ORIGINAL ARTICLE



Head-to-head bisbenzazole derivatives as antiproliferative agents: design, synthesis, in vitro activity, and SAR analysis

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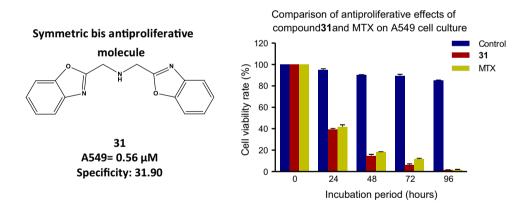
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Abstract

In the present work, a series of bisbenzazole derivatives were designed and synthesized as antiproliferative agents. The antiproliferative activity of these compounds was investigated using MTT assay. Bisbenzazole derivatives showed significant antiproliferative activity against all the four tested cancer cell lines. Among the various bisbenzazole derivatives, bisbenzoxazole derivatives exhibited the most promising anticancer activity followed by bisbenzimidazole and bisbenzothiazole derivatives. All the derivatives were found to be less toxic as compared to methotrexate (positive control) in normal human cells, indicating selective and efficient antiproliferative activity of these bisbenzazole derivatives. The structure–activity relationships of heteroaromatic systems and linkers present in bisbenzazole derivatives were analyzed in detail. In silico ADMET prediction revealed that bisbenzazole is a drug-like small molecule with a favorable safety profile. Compound **31** is a potential antiproliferative hit compound that exhibits unique cytotoxic activity distinct from methotrexate.

Graphic abstract

Twenty-one bisbenzoxazole derivatives have been designed synthesized and evaluated to be an antiproliferative activity against four human tumor cell lines.



Keywords Bisbenzimidazole · Bisbenzoxazole · Bisbenzothiazole · Antiproliferative activity · SAR · ADMET

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Extended author information available on the last page of the article

Introduction

Benzazoles are a family of heterocyclic compounds having a chemical skeleton consisting of a benzene ring fused with azole rings. Benz-fused azoles are among the most imperative class of molecules having a common heterocyclic

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scaffold found in several biologically active and medicinally significant compounds.

These compounds are known to exhibit several therapeutic activities, including anticancer, antimicrobial, antiparasitic, antiviral, antihistamine, fungicidal, and antitubercular activity [1-10]. Some of the well-known drugs with benzazole rings that are used in clinical applications are omeprazole, emedastine, candesartan, astemizole, bezitramide, domperidone, lansoprazole, flunoxaprofen, and riluzole.

Bisbenzazoles consist of two benzazole nuclei fused together using a variety of linkers. The wide spectrum of pharmacological activities displayed by bisbenzazole derivatives makes them a highly important scaffold from drug development perspective. There are several reports on the medicinal activities/properties of this class of compounds. Various studies have revealed that bis-derivatives show better antiproliferative activity than the monomeric compounds [11]. Ueki et al. and Sato et al. demonstrated the cytotoxic activity of UK-1 and AJI95618 against B-16, HeLa, and P-338 cancer cell lines [12, 13]. Hoechst 33258 has undergone phase I clinical evaluation as an anticancer agent. It has been proposed that Hoechst 33258 acts via inhibition of topoisomerase and DNA helicase [14, 15]. Rance et al. demonstrated the antiproliferative activity of novel bisbenzothiazole analogs [16]. Among them, bisbenzazole moiety is one of the most important structures in drug discovery. A number of studies have been conducted in past that involved design and synthesis of novel bioactive bisbenzazole derivatives and evaluation of their antiproliferative activity against different human cancer cell lines (Fig. 1) [17–19].

All these studies indicated the therapeutic significance of bisbenzazole derivatives. In the present study, head-tohead bisbenzazole derivatives were designed and synthesized by accumulation of two benzazole units with different aliphatic and heteroaliphatic linkers with the aim to obtain/achieve better antiproliferative agents.

As shown in Fig. 2, the designing of bisbenzazole derivatives with different linkers was broadly divided into two segments. The first segment involves main backbone

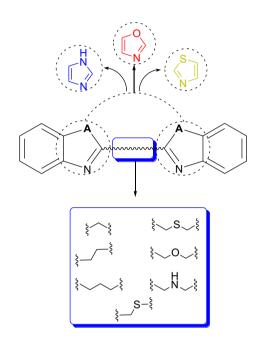
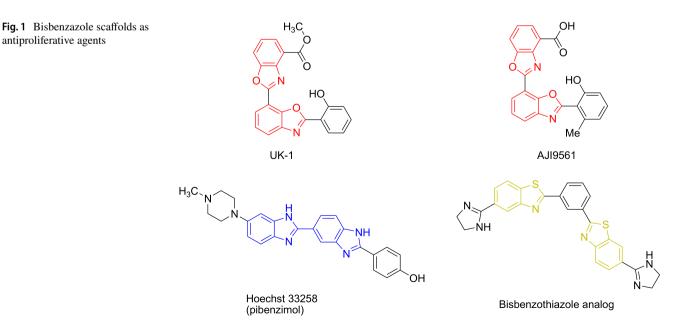
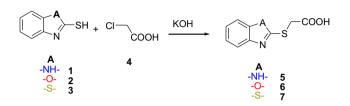


Fig. 2 Molecular design strategy for the synthesis novel bisbenzazole derivatives connected via aliphatic and heteroaliphatic linkers



antiproliferative agents

of the design, a benzazole unit that acts as a source of hydrogen-bond acceptor and donor. This would help to enhance the pharmacophoric properties as they exhibit drug-like properties. The second segment included aliphatic linkages with or without heteroatoms, which were generally used to control the lipophilicity and flexibility of the skeleton, and in silico ADMET prediction was also performed using the compounds.



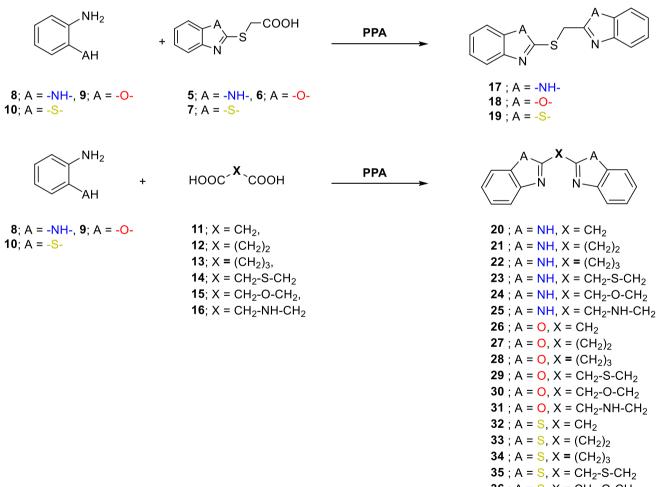
Scheme 1 Synthesis 2-((1H-benzazol-2-yl)thio)acetic acid (5-7) from 2-mercaptobenzazoles (1-3) and chloroacetic acid (4)

Results and discussion

Chemistry

The symmetric and asymmetric bisbenzazole derivatives (17–37) were synthesized using one-step and multi-step synthetic pathways as illustrated in Schemes 1 and 2. Polyphosphoric acid (PPA) method was used for the synthesis of desired compounds (20–37) (Scheme 2). For the synthesis of asymmetric compounds (17–19), 2-((1*H*-benzazol-2-yl)thio)acetic acid derivatives (5–7) were used as the starting material. These were prepared from 2-mercaptobenzazoles (1–3) and chloroacetic acid, followed by treatment with potassium hydroxide (Scheme 2).

The structures of all the designed and synthesized bisbenzazole derivatives (17–37) are shown in Scheme 2. All synthetic analogs (17–37) were characterized by physical and spectral analysis (FTIR, ¹H NMR, ¹³C NMR) and



36 ; A = S, X = CH₂-O-CH₂ **37** ; A = S, X = CH₂-NH-CH₂

Scheme 2 Synthesis of symmetric and asymmetric bisbenzazole derivatives using PPA method (20–37)

elemental analysis. (For details, please see the Supplementary file.)

