

Asymmetric Synthesis of Oxygenated Monoterpenoids of Importance for Bark Beetle Ecology

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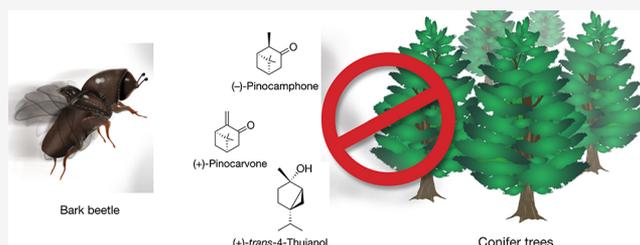


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ABSTRACT: Herein we report the asymmetric syntheses of a number of oxygenated terpenoids that are of importance in the chemical ecology of bark beetles. These are pinocamphones, isopinocamphones, pinocarvones, and 4-thujanols (= sabinene hydrates). The camphones were synthesized from isopinocampheol, the pinocarvones from β -pinene, and the thujanols from sabinene. The NMR spectroscopic data, specific rotations, and elution orders of their stereoisomers on a chiral GC-phase (β -cyclodextrin) are also reported. This enables facile synthesis of pure compounds for biological activity studies and identification of stereoisomers in mixed natural samples.



The colonization of trees by bark beetles is generally influenced by an intricate release of chemical signals in strict chronological order. The signals recruit conspecifics to a suitable host tree, and later in the colonization, other compounds are produced to convey to conspecifics that this tree is becoming overexploited.¹ Thereby competition for food and larvae is avoided. These attractant chemical signals can originate from metabolized monoterpenoids, like for the noxious larger spruce bark beetle, *Ips typographus*, where *cis*-verbenol is converted to verbenone, but the pheromones can also be synthesized *de novo*.² In recent work with semi-chemicals for tree-killing bark beetles we have encountered a number of oxygenated monoterpenoids that are physiologically active (antiattractive, i.e., reduce the effect of aggregation pheromone) in these beetles (Figure 1).^{3,4}

We recently published that the production of oxygenated monoterpenoids is related to tree stress and that it might be a signal for a suitable or unsuitable host for bark beetles.³ Investigations by gas chromatographic electroantennographic detection (GC-EAD) of monoterpenones by us³ and Kalinova et al.⁵ revealed that both isopinocamphones and pinocamphones elicit antennal responses in *I. typographus*. There are relatively few syntheses of pinocamphone and isopinocampheol reported. In one report in Chinese, Wang et al. reacted α -pinene with borane to obtain diisopinocampheylborane, which was oxidized to isopinocampheol by sodium perborate to afford isopinocampheol.⁶ The isopinocampheol was finally oxidized by H₂O₂ with vanadium phosphorus oxide as catalyst to yield isopinocampheol. Pitinová-Šteková and co-workers utilized different titanosilicate catalysts to convert α -pinene to obtain campholenic aldehyde.⁷ Some of these catalysts produced pinocamphone as side-product. In another report, thermolysis of α -pinene epoxide in supercritical anhydrous isopropanol afforded up to ~25% pinocamphone, but in an

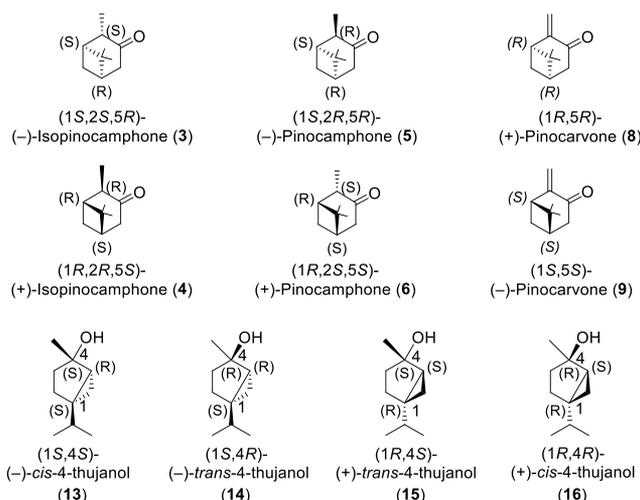


Figure 1. Absolute configuration of the synthesized oxygenated terpenoids.

inseparable mixture of oxygenated monoterpenoids.⁸ These syntheses do not yield pure stereoisomers or are tedious and have low yields. For short and convenient synthesis without heating, we developed a simple method using pure isopinocampheol stereoisomers that are commercially available.

