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# A new chemotype with promise against Trypanosoma cruzi

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ARTICLE INFO	A B S T R A C T
Keywords: Chagas disease Trypanosoma cruzi SAR	Pyridyl benzamide <b>2</b> is a potent inhibitor of <i>Trypanosoma cruzi</i> , but not other protozoan parasites, and had a selectivity-index of $\geq 10$ . The initial structure–activity relationship (SAR) indicates that benzamide and sulfonamide functional groups, and <i>N</i> -methylpiperazine and sterically unhindered 3-pyridyl substructures are required for high activity against <i>T. cruzi</i> . Compound <b>2</b> and its active analogs had low to moderate metabolic stabilities in human and mouse liver microsomes.

Chagas disease, endemic in over 20 Latin American countries, is a vector-borne infection with the protozoan parasite *Trypanosoma cruzi.*<sup>1,2</sup> The nitroaromatic drugs nifurtimox and benznidazole are effective against the acute phase of the disease, but are less useful against the chronic stage.<sup>1</sup> Both of these drugs have relatively low therapeutic indices and are poorly tolerated. Fexinidazole, another nitroaromatic compound, and two azole antifungal drugs, posaconazole and fosravuconazole, are in clinical trials for Chagas disease.<sup>3</sup> However, for the latter two drugs, there was recrudescence after treatment ceased, indicating a lack of trypanocidal activity. Thus, we need better drugs for this neglected parasitic disease.

We were interested in identifying new hit compounds against pathogenic protozoa by starting with known glucose transporter (GLUT) inhibitors<sup>4</sup> and modifying structure to enable selective inhibition of protozoal hexose transporters<sup>5</sup> and parasite growth. We chose to start with pyridyl benzamide **2**,<sup>6</sup> an optimized analog of the reported GLUT1 inhibitor STF-31 (1)<sup>7</sup> Fig. 1). Our initial screen of **2** revealed that it was a potent inhibitor of *T. cruzi*, but not other protozoan parasites. We now describe physicochemical profiling, in vitro ADME, and antiprotozoal activity for **2** and analogs **3–19** designed to establish a baseline structure–activity relationship (SAR) for this chemotype.

Target compounds 2–19 were prepared (Supporting Information) by a variety of reactions described in Schemes 1–4. Compounds 3–9 were prepared in nucleophilic aromatic substitution reactions between aryl fluoride  $20^6$  and the requisite secondary amines in moderate (42–64%) yields (Scheme 1). For 3, a subsequent Boc deprotection with MsOH was required.

Compounds **10–16** were in a two-step sequence starting with carboxylic acid **21<sup>6</sup>** (Scheme 2). The first step was in situ formation of the acid chloride or HOBt active ester of **21** followed by amide bond formation to afford intermediates **22–28**. This was followed by nucleophilic aromatic substitution reactions with 1-methylpiperazine to afford **10–16** in moderate to good (33–72%) yields.

Compounds 17 and 18 were obtained in a two-step reaction sequence (Scheme 3). Reactions of the HOBt active esters of  $29^8$  and  $30^9$  with 3-aminopyridine afforded 31 (96%) and 32 (97%), respectively. Nucleophilic aromatic substitution reactions of 31 and 32 with 1-methylpiperazine afforded 17 and 18 in moderate yields.

Compound **19** was prepared in moderate yield by converting **21** to **33** by nucleophilic aromatic substitution reaction with 1-methylpiperazine followed by 1,2,4-oxadiazole formation by condensation with *N*hydroxynicotinimidamide (Scheme 4). Target compounds **2**, **5**, and **6** were prepared following procedures described by Sutphin et al.<sup>6</sup>

In vitro assays with the Tulahuen C4 strain of *Trypanosoma cruzi* were performed as previously described.<sup>10</sup> Data for **2–9** in Table 1 reveal the SAR of the *N*-methylpiperazine substructure of **2**. First, we note that **2** has potent activity against *T. cruzi* with an IC<sub>50</sub> of 0.007  $\mu$ M; this compares with an IC<sub>50</sub> value of 1.2 to 4.2  $\mu$ M for the benznidazole control. Second, relatively minor structural changes to the *N*-methylpiperazine substructure decreased activity against *T. cruzi* significantly.

