Eur J Med Chem (1995) 30, 85–94 © Elsevier, Paris

Synthesis and activity of 2-methyl-3-aminopropiophenones as centrally acting muscle relaxants*

A Shiozawa, K Narita, G Izumi, S Kurashige, K Sakitama, M Ishikawa

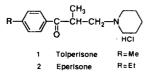
Research Laboratories. Pharmaceuticals Group, Nippon Kayaku Co Ltd, 31-12, Shimo 3-chome, Kita-ku, Tokyo 115, Japan

(Received 20 June 1994; accepted 3 October 1994)

Summary — Some novel 2-methyl-3-aminopropiophenones were synthesized and their centrally acting muscle relaxant activities were evaluated for an inhibitory effect on the flexor reflex in rats. The structure-activity relationships are discussed. In this series, 2-methyl-3-pyrrolidino-1-(4-trifluoromethylphenyl)-propan-1-one (**28**) showed significant centrally acting muscle relaxant activity. In addition, the activities of each enantiomer (**28**-(S) and (R)) were studied along with their acute toxicities. Compound **28**-(R) was found to exhibit more potent activity and weaker acute toxicity than **28**-(S). Accordingly, compound **28**-(R) (NK433) is under development as a novel centrally acting muscle relaxant.

centrally acting muscle relaxant / 2-methyl-3-aminopropiophenone / flexor reflex / optical resolution

Centrally acting muscle relaxants with few side effects have been an attractive target for drug research in recent years. Tolperisone 1 [1] and eperisone 2 [2] are a class of potent centrally acting muscle relaxants structurally characterized by a 2-methyl-3-aminopropiophenone moiety. They are of clinical importance in the treatment of gait or posture disturbance, hypertonea, cervicodynia, tremor caused by spasticity, and are also useful for lower back pain and shoulder stiffness.



Despite showing lower potency than agents of other classes, such as baclofen [3], diazepam [4], and tizanidine [5], involved in centrally acting muscle relaxants, drugs such as 1 and 2 have been clinically preferred and widely used because of a lack of severe adverse effects.

As part of a program directed to the search for new highly potent centrally acting muscle relaxants with fewer side effects, our interest was focused on 2methyl-3-aminopropiophenones. In the present study, we describe the synthesis of 2-methyl-3-aminopropiophenones (33-35) and the results obtained in the evaluation of their centrally acting muscle relaxant activities.

On the other hand, although the 2-methyl-3-aminopropiophenone structure has an asymmetric carbon at the 2-position of the propanone, drugs **1** and **2** are racemates. We were interested in the different biological activities of each enantiomer because there are few reports [5] dealing with centrally acting muscle relaxant activities of 2-methyl-3-aminopropiophenone enantiomers. We report the activities of each enantiomer of the most interesting compound in this series.

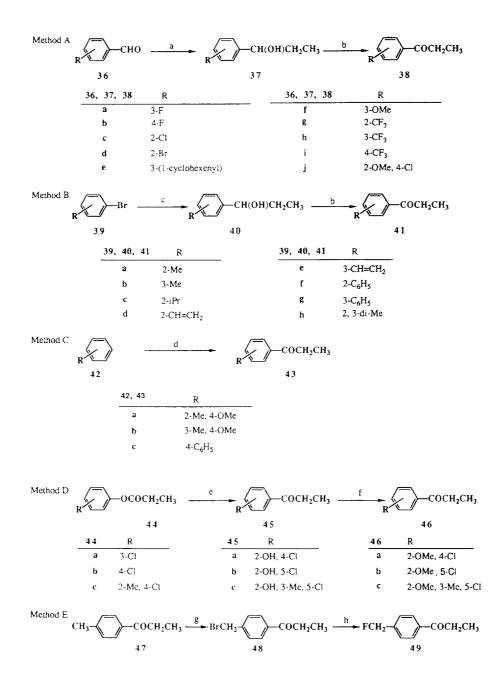
Chemistry

The 2-methyl-3-aminopropiophenones (3-35) were synthesized by the Mannich reaction [6] of the appropriate propiophenones with paraformaldehyde and pyrrolidine HCl in the presence of a small amount of hydrochloric acid (scheme 1).



Scheme 1. Reagents: a) paraformaldehyde, pyrrolidine-HCl.

^{*}Preliminary reports of our work were presented at the XIIth International Symposium on Medicinal Chemistry, Basel, Switzerland, September, 1992; Abstracts P-079 C.



Scheme 2. Reagents: a) EtMgBr; b) CrO₃, H₂SO₄; c) Mg, EtCHO; d) EtCOCl; e) AlCl₃; f) MeONa, MeI; g) NBS; h) KF.

The Grignard reaction [7] of appropriate benzaldehydes (**36a**-**j**) with ethylmagnesium bromide gave the corresponding alcohols (**37a**-**j**), followed by oxidation with chromium trioxide [8] in dilute sulfuric acid to afford propiophenones **38a**-**j** in 35–88% yields from **36a**-**j**, respectively (scheme 2, *Method A*).

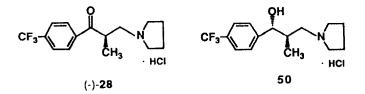
Propiophenones 41a-h were prepared in 20-81% yields from 39a-h, respectively (Method B), by the Grignard reaction of propionyl aldehyde with the phenylmagnesium bromide obtained from the appropriate phenyl bromide with magnesium, followed by oxidation with chromium trioxide. The Friedel-Crafts reaction [9] of appropriately substituted benzenes (42a-c) with propionyl chloride in the presence of aluminum chloride gave propiophenones **43a-c** in 20-81% yields, respectively (*Method C*). The Fries rearrangement [10] of propionates 44a-c in the presence of aluminum chloride at 130°C gave the corresponding phenols (45a-c). The sodium salts of the phenols were then methylated with methyl iodide to give the required propiophenones (46a-c) in 54-88% yields (Method D). The bromination of 4methylpropiophenone 47 with N-bromosuccinimide and perbenzoic acid gave 4-bromomethylpropiophenone 48, which was converted into 4-fluoromethylpropiophenone 49 in a 15.5% yield from 47 (Method E).