The chemical shifts of the ¹H NMR signals of bisbenzazole derivatives were investigated, and the signal of two protons of -CH2- linker group and four protons of -CH₂-CH₂- linker group between benzazole rings is observed as a singlet with chemical shift values 3.35–4.88 ppm, while the signal of six protons of -CH₂-CH₂-CH₂- linker group between benzazole rings is observed as a triplet of four protons of two terminals -CH₂- attached to benzazole rings with chemical shift values 2.88-3.21 ppm and pentet or multiplet of two protons of -CH₂- center of symmetry with chemical shift values 2.26-2.45 ppm. The signal of two protons of -CH₂-S- linker group and four protons of -CH₂-S-CH₂-, -CH₂-NH-CH₂-, -CH₂-O-CH₂- linker groups between benzazole rings is observed as a singlet with chemical shift values 3.58-5.20 ppm. The signal of aromatic -NH- protons of bisbenzimidazole rings is observed as a singlet with chemical shift values 12.70-13.69 ppm. The signal of aromatic protons of benzazole rings is observed as (s, d, dd, ddd, td, t, and m) with chemical shift values 6.87-8.17 ppm. The chemical shifts of the ¹³C NMR signals of bisbenzazole derivatives were investigated; the values were 22.6–69.9 ppm for aliphatic carbons and 169.3–109.3 ppm for aromatic carbons.

Antiproliferative Activity

The in vitro antiproliferative activities of all the synthesized compounds (17-37) were evaluated in four human cancer cell lines, lung cancer (A549), kidney cancer (A498), cervical cancer (HeLa), and liver cancer (HepG2), using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT assay) [20–22]. An antiproliferative drug, methotrexate, was used as the standard for comparison. The results of the MTT assay are summarized in Table 1. The antiproliferative activity of each analog was presented as the concentration of the compound that led to a 50% inhibition (IC_{50}) of cancer cell growth. These compounds were further screened for their cytotoxic nature using Vero cells (normal cell line). This was used to establish the selectivity of these active compounds toward cancerous cells. A potent anticancer compound should be more selective toward cancer cells and less toxic toward the normal cells.

To assess the specificity of the compounds, their toxicity levels were tested against Vero cells. The specificity of the compounds was calculated as their IC_{50} values for normal cells divided by their IC_{50} values for the specific cancer cells. Table 1 shows the specificity of all compounds.

Additionally, antiproliferative activity studies were performed at non-toxic concentrations of the compounds that were determined on the Vero cell line. The most specific compound **31** and MTX were exposed for a period of 0–96 h, and A549 cells were evaluated and subjected to an assessment of their cytotoxicity responses using an MTT assay (Fig. 3). The proliferation of cancer cells treated with a non-toxic and lower-concentration dose of the compound was inhibited in a time-dependent manner. After 24, 48, and 72 h of treatment, compound **31** was more effective, and the number of live cells was lower than was observed with MTX. The antiproliferative activity of the compound increased as the exposure time was extended, and after 96-h treatment, the tested compounds had an activity equal to that of MTX.

As shown in Table 1, all the synthesized compounds showed significant antiproliferative activity against all the tested cancer cell lines. Among the synthesized compounds, bisbenzimidazole compounds 20, 21, and 23; bisbenzoxazole compounds 18, 27, 30, and 31; and bisbenzothiazole compounds 32 and 33 showed promising antiproliferative activity against all the tested cancer cell lines. Among the asymmetric compounds bisbenzoxazole 18 and among the symmetric compounds bisbenzoxazole derivatives 30 and 31, and bisbenzothiazole 32 were found to be most potent as antiproliferative agents and showed low toxicity and high specificity.

As most of the compounds from this series showed potency against tested human cancer cell lines, they can serve as attractive lead molecules for the discovery of novel antiproliferative agents in future. When the compounds were evaluated according to selectivity, for A549 cells, 18, 30, and 31 were found to be most potent antiproliferative agents with IC₅₀ values of 0.28, 0.28, and 0.56 µM, respectively. The compounds 18, 30, 32, and 33 showed good antiproliferative activities against both A498 and HeLa cells. For these two cell lines, 18, 30, and 33 displayed IC_{50} values of 0.28 µM, 0.56 µM, and 0.52 µM, respectively. Among the various compounds screened, 27 showed better specificity against A498, HeLa, and HepG2 cell lines. For HepG2 cells, **30** and **33** were found to be most potent as indicated by IC_{50} values of 0.28 µM. In addition to this, the compounds 18, 30, and 33 displayed potency against all the four cell lines. The compounds 18, 30, and 33 showed IC_{50} values of 0.28, 0.28, and 0.68 µM, respectively, in A549; IC₅₀ values of 0.28, 0.56, and 0.52 μ M, respectively, in A498; IC₅₀ values of 0.56, 0.56, and 0.56 μ M, respectively, in HeLa; and IC₅₀ values of 0.56, 0.28, and 0.28 µM, respectively, in HepG2 cells.

For cytotoxicity test in Vero cells, **31** showed IC₅₀ value of 17.90 μ M, indicating 32 times more selectivity toward A549 and eight times more selectivity toward A498 and HeLa cancer cell lines. Similarly, **27** displayed IC₅₀ value of 18.92 μ M against Vero cells. Thus, **27** was found to be 16 times more selective toward A498, HeLa, and HepG2 cells and eight times more selective toward A549 cancer cell line.

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HN'		Vero ⁰	$A549^{c}$	$A498^{c}$	HeLa ^c	HepG2 ^c	A549	A498	HeLa	HepG2
		27.66±2.19	221.38 ± 10.7	221.38 ± 5.36	442.70 ± 5.12	221.38 ± 7.06	0.12	0.12	0.06	0.12
		1.13 ± 0.84	0.28 ± 0.12	0.28 ± 0.63	0.56 ± 0.79	0.56 ± 0.41	4.00	4.00	2.00	2.00
- HN' - HN' - HN' - HN' - HN' - HN' - HN' - HN'		12.44 ± 0.86	48.60 ± 2.24	49.52 ± 2.57	49.35 ± 2.81	49.24 ± 3.56	0.26	0.25	0.25	0.25
HN' - HN'		5.030 ± 0.07	1.26 ± 1.55	1.26 ± 0.07	2.52 ± 1.77	2.52 ± 0.78	4.01	4.01	2.00	2.00
	H ₂	0.60 ± 0.25	0.60 ± 0.38	1.19 ± 0.27	0.60 ± 0.29	0.60 ± 0.13	1.00	0.50	1.00	1.00
-HN- -HN- -HN-	H_2 - CH_2 -	36.19 ± 1.17	18.10 ± 2.22	36.19 ± 2.88	36.18 ± 2.65	36.19 ± 2.43	2.00	1.00	1.00	1.00
-HN- -HN-	$-CH_2$	2.12 ± 0.22	2.12 ± 0.45	1.06 ± 0.15	2.12 ± 0.54	$2.12 \pm 0.01.89$	1.00	2.00	1.00	1.00
-HN-	$-CH_2$	17.970 ± 3.41	8.98 ± 1.35	4.49 ± 1.02	8.98 ± 1.56	4.49 ± 0.96	2.00	4.00	2.00	4.00
-0-	H-CH ₂	14.10 ± 4.32	55.17 ± 3.52	56.36 ± 5.22	56.35 ± 2.63	56.36 ± 3.31	0.26	0.25	0.25	0.25
		9.99 ± 1.28	4.99 ± 0.98	4.99 ± 1.67	4.99 ± 1.48	9.99 ± 2.57	2.00	2.00	2.00	1.00
27 $-0 -CH_2-CH_2-$	H ₂ -	18.92 ± 3.67	2.37 ± 0.85	1.18 ± 0.66	1.18 ± 0.29	1.18 ± 0.45	8.00	16.03	16.03	16.03
28 –0– –CH ₂ –CH ₂	H_2 - CH_2 -	35.93 ± 2.98	35.93 ± 1.88	17.97 ± 2.33	17.96 ± 2.53	35.93 ± 2.49	1.00	2.00	2.00	1.00
29 –0– –CH ₂ –S–CH ₂	$-CH_2$	33.75 ± 1.02	16.87 ± 1.05	16.87 ± 2.63	16.87 ± 0.36	16.87 ± 2.33	2.00	2.00	2.00	2.00
30 –O– –CH ₂ –O–CH ₂	$-CH_2$	2.23 ± 0.62	0.28 ± 0.37	0.56 ± 0.25	0.56 ± 0.36	0.28 ± 0.10	8.01	4.01	4.01	8.01
31 –0– –CH ₂ –NH–CH ₂	$H-CH_2$	17.90 ± 0.71	0.56 ± 0.38	2.24 ± 0.84	2.24 ± 0.45	2.24 ± 0.63	31.90	8.00	8.00	8.00
32 –S– –CH ₂ –		4.42 ± 0.83	1.11 ± 0.52	0.56 ± 0.96	1.11 ± 0.32	1.11 ± 0.06	4.01	8.01	4.01	4.01
33 –S– –CH ₂ –CH ₂ –	$H_{2}-$	1.05 ± 0.16	0.68 ± 0.55	0.52 ± 0.39	0.56 ± 0.72	0.28 ± 0.96	2.00	2.00	2.00	4.00
34 –S– –CH ₂ –CH ₂ –CH ₂ –	H_2 - CH_2 -	32.21 ± 2.97	16.11 ± 1.49	16.11 ± 5.33	16.10 ± 1.43	16.11 ± 0.85	2.00	2.00	2.00	2.00
35 –S– –CH ₂ –S–CH ₂	$-CH_2$	7.61 ± 1.04	7.61 ± 1.56	3.81 ± 0.44	3.81 ± 0.36	7.61 ± 1.79	1.00	2.00	2.00	1.00
36 –S– –CH ₂ –O–CH ₂	$-CH_2$	4.00 ± 1.73	4.00 ± 1.46	2.00 ± 1.01	4.00 ± 0.75	4.00 ± 1.39	1.00	2.00	1.00	1.00
37 –S– –CH ₂ –NH–CH ₂	$H-CH_2$	32.11 ± 3.55	16.10 ± 1.86	16.10 ± 2.57	16.05 ± 3.55	16.10 ± 2.84	2.00	2.00	2.00	2.00
Methotrexate ^e		0.030 ± 0.41	0.025 ± 0.53	0.016 ± 0.28	0.022 ± 0.36	0.041 ± 0.12	1.20	1.87	1.36	0.73
Abbreviation Vero; African green monkey kidney epithelial cell, line HepG2; human hepatocellular cell line. ^a The reported values	ey kidney epitl ine. ^a The repo		human lung adeno ent the mean±SD	carcinoma epitheli for each compoun	al cell line, A498; d based on three ir	A549; human lung adenocarcinoma epithelial cell line, A498; human renal cancer cell line HeLa; human cervical cancer cell represent the mean \pm SD for each compound based on three independent experiments, ^b normal kidney epithelial cell, ^c cancer	cell line H ants, ^b norm	eLa; huma al kidney e	n cervical pithelial ce	cancer ell, ^c cai