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# R = \* - N N R R = 1 (STF-31)

Fig. 1. Structures of STF-31 (1) and its analog 2.



**Scheme 1.** Reagents and conditions: (a)  $K_2CO_3$ , DMA, 110 °C, 20 h and 2,6-*cis*dimethylpiperazine (4), morpholine (5), piperidine (6), 4-hydroxypiperidine (7), ethyl piperazine-1-carboxylate (8), or azetidine hydrochloride (9); (b) *tert*butyl piperazine-1-carboxylate (3),  $K_2CO_3$ , DMA, 110 °C, 20 h, then MsOH, rt, 24 h followed by aq. NaHCO<sub>3</sub>.



**Scheme 2.** Reagents and conditions: (a) SOCl<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to 50 °C, 3 h, then 4-aminopyridine (**22**), 2-aminopyrazine (**23**), 5-aminopyrimidine (**24**), or 5-amino-2-pyridinecarbonitrile (**25**); (b) HOBt, EDCl, TEA, DMA, rt, 24 h and 3-amino-2-methylpyridine (**26**), 3-amino-6-methylpyridine (**27**), or 3-amino-2,6-dimethylpyridine (**28**); (c) 1-methylpiperazine, K<sub>2</sub>CO<sub>3</sub>, DMA, 110 °C, 20 h.

For example, *N*-desmethylpiperazines **3** and **4**, piperidine **6**, piperidinol **7**, and piperazine carbamate **8** were three-orders of magnitude less potent than **2**. Although morpholine **5** and azetidine **9** were also less potent than **2**, they had *T. cruzi*  $IC_{50}$  values of 2.1 and 8.3  $\mu$ M, respectively. For **2–9**, we could discern no clear SAR trends except that a *N*-

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**Scheme 3.** Reagents and conditions: (a) 3-aminopyridine, HOBt, EDCl, TEA, DMA, rt, 24 h; (b) 1-methylpiperazine, K<sub>2</sub>CO<sub>3</sub>, DMA, 110 °C, 20 h.



Scheme 4. Reagents and conditions: (a) 1-methylpiperazine,  $K_2CO_3$ , DMA, 110 °C, 20 h; (b) *N*-hydroxynicotinimidamide, EDCl, DMA, MW 150 °C, 10 min.

methylpiperazine was the optimal substructure for T. cruzi potency.

Data for **10–16** (Table 1) reveals that any change to the 3-pyridyl substructure of **2** reduces potency by three-orders of magnitude. These include other heterocycles (pyrazine **11**, pyrimidine **12**) or substitution with cyano (**13**) or methyl (**14–16**) groups. Finally, the weak activities of **17** and **18** (Table 1) indicate that the sulfonamide and benzamide substructures of **2** are required for high activity against *T. cruzi*. Although **19**, with its 1,2,4-oxadiazole carboxamide isostere, was less potent than **2**, it had a *T. cruzi* IC<sub>50</sub> value of 1.2  $\mu$ M.

To assess host cell cytotoxicity, the three most potent compounds **2**, **5**, and **19** were tested for growth inhibition of four human cell lines.<sup>11</sup> foreskin fibroblast (HFF), osteosarcoma (U-2 OS), kidney (HEK 293 T), and hepatocyte (HC-04) (Table 2). Only **5** inhibited the HFF cell line, whereas both **2** and **5** inhibited the U-2 OS and HEK 293 T cell lines with IC<sub>50</sub> values < 1  $\mu$ M; the HC-04 cell line was unaffected by all three compounds at concentrations up to 50  $\mu$ M. Compound **19** was the least cytotoxic of the three. These values compare with previously reported<sup>12</sup> IC<sub>50</sub> values for **2** ranging from 0.01 to 1  $\mu$ M against a panel of cancer cell lines.