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Pinocarvone is reported as a pheromone for the southern pine beetle (*Dendroctonus frontalis*)⁹ and has been found in the hindguts of male white pine cone beetles, *Conophthorus coniperda*.¹⁰ In the GC-EAD analysis of *I. typographus*, pinocarvone gave strong antennal responses, indicating biological activity.³ Pinocarvone has been isolated from Eucalyptus oil and has been produced by oxidation of β -pinene with SeO_2 . In this reaction, myrtenal is formed by a rearrangement, and this byproduct was either overlooked or lost during spinning band distillation.^{11,12}

In previous reports, we reported that one of the two *trans*-4-thujanol stereoisomers showed strong GC-EAD activity for bark beetles³ and that this (+)-*trans*-thujanol is a field-active semiochemical for the bark beetle, *I. typographus*.¹³ Blazytė-Cereskienė et al. reported that young spruce trees release more 4-thujanol than older trees and that 4-thujanol plays an important role in both host defense and tree choice by bark beetles.¹⁴ Thus, 4-thujanol is seemingly an indicator of healthy strong trees which should be avoided and could be of interest in forest protection. Several publications on the synthesis of 4-thujanol have been published, including the biotransformation of α -pinene to *cis*-4-thujanol using the microorganism *Fusarium saloni*,¹⁵ Bäckström's synthesis of *trans*-4-thujanol from 3-thujol,¹⁶ Galopin's synthesis of the *trans* isomers from methyl vinyl ketone,¹⁷ Cheng's syntheses of *cis*-thujanol,^{18,19} and Fanta's synthesis of *trans*-thujanol.²⁰ However, all these synthetic procedures involve many steps and/or expensive starting materials, and there are no effective synthetic routes for all possible stereoisomers. In order to develop a short synthesis of all stereoisomers of 4-thujanol for the investigation of GC-EAD activity, herein these were synthesized from commercial sabinene. The absolute configuration of each stereoisomer was unambiguously assigned by the deduction from the original sabinene in combination with NMR spectroscopy and chiral-phase GC-MS analysis.

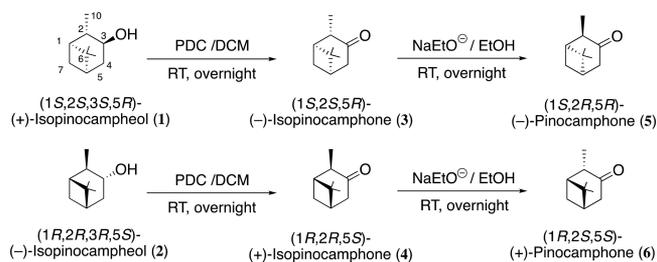
These compounds are obviously important in bark beetle ecology, and some of them act as indicators of tree health; they are also of interest for managing bark beetle populations. It is well known that the stereochemistry of pheromones and other semiochemicals is often extremely important.^{21–24} Thus, there is a need to develop analytical procedures to be able to use enantioselective gas chromatography to differentiate between the stereoisomers of these semiochemicals in a biological sample, as well as their facile synthesis. We herein report the syntheses, specific rotations, and elution orders on a chiral GC phase (β -cyclodextrin) of pinocamphones, isopinocamphones, pinocarvones, and 4-thujanols (sabinene hydrate).