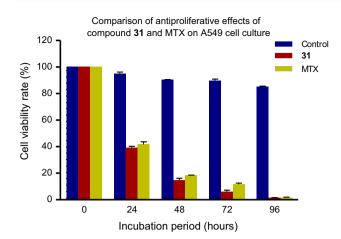


Fig. 3 MTT assay on A549 cell line after 96 h with MTX and compound **31**. The absorbance values were selected as 570 nm for the MTT method. Control cells not containing compounds and MTX were incubated same conditions. Cell viability was calculated as the ratio of absorbance of treated cells with compound or MTX to untreated cells. Given values show the mean standard deviations from three independent experiments carried out in triplicate. *Note: p* values: 24 h: control—31; *p*<.000; control—MTX; *p*<.000; control—MTX

Furthermore, compound **30** with IC_{50} value 2.23 µM against Vero cells showed eight times more selectivity toward A549 and HepG2. The results of this comparative analysis suggested that the most potent compounds from this newly synthesized series were characterized by more selectivity toward cancerous cells and very less toxicity toward the normal cells.

SAR analysis

Most of the compounds from the series displayed moderate to good antiproliferative activity toward cancer cell lines. A careful examination of the data helped to establish a significantly regular SAR. The synthesized compounds were analogs of bisbenzazoles with varying alkyl or heteroalkyl linkers. Accordingly, SAR was established by the comparison of the antiproliferative activities of the different linker and bisbenzazole groups. Among these compounds, bisbenzimidazole derivatives 20, 21, and 23; bisbenzoxazole derivatives 18, 26, 27, 30, and 31; and bisbenzothiazole derivatives 32 and 33 exhibited the most potent antiproliferative activity. In contrast, compounds 22, 24, 25, 28, 29, and 34-37 with different linker and bisbenzazole groups showed the lowest antiproliferative activity against all cancer cells. In particular, the 1C linker in bisbenzothiazole 32 and 2C linker in bisbenzimidazole and bisbenzothiazole 21 and 33 were found to be associated with increased potency of the compounds, as compared to the

bisbenzoxazoles **26** and **27**, respectively. In addition, the threeatom linker groups ($-CH_2-CH_2-CH_2-$, $-CH_2-O-CH_2-$, and $-CH_2-NH-CH_2-$) in the benzazole derivatives (**22–25**, **28**, **29**, and **34–37**) did not significantly improve the activities against cancer cell lines, except for compound **31** against A549 and compound **30** in all cancer cell lines. In general, the 3C linkers in all benzazole scaffolds were inactive compounds for antiproliferative activity.

The resulting data are presented in Table 1. It shows that all the active compounds had generally considerable activity against cancer cell lines with IC_{50} values of 0.28–2.12 μ M. Compound **30** was the most active agent against A549 and HepG2 cells with an IC_{50} value of 0.28 μ M.

Physicochemical and toxicology properties

After completing structural characterization and biological activity studies, important points to know about these compounds were their physicochemical and toxicological properties. (Details are provided in supplementary content of Table S1.) All compounds were analyzed using PRE-ADMET and DATAWARRIOR 4.07.02 software. (Details are provided in supplementary content of Table S2.) With respect to the toxicological parameters, all compounds were evaluated as good, as none showed the potential to be carcinogenic or mutagenic, and they showed a medium-level ability to inhibit the hERG potassium channel and CYP450. For ADME properties, in general, the compounds' inhibitors were well evaluated; only compound 31 presented a weak binding to plasma proteins, low absorption in the brain-blood barrier, and good absorption in the intestinal system. In the case of drug-like parameters, almost the all of the compounds reached values allowing them to be considered as potential oral drugs.

All of the synthesized compounds were found to comply with Lipinski's rule of five. With respect to the toxicological parameters, none of the synthesized compounds had estimated mutagenic, carcinogenic, irritant, or reproduction effects. In the case of drug-like parameters, all compounds can be considered as potential oral drugs because they are well absorbed in the human intestine. Except for compounds **29** and **35**, all compounds inhibit the hERG potassium channel. Only **25** of the compounds does not inhibit CYP450. In addition, compound **25**, unlike most of the others, follows the lead-like rule and is weakly bound to plasma proteins. (Details are provided in supplementary content of Table S2.)

Conclusion

Bisbenzazole analogs (17–37) were synthesized, characterized, and evaluated for their antiproliferative activities against different cell lines (A549, A498, HeLa, and HepG2). Having a $-CH_2-O-CH_2$ - linker at the 2-position of the benzoxazole moiety enhanced the cytotoxic activity against the designated cell lines at low micromolar concentrations. The structure-activity relationships of heteroaromatic systems and linkers present in bisbenzazole derivatives were analyzed in detail.

Compounds 18, 21, 30, and 33 displayed the highest antiproliferative activity and lower IC₅₀ (μ M) values against all cancer cell lines, compared to MTX. More remarkably, compound 31 showed approximately 16-fold better cytotoxicity on A549 cells, with approximately 13-fold high specificity, compared to MTX. The results also demonstrated that compound 27 has significant antiproliferative effects on A498, HeLa, and HepG2 cells, in comparison with MTX. Moreover, compound 27 showed more selective but much higher IC₅₀ values than other active compounds and MTX; therefore, it was a less active compound. As a result, we can conclude that the linker group consists of two or three atoms and that one of the atoms should be a heteroatom.

The clinical application of most anticancer drugs has been found to be associated with certain side effects or toxicity. These limitations of anticancer drugs are mostly attributed to the poor selectivity of drugs toward cancer cells. The selectivity of drugs is an important factor affecting their utility. In the present study, a pharmacophore hypothesis was developed to analyze SARs between the molecular structures of the synthesized bisbenzazole derivatives and observed biological activity in the A549 cell line. At the same time, the physicochemical properties of the compounds play an important role in reaching the active region of the cell and the interactions with the active region.

In conclusion, our studies have shown that bisbenzazole scaffolds have good antiproliferative effects on all cancer cell lines. However, linkers are also important for changing antiproliferative activities. SAR studies of symmetrical derivatives showed that 1C linker compounds have moderate activity, although 2C linker compounds displayed more improved antiproliferative activity against all tested cancer cell lines. Interestingly, all three-atom linker compounds were inactive. Also, in general, three-atom linker compounds are less active than 2C linker compounds, except for compound **30**, which showed good antiproliferative activity against all cancer cell lines.

For asymmetrical bisbenzoxazole derivative, compound **18** exhibited good antiproliferative activity compared to other benzazoles (**17** and **19**).

