hh: Hedgehog; Gli: glioblastoma associated oncogene; Ptch: patched; BCC: Basal cell carcinoma; MB: Medulloblastoma; Smo: Smoothened; THF: Tetrahydrofuran; OXZ: (4R)-(+)-4-benzyl-2-oxazolidinone; Ptch-CKO: Patched knockout; ECD: Extracellular domain; CRD: Cysteine rich domain; ELD: Linker domain; 7TM: Seven transmembrane domain; ECL2: Extracellular loop 2

References

[1] P.W. Ingham, A.P. McMahon, Hedgehog signaling in animal development: paradigms and principles. Genes Dev. 15 (2001) 3059-3087.

[2] J. Briscoe, P.P. Thérond, The mechanisms of hedgehog signalling and its roles in development and disease. Nat. Rev. Mol. Cell Biol. 14 (2013) 416-429.

[3] D. Amakye, Z. Jagani, M. Dorsch, Unraveling the therapeutic potential of the hedgehog pathway in cance. Nat. Med. 19 (2013) 1410-1422.

[4] L.A. Fecher, W.H. Sharfman, Advanced basal cell carcinoma, the hedgehog pathway, and treatment options – role of smoothened inhibitors. Biologics 9 (2015) 129-140.

[5] M.W. Kieran Targeted treatment for sonic hedgehog-dependent medulloblastoma. Neuro-Oncol. 16 (2014) 1037-1047.

[6] F. Wu, Y, Zhang, B. Sun, A.P. McMahon, Y. Wang, Hedgehog signaling: From basic biology to cancer therapy. Cell Chem. Biol. 24 (2017) 252-280.

[7] C.M. Rudin, C.L. Hann, J. Laterra, R.L. Yauch, C.A. Callahan, L. Fu, T. Holcomb, J. Stinson, S. E. Gould, B. Coleman, P.M. LoRusso, D.D. Von Hoff, F.J. de Sauvage, J. A. Low, Treatment of medulloblastoma with hedgehog pathway inhibitor GDC-0449. N. Engl. J. Med. 361 (2009) 1173-1178. [8] R.L. Yauch, G.J. Dijkgraaf, B. Alicke, T. Januario, C.P. Ahn, T. Holcomb, K. Pujara, J. Stinson, C.A. Callahan, T. Tang, J.F. Bazan, Z. Kan, S. Seshagiri, C.J. Hann, S.E. Gould, J.A. Low, C.M. Rudin, F.J. de Sauvage, Smoothened mutation confers resistance to a hedgehog pathway inhibitor in medulloblastoma. Science 326 (2009) 572-574.

[9] S.X. Atwood, K.Y. Sarin, R.J. Whitson, J.R. Li, G. Kim, M. Rezaee, M.S. Ally, J. Kim, C. Yao, A.L.S. Chang, A.E. Oro, J.Y. Tang, Smoothened variants explain the majority of drug resistance in basal cell carcinoma. Cancer Cell 27 (2015) 342-353.

[10] H.J. Sharpe, G. Pau, G.J. Dijkgraaf, N. Basset-Seguin, Z. Modrusan, T. Januario, V. Tsui,
A. B. Durham, A.A. Dlugosz, P.M. Haverty, R. Bourgon, J.Y. Tang, K.Y. Sarin, L. Dirix, D.C.
Fisher, C.M. Rudin, H. Sofen, M.R. Migden, R.L. Yauch, F.J. de Sauvage, Genomic analysis of smoothened inhibitor resistance in basal cell carcinoma. Cancer Cell 27 (2015) 327-341.

[11] G.J. Dijkgraaf, B. Alicke, L. Weinmann, T. Januario, K. West, Z. Modrusan, D. Burdick, R. Goldsmith, K. Robarge, D. Sutherlin, S.J. Scales, S.E. Gould, R.L. Yauch, F.J. de Sauvage, Small molecule inhibition of GDC-0449 refractory smoothened mutants and downstream mechanisms of drug resistance. Cancer Res. 71 (2011) 435-444.

[12] J. Wen, M. K. Hadden, Structure-activity relationship studies of vitamin D3 analogues containing an ether or thioether linker as Hedgehog pathway inhibitors. ChemMedChem 13 (2018) 748-753.

[13] G. B. Carballo, J. R. Honorato, G. P. Farias de Lopes, T. C. Leite de Sampaio e Spohr, A highlight on sonic hedgehog pathway. Cell Communication and Signaling 16 (2018) 1-15.

[14] X. Dong, C. Wang, Z. Chen, W. Zhao, Overcoming the resistance mechanisms of Smoothened inhibitors. Drug Discovery Today 23 (2018) 704-710.

[15] F. Ghirga, M. Mori, P. Infante, Current trends in Hedgehog signaling pathway inhibition by small molecules. Bioor. Med. Chem. Lett. 28 (2018) 3131-3140.

[16] M. Xin, X. Ji, L. K. De La Cruz, S. Thareja, B. Wang, Strategies to target the Hedgehog signaling pathway for cancer therapy. Med. Res. Rev. 38 (2018) 870-913.

[17] J. Kim, J.Y. Tang, R. Gong, J. Kim, J.J. Lee, K.V. Clemons, C.R. Chong, K.S. Chang, M. Fereshteh, T. Reya, J.O. Liu, E.H. Epstein, D.A. Stevens, P.A. Beachy, Itraconazole, a commonly used anti-fungal that inhibits Hedgehog pathway activity and cancer growth Cancer Cell 17 (2010) 388–399.