Optical resolution of 28 into its enantiomers ((-) and (+)) was achieved by fractional crystallization of the salts prepared from the free base of 28 with (L)and (D)-N-acetylphenyl glycine, respectively. The absolute configuration of (-)-28 was determined on the basis of X-ray crystallographic analysis of its alcohol derivative (50) by Iidaka *et al* (unpublished results) to show unambiguously that (-)-28 has an *R* configuration at C-2.

Pharmacological results and discussion

The centrally acting muscle relaxant activities 2methyl-3-aminopropiophenones were evaluated for their inhibitory effects on the flexor reflex [11] in rats.

The activities were expressed as inhibitory percentages (%) and compared with the activities of tolperisone 1 and eperisone 2. Table I shows the results obtained at a dose of 5 mg/kg intravenously administered in rats. The replacement of a piperidino group on the amine moiety of 1 or 2 with a pyrrolidino group (12) showed a more potent activity than 1. Thus, a series of derivatives fixed with a pyrrolidino group as an amine moiety of 2-methyl-3-aminopropiophenones was examined. Table I indicates that the type and position of the substituents on the phenyl ring of the 2-methyl-3-pyrrolidinopropiophenones remarkably contributes to the activity. Introduction of a methyl group (10, 11, and 12) into the phenyl ring



significantly enhanced the activities compared with the unsubstituted compound **3**, and the 2-methyl compound **10** was more potent than the 3- (**11**) and 4isomers (**12**). The activities decreased in the order **10** > **11** > **12**. However, the replacement of the 2-methyl group (**10**) with a 2-ethyl group (**13**), a 2-isopropyl group (**14**), a 2-vinyl group (**15**), or a 2-phenyl group (**18**) decreased the activity in the order 2-methyl (**10**) > 2-vinyl (**15**) > 2-ethyl (**13**) > 2-phenyl (**18**).

These results suggest that an increase in the bulkiness of the *ortho* substituents might lead to a decrease in the activity. Similarly, when the 3-methyl group (11) was replaced with a 3-vinyl group (16), a 3-(1-cyclohexenyl) group (17), or a 3-phenyl group (19), the activities decreased in the order 3-methyl (11) > 3-vinyl (16) > 3-(1-cyclohexenyl) (17) > 3-phenyl (19). The activity of the 4-phenyl compound 20 was also lower than that of the 4-methyl compound 12.

In the series of methoxy derivatives, the 2- (22), 3-(23), and 4-methoxy (24) analogs were more potent than the unsubstituted compound 3, but they were also less potent than the corresponding 2- (10), 3- (11), and 4-methyl (12) derivatives, respectively. The 2-hydroxy compound 21 had slightly enhanced activity compared with 22.

When the halogenated analogs (4-9 and 25-28) were examined, the 3-fluoro compound 5 was more potent than the 2- (4) and 4-isomers (6), and 5 exhibited an approximately five times more potent activity than 3. The activity of the 3-bromo compound 9 was slightly more potent than that of the 2-isomer 8. The activity of the 2-chloro compound 7 was approximately comparable to those of the 2-fluoro (4) and 2bromo (8) analogs. In addition, the trifluoromethyl analogs (26-28) were examined. The 4-trifluoromethyl compound 28 was considerably more potent in comparison to the 2- (26) and 3- (27) congeners and the activity decreased in the order 28 > 26 > 27. The replacement of the 4-trifluoromethyl group (28) by a 4-fluoromethyl group (25) led to a dramatic decrease in the activity.

When disubstituted compounds were examined, compound 34 (2,3-diMe) exhibited a more potent activity than 10 (2-Me) or 11 (3-Me). Compound 34 was found to have the most potent activity in this study. Introduction of a 4-methoxy group into the 2- (10) and 3-methyl (11) compounds decreased the activities as

Table I. The physical properties of 3-pyrrolidinopropiophenones and their centrally acting muscle relaxant activities.

сн ₃	12-N
н — О	HCI
3-35	

Comp	od R	Yield	•	Recrystn	Formula	FR ^a	LD _{so} °
No.		(%)	(°C)	Solvent		I(%)	(mg/kg)
1	Tolperisone					31.3	
2	Eperisone					34.5	
3	н	80	148-149	CH ₂ Cl ₂ -CH ₃ COCH ₃	$C_{14}H_{19}NO \cdot HCl$	7.7	
4	2-F	33	121-122	CH ₃ COCH ₃	$C_{14}H_{18}FNO \cdot HCl$	23.2	
5	3-F	65	152-153.5	CH3COCH3	C ₁₄ H ₁₈ FNO · HCl		41.2
6	4-F	23	148-149	Et ₂ O-CH ₃ COCH ₃	C ₁₄ H ₁₈ FNO · HCl	25.8	
7	2-Cl	62	163-164	CH ₂ Cl ₂ -AcOEt	$C_{14}H_{18}CINO \cdot HCI$	24.1	
8	2-Br	14	134-136	MeOH-CH ₃ COCH ₃	C14H18BrNO · HCl	22.8	
9	3-Br	80	152.5-154	MeOH-CH ₃ COCH ₃	$C_{14}H_{18}BrNO \cdot HCl$	29.9	
10	2-Me	73	144-145.5	MeOH-CH ₃ COCH ₃	$C_{15}H_{21}NO \cdot HCI$	51.3	30.6
11	3-Me	98	166-168	MeOH-CH ₃ COCH ₃	$C_{15}H_{21}NO \cdot HCl$	44.6	25.9
12	4-Me	90	168-169	MeOH-CH ₃ COCH ₃	$C_{15}H_{21}NO \cdot HCI$	37.2	
13	2-Et	75	138-139	MeOH-CH ₃ COCH ₃	C ₁₆ H ₂₃ NO · HCI	33.7	
14	2-іРт	58	116-118	CH ₃ COCH ₃	C ₁₇ H ₂₅ NO · HCl	25.3	
15	2-CH=CH ₂	73	143-144	MeOH-CH ₃ COCH ₃	$C_{16}H_{21}NO \cdot HCI$	38.9	
16	3-CH=CH ₂	68	151-153	MeOH-CH3COCH3	$C_{16}H_{21}NO \cdot HCl$	34.7	
17	3-(1-cyclohexenyl)	51	142-144	CH ₃ COCH ₃	C ₂₀ H ₂₇ NO · HCl	30.1	
18	2-Ph	51	138-139	MeOH-CH,COCH,	C ₂₀ H ₂₃ NO · HCl	9.0	
19	3-Ph	48	157.5-158.5	MeOH-CH ₃ COCH ₃	C ₂₀ H ₂₃ NO · HCl	28.0	
20	4-Ph	61	156-158.5	MeOH-CH ₃ COCH ₃	C ₂₀ H ₂₃ NO · HCl	7.1	
21 ·	2-OH	91	104-106	CH ₃ COCH ₃	C14H19NO2 · HCl	26.3	
22	2-OMe	89	138-140	CH ₂ Cl ₂ -CH ₁ COCH ₁	$C_{15}H_{21}NO_2 \cdot HCI$	20.4	
23	3-OMe	87	130-132	CH ₃ COCH ₃	C ₁₅ H ₂₁ NO ₂ ·HCl	10.8	
24	4-OMe	63	154-156	CH ₃ COCH ₃	$C_{15}H_{21}NO_2 \cdot HCl$	22.8	
25	4-CH ₂ F	74	151-152	CH ₂ Cl ₂ -AcOEt	C ₁₅ H ₂₀ FNO · HCl	15.5	
26	2-CF ₃	19	148-149	MeOH-CH,COCH,	C ₁₅ H ₁₈ F ₃ NO · HCl		
27	3-CF	33	144-145	CH ₁ COCH ₁ -ELO	C ₁₅ H ₁₈ F ₃ NO · HCl		
28	4-CF	94	154-156	MeOH-CH,COCH,	C ₁₅ H ₁₈ F ₃ NO · HCl		61.1
29	2-Me,4-OMe	52	142-143	CH ₁ COCH ₁ -AcOEt	C ₁₆ H ₂₃ NO ₂ · HCl	25.5	
30	3-Me,4-OMe	69	163-165	AcOEt	$C_{16}H_{23}NO_2 \cdot HCI$	29.4	
31	2-OMe,4-Cl	64	140-141.5	CH ₃ COCH ₃ -AcOEt	$C_{15}H_{20}CINO_2$ HCl		
32	2-OMe,5-Cl	65	166-167	CH ₃ COCH ₃	$C_{15}H_{20}CINO_2 \cdot HCI$		
33	2-OMe,3-Cl	59	125-127	CH ₁ COCH ₁	$C_{15}H_{20}CINO_2 \cdot HCI$		
34	2,3-di-Me	52	149-151	McOH-CH ₃ COCH ₃	$C_{16}H_{23}NO \cdot HCl$	53.9	33.9
35	2-OMe,3-Me,5-Cl	64	149-150	CH ₃ COCH ₃	$C_{16}H_{22}CINO_2 \cdot HCI$		2217