Finally, in silico ADMET prediction highlighted that all compounds have desirable drug-like properties, favorable safety profiles, and comply with Lipinski's rule of five.

Experimental

Chemistry

All reagents used were commercially available unless otherwise specified, and all solvents were distilled before use. The precursor (5-7) was synthesized using the reported method in the literature [23]. Melting points (mp) were determined with Mettler Toledo MP90 melting point device. General reaction visualization was achieved by thin-layer chromatography (TLC) purchased from Merck KGaA (silica gel 60 F254) based on Merck DC plates (aluminum based) by using UV light (254 nm). Chromatographic separations were carried out using silica gel 60 (Merck, 63-200 µm particle size, 60-230 mesh). FTIR spectra were recorded as ATR on a Perkin Elmer Spectrum One FTIR spectrometer. All the nuclear magnetic resonance spectra of the analogs obtained on Bruker spectrometers (400 MHz for ¹H NMR and 100 MHz for ¹³C NMR) in ppm (δ) refer to the solvent signal center at δ (7.19 and 76.0) ppm, δ (2.52 and 39.5) ppm, and δ (3.34 and 49.0) ppm for CDCl₃, d₆-DMSO, and d₄-CH₃OH, respectively. Chemical shifts (δ) are reported in ppm. Coupling constants (J) are reported in Hz. Standard abbreviations indicating multiplicity were used as follows: br s (broad singlet), s (singlet), d (doublet), t (triplet), q (quartet), p (pentet), m (multiplet), and dd (doublet of doublets).

General procedure for preparation of compounds (20-37) 1,2-Phenylenediamine (8), 2-aminophenol (9), or 2-mercaptoanilin (10) (2 eq) and the corresponding dicarboxylic acid derivatives (11, 12, 13, 14, 15, and 16) (1 eq) are heated for a period of 13-15 h in PPA at 180 °C. The reaction was monitored by thin-layer chromatography (TLC). UV (ultraviolet) light was used in the determination of stains in the works of TLC (Kieselgel 60 F254, ready-touse aluminum plate coated with 0.2 mm thickness) which was made by using ready-made plates. After cooling, the reaction mixture was poured into ice water and neutralized by mixing with 5 M NaOH till slightly basic pH (8–9) to get the precipitate. The resulting precipitate was filtered off, washed with cold water, and crystallized with a suitable solvent. The resulting crystalline compounds were filtered, and the vacuumed product was dried.

Bis(1H-benzo[d]imidazol-2-yl)methane (20) The above procedure was followed with **8** and **11** to yield **20** as a brown powder solid (36% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1)=0.60; **mp**=119–123 °C; **IR** (KBr, cm⁻¹) Vmax 3058, 2962, 1656, 1314, 756, 672. The ¹H NMR and ¹³C NMR spectra are in agreement with the reported data [24, 25]. **Anal. calcd.** for C₁₅H₁₂N₄: C, 72.56; H, 4.87; N, 22.57. Found: C, 72.44; H, 4.95; N, 22.63.

1,2-Bis(1H-benzo[d]imidazol-2-yl)ethane (21) [26, 27] The above procedure was followed with **8** and **12** to yield **21** as a white powder solid (55% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1)=0.58; **mp**=119–123 °C; **IR** (KBr, cm⁻¹) Vmax 3387, 3200, 2719, 2608, 1626, 1574, 814, 764; ¹H NMR (400 MHz, d₆-DMSO) δ 7.84–7.76 (m, 4H, Ar–H), 7.60–7.49 (m, 4H, Ar–H), 3.91 (s, 4H, CH₂); ¹³C NMR (100 MHz, d₆-DMSO) δ 151.5, 131.3, 125.3, 113.8, 23.7. **Anal. calcd.** for C₁₆H₁₄N₄: C, 73.26; H, 5.38; N, 21.36. Found: C, 73.34; H, 5.29; N, 21.25.

1,3-Bis(1H-benzo[d]imidazol-2-yl)propane (22) The above procedure was followed with **8** and **13** to yield **22** as a brown powder solid (65% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1)=0.48; **mp**=270–275 °C; **IR** (KBr, cm⁻¹) Vmax 3048, 2952, 1547, 1435, 735. The ¹H NMR and ¹³C NMR spectra are in agreement with the reported data [28, 29]. **Anal. calcd.** for C₁₇H₁₆N₄: C, 73.89; H, 5.84; N, 20.27. Found: C, 73.74; H, 5.79; N, 20.25.

Bis((1H-benzo[d]imidazol-2-yl)methyl)sulfane (23) The above procedure was followed with **8** and **14** to yield **23** as a brown powder solid (47% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1)=0.71; **mp**=119–122 °C; **IR** (KBr, cm⁻¹) Vmax 3055, 2781, 1510, 1437, 725. The ¹H NMR and ¹³C NMR spectra are in agreement with the reported data [30–32]. **Anal. calcd.** for C₁₆H₁₄N₄S: C, 65.28; H, 4.79; N, 19.03; S, 10.89. Found: C, 65.14; H, 4.89; N, 19.15; S, 10.95.

2,2'-(**Oxybis(methylene))bis(1H-benzo[d]imidazole) (24)** The above procedure was followed with **8** and **15** to yield **24** as an orange powder solid (70% yield). The crystallization solvent is ethanol–water. **R**_f (chloroform/methanol 9:1)=0.72; **mp** = 122–126 °C; **IR** (KBr, cm⁻¹) Vmax 3056, 2915, 1439, 731. The ¹H NMR spectrum is in agreement with the reported data [33, 34]. ¹³C NMR (100 MHz, DMSO-d₆) δ 157.5, 151.3, 136.6, 134.3, 129.6, 127.8, 123.8, 121.6, 116.1, 114.8, 114.6, 62.5. **Anal. calcd.** for C₁₆H₁₄N₄O: C, 69.05; H, 5.07; N, 20.13. Found: C, 69.14; H, 4.97; N, 20.25.

Bis((1H-benzo[d]imidazol-2-yl)methyl)amine (25) The above procedure was followed with 1 and 16 to yield 25 as

a brown powder solid (47% yield). The crystallization solvent is ethanol-water. **R**_f (ethyl acetate/hexanes 1:1)=0.60; **mp**=170-175 °C; **IR** (KBr, cm⁻¹) Vmax 3469, 3139, 2991, 1506, 1422, 747. The ¹H NMR and ¹³C NMR spectra are in agreement with the reported data [35–37]. **Anal. calcd.** for C₁₆H₁₅N₅: C, 69.29; H, 5.45; N, 25.25. Found: C, 69.32; H, 5.51; N, 25.37.

Bis(benzo[d]oxazol-2-yl)methane (26) The above procedure was followed with **9** and **11** to yield **26** as a brown powder solid (62% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1) = 0.86; **mp** = 121–123 °C; **IR** (KBr, cm⁻¹) Vmax 3093, 3066, 2956, 1614, 1570, 1239, 833, 742. The ¹H NMR and ¹³C NMR spectra are in agreement with the reported data [38, 39]. **Anal. calcd.** for $C_{15}H_{10}N_2O_2$: C, 71.99; H, 4.03; N, 11.19. Found: C, 72.12; H, 3.92; N, 11.15.

1,2-Bis(benzo[d]oxazol-2-yl)ethane (27) [40, 41] The above procedure was followed with 9 and 12 to yield 27 as an orange powder solid (60% yield). The crystallization solvent is ethanol–water. \mathbf{R}_{f} (chloroform/methanol 95:05) = 0.59; \mathbf{mp} = 135–138 °C; IR (KBr, cm⁻¹) Vmax 3098, 3059, 2926, 1611, 1569, 1242, 831, 752. ¹H NMR (400 MHz, CDCl₃) δ 7.66–7.57 (m, 2H, Ar–H), 7.47–7.38 (m, 2H, Ar–H), 7.26–7.23 (m, 4H, Ar–H), 3.51 (s, 4H, CH₂); ¹³C NMR (100 MHz, CDCl₃) δ 163.9, 160.0, 140.2, 123.8, 123.2, 118.8, 109.4, 24.5. Anal. calcd. for C₁₆H₁₂N₂O₂: C, 72.72; H, 4.58; N, 10.60. Found: C, 72.61; H, 4.63; N, 10.49.