RESULTS AND DISCUSSION

Synthesis of Isopinocamphones and Pinocamphones. Scheme 1 summarizes the syntheses of the four stereoisomers of pinocamphone. The pure enantiomers of isopinocamphol (1 and 2) were separately oxidized with pyridinium dichromate (PDC) to obtain both enantiomers of isopinocamphone in >98% optical purity. The oxidation was improved by adding silica gel to the reaction mixture, which prevents the formation of lumps and tar and in turn leads to higher yields and easier filtration at workup.

To produce pinocamphones, NaOEt was used to epimerize C-2 of isopinocamphone. The thermodynamic equilibrium seems to be 4:1 in favor of pinocamphone, and thus, 20% of isopinocamphone had to be removed by chromatography to obtain pure pinocamphone (Scheme 1).

Scheme 1. Synthesis of the Four Stereoisomers of Pinocamphone



Specific Rotation of Isopinocamphones and Pinocamphones. The sign of the specific rotation changed when going from isopinocamphols (1 and 2) to isopinocamphones (3 and 4), but not during epimerization from isopinocamphones to pinocamphones (5 and 6). The specific rotations are listed in Table 1.

Table 1. Enantioselective GC-FID and Specific Rotations of Isopinocamphols, Isopinocamphones, and Pinocamphones

s. no	compound	t_R	specific rotation	class
1	(1S,2S,3S,5R)-(+)-isopinocamphol		+34 (c 1.0, DCM)	isopinocamphols
2	(1R,2R,3R,5S)-(-)-isopinocamphol		-36 (c 1.0, DCM)	
3	(1S,2S,5R)-(-)-isopinocamphone	11.69	-11.4 (c 1.0, EtOH)	isopinocamphones (IPC)
4	(1R,2R,5S)-(+)-isopinocamphone	11.61	+11.2 (c 1.0, EtOH)	
5	(1S,2R,5R)-(-)-pinocamphone	11.37	-20.2 (c 1.0, EtOH)	pinocamphones (PC)
6	(1R,2S,5S)-(+)-pinocamphone	11.53	+22.7 (c 1.0, EtOH)	

GC Elution Order of Isopinocamphones and Pinocamphones. In the analysis by GC equipped with HP-SMS or DB-SMS columns, the pinocamphones eluted before the isopinocamphones. In the GC analysis using a chiral-phase column (Cyclosil B), (-)-pinocamphone (5) eluted before (+)-pinocamphone (6) and (+)-isopinocamphone (4) before (-)-isopinocamphone (3) (Table 1 and Figure 2).

Synthesis of Pinocarvone Stereoisomers. There are only a few syntheses of pinocarvone published, and usually

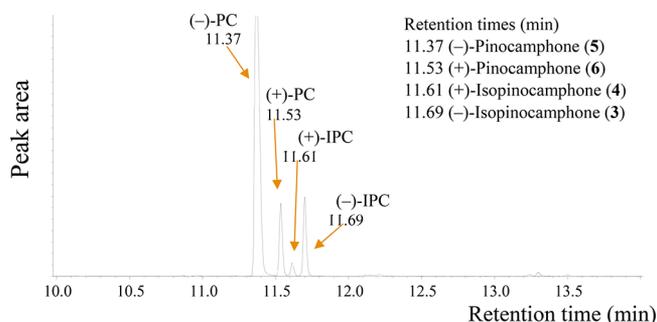
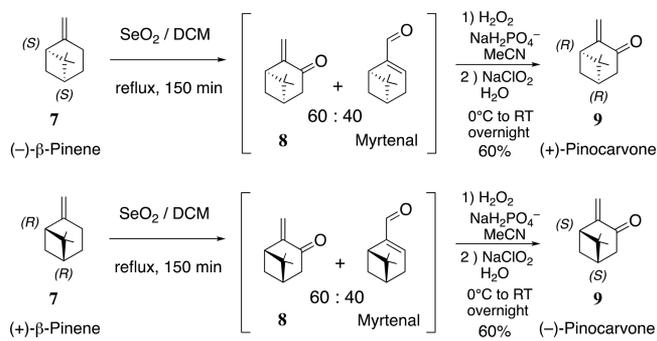