^aFR: flexor reflex. The compounds were intravenously administered at 5 mg/kg dose to rats (see *Experimental protocols*). ^bThe values of LD_{50} were determined by the up-and-down method (*iv*, mice) (see *Experimental protocols*).

Table II. The inhibitory effects of 1, 2, 5, 28 and 34 on anemic decerebrate rigidity and the flexor reflex, and their acute toxicities.

Compound	Rigidity (iv) ^a I (%)	FR (po) ^b I (%)	LD ₅₀ (po) ^c mg/kg
1	15.4	35.3	320
2	24.2	36.7	326
5	16.0		
28	23.6	72.9	800
34	53.9	58.1	280

^aRigidity means anemic decerebrate rigidity [13] and each compound was administered at 3.5 mg/kg dose to rats. ^bEach compound was orally administered at a dose of 50 mg/kg. ^cThe LD₅₀ value of each compound was determined by Van der Waerden's method in mice.

shown by a comparison of 29 (2-Me, 4-OMe) and 30 (3-Me, 4-OMe) with 10 and 11, respectively. On the other hand, the introduction of a 3- or 5-chloro group into 2-methoxy compound 22 slightly increased the activity as shown by a comparison of 32 (2-OMe, 5-Cl) and 33 (2-OMe, 3-Cl) with 22 (2-OMe), but compound 31 (2-OMe, 4-Cl) with a 4-chloro group was less potent than 22. Furthermore, when a 3-

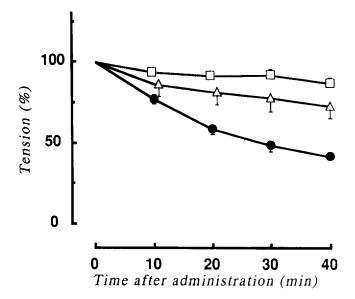


Fig 1. Effects of optical isomers and racemate (100 mg/kg, po) on anemic decerebrate rigidity in rats. The ordinate represents the mean amplitudes of the flexor reflex, as percentages of the value just prior to drug administration, with SEM indicated. •: 28-(R)(NK433); \Box : 28-(S); \triangle : racemate.

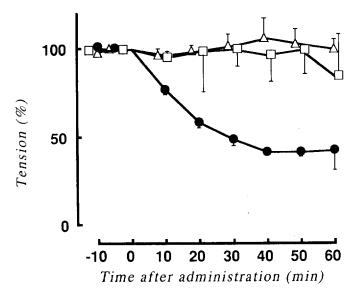


Fig 2. Effects of 1, 2, and NK433 (100 mg/kg, po) on anemic decerebrate rigidity in rats. The ordinate represents the mean amplitudes of the flexor reflex, as percentages of the value just prior to drug, administration, with SEM indicated. •: NK433; \Box : 1 tolperisone; \triangle : 2 eperisone.

methyl group was introduced into compound **32**, compound **35** (2-OMe, 3-Me, 5-Cl) showed less potent activity.

Among the compounds described above, compounds 5, 28 and 34, which show the most potent activities and the lowest toxicity, were selected for further evaluation of centrally acting muscle relaxant activity. Thus, the inhibitory effects of the intravenously administered compounds on anemic decerebrate rigidity [12] and that of the orally administered compounds on the flexor reflex in rats were evaluated. Table II shows the results and LD_{50} (mg/kg) values. Compound 28 was selected on the basis of these pharmacological activities and safety evaluations.

As the compound **28** has an asymmetric carbon in the molecule, we were interested in the pharmacological action of each of enantiomer (**28**-(S) and -(R)). The activity of each of enantiomer (**28**-(S) and -(R)) was studied in relation to its acute toxicity. These results are shown in figure 1 and 2. Compound **28**-(R) exhibited a more potent activity and weaker acute toxicity than **28**-(S). Accordingly, compound **28**-(R) (NK433) has been selected for development as a centrally acting muscle relaxant.