1,3-Bis(benzo[d]oxazol-2-yl)propane (28) [**42, 43**] The above procedure was followed with **9** and **13** to yield **28** as a pink powder solid (70% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1)=0.63; **mp**=146–150 °C; **IR** (KBr, cm⁻¹) Vmax 3051, 2997, 1571, 1450, 1240, 756; ¹H NMR (400 MHz, CDCl₃) δ 7.62–7.54 (m, 2H, Ar–H), 7.43–7.36 (m, 2H, Ar–H), 7.26–7.20 (m, 4H, Ar–H), 3.06 (t, *J*=7.34 Hz, 4H, –CH₂), 2.45 (p, *J*=7.34 Hz, 2H, –CH₂); ¹³C NMR (100 MHz, CDCl₃) δ 165.0, 149.9, 140.3, 123.6, 123.2, 109.3, 26.8, 22.6. **Anal. calcd.** for C₁₇H₁₄N₂O₂: C, 73.37; H, 5.07; N, 10.07. Found: C, 73.41; H, 4.98; N, 10.21.

Bis(benzo[d]oxazol-2-ylmethyl)sulfane (29) [44, 45] The above procedure was followed with 9 and 14 to yield 29 as an orange powder solid (55% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1)=0.50; **mp** = 189–193 °C; **IR** (KBr, cm⁻¹) Vmax 3131, 2900, 1548, 1455, 1278, 744; ¹H NMR (400 MHz, d₄-CH₃OH) δ 7.78 (dd, J = 1.44, 8.01 Hz, 2H, Ar–H), 7.00 (td, J = 1.53, 8.06 Hz, 2H, Ar–H), 6.87 (dd, J = 1.24, 8.08 Hz, 2H, Ar–H), 6.87 (td, J = 1.44, 8.03 Hz, 2H, Ar–H), 3.63 (s, 4H, –CH₂); ¹³C NMR (100 MHz, DMSO-d₆) δ 157.6, 130.9, 128.8,

128.6, 116.6, 35.4. Anal. calcd. for $C_{16}H_{12}N_2O_2S$: C, 64.85; H, 4.08; N, 9.45; S, 10.82. Found: C, 64.96; H, 4.03; N, 9.38; S, 10.95.

2,2[']-(Oxybis(methylene))bis(benzo[d]oxazole) (30) [46] The above procedure was followed with 9 and 15 to yield 30 as a brown powder solid (35% yield). The crystallization solvent is ethanol–water. $\mathbf{R}_{\mathbf{f}}$ (ethyl acetate/hexanes 1:1)=0.50; \mathbf{mp} =191–194 °C; IR (KBr, cm⁻¹) Vmax 3048, 2948, 1589, 1452, 1251, 795; ¹H NMR (400 MHz, CDCl₃) δ 8.00 (d, J=8.14 Hz, 2H, Ar–H), 7.83 (d, J=7.94 Hz, 2H, Ar–H), 7.47 (t, J=7.74 Hz, 2H, Ar–H), 7.36 (t, J=7.19 Hz, 2H, Ar–H), 3.74 (s, 4H, –CH₂); ¹³C NMR (100 MHz, DMSO-d₆) δ 159.7, 151.3, 129.3, 126.3, 125.0, 122.2, 116.6, 63.9. Anal. calcd. for C₁₆H₁₂N₂O₃: C, 68.56; H, 4.32; N, 9.99. Found: C, 68.65; H, 4.43; N, 10.11.

Bis(benzo[d]oxazol-2-ylmethyl)amine (31) The above procedure was followed with 9 and 16 to yield 31 as a red powder solid (30% yield). The crystallization solvent is ethanol–water. **R**_f (chloroform/methanol 95:05) = 0.25; **mp** = 132–136 °C; **IR** (KBr, cm⁻¹) Vmax 3453, 3058, 2930, 1569, 1459, 1243, 783; ¹H NMR (400 MHz, CDCl₃) δ 7.68 (dd, J = 3.35, 5.66 Hz, 2H, NH), 7.52–7.46 (m, 2H, Ar–H), 7.32 (t, J = 6.43 Hz, 4H, Ar–H), 3.58 (s, 4H, –CH₂); ¹³C NMR (100 MHz, CDCl₃) δ 149.3, 127.0, 126.7, 123.5, 120.6, 116.7, 37.2. **Anal. calcd.** for C₁₆H₁₃N₃O₂: C, 68.81; H, 4.69; N, 15.05. Found: C, 68.75; H, 4.59; N, 15.17.

Bis(benzo[d]thiazol-2-yl)methane (32) The above procedure was followed with **10** and **11** to yield **32** as a green powder solid (49% yield). The crystallization solvent is ethanol–water. \mathbf{R}_{f} (chloroform) = 0.33; $\mathbf{mp} = 170-174$ °C; **IR** (KBr, cm⁻¹) Vmax 3053, 2951, 1592, 1590, 1503, 1062, 854, 729. The ¹H NMR and ¹³C NMR spectra are in agreement with the reported data [24, 25]. **Anal. calcd.** for C₁₅H₁₀N₂S₂: C, 63.80; H, 3.57; N, 9.92; S, 22.71. Found: C, 63.71; H, 3.66; N, 9.81; S, 22.64.

1,2-Bis(benzo[d]thiazol-2-yl)ethane (33) The above procedure was followed with **10** and **12** to yield **33** as a yellow powder solid (55% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1)=0.37; **mp**=137–140 °C; **IR** (KBr, cm⁻¹) Vmax 3055, 2993, 1591, 1510, 1088, 877, 724. The ¹H NMR and ¹³C NMR spectra are in agreement with the reported data [47, 48]. **Anal. calcd.** for $C_{16}H_{12}N_2S_2$: C, 64.83; H, 4.08; N, 9.45; S, 21.64. Found: C, 64.95; H, 3.96; N, 9.56; S, 21.76.

1,3-Bis(benzo[d]thiazol-2-yl)propane (34) [49, 50] The above procedure was followed with **10** and **13** to yield **34** as a green powder solid (61% yield). The crystallization solvent is ethanol–water. \mathbf{R}_{f} (chloroform)=0.61; $\mathbf{m}p$ =199–202 °C;

IR (KBr, cm⁻¹) Vmax 3050, 2981, 1515, 1436, 1050, 760; ¹**H NMR** (400 MHz, CDCl₃) δ 7.91 (d, J = 8.13 Hz, 2H, Ar–H), 7.77 (d, J = 7.99 Hz, 2H, Ar–H), 7.39 (t, J = 8.10, 2H, Ar–H), 7.29–7.27 (m, 2H, Ar–H), 3.21 (t, J = 7.05, 4H, –CH₂), 2.43 (m, 4H, –CH₂); ¹³**C NMR** (100 MHz, CDCl₃) δ 170.6, 153.3, 135.2, 126.0, 124.8, 122.7, 121.5, 33.4, 29.0. **Anal. calcd.** for C₁₇H₁₄N₂S₂: C, 65.77; H, 4.55; N, 9.02; S, 20.66. Found: C, 65.85; H, 4.66; N, 9.17; S, 20.47.

Bis(benzo[d]thiazol-2-ylmethyl)sulfane (35) [51, 52] The above procedure was followed with 10 and 14 to yield 35 as a brown powder solid (49% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1)=0.54; **mp**=189–193 °C; **IR** (KBr, cm⁻¹) Vmax 3059, 2966, 1513, 1433, 1095, 762; ¹H NMR (400 MHz, CDCl₃) δ 7.88 (d, J=8.12 Hz, 2H, Ar–H), 7.76 (d, J=7.95 Hz, 2H, Ar–H), 7.40–7.27 (m, 4H, Ar–H), 4.17 (s, 4H, –CH₂); ¹³C NMR (100 MHz, DMSO-d₆) δ 155.9, 135.5, 135.1, 129.0, 128.7, 127.2, 125.8, 115.8, 33.6. Anal. calcd. for C₁₆H₁₂N₂S₃: C, 58.50; H, 3.68; N, 8.53; S, 29.29. Found: C, 58.63; H, 3.74; N, 8.67; S, 29.12.

2,2[']-(Oxybis(methylene))bis(benzo[d]thiazole) (36) The above procedure was followed with 10 and 15 to yield 36 as a cream powder solid (67% yield). The crystallization solvent is ethanol–water. \mathbf{R}_{f} (ethyl acetate/hexanes 1:1)=0.72; \mathbf{mp} =103–106 °C; IR (KBr, cm⁻¹) Vmax 3062, 2884, 1528, 143, 1038, 757. The ¹H NMR spectrum is in agreement with the reported data [53, 54]. ¹³C NMR (100 MHz, DMSOd₆) δ 169.0, 152.6, 134.5, 126.3, 125.3, 122.7, 122.4, 69.9. Anal. calcd. for C₁₆H₁₂N₂OS₂: C, 61.51; H, 3.87; N, 8.97; S, 20.53. Found: C, 61.42; H, 3.73; N, 8.84; S, 20.44.