Figure 2. Mix of pinocamphones and isopinocamphones separated by enantioselective GC. The temperature program was isothermal 110 °C on a Cyclosil B column.

pinocarvone has been synthesized by oxidation of α -pinene or β -pinene (Scheme 2).^{25,26} One example is the Crich synthesis

Scheme 2. Synthesis of Pinocarvones



of pinocarvone from β -pinene (7) using perfluorooctyl selenic acid, where focus was on the preparation of the catalyst, and the yield was ca. 40%.²⁷ However, a serious drawback with pinene as starting material is the formation of myrtenal, which cannot be removed by silica column chromatography. Here we report a process where each pinocarvone enantiomer was separately synthesized from enantiopure β -pinene isomers (7) by oxidation with SeO_2 (Scheme 2). The mixture of pinocarvone and myrtenal was subsequently oxidized with $\text{H}_2\text{O}_2/\text{NaH}_2\text{PO}_4$ and NaClO_2 to remove the myrtenal in the form of myrtenic acid by silica gel chromatography (see GC-chromatogram, Figure 3). On the enantioselective GC-column

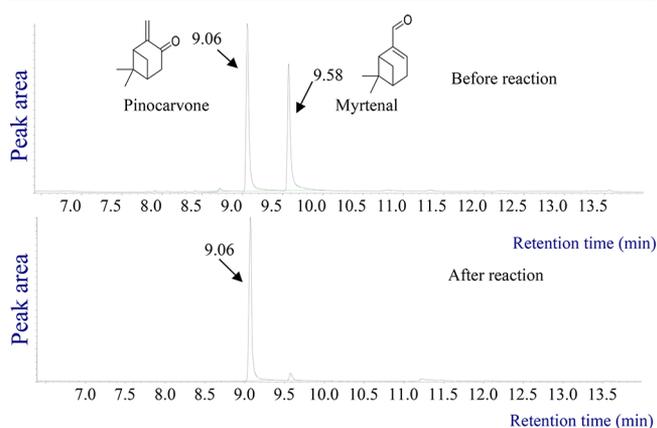


Figure 3. Gas chromatograms before and after oxidation of myrtenal by $\text{NaClO}_2/\text{H}_2\text{O}_2$.

phase (–)-pinocarvone (9) elutes before (+)-pinocarvone (8) (Figure 4). The ^1H NMR spectroscopic data of pinocarvone have been published,²⁸ but here we also provide ^{13}C NMR spectroscopic data.

Specific Rotation of Pinocarvone Stereoisomers. The sign of specific rotation changed when going from pinene to pinocarvone, i.e., (–)- β -pinene yielded (+)-pinocarvone (8) and (+)- β -pinene yielded (–)-pinocarvone (9). The specific rotations were as follows: (+)-pinocarvone (8) $[\alpha]_D^{23} +30.8$ (c 1.0, EtOAc); (–)-pinocarvone (9) $[\alpha]_D^{23} -29.6$ (c 2.0, EtOAc).

Synthesis of 4-Thujanol (Sabinene Hydrate) Stereoisomers. (–)-Sabinene (10) (86% ee) was subjected to a mild permanganate oxidation yielding sabinenediol (11) (Scheme 3). The diol was cleaved using periodate to yield sabinone (12).

Chirality of the sabinone was confirmed by use of specific rotation and a GC column (β -cyclodextrin phase).²⁹ The ketone was reacted with MeLi. The methyl group attacked stereoselectively from the sterically less hindered side of the carbonyl, resulting in a 10:1 excess of *cis*-forms (13 plus enantiomer 16) over the corresponding *trans*-forms of thujanol (14 plus enantiomer 15) (i.e., *cis*-4-thujanol is the major diastereomer formed).