Compound **2**, **5**, and **19** were tested against a panel of protozoan parasites (Table 3) using the methods of Orhan et al.<sup>10</sup> and Sanford et al.<sup>11</sup> Compound **2** was three-orders of magnitude less potent against *Plasmodium falciparum* and *Trypanosoma brucei rhodesiense* than it was against *T. cruzi* (Table 1) and it had no effect on the growth of

Table 2 Cvtotoxicity screen of 2. 5. and 19.

Compd	Cytotoxicity	Cytotoxicity IC <sub>50</sub> (μM)			
	HFF	U-2 OS	HEK 293 T	HC-04	
2	> 50	0.15	0.36	> 50	
5	2.0	0.16	0.95	> 50	
19	> 50	28	42	> 50	

Table 3

Activity of 2, 5, and 19 against a panel of protozoan parasites.

Compound	IC <sub>50</sub> (µM) <sup>a</sup>			
	P. falciparum NF54	T. b. rhodesiense STIB 900	L. donovani <sup>b</sup> MHOM-ET 67/L82	<i>T. gondii</i> <sup>c</sup> RH-dTom
2	10	32	> 200	> 100
5	21	25	180	> 100
19	5.8	3.4	49	21
Drug Standards <sup>d</sup>	0.0042	0.0070	0.99	0.80

<sup>a</sup> Mean from  $n \ge 2$ , individual measurements differed by less than 50%.

<sup>b</sup> The compounds were tested against *L. donovani* amastigotes in an axenic assay.

<sup>c</sup> RH (Type I) strain *T. gondii* with inserted fluorescent transgene dimerized Tomato (dTom).

<sup>d</sup> Chloroquine for *P. falciparum*; melarsoprol for *T. b. rhodesiense*; miltefosine for *L. donovani*; pyrimethamine for *T. gondii*.

### Table 4

Physicochemical properties and in vitro metabolic stability of 2, 5, and 19.

Compd	gLogD <sub>7.4</sub> <sup>a</sup>	PSA (Å <sup>2</sup> ) <sup>b</sup>	$Sol_{2.0}/Sol_{6.5}$ (µg/mL) <sup>c</sup>	h/m CL <sub>int</sub> (μL/min/mg protein) <sup>d</sup>	cPPB (%) <sup>e</sup>
2	2.8	95.8	> 100/25–50	45/112	85.2
5	2.6	100.6	50–100/12.5–25	69/200	76.2
19	4.1	105.7	> 100/25–50	535/677	not assessed

 $^{\rm a}$  LogD values were estimated by correlation of their chromatographic retention properties using a modified gradient HPLC method adapted from Lombardo et al.  $^{13}$ 

<sup>b</sup> Calculated using ChemAxon JChem for Excel.

<sup>c</sup> Compounds in DMSO were spiked into either pH 6.5 phosphate buffer or 0.01 M HCl (approx. pH 2.0) and analyzed by nephelometry<sup>14</sup> to determine a concentration range.

<sup>d</sup> In vitro intrinsic clearance measured in human and mouse liver microsomes.

<sup>e</sup> Plasma protein binding was estimated using a gradient HPLC method<sup>15</sup> where the chromatographic retention on a human albumin column was compared against the properties of standard compounds with known binding values.

*Leishmania donovani* and *Toxoplasma gondii*. Compound **5** was 10-fold less active against *P. falciparum* and *T. b. rhodesiense* than it was against *T. cruzi* (Tables 1 and 3) and it had weak to no activity against the other two protozoans. In contrast, there was little difference in the potency of **19** against *T. cruzi* (Tables 1 and 3), *P. falciparum* and *T. b. rhodesiense*, and it was an order of magnitude less potent against the other two protozoans.