Bis(benzo[d]thiazol-2-ylmethyl)amine (37) The above procedure was followed with **10** and **16** to yield **37** as a yellow powder solid (40% yield). The crystallization solvent is ethanol–water. **R**_f (chloroform/methanol 95:05) = 0.27; **mp** = 101–104 °C; **IR** (KBr, cm⁻¹) Vmax 3055, 2995, 1510, 1437, 1012, 724. The ¹H NMR and ¹³C NMR spectra are in agreement with the reported data [55, 56]. **Anal. calcd.** for $C_{16}H_{13}N_3S_2$: C, 61.71; H, 4.21; N, 13.49; S, 20.59. Found: C, 61.82; H, 4.19; N, 13.37; S, 20.48.

Synthesis of 2-((benzazol-2-yl)thio)acetic acid (5–7) A mixture of 2-mercaptobenzimidazole (1), 2-mercaptobenzoxazole (2), 2-mercaptobenzothiazole (3) (1 eq), and potassium hydroxide (1.2 eq) in methanol (20 mL) was stirred for 1 h. Then, 2-chloroacetic acid (4) (1.1 eq) was added into the mixture and the reaction mixture was refluxed for 8 h. After monitoring the reaction with TLC, the solvent was removed under reduced pressure, washed with cold water, dried, and recrystallized in ethanol to furnish the desired compounds (5–7). 2-((1H-benzo[d]imidazol-2-yl)thio)acetic acid (5) [23] The above procedure was followed with 1 and 4 to yield 5 as a white powder (91% yield). The crystallization solvent is ethanol. $\mathbf{R}_{\mathbf{f}}$ (ethyl acetate/hexanes 2:1)=0.16; \mathbf{mp} =120– 123 °C; ¹H NMR (400 MHz, d₆-DMSO) δ 7.46–7.44 (m, 2H, Ar–H), 7.15–7.12 (m, 2H, Ar–H), 4.15 (s, 2H, –CH₂).

2-(*Benzo[d]oxazol-2-ylthio)acetic acid* (6) [23] The above procedure was followed with 2 and 4 to yield 6 as a pink crystal (75% yield). The crystallization solvent is ethanol. $\mathbf{R_f}$ (ethyl acetate/hexanes 2:1)=0.2; \mathbf{mp} =110–113 °C; ¹H NMR (400 MHz, d₆-DMSO) δ 7.66–7.63 (m, 2H, Ar–H), 7.36–7.32 (m, 2H, Ar–H), 4.22 (s, 2H, –CH₂).

2-(benzo[d]thiazol-2-ylthio)acetic acid (7) [23] The above procedure was followed with 3 and 4 to yield 7 as a white powder (83% yield). The crystallization solvent is ethanol. $\mathbf{R_f}$ (ethyl acetate/hexanes 2:1)=0.16; \mathbf{mp} =118–121 °C; ¹H NMR (400 MHz, d₆-DMSO) δ 8.03 (d, J=7.71 Hz, 1H, Ar–H), 7.87 (d, J=7.14 Hz, 1H, Ar–H), 7.51–7.41(m, 1H, Ar–H), 7.41–7.36 (m, 1H, Ar–H), 4.12 (s, 2H, –CH₂).

General Procedure for preparation of compounds (17-19) A mixture of *o*-phenylenediamine (8), 2-aminophenol (9), 2-mercaptoaniline (10) (1 eq), and 2-((benzazol-2-yl)thio) acetic acid (5-7) (1 eq) in PPA (5-7 g) was heated for 12 h in an oil bath at 150 °C. The reaction mixture was poured into ice water and neutralized by mixing with 5 M NaOH till slightly basic pH (8–9) to get the precipitate. The resulting precipitate was filtered off, washed with cold water, and recrystallized with ethanol–water.

2-(((1H-benzo[d]imidazol-2-yl)methyl)thio)-1H-benzo[d]imidazole (17) [57, 58] The above procedure was followed with 8 and 5 to yield 17 as a white powder (65% yield). The crystallization solvent is ethanol–water. \mathbf{R}_{f} (ethyl acetate/hexanes 1:1)=0.67; \mathbf{mp} =258–262 °C; \mathbf{IR} (KBr, cm⁻¹) Vmax 3053, 2931, 1504, 1407, 849, 741. The ¹H NMR and ¹³C NMR spectra are in agreement with the reported data. Anal. calcd. for $C_{15}H_{12}N_{4}S$: C, 64.26; H, 4.31; N, 19.98; S, 11.44. Found: C, 64.22; H, 4.19; N, 20.07; S, 11.58.

2-((Benzo[d]oxazol-2-ylmethyl)thio)benzo[d]oxazole (18) [57, 58] The above procedure was followed with 9 and 6 to yield 18 as a pink powder (56% yield). The crystallization solvent is ethanol–water. $\mathbf{R}_{\mathbf{f}}$ (ethyl acetate/hexanes 1:1)=0.63; \mathbf{mp} =146–150 °C; IR (KBr, cm⁻¹) Vmax 3093, 2995, 1501, 1445, 1238, 743; ¹H NMR (400 MHz, CDCl₃) δ 7.74–7.69 (m, 1H, Ar–H), 7.66–7.61 (m, 1H, Ar–H), 7.54–7.45 (m, 2H, Ar–H), 7.32 (m,,4H, Ar–H), 4.84 (s, 2H, –CH₂); ¹³C NMR (100 MHz, DMSO-d₆) δ 156.1, 131.0, 115.9, 115.5, 115.3, 33.0. Anal. calcd. for C₁₅H₁₀N₂O₂S: C, 63.81; H, 3.57; N, 9.92; S, 11.36. Found: C, 63.92; H, 3.53; N, 10.01; O, S, 11.30.

2-((Benzo[d]thiazol-2-ylmethyl)thio)benzo[d]thiazole (19) [**59**] The above procedure was followed with **10** and **7** to yield **19** as a green powder (55% yield). The crystallization solvent is ethanol–water. **R**_f (ethyl acetate/hexanes 1:1)=0.69; **mp**=202–206 °C; **IR** (KBr, cm⁻¹) Vmax 3153, 2878, 1656, 1462, 1212, 768; ¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, *J* = 8.16 Hz, 1H, Ar–H), 7.93 (d, *J* = 8.10 Hz, 1H, Ar–H), 7.82–7.75 (m, 2H, Ar–H), 7.50–7.29 (m, 4H, Ar–H), 5.05 (s, 2H, –CH₂); ¹³C NMR (100 MHz, DMSO-d₆) δ 163.4, 161.0, 155.5, 138.4, 135.9, 130.5, 125.2, 116.0, 115.9, 115.8, 114.0, 113.8, 33.5. **Anal. calcd.** for C₁₅H₁₀N₂S₃: C, 57.29; H, 3.21; N, 8.91; S, 30.59. Found: C, 57.18; H, 3.28; N, 8.87; S, 30.44.

Biochemistry

Cell culture Studies

All the synthesized compounds (17–37) were evaluated for their in vitro antiproliferative activity against four human cancer cell lines by comparing the results with the standard antiproliferative drug, methotrexate (MTX). In vitro antiproliferative screening of titled compounds against human hepatocellular cell line (HepG2), human renal cancer cell line (A498), human lung adenocarcinoma epithelial cell line (A549), and human cervical cancer cell line (HeLa) were performed using MTT assay [22]. The obtained results of in vitro antiproliferative activities are summarized in Table 1. The selectivity of these compounds toward cancerous cells is evaluated against Vero (African green monkey kidney epithelial cell) to determine the non-toxic concentrations of the compounds on relatively healthy cells. All cell lines were obtained from the cell culture collections of Mustafa Kemal University. Cell incubations were done at 37 °C in 5% (v/v) CO₂.

MTT cell viability assay

The cytotoxic activity of the compounds was determined by using MTT assay [20, 21]. The compounds were dissolved in dimethyl sulfoxide (DMSO) and then diluted to the appropriate concentrations in Dulbecco's modified Eagle's medium (DMEM). Concentration of DMSO was less than 1% in the culture medium. Subsequently, the cells were seeded at 104 cells/well in DMEM, supplemented with 10% fetal bovine serum (FBS), 100 units/mL penicillin G, and 100 µg/mL streptomycin in each well of 96-well microculture plates. The cells were cultured at 37 °C for 24, 48, 72, and 96 h in an incubator containing 5% CO₂.