As the ratio of stereoisomers in the sabinone (10) was (–)-93:(+)-7, it was easy to differentiate the (+)-(1*R*)- and (–)-(1*S*)-forms of sabinone by enantioselective GC. The reaction with MeLi yielded a mixture of all four 4-thujanol stereoisomers, which could be defined as (+)-*trans*-(1*R*,4*S*)-4-thujanol (15), (–)-*trans*-(1*S*,4*R*)-4-thujanol (14), (+)-*cis*-(1*R*,4*R*)-4-thujanol (16), and (–)-*cis*-(1*S*,4*S*)-4-thujanol (13), in a ratio of 1:9:4:86, by use of the enantioselective GC column.

The NMR spectrum and specific rotation proved that the isomer purchased from Sigma-Aldrich was the (+)-*trans*-isomer, and the major product could be assigned as (–)-*cis*-4-thujanol (13), based on the retention of ring configuration in the synthesis sequence, as well as regioselective considerations and reported NMR spectroscopic data.^{30,31}

GC Elution Order of 4-Thujanol Stereoisomers. On the HP-5MS GC column, the *trans* diastereomers eluted first. On the β -cyclodextrin column the first peak of four synthetic isomers coeluted with the commercial (+)-*trans*-4-thujanol stereoisomer (15) purchased from Sigma-Aldrich, and the last peak coeluted with the isolated (–)-*cis*-thujanol (13). The elution order of all isomers was (+)-*trans*, (–)-*trans*, (+)-*cis*, (–)-*cis* (Figure 5). The elution order is in accordance with those reported by Larkov et al.³¹ and Marriott et al.³²

Specific Rotation of Sabinone and 4-Thujanol Stereoisomers. It should be noted that (–)-sabinone (10) yields (+)-sabinone (12), which is subsequently transformed to thujanol with (–)-*cis*-thujanol (13) as the major isomer (Scheme 3). The commercial sabinone (apparently from a natural source) has a specific rotation of -73 (c 1.0, EtOH) and -81 (c 1.0, DCM). Moreover, the chemical purity of the commercial sabinone (10) was only 75%, with 25% β -pinene as an impurity and with an ee of 86%. Sabinone (12) was obtained in 86% ee and with specific rotations of $[\alpha]_D^{23} +24$ (c 1.0, EtOH) and $[\alpha]_D^{23} +33.5$ (c 1.0, EtOAc) after removal of byproducts (pinene ketones) by chromatography. The (–)-*cis*-thujanol isomer (13) produced in the last step had, after chromatography, an optical purity of 91% ee and a specific rotation of -40 (c 0.5, DCM). The commercial (+)-*trans*-4-thujanol (15) (Sigma-Aldrich) had a specific rotation of $+29.8$ (c 0.5, DCM).

EXPERIMENTAL SECTION

General Experimental Procedures. Optical rotations were recorded in EtOH, EtOAc, and DCM on a 2019 model Rudolph automatic polarimeter (APIII) manufactured by Rudolph Research Analytical (Hackettstown, NJ, USA). NMR spectra were recorded in CDCl_3 on Bruker 400 and Varian 500 MHz spectrometers. The GC-MS instrument was an Agilent 6890 GC and 5973 mass detector and a Hewlett-Packard with a FID detector (Palo Alto, CA, USA). Helium was used as carrier gas. Two types of columns, a nonpolar column (HP-5MS, film thickness = 0.25 μm ; Agilent Technologies 19091S-433) and a chiral-phase capillary column (Cyclosil-B, 30 m \times 0.25 μm , i.d. 0.25 mm, J&W Scientific, via Scantech Nordic AB, Jonsered, Sweden), were used. Mass spectra were obtained by electron impact ionization (70 eV). The general gas chromatography temperature