Experimental protocols

All melting points were determined on a Yanaco micro melting point apparatus and are uncorrected. The structures of all compounds were supported by the infrared (IR), proton magnetic resonance (¹H-NMR), and mass spectra (MS). IR spectra were measured on a JASCO IR-G spectrometer. ¹H-NMR spectra were recorded on a JEOL JNM-PMX60 spectrometer using Me₄Si as an internal standard. MS were determined with a Shimadzu GC MS-7000 Spectrometer (electron impact ionization mode). Elemental analysis (C, H, N and halogen) were in agreement with calculated values (within $\pm 0.4\%$).

Chemistry

General procedures for the preparation of the propiophenones Method A. A typical example: 4-trifluoromethylpropiophenone 38i [13]. A solution of ethyl bromide (37.6 g, 0.345 mol) in dry Et₂O (50 ml) was dropped into a suspension of Mg (10.1 g. 0.414 mol in dry Et₂O (50 ml) under stirring to give a Grignard solution. To the Grignard solution, a solution of 4-trifluoromethylbenzaldehyde (20.0 g, 0.115 mol) in Et₂O (150 ml) was added, and the mixture was then stirred for 2 h at room temperature. Water was added to the mixture to decompose the Grignard reagent and the mixture was extracted with AcOEt three times. The organic layer was dried over MgSO₄ and evaporated in vacuo to give a residual oil. The oil was purified by SiO₂ column chromatography using CH₂Cl₂/*n*-hexane (50:50) as an eluent to afford **37i** (22.0 g, 93.7%) as an oil. A solution of 37i (22 g, 0.108 mol) in CH₃COCH₃ (70 ml) was added to a mixture of chromium trioxide [8] (22.0 g, 0.108 mol) and sulfuric acid (18.5 ml) in H₂O (205 ml) under ice cooling. The mixture was stirred for 2 h and then evaporated in vacuo to remove CH₃COCH₃. The mixture was then extracted with AcOEt and the AcOEt layer was dried over MgSO₄ and evaporated in vacuo to give a residual oil. The oil was distilled to give 38i (11.6 g, 53.1%) as an oil, bp 76°C (2 mmHg). IR v (CHCl₃) cm⁻¹: 1690 (C=O). ¹H-NMR δ (CDCl₃): 1.2 (t, 3H; J = 7.0 Hz), 3.0 (q, 2H; J = 7.0 Hz), 7.7 (dd, 2H; J = 8.0 Hz), 8.0 (dd, 2H; J = 8.0 Hz).

Compounds **38a-h** and **j** were prepared by a similar method from appropriate benzaldehydes **36a-h** and **j**, respectively: **38a**: oil, bp 89°C (8 mmHg) [14]; **38b**: oil, bp 105–107°C (22 mmHg) [15]; **38c**: oil, bp 82–87°C (1.5 mmHg) [16]; **38d**: oil, bp 116–118°C (10–11 mmHg) [14]; **38e**: oil (the product was purified by chromatography on silica gel with CH₂Cl₂/*n*hexane (50:50) as eluent); **38f**: oil, bp 129°C (14 mmHg) [17]; **38g**: oil, bp 55–58°C (0.5 mmHg) [18]; **38h**: oil, bp 100–102°C (16–18 mmHg) [19]; **38j**: oil, bp 170°C (50 mmHg) [20].

Method B. A typical example: 2-methylpropiophenone 41a A solution of 2-tolylbromide (17.1 g, 0.10 mol) in dry [21]. THF (30 ml) was added to a suspension of Mg (2.92 g, 0.12 mol) in dry THF (15 ml). The mixture was stirred at room temperature for 2 h and then refluxed for 0.5 h to give a 2-tolylmagnesium bromide solution. To this Grignard solution, a solution of propionaldehyde (6.97 g, 0.12 mol) in dry THF (5 ml) was added under ice cooling, and the mixture was stirred for 1 h under ice cooling. The mixture was then poured into a saturated NH₄Cl aqueous solution (100 ml) and extracted with Et₂O. The Et₂O layer was washed with H₂O and dried over MgSO₄. The Et₂O was then evaporated in vacuo to give a residual oil. The oil was distilled to give 40a (12.3, 81.8%) as an oil, bp 107°C (15 mmHg). A solution of 40a (10.5 g, 70 mmol) in CH₃COCH₃ (47 ml) was added dropwise under ice cooling into a mixture of chromium trioxide (7.70 g, 77 mmol) in sulfuric acid (10.7 ml) and H₂O (95.0 ml) over 0.5 h. The mixture was stirred for 2 h, and then evaporated in vacuo to remove CH₃COCH₃. The mixture was extracted with AcOEt. The organic layer was washed with H₂O three times, dried over MgSO₄, and then evaporated *in vacuo* to give a residual oil. The oil was distilled to give **41a** (8.22 g, 79.2%) as an oil, bp 70–72°C (3 mmHg). IR v (CHCl₃) cm⁻¹: 1685 (C=O). ¹H-NMR δ (CDCl₃) 1.18 (t, 3H, *J* = 7.0 Hz), 2.46 (s, 3H), 2.90 (q, 2H, *J* = 7.0 Hz), 7.1–7.4 (m, 3H), 7.4–7.7 (m, 1H).

Compounds **41b**-h were prepared by a similar method from appropriate phenylbromides **39b**-h, respectively: **41b**; oil, bp 134°C (32 mmHg) [21]; **41c**: oil (the product was purified by chromatography on silica gel with *n*-hexane/AcOEt (50:50) as eluent); **41d**: oil, bp 74°C (5 mmHg) [22]; **41e**: oil (the product was purified by chromatography on silica gel with *n*-hexane/ AcOEt (50:50) as eluent); **41f**: bp 120-121°C (6 mmHg) [23]; **41g**: bp 164-167°C (7 mmHg); **41h**: oil (the product was purified by chromatography on silica gel with *n*-hexane/AcOEt (50:50) as eluent) [24].