[18] J. Kim, B.T. Aftab, J.Y. Tang, D. Kim, A.H. Lee, M. Rezaee, J. Kim, B. Chen, E.M. King, A. Borodovsky, G.J. Riggins, E.H. Epstein Jr., P.A. Beachy, C.M. Rudin, Itraconazole and arsenic trioxide inhibit Hedgehog pathway activation and tumor growth associated with acquired resistance to smoothened antagonists. Cancer Cell 23 (2013) 23-34.

[19] J.R. Pace, A.M. DeBerardinis, V. Sail, S.K. Tacheva-Grigorova, K.A. Chan, R. Tran, D.S. Raccuia, R.J. Wechsler-Reya, M.K. Hadden, Repurposing the clinically efficacious antifungal agent itraconazole as an anticancer chemotherapeutic. J. Med. Chem. 59 (2016) 3635–3649.

[20] Pace, J. R.; Teske, K. A.; Chou, L. Q.; Wechsler-Reya, R. J.; Hadden, M. K. Structureactivity relationships for itraconazole-based triazolone analogues as hedgehog pathway inhibitors. *J. Med. Chem.* Under Revision.

[21] B. Chen, V. Trang, A. Lee, N.S. Williams, A.N. Wilson, E.H. Epstein, J. Y. Tang, J. Kim, Posaconazole, a second-generation triazole antifungal drug, inhibits the hedgehog signaling and progression of basal cell carcinoma. Mol. Cancer Ther. 15 (2016) 866-876.

[22] L. Hoch, H. Faure, H. Roudaut, A. Schoenfelder, A. Mann, N. Girard, L. Bihannic, O. Ayrault, E. Petricci, M. Taddei, D. Rognan, M. Raul, MRT-92 inhibits hedgehog signaling by

blocking overlapping binding sites in the transmembrane domain of the smoothened receptor. FASEB J. 29 (2015) 1817-1829.

[23] A.K. Saksena, V.M. Girijavallabhan, H. Wang, Y.–T. Liu, R.E. Pike, A.K. Ganguly, Concise asymmetric routes to 2,2,4-trisubstituted tetrahydrofurans via chiral titanium imide enolates: Key intermediates towards synthesis of highly active azole antifungals SCH 51048 and SCH 56592. Tet. Lett. 37 (1996) 5657-60.

[24] P.–L. So, A.W. Langston, N. Daniallinia, J.L. Hebert, M.A. Fujimoto, Y. Khaimskiy, M. Asxterbaum, E.H. Epstein, Long-term establishment, characterization and manipulation of cell lines from mouse basal cell carcinoma tumors. Exp. Dermatol. 15 (2006) 742-750.

[25] S.N. Brun, S.L. Markant, L.A. Esparza, G. Garcia, D. Terry, J.–M. Huang, M.S. Pavlyukov, X.–N. Li, G.A. Grant, J.R. Crawford, M.L. Levy, E.M. Conway, L.H. Smith, I. Nakano, A. Berezov, M.I. Greene, Q. Wang, R.J. Wechsler-Reya, Survivin as a therapeutic target in Sonic hedgehog-driven medulloblastoma. Oncogene 34 (2015) 3770–3779.

[26] C. Wang, H. Wu, V. Katritch, G.W. Han, X.–P. Huang, W. Liu, F.Y. Siu, B.L. Roth, V. Cherezov, R.C. Stevens, Structure of the human smoothened receptor bound to an antitumour agent. Nature 497 (2013) 338-343.

[27] P. Huang, S. Zheng, B.M. Wierbowski, Y. Kim, D. Nedelcu, L. Aravena, J. Liu, A.C. Kruse and A. Salic, Structural basis of smoothened activation in hedgehog signaling. Cell 174 (2018) 312-324.

[28] X. Zhang, F. Zhao, Y. Wu, J. Yang, G.W. Han, S. Zhao, A. Ishchenko, L. Ye, X. Lin, K. Ding, V. Dharmarajan, P.R. Griffin, C. Gati, G. Nelson, M.S. Hunter, M.A. Hanson, V. Cherezov, R.C. Stevens, W. Tan, H. Tao and F. Xu, Crystal structure of a multi-domain human smoothened receptor in complex with a super stabilizing ligand. Nat. Commun. 8 (2017) 15383.

[29] E.F.X. Byrne, R. Sircar, P.S. Miller, G. Hedger, G. Luchetti, S. Nachtergaele, M.D. Tully,
L. Mydock-McGrane, D.F. Covey, R.P. Rambo, M.S.P. Sansom, S. Newstead, R. Rohatgi, C.
Siebold, Structural basis of Smoothened regulation by its extracellular domains. Nature 535
(2016) 517-522.



Graphical Abstract:

Highlights

1. A des-triazole posaconazole (PSZ) analogue maintained potent inhibition of the hedgehog (Hh) signaling pathway with enhanced drug-like properties.

2. Docking studies provide insight into the binding orientation these compounds adopt when complexed with their molecular target Smoothened.

3. Structure-based design of PSZ analogues resulted in compounds with improved potency and stability.

4. Molecular dynamics simulations demonstrate that PSZ and analogues stabilize an inactive conformation of Smo upon binding.