After incubation, cells were treated with test compounds of appropriate concentrations for 24, 48, 72, and 96 h. After incubation period, 10 μ L MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) (5 mg/mL) was added to each well and the plates were further incubated for 4 h. Then, the medium of each well was carefully removed and formazan crystals were dissolved in 100 μ L of DMSO. Absorbance was determined at 570 nm for each well using a microplate reader (Bioteck) [60].

The cell viability was expressed as percentage of the viable cells in each sample with respect to the control wells. Three independent experiments in triplicates were done for the determination of the growth inhibition of each compound. The IC_{50} values were calculated from concentration–response curves using the SPSS (SPSS Inc., Chicago) software.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Spasov AA, Yozhitsa IN, Bugaeva LI (1999) Benzimidazole derivatives: spectrum of pharmacological activity and toxicological properties. Pharm Chem J 33:232–243. https://doi. org/10.1007/BF02510042
- Gupta M, Paul S, Gupta R (2009) General characteristics and applications of microwaves in organic synthesis. Acta Chim Slov 56:749–764
- Piscitelli F, Ballatore C, Smith A (2010) Solid phase synthesis of 2-aminobenzothiazoles. Bioorg Med Chem Lett 20:644–648. https ://doi.org/10.1016/j.bmcl.2009.11.055
- Balakumar C, Kishore DP, Rao KV, Narayana BL, Rajwinder K, Rajkumar V, Rao AR (2012) Design, microwave-assisted synthesis and in silico docking studies of new 4H-pyrimido[2,1-b] benzothiazole-2-arylamino-3-cyano-4-ones as possible adenosine A2B receptor antagonists. Indian J Chem 51B:1105–1113
- Patil A, Ganguly S, Surana S (2008) A systematic review of benzimidazole derivatives as an antiulcer agent. RJC 1:447–460
- Narasimhan B, Sharma D, Kumar P (2012) Benzimidazole: a medicinally important heterocyclic moiety. Med Chem Res 21:269–283. https://doi.org/10.1007/s00044-010-9533-9
- Kohli P, Srivastava SD, Srivastava SK (2007) Synthesis and biological activity of mercaptobenzoxazole based thiazolidinones and their arylidenes. J Chin Chem Soc 54:1003–1010. https:// doi.org/10.1002/jccs.200700144
- Sivakumar R, Pradeepchandran R, Jayaveera KN, Kumarnallasivan P, Vijaianand PR, Venkatnarayanan R (2011) Benzimidazole: an attractive pharmacophore in medicinal chemistry. Int J Pharm Sci Res 3:19–31
- 9. Deb PK, Kaur R, Chandrasekaran B, Bala M, Gill D, Kaki VR, Akkinepalli RR, Mailavaram R (2014) Synthesis,

anti-inflammatory evaluation, and docking studies of some new thiazole derivatives. Med Chem Res 23:2780–2792. https://doi.org/10.1007/s00044-013-0861-4

- Palmer FJ, Trigg RB, Warrington JV (1971) Benzothiazolines as antituberculous agents. J Med Chem 14:248–251. https://doi. org/10.1021/jm00285a022
- Hadden MK, Blagg BS (2008) Dimeric approaches to anti-cancer chemotherapeutics. Anticancer Agents Med Chem 8:807– 816. https://doi.org/10.2174/187152008785914743
- Ueki M, Ueno K, Miyadoh S, Abe K, Shibata K, Taniguchi M, Oi S (1993) A novel cytotoxic metabolite from Streptomyces sp. 517-02. J Antibiot 46:1089–1094. https://doi.org/10.7164/ antibiotics.46.1089
- Sato S, Kajiura T, Noguchi M, Takehana K, Kobayashi T, Tsuji T (2001) A new cytotoxic benzoxazole derivative produced by Streptomyces sp. J Antibiot 54:102–104. https://doi. org/10.7164/antibiotics.54.102
- Jenkins TC (2000) Targeting multi-stranded DNA structures. Curr Med Chem 7:99–115. https://doi.org/10.2174/0929867003 375551
- 15. Singh MP, Joseph T, Kumar S, Bathini Y, Lown JW (1992) Synthesis and sequence-specific DNA binding of a topoisomerase inhibitory analog of Hoechst 33258 designed for altered base and sequence recognition. Chem Res Toxicol 5:597–607. https ://doi.org/10.1021/tx00029a003
- Racane L, Kraljevic PS, Ratkaj I, Stepanic V, Pavelic K, Tralic-Kulenovic V, Karminski-Zamola G (2012) Synthesis and antiproliferative evaluation of some new amidino-substituted *bis*-benzothiazolyl-pyridines and pyrazine. Eur J Med Chem 55:108–116. https://doi.org/10.1016/j.ejmech.2012.07.005
- Kumbhare RM, Dadmal T, Kosurkar U, Sridhar V, Rao JV (2012) Synthesis and cytotoxic evaluation of thiourea and N-bisbenzothiazole derivatives: a novel class of cytotoxic agents. Bioorg Med Chem Lett 22:453–455. https://doi.org/10.1016/j. bmcl.2011.10.106
- Gravatt GL, Baguley BC, Wilson WR, Denny WA (1994) DNAdirected alkylating agents. 6. Synthesis and antitumor activity of DNA minor groove-targeted aniline mustard analogs of pibenzimol (Hoechst 33258). J Med Chem 37:4338–4345. https://doi. org/10.1021/jm00051a010
- Shi Z, Zhao D, Huang Y, Du Y, Cao X, Gong Z, Zhao R, Li J (2012) Discovery, synthesis, and evaluation of small-molecule signal transducer and activator of transcription 3 inhibitors. Chem Pharm Bull 60:1574–1580. https://doi.org/10.1248/cpb. c12-00745
- McGahon AJ, Martin SJ, Bissonnette RP, Mahboubi A, Shi Y, Mogil RJ, Nishioka WK, Green DR (1995) The end of the (cell) line: methods for the study of apoptosis in vitro. Methods in Cell Biology: Cell Death 46:153–185. https://doi.org/10.1016/s0091 -679x(08)61929-9
- Mosmann T (1983) Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. J Immunol Methods 65:55–63. https://doi.org/10.1016/0022-1759(83)90303-4
- van Meerloo J, Kaspers GJL, Cloos J (2011) Cell sensitivity assays: the MTT assay. Methods Mol Biol 731:237–245. https:// doi.org/10.1007/978-1-61779-080-5_20
- 23. Lan P, Romero FA, Wodka D, Kassick AJ, Dang Q, Gibson T, Cashion D, Zhou G, Chen Y, Zhang X, Zhang A, Li Y, Trujillo ME, Shao Q, Wu M, Xu S, He H, MacKenna D, Staunton J, Chapman KT, Weber A, Sebhat IK, Makara GM (2017) Hit-to-lead optimization and discovery of 5-((5-([1, 1'-Biphenyl]-4-yl)-6chloro-1 H-benzo [d] imidazol-2-yl) oxy)-2-methylbenzoic Acid (MK-3903): A novel class of benzimidazole-based activators of AMP-activated protein kinase. J Med Chem 60:9040–9052. https ://doi.org/10.1021/acs.jmedchem.7b01344