Method C. A typical example: 2-methyl-4-methoxypropiophenone 43a [25]. A solution of propionyl chloride (8.3 g, 90.0 mmol) in CH₂Cl₂ (10 ml) was added dropwise to a mixture of aluminum chloride (13.1 g, 98.2 mmol) in CH₂Cl₂ (50 ml) under ice cooling and then stirred for 0.5 h. The mixture was added to a solution of 3-methylanisole (10.0 g, 81.9 mmol) in CH₂Cl₂ (50 ml) under ice cooling and then stirred for 1 h. This mixture was poured into aqueous ammonia solution (10%) under ice cooling and then extracted with CH₂Cl₂. The CH₂Cl₂ layer was washed with H₂O, and then dried over MgSO₄. The solution was evaporated *in vacuo* to give a residual oil. The oil was distilled to give 43a (13.25 g, 74.3%) as an oil, bp 110–112°C (8 mmHg). IR v (neat) cm⁻¹: 1680 (C=O). 'H-NMR δ (CDCl₃),: 1.06 (t, 3H, J = 7.1 Hz), 2.33 (s, 6H), 3.00 (q, H, J = 7.0 Hz), 7.2 (d, 1H, J = 8.0 Hz); 7.7 (m, 2H).

Compounds **43b** and **43c** were prepared by a similar method from the appropriately substituted benzenes **41b** and **41c**, respectively: **43b**: colorless prisms, mp 50°C (pet ether) [15]; **43c**: mp 98°C (pet ether) [26].

Method D. A typical example: 4-chloro-2-hydroxypropiophenone 45a. A solution of propionyl chloride (9.25 g, 0.1 mol) in dry benzene (5 ml) was added dropwise to a mixture of 3-chlorophenol (12.9 g, 0.1 mol) and Mg (1.2 g, 0.1 mol) in dry benzene (25 ml) at room temperature and the mixture was stirred for 0.5 h. The mixture was then heated at 90°C for 2 h. The mixture was filtered and the filtrate was washed with 1%aqueous NaOH solution and then H₂O. The organic layer was dried over MgSO4 and evaporated in vacuo to give 3-chlorophenyl propionate (16.4 g, 88.6%) as an oil, bp 90°C (0.8 mmHg). A mixture of 3-chlorophenyl propionate **44a** (5.53 g, 30 mmol) and AlCl₃ (6.67 g, 50 mmol) was heated at 140°C for 2 h. The resulting solid was decomposed with 3 N HCl and then extracted with Et₂O and AcOEt. The combined organic layers were dried over MgSO₄ and then evaporated *in vacuo* to give crude **45a** (4.96 g, 89.2%) as colorless prisms. IR v (CHCl₃) cm⁻¹: 1680 (C=O). ¹H-NMR δ (CDCl₃): 1.23 (t, 3H, J = 8.0 Hz), 3.02 (q, 2H, J = 8.0 Hz), 6.95 (dd, 2H, J =9.0 Hz, J = 2.0 Hz), 7.74 (d, 1H, J = 9.0 Hz), 10.0–11.5 (brs, 1H).

4-Chloro-2-methoxypropiophenone **46a** [20]. A solution of **45a** (4.9 g, 26.7 mmol) in EtOH (10 ml) was added dropwise to a suspension of NaOMe (1.4 g, 26.7 mmol) in EtOH (100 ml) at room temperature for 0.5 h. Methyl iodide (4.54 g, 32 mmol) was added to the mixture. The mixture was stirred for 1 h at room temperature, and then warmed at 50°C for 6 h. The reaction mixture was evaporated *in vacuo* to give a residue, which was partitioned between H₂O and Et₂O. The Et₂O layer was dried over MgSO₄ and then evaporated *in vacuo* to give a residue.

due, which was chromatographed over silica gel using CH₂Cl₂/ *n*-bexane (40:60) as an eluent solvent to give **46a** (3.21 g, 60.6%) as an oil. IR v (CHCl₃) cm⁻¹: 1680. ¹H-NMR δ (CDCl₃): 1.13 (t, 3H, J = 7.8 Hz), 2.96 (q, 2H, J = 7.8 Hz), 3.93 (s, 3H), 6.7–7.2 (m, 2H), and 7.7 (d, 1H). Compounds **46b** and **46c** were prepared by a similar method using the appropriate phenol derivatives **44b** and **44c** respectively: **46b**: oil, bp 137°C (16.5 mm Hg) [27]; **46c**: oil, bp 139°C (8.0 mm Hg) [10].

Method E. Preparation of 4-bromomethylpropiophenone 48. A mixture of 4-methylpropiophenone 47 (21.7 g, 146.4 mmol), N-bromosuccimide (26.1 g, 146.4 mmol), and perbenzoic acid (0.25 g, 1.81 mmol) in CCl₄ (120 ml) was refluxed for 1 h. After the mixture was filtered, the filtrate was evaporated *in* vacuo to give a residue. The residue was chromatographed over SiO₂ using *n*-hexane/AcOEt (90:10) as an eluent solvent to give 48 (10.2 g, 31%) as an oil. IR v (KBr) cm⁻¹: 1685 (C=O). ¹H-NMR δ (CDCl₃): 1.27 (t, 3H, J = 7.0 Hz), 3.03 (q, 2H, J =7.0 Hz), 4.56 (s, 2H), 7.56 (d, 2H, J = 8.0 Hz), 8.05 (d, 2H, J =8.0 Hz).

4-Fluoromethylpropiophenone **49** [28]. A mixture of **48** (10.0 g, 44.0 mmol) and KF (12.7 g, 219 mmol) in diethyleneglycol (25 ml) was heated at 140°C for 1 h. Water was added to the mixture and then the mixture was extracted with Et₂O. The Et₂O layer was washed with H₂O and dried over MgSO₄, then evaporated *in vacuo* to give a residue, which was distilled to give **49** (3.62 g, 50.0%) as an oil, bp 99–102°C (6 mmHg). IR v (neat) cm⁻¹: 1690 (C=O). ¹H-NMR δ (CDCl₃): 1.22 (t, 3H, J = 0.7 Hz); 3.03 (t, 2H, J = 7.0 Hz), 5.46 (d, 2H, J = 56 Hz), 7.46 (d, 2H, J = 8.0 Hz), 8.01 (d, 2H, J = 8.0 Hz).