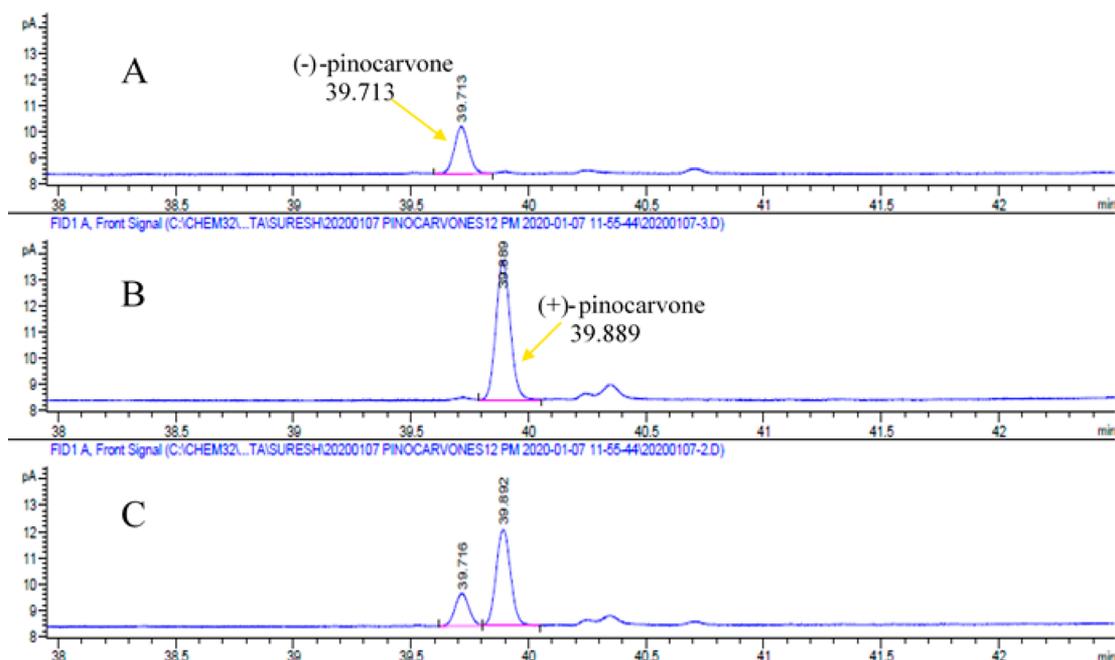


Figure 4. GC elution order of pinocarvone stereoisomers. Separated on GC equipped with a Cyclosil B column. In the chiral analysis (–)-pinocarvone eluted earlier than (+)-pinocarvone. (A) (–)-Pinocarvone (9), (B) (+)-pinocarvone (8), and (C) mix of pinocarvone enantiomers. The temperature program: Initial oven temperature was 40 °C (held for 5 min) increased to 150 °C at 3 °C/min and finally increased to 220 °C at 10 °C/min (held for 5 min at the final temperature).

Scheme 3. Synthesis of 4-Thujanol Stereoisomers

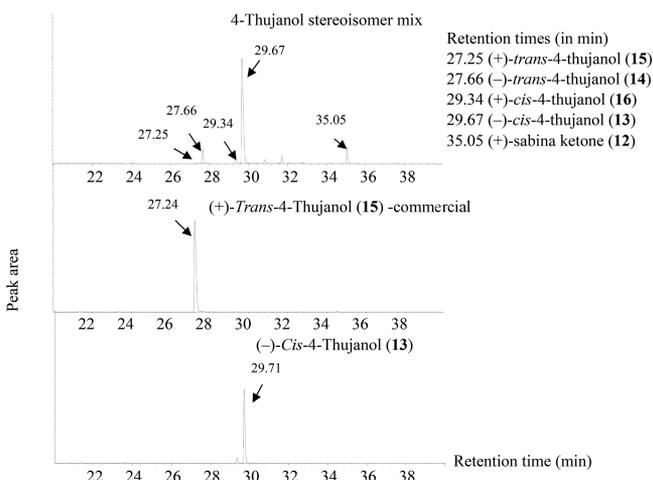
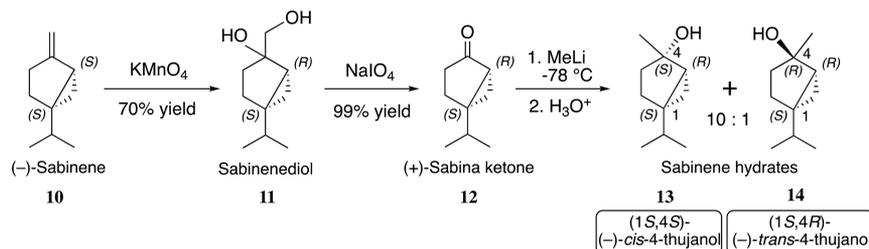