Physicochemical and in vitro ADME properties of **2** and its most potent analogs **5** and **19** are shown in Table 4. The  $gLogD_{7.4}$  values ranging from 2.6 to 4.1, calculated polar surface area (PSA) values<sup>16</sup> ranging from 95 and 106 Å<sup>2</sup>, aq. solubilities ranging from 12.5 to > 100 µg/mL, and plasma protein binding < 90% suggest that these compounds would be expected to have relatively good biopharmaceutical properties. However, all of the compounds had relatively high intrinsic clearance values in human and mouse liver microsomes indicating that each would be expected to be rapidly metabolized. Metabolic stabilities were higher in human vs. mouse liver microsomes. Compound **19**, the most lipophilic of the three, had the lowest metabolic stability.

In this SAR scoping study, we started with **2**, a known<sup>7</sup> GLUT1 inhibitor. As we did not assess inhibition of glucose transport in T. cruzi<sup>17</sup> by 2-19, we cannot confirm that the T. cruzi hexose transporter is a potential cellular target for these compounds. In this respect, a recent study<sup>12</sup> claimed that nicotinamide phosphoribosyltransferase (NAMPT), not GLUT-1, is the target of 1 and 2 in cancer cell lines. In addition, posaconazole and a high proportion of hits identified in HTS campaigns against *T. cruzi* appear to inhibit sterol  $14\alpha$ -demethylase (*T.* cruzi CYP51), and most of these hits contain imidazole or pyridyl substructures.<sup>1,18–21</sup> Of the pyridyl-containing T. cruzi CYP51 inhibitors, a sterically unhindered 3-pyridyl substructure was predominant. Thus, we wondered whether T. cruzi CYP51 might be a potential target of this compound series based on our observed SAR - particularly the dramatic (three orders of magnitude) loss of potency for 14-16 when methyl groups were introduced adjacent to the nitrogen atom in the 3-pyridyl substructure of 2.

To test this hypothesis, 2 and 14 were analyzed as T. cruzi CYP51 heme binding ligands<sup>22</sup> and for inhibition of enzyme activity in a reconstituted *T. cruzi* CYP51 activity assay.<sup>23</sup> While **2** induced a moderate type II spectral response in the Soret band (K<sub>d</sub> 1.32 µM, Fig. 2a), 14 did not cause any changes (not shown). The IC<sub>50</sub> values, determined in a 30 min reconstituted enzyme reaction, were 49 and 487 µM, respectively (Fig. 2b). These data suggest that CYP51 could be a target for 2 (presumably mediated by binding of pyridine moiety to the heme cofactor of CYP51),<sup>22</sup> although the inhibition at the enzyme level (IC<sub>50</sub> of 49 µM) does not seem sufficient to fully account for the potency at the cellular level (IC<sub>50</sub> of 7 nM). For comparison, posoconazole similarly inhibits the growth of T. cruzi (IC<sub>50</sub> of 5 nM), but has a T. cruzi CYP51 K<sub>d</sub> of 18 nM.<sup>22,24</sup> Thus, the mechanism of action of **2** probably includes another target(s) which is also suggested by the dramatic loss of potency against T. cruzi observed with small variations in the N-methylpiperazine substructure - e.g. piperazine 3 and morpholine 5. Finally, the potent activity of 2 on T. cruzi and the weak activity against the other protozoa suggest that the target may be highly T. cruzi specific.

### **Declaration of Competing Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Fig. 2. Ligand binding (a) and inhibition (b) of T. cruzi CYP51.

### Table 1

In vitro antitrypanosomal activity of 2-19 against the Tulahuen C4 strain of T. cruzi.



Compd	<i>T. cruzi</i> IC <sub>50</sub> (μM) <sup>a</sup>	Compd	<i>T. cruzi</i> $IC_{50}$ ( $\mu$ M) <sup>a</sup>
2	0.007	11	17
3	69	12	96
4	130	13	100
5	2.1	14	65
6	110	15	88
7	120	16	110
8	46	17	120
9	8.3	18	120
10	28	19	1.2

<sup>a</sup> Mean from  $n \ge 2$ , individual measurements differed by less than 50%.

### Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.bmcl.2019.126778.

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