2-Methyl-3-pyrrolidinopropiophenones 3–35. A typical example: 2-methyl-3-pyrrolidino-1-(4-trifluoromethylphenyl)-propan-1one hydrochloride 28. A mixture of 4-trifluoromethylpropiophenone (4.0 g, 20 mmol), paraformaldehyde (1.8 g, 60 mmol), pyrrolidine hydrochloride (3.2 g, 30 mmol), and HCl (0.2 ml) in *i*PrOH (35 ml) was refluxed for 7 h. The reaction mixture was evaporated *in vacuo*. The residue was made alkaline with a saturated aqueous NaHCO₃ solution. The mixture was then extracted with toluene. The extract was dried over MgSO₄, and evaporated *in vacuo* to afford the free base of **28** (5.4 g, 94%) as an oil. IR v (neat): 1690 cm⁻¹ (C=O). ¹H-NMR (CDCl₃) δ : 1.25 (d, 3H, J = 7.0 Hz), 1.4–2.1 (m, 4H), 2.3–3.2 (m, 6H), 3.65 (m, 1H), 7.70 (d, 2H, J = 8.0 Hz), 8.05 (d, 2H, J = 8.0 Hz). Mass m/z (relative intensity %): 285 (2.21, M⁺), 214 (100), 173 (100), 145 (100), 95 (29.7), 84 (100).

This free base was converted into its hydrochloride with dry HCl gas in Et₂O. The resulting salt was recrystallized from MeOH/CH₃COCH₃ to give **28** as colorless prisms, mp 154–156°C. Anal ($C_{15}H_{18}F_{3}NO$ HCl) C, H, N, Cl, F. Compounds **3–27**, **29–35** were prepared by a method similar to that described above. Yields were calculated on the basis of propiophenones. The yields, recrystallization solvents, and melting points are listed in table I. The ¹H-NMR spectral data are shown in table III.

Optical resolution. (+)-(R)-2-Methyl-3-pyrrolidino-1-(4-trifluoromethylphenyl)-propan-1-one hydrochloride **28**-(R)(NK433)

(*RS*)-2-Methyl-3-pyrrolidino(4-trifluoromethylphenyl)propan-1-one (30.6 g, 107 mmol) and (L)-*N*-acetylphenylglycine (21.0 g, 109 mmol, $[\alpha]_D^{20}$ +212.8°, c = 1.0, MeOH) were dissolved in AcOEt. The solution was allowed to stand overnight at room temperature, and then was stirred at about 5°C for 3 h. The resulting precipitate was filtered off as first crude crystals (21.9 g). The filtrate was then reduced in weight to 5.74 g *in* vacuo, allowed to stand at room temperature for 2 d, and then stirred at $ca 5^{\circ}$ C for 3 h. The deposited crystals were filtered off as second crude crystals (17.5 g).

The first and second crude crystals prepared above were mixed and added to a mixture of 10% aqueous NaCl solution (63 ml) and AcOEt (95 ml). The mixture was made alkaline with saturated aqueous ammonia solution (28%) and then the AcOEt was extracted. The AcOEt was washed with 10% aqueous NaCl solution and then dried over MgSO₄. Dry HCl gas (2.9 g) was introduced to the AcOEt solution below 25°C and then the solution was stirred at 5°C for 2 h. The crystals were filtered off and washed with cold AcOEt (37 ml) to give (R)-2-methyl-3-pyrrolidino-1-(4-trifluoromethylphenyl)propan-1-one hydrochloride (21.5 g, optical purity 97%). The crude salts were added to a mixture of 10% aqueous NaCl solution (32 ml) and AcOEt (48 ml) similar to the previous treatment. The mixture was made alkaline with saturated aqueous ammonia solution (28%) and then the AcOEt was extracted. The AcOEt was washed with 10% aqueous NaCl solution and then dried over MgSO₄. Dry HCl gas (2.9 g) was introduced to the AcOEt solution below 25°C and then the solution was stirred at 5°C for 2 h to give (R)-2-methyl-3-pyrrolidino-1-(4-trifluoromethylphenyl)-1-propanone hydrochloride (17.4 g, optical purity 99.5%), $[\alpha]_D^{20} = 1.0, -45^{\circ}(c = 1.0, MeOH);$ mp 156-159°C.

Pharmacology

Flexor reflex inhibitory action

The flexor reflex was recorded according to the methods of Sakitama and Ishikawa [11]. In brief, animals (male Wistar rats) were anesthetized with urethane and α -chloralose, and the tibial nerve was dissected and stimulated (0.1 ms, 0.1 Hz, supra-maximum stimulation) by a stimulator (Model MSE-3; Nihon Kohden). The evoked electromyogram (EMG) recorded through a silver ball electrode placed on the ipsilateral muscle tibialis anterior was amplified and displayed on a memory-oscilloscope (VC-10; Nihon Kohden). The amplitude of this evoked EMG was recorded on a pen recorder through a peak holder. The activity of each of the compounds was expressed by the flexor reflex inhibition rate. The flexor reflex inhibition rate was calculated from the following equation:

Inhibition rate =
$$(A - B)/A \times 100$$
 (%)

where A is the average amplitude of the EMG in the period of 10 min before the administration of one of the abovementioned compounds, and B is the average amplitude of the EMG in the period of 30 min after intravenous (iv) administration of 5 mg/kg of the compound dissolved in physiological saline solution. In the case of the per os (po) study, B is the average amplitude of the EMG in the period of 60 min after the administration of 50 mg/kg of the compound dissolved in distilled water.

Action on anemic decerebrate rigidity of rats

This rigidity is caused by hyperactivity of α -motoneurons, and is considered a good experimental model for spasticity in humans. Anemic decerebrate rigidity was produced according to the method of Fukuda *et al* [12]. A tracheal cannula was inserted into an animal under ether anesthesia. Both of the common carotid arteries were ligated and the basilar artery was cauterized with a bipolar coagulator to block blood circulation to produce rigidity. The rigidity was recorded as described below. A rat was fixed on its back on a fixing stand and its