- Elagab HA, Alt HG (2015) Structure–property-relationship studies with ethylene polymerization catalysts of Ti, Zr and V containing heterocyclic ligands. Inorgan Chim Acta 437:26–35. https:// doi.org/10.1016/j.ica.2015.08.002
- Qian J, Zhang Y, Yin X (2012) The synthesis and study of bis(1octylbenzimidazoi-2-yl)alkane oil-soluble corrosion inhibitor. Huaxue Tongbao 75:88–91
- 26. Heinrich J, Koenig NF, Sobottka S, Sarkar B, Kulak N (2019) Flexible vs. rigid bis(2-benzimidazolyl) ligands in Cu(II) complexes: impact on redox chemistry and oxidative DNA cleavage activity. J Inorg Biochem 194:223–232. https://doi. org/10.1016/j.jinorgbio.2019.01.016
- Yilmaz U, Kucukbay H (2016) Synthesis and characterization of novel phosphoramidates containing benzimidazole moiety. Phosphorus, Sulfur Silicon Relat Elem 191:140–143. https:// doi.org/10.1080/10426507.2015.1067209
- Inamdar SM, More VK, Mandal SK (2013) CuO nano-particles supported on silica, a new catalyst for facile synthesis of benzimidazoles, benzothiazoles and benzoxazoles. Tetrahedron Lett 54:579–583. https://doi.org/10.1016/j.tetlet.2012.11.091
- 29. Berends HP, Stephan DW (1984) Copper (I) and copper (II) complexes of biologically relevant tridentate ligands. Inorgan Chim Acta 93:173–178. https://doi.org/10.1016/S0020 -1693(00)88159-1
- Mao S, Shen K, Shi X, Wu H, Han X, Li C, Huang G (2018) Synthesis, crystal structure and biological activity of two binuclear Ag(I) complexes with bis-benzimidazole thioether ligands. Inorgan Chim Acta 471:82–90. https://doi.org/10.1016/j. ica.2017.10.038
- 31. Rao SS, Reddy CV, Dubey PK (2014) Synthesis of *N*,*N*-disubstitutedbisbenzimidazolesulphides of Potential Pharmacological Interest. JCHPS 6:1199–1204
- 32. Xu Y, Wu H, Zhang H, Aderinto SO, Yang Z (2016) Synthesis, crystal structures, and DNA-binding studies of two silver (I) complexes with 1, 3-bis(1-ethylbenzimidazol-2-yl)-2-thiapropane. J Coord Chem 69:2988–2998. https://doi. org/10.1080/00958972.2016.1218484
- Mao S, Shen K, Shi X, Xu Y, Wu H (2017) Two silver (I) complexes with bis (benzimidazole)-2-oxopropane ligands: syntheses, crystal structures and DNA binding studies. APPI 31:e3747. https://doi.org/10.1002/aoc.3747
- 34. Wu HL, Yun RR, Wang KT, Li K, Huang XC, Sun T (2010) Synthesis, crystal structure, and spectrum properties of Cobalt (II) complexes based on tridentate 1, 3-bis(benzimidazol-2-yl)-2-oxopropane ligand and derivative. Z Anorg Allg Chem 636:629–633. https://doi.org/10.1002/zaac.200900298
- 35. Feng R, Hou Y, Wu Z, Yang Y, Nie F (2015) Structures and fluorescent properties of Cadmium (II) complexes with 1D and 2D Structures based on tridentate benzimidazole ligands. Z Anorg Allg Chem 641:1918–1925. https://doi.org/10.1002/zaac.20140 0511
- Kopel P, Wawrzak D, Langer V, Cihalova K, Chudobova D, Vesely R, Adam V, Kizek R (2015) Biological activity and molecular structures of bis(benzimidazole) and trithiocyanurate complexes. Molecules 20:10360–10376. https://doi.org/10.3390/molecules200610360
- Chen B, Morlanes N, Adogla E, Takanabe K, Rodionov VO (2016) An efficient and stable hydrophobic molecular cobalt catalyst for water electro-oxidation at neutral pH. ACS Catal 6:4647–4652. https://doi.org/10.1021/acscatal.6b01237
- Dauer D, Stalke D (2014) Heterocyclic substituted methanides as promising alternatives to the ubiquitous nacnac ligand. Dalton Trans 43:14432–14439. https://doi.org/10.1039/C4DT0 1008F
- Kretsch J, Kreyenschmidt A, Herbst-Irmer R, Stalke D (2018) Alkali metal complexes based on bisheterocyclomethanide

ligands. Dalton Trans 47:12606–12612. https://doi.org/10.1039/ C8DT01678J

- Kumar R, Selvam C, Kaur G, Chakraborti AK (2005) Microwave-assisted direct synthesis of 2-substituted benzoxazoles from carboxylic acids under catalyst and solvent-free conditions. Synlett 9:1401–1404. https://doi.org/10.1055/s-2005-868509
- Shirakawa Y, Masuda T (2013) Coupling agents for rubber/carbon black and rubber compositions containing them. PCT Int Appl WO 2013015425
- 42. Cakir B, Ucucu U, Buyukbingol E, Abbasoglu U (1989) Benzoxazoles: bis-benzoxazole derivatives, synthesis, antifungal activities and QSARs. J Faculty Pharm Gazi Uni 6:15–21
- Terao H, Ono Y, Ito Y, Isogai M, Hamada T, Imanishi, Tsunoda A (1991) Organic nonlinear optical device. Japanese Kokai Tokkyo Koho JP 03188425
- 44. Hao Y, Chen Y (2016) Excited-state intramolecular single and double proton transfer emission of 2,5-bis(benzoxazol-2-yl) thiophene-3,4-diol. Dyes Pigme 129:186–190. https://doi. org/10.1016/j.dyepig.2016.03.002
- Hao Y, Zheng M, Chen Y (2014) A highly stable and watersoluble fluorescent dye for fluorescence imaging of living cells. J Mater Chem B 2:7369–7374. https://doi.org/10.1039/C4TB0 1210K
- Zhou H, Wang L, Yin B (2004) Synthesis of bis(2-benzoxazolylmethyl) ether by dry reaction under microwave irradiation. Huaxue Shiji 26:308–311
- Alt H, Elagab H, Al-Humydi A (2011) Ethylene Polymerisation. PCT Int Appl WO 2011088990
- Chakraborti AK, Selvam C, Kaur G, Bhagat S (2004) An efficient synthesis of benzothiazoles by direct condensation of carboxylic acids with 2-aminothiophenol under microwave irradiation. Synlett 5:851–855. https://doi.org/10.1055/s-2004-820012
- Katritzky AR, Liang D, Fan W (1988) Bridged cyanine dyes. Part 2 [1].1-(*N*-methyl-2-benzothiazolylinio)-3-(*N*-methyl-2-benzothiazolylene) and 1-(*N*-methyl-4-pyridinio)-3-(*N*-methyl-4-pyridylene)cyclopenta-1,4-dienes with fused rings. J Heterocycl Chem 25:1315–1319. https://doi.org/10.1002/jhet.5570250509
- Rai C, Braunwarth JB (1961) Synthesis of bisbenzothiazoles1. J Org Chem 26:3434–3436. https://doi.org/10.1021/jo01067a100
- Strasser CE, Jongh LAD, Raubenheimer HG, Cronje S (2011) 2,2'-(Sulfanediyldimethylene)bis(1,3-benzothiazole). Acta Cryst E 67:o622. https://doi.org/10.1107/S1600536811004478
- Sih JC, Graber DR (1983) 2-Mercapto-1,3-benzoxazole: a useful reagent for the preparation of symmetrical and unsymmetrical sulphides. J Org Chem 48:3842–3845. https://doi.org/10.1021/ jo00169a058
- Das SK, Mathur P (1999) Copper (II) complexes with bis thiazole based ligands: spectral, cyclic voltammetric and EPR studies. Indian J Chem A 38A:1277–1282
- 54. Ushenko IK (1952) α,γ-Epoxythiacarbocyanines. I Zh Obshch Khim 22:711–715
- Finn MG, Rodionov VO (2009) Ligands for copper-catalyzed azide-alkyne cycloaddition reactions. PCT Int Appl WO 2009038685
- Buehrdel G, Beckert R, Herzigova P, Petrlikova E, Schuch D, Birckner E, Goerls H (2009) A new synthesis of push-pull pyrroles, their oxidation to stable 3H-pyrroles and an unexpected anellation reaction. Eur J Org Chem 20:3404–3412. https://doi. org/10.1002/ejoc.200900295
- Akpa SJ, Say MV, Zoakouma SPR, Fante B, Sissouma D, Adjou A (2016) Synthesis of 2-(benzylthio) benzimidazole, 2-[(benzimidazol-2-yl)methylthio]benzimidazole and structural analogues against Haemoncus contortus. AFR J Pharm Pharmacol 10:670– 680. https://doi.org/10.5897/AJPP2016.4557
- 58. Bouchouit M, Said M, Kara M, Bouacida S, Merazig H, Kacem-Chaouche N, Chibani A, Zouchoune B, Belfaitah A, Bouraiou

A (2016) Synthesis, X-ray structure, theoretical investigation, corrosion inhibition and antimicrobial activity of benzimidazole thioether and theirs metal complexes. Polyhedron 119:248–259. https://doi.org/10.1016/j.poly.2016.08.045

- 59. Zubarovskii VM (1951) 2-(Hydroxymethyl)benzothiazole and its transformations. Zh Obshch Khim 21:2055–2064
- 60. Sabina XJ, Karthikeyan J, Velmurugan G, Tamizh MM, Shettyd AN (2017) Design and in vitro biological evaluation of substituted

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chalcones synthesized from nitrogen mustards as potent microtubule targeted anticancer agents. New J Chem 41:4096–4109. https://doi.org/10.1039/C7NJ00265C

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