Compd No	$\delta(CDCl_3)$
3	1.23 (d, 3H, J = 7.0 Hz), 1.4-2.1 (m, 4H), 2.1-3.2 (m, 6H), 3.34.1 (m, 1H), 7.2-7.7 (m, 3H), 7.7-8.2 (m, 2H)
4	1.23 (d, $3H$, $J = 7.0Hz$), 1.3-2.1 (m, $4H$), 2.0-3.2 (m, $6H$), 3.2-3.8 (m, $1H$), 6.8-8.0 (m, $4H$)
5	1.22 (d, 3H, J = 7.0Hz), 1.4-2.1 (m, 4H), 2.1-3.2 (m, 6H), 3.3-3.9 (m, 1H), 7.0-7.5 (m, 4H)
6	1.21 (d, 3H, J = 7.0Hz), 1.4-2.1 (m, 4H), 2.1-3.2 (m, 6H), 3.3-4.0 (m, 1H), 6.9-7. (m, 2H), 7.8-8.3 (m, 2H)
7	1.23 (d, 3H, J = 7.0Hz), 1.4-2.0 (m, 4H), 2.1-3.2 (m, 6H), 3.2-3.8 (m, 1H), 7.1-7. (m, 4H)
8	1.25 (d, 3H, J = 7.0Hz), 1.4-2.1 (m, 4H), 2.2-3.1 (m, 6H), 3.2-3.8 (m, 1H), 7.0-7. (m, 4H)
9	1.20 (d, 3H, J = 7.0Hz), 1.4-2.0 (m, 4H), 2.1-3.1 (m, 6H), 3.2-3.9 (m, 1H), 7.2-8 (m, 4H)
10	1.20 (d, 3H, J = 7.0Hz), 1.4-2.0 (m, 4H), 2.2-3.1 (m, 6H), 2.43 (s, 3H), 3.2-3.9 (m, 1H), 7.0-7.4 (m, 3H), 7.4-7.7 (m, 1H)
11	1.20 (d, $3H$, $J = 7.0Hz$), 1.5-2.1 (m, $4H$), 2.2-3.2 (m, $6H$), 2.42 (s, $3H$), 3.2-3.9 (n 1H), 7.3-7.6 (m, 2H), 7.6-8.0 (m, 2H)
12	1.22 (d, $3H$, $J = 7.0Hz$), 1.5-2.1 (m, $4H$), 2.2-3.2 (m, $6H$), 2.40 (s, $3H$), 3.3-4.0 (n 1H), 7.0-7.4 (m, 2H), 7.7-8.1 (m, 2H)
13	1.20 (d, $3H$, $J = 7.0Hz$), 1.25 (t, $3H$, $J = 8.0Hz$), 1.4-1.9 (m, $4H$), 2.2-3.1 (m, $8H$), 3.1-3.8 (m, $1H$), 7.1-7.7 (m, $4H$)
14	1.22 (d, 3H, J = 7.0Hz), 1.25 (d, 6H, J = 7.0Hz), 1.4-2.0 (m, 4H), 2.2-3.7 (m, 8H 7.1-7.5 (m, 4H))
15	1.18 (d, $3H$, $J = 7.0Hz$), 1.2-2.0 (m, $4H$), 2.2-3.1 (m, $6H$), 3.2-3.8 (m, $1H$), 5.1-5. (m, $2H$), 6.7-7.2 (m, $1H$), 7.2-7.8 (m, $4H$)
16	1.23 (d, $3H$, $J = 7.0$ Hz), 1.4-2.0 (m, 4H), 2.2-3.2 (m, 6H), 3.3-4.0 (m, 1H), 5.2-6. (m, 2H), 6.5-7.1 (m, 1H), 7.2-8.1 (m, 4H)
17	1.22 (d, 3H, J = 7.0Hz), 1.3-2.1 (m, 8H), 2.1-3.2 (m, 10H), 3.3-4.1 (m, 1H), 6.0-6.3 (m, 1H), 7.2-8.1 (m, 4H)
18	0.88 (d, 3H, J = 7.0Hz), 2.3-2.9 (m, 4H), 2.9-3.0 (m, 7H), 7.1-7.6 (m, 9H)
19	1.22 (d, 3H, J = 7.0Hz), 1.4-1.9 (m, 4H), 2.1-3.2 (m, 6H), 3.3-4.0 (m, 1H) 7.1-8.3 (m, 9H)
20	1.26 (d, 3H, J = 7.0Hz), 1.5-2.1 (m, 4H), 2.2-3.2 (m, 6H), 3.4-4.1 (m, 1H), 7.2-8. (m, 9H)
21	1.25 (d, 3H, J = 7.0Hz), 1.5-2.1 (m, 4H), 2.3-3.2 (m, 6H), 3.4-4.3 (m, 1H), 6.7-8. (m, 4H), 10.0-11.5 (brs, 1H)

Table III. ¹H-NMR spectra of 3-pyrrolidinopropiophenones (3–27, 29–35).

Compd No	$\delta(CDCl_{\beta})$
22	1.18 (d, 3H, J = 7.0Hz), 1.3-2.1 (m, 4H), 2.2-3.1 (m, 6H), 3.1-3.9 (m, 1H), 3.90
	(s, 3H), 6.7-7.7 (m, 4H)
23	1.23 (d, 3H, J = 7.0Hz), 1.4-2.1 (m, 4H), 2.1-3.2 (m, 6H), 3.3-4.0 (m, 1H), 3.87 (s, 3.4)
	3H), 6.9-7.7 (m, 4H)
24	1.23 (d, 3H, J = 7.0 Hz), 1.4-2.0 (m, 4H), 2.2-3.2 (m, 6H), 3.3-3.9 (m, 1H), 3.86
	(s, 3H), 6.97 (d, 2H, $J = 8.0$ Hz), 8.03 (d, 2H, $J = 8.0$ Hz)
25	1.10 (d, 3H, J = 7.0Hz), 1.2-2.0 (m, 4H), 2.1-2.9 (m, 6H), 3.0-3.9 (m, 1H), 4.90 (d, 3H) = 0.000 (m, 1H), 0.0
	2H, J = 43.0 Hz), 6.5-6.9 (m, 2H), 7.0-7.5 (m, 2H)
26	1.18 (d, 3H, J = 7.0Hz), 1.5-2.0 (m, 4H), 2.2-3.1 (m, 6H), 3.1-3.7 (m, 1H), 7.4-7.9
	(m, 4H)
27	1.22 (d, 3H, J = 7.0Hz), 1.3-2.1 (m, 4H), 2.2-3.1 (m, 6H), 3.2-4.0 (m, 1H), 7.5-8.0
	(m, 2H), 8.0-8.4 (m, 2H)
28	1.25 (d, 3H, J = 7.0 Hz), 1.4-2.1 (m, 4H), 2.3-3.2 (m, 6H), 3.3-4.0 (m, 1H), 7.5-7.9
	(m, 2H), 7.9-8.3 (m, 2H)
29	1.16 (d, 3H, J = 7.0 Hz), 1.4-1.9 (m, 4H), 2.1-3.2 (m, 6H), 2.47 (s, 3H), 3.2-3.9
	(m, 1H), 3.85 (s, 3H), 6.6-6.9 (m, 2H), 7.4-7.8 (m, 1H)
30	1.20 (d, 3H, J = 7.0Hz), 1.5-2.1 (m, 4H), 2.1-3.2 (m, 6H), 2.25 (s, 3H), 3.3-4.1 (m, 6H), 2.25 (s, 3H), 3.3-4.1 (m, 6H), 2.25 (s, 3H), 3.3-4.1 (m, 6H), 3.3-4
	1H), 3.90 (s, 3H), 6.7-7.1 (m, 1H), 7.6-8.1 (m, 2H)
31	1.15 (d, 3H, J = 7.0Hz), 1.4-2.0 (m, 4H), 2.1-3.1 (m, 6H), 3.2-4.2 (m, 1H), 3.88 (s, 1.15) (m,
	3H), 6.7-7.3 (m, 2H), 7.3-7.8 (m, 1H)
32	1.18 (d, 3H, J = 7.0 Hz), 1.4-2.0 (m, 4H), 2.1-3.1 (m, 6H), 3.3-4.2 (m, 1H),
	3.88 (s, 3H), 6.8-7.1 (m, 1H), 7.2-7.8 (m, 2H)
33	1.17 (d, 3H, J = 7.0Hz), 1.4-2.0 (m, 4H), 2.1-3.1 (m, 6H), 3.2-3.8 (m, 1H), 3.86 (s,
	3H), 6.8-7.7 (m, 3H)
34	1.20 (d, 3H, J = 7.0Hz), 1.5-2.0 (m, 4H), 2.1-3.1 (m, 6H), 2.66 (s, 6H), 3.1-3.8 (m, 6H), 2.66 (s, 6H), 3.1-3.8 (m, 6H), 3.
	1H), 6.9-7.5 (m, 3H)
35	1.18 (d, 3H, J = 7.0Hz), 1.5-2.1 (m, 4H), 2.2-3.1 (m, 6H), 2.30 (s, 3H), 3.3-4.0 (m, 6H), 2.30 (s, 3H), 3.3-4.0 (m, 6H), 2.30 (s, 3H), 3.3-4.0 (m, 6H), 3.3-4
	1H), 3.73 (s, 3H), 7.2-7.5 (m, 2H)

forepaws were allowed to grasp an end of a celluloid plate provided with strain gauges on both sides. A change of the resistance observed when the celluloid plate was forced up by the rigidity of the forepaws was recorded as a tension through a bridge circuit on a recorder. A rigidity inhibition rate was calculated according to the following equation:

Rigidity inhibition rate = $(C - D) / C \times 100 (\%)$

where C represents the average tension (g) for 10 min before the administration of one of the above-mentioned compounds, and D represents the average for 10 min at peak period tension after iv administration of 3.5 mg/kg of the compound. The results are shown in table II.

Acknowledgments

We thank Y lidaka and H Nakamura for the X-ray analysis of compound 50.

References

- 1 CèrnaLcèk J, JaLgr J (1966) In: IV Conferentia Hungarica pro Therapia et investigatione in Pharmacologia, Akado, Kiado, Budapest, Hungary, 557–561
- 2 Kuroiwa Y, Sobue I, Tazaki Y, Nakanishi T, Ohtomo E, Itahara K (1981) Clin Eval 9, 391 (abstract in English)
- 3 Hudgson P, Weightman D (1971) Br Med J IV, 15-17
- 4 Kaurimsky ZF, Fassbender HM (1981) J Int Med Res 9, 501-505

- 94
- 5 Nippon Kayaku Co Ltd (1978) Jpn Kokai 7840779; Chem Abstr (1978) 109128m
- 6 Blicke FF (1942) In: Organic Reactions (Adams R, ed) John Wiley & Sons Inc, New York, USA, vol 1, Ch 10, 303-341
- 7 Ropp GA, Coyner EC (1950) J Am Chem Soc 72, 3960-3963
- 8 Bull JR (1972) J Chem Soc Perkin Trans 1, 627-632
- 9 Olah GA (1964) In: Friedel Crafts and Related Reactions. John Wiley & Sons Inc, New York, USA, Part 1, 2;
- 10 Chakravarti, Dutta (1940) J Ind Chem Soc 17, 65-71
- 11 Sakitama K, Ishikawa M (1992) Jpn J Pharmacol 60, 127-131
- 12 Fukuda H, Ito T, Hashimoto S, Kudo Y (1974) Jpn J Pharmacol 24, 810-813
- 13 Capillon J, Guétté JP (1979) Tetrahedron 35, 1817-1820
- 14 Zenitz BL, Hartung WL (1946) J Org Chem 11. 444-450
- 15 Kindler K, Trauping Li (1941) Chem Ber 74, 321-324
- 16 Jensen BL, Burke SE, Thomas SE (1978) Tetrahedron 34, 1627-1631

- 17 Johnson WS. Erickson CA, Ackerman J (1952) J Am Chem Soc 74, 2251– 2253
- 18 Mc Neil Lab (1967) US Pat No 3308121; Chem Abstr 67, 32693
- 19 Eistert B, Schade W, Meck N (1968) Justus Liebigs Ann Chem 717, 80-90
- 20 Sen AB, Tiwari SS (1952) J Ind Chem Soc 29, 419-424
- 21 Birch SF, Dean RA, Fidler FA, Lowry RA (1949) J Am Chem Soc 71, 1362-1369
- 22 Reid AA, Sharp JT, Sood HR, Thorogood PB (1973) J Chem Soc Perkin Trans 1, 2543-2551
- 23 Stiles M, Sisti AJ (1961) J Org Chem 26, 3639-3644
- 24 Leont'eva LI, Yuldashev KhY, Sidorova NG (1971) Uzb Khim Zh 15, 54-57
- 25 Miquel JF, Buu-Hoï NP, Royer R (1955) J Chem Soc 3417-3420
- 26 Mann N, Back W, Mutscher E (1973) Arch Pharm 306, 625-631
- 27 Chakravarti D, Majumdar B (1938) J Ind Chem Soc 15, 136-138
- 28 Hokuriku Pharmaceutical Co Ltd (1985) Jpn Kokai Tokkyo Koho JP 59 190 982; Chem Abstr (1985) 102, 166464d