Figure 5. Chromatograms of chiral-phase GC of 4-thujanol isomers in the mixture and chromatograms of isolated (+)-*trans*- and (–)-*cis*-4-thujanol isomers. The temperature program: Initial oven temp 40 °C (hold for 3 min) and increased to 150 °C at 3 °C/min and finally increased to 250 °C at 15 °C/min. It was kept for 10 min at the final temperature of 250 °C.

program for both columns was as follows: initial temperature 50 °C (hold for 2 min), raised to 200 °C with 10 °C/min (hold for 15 min) (splitless). When a different temperature program was used for resolution of enantiomers (on the Cyclosil-B column), the temperature program is described in the figure legends of the GC chromatograms. The synthesized compounds were purified on silica gel column chromatography using 230–400 mesh ultra pure silica.

Synthesis of the Four Stereoisomers of Pinocampheol (Scheme 1). *Synthesis of (–)-Isopinocampheol (3).* (1*S*,2*S*,3*S*,5*R*)-2,6,6-Trimethylbicyclo[3.1.1]heptan-3-ol [(1*S*,2*S*,3*S*,5*R*)-(+)-isopinocampheol] (1) (Sigma-Aldrich, Schnelldorf, Germany) (12.32 g, 80.0 mmol, chemical purity 98% and optical purity 95% ee) was dissolved in CH₂Cl₂ (150 mL).³² Silica gel (18 g) and PDC (60 g, 160 mmol) were added, and the mixture was stirred for 3.5 h at RT before leaving it in a fridge overnight. The slurry was diluted with cyclohexane and filtered. The solid material was washed twice with 1:1 cyclohexane/CH₂Cl₂ (50 mL). The filtrate was concentrated and subjected to MPLC, yielding 87% (10.6 g, 69.7 mmol) of (1*S*,2*S*,5*R*)-2,6,6-trimethylbicyclo[3.1.1]heptan-3-one ((1*S*,2*S*,5*R*)-(–)-isopinocampheone) (3). The chemical purity was 99% with 98% ee.

Specific rotation [α]_D²³ = –11.4 (*c* 1.0, EtOH); ¹H NMR (CDCl₃, 500 MHz) δ (in ppm) 2.68–2.58 (2H, m), 2.54–2.45 (2H, m), 2.15–2.10 (1H, m), 2.08–2.05 (1H, m), 1.42 (1H, br s), 1.33 (3H, s, –CH₃), 1.21 (3H, d, –CH₃), 0.87 (3H, s, –CH₃); ¹³C NMR (CDCl₃, 125 MHz) δ (in ppm) 215.3, 51.4, 45.1, 45.0, 39.1, 34.5, 27.2, 27.1, 22.1, 17.0; GC-MS *m/z* 83 (100%), 69, 55, 95, 41, 81, 97, 67, 152 (*M*⁺), 110 (decreasing order of intensity).

(+)-*Isopinocampnone* (**4**). In analogy with the procedure used for the (−)-antipode, (1*R*,2*R*,3*R*,5*S*)-2,6,6-trimethylbicyclo[3.1.1]heptan-3-ol [(1*R*,2*R*,3*R*,5*S*)-(−)-isopinocampheol] (**2**) (12.21 g, 79.3 mmol) was oxidized with PDC (60 g, 160 mmol) to yield 75% (9.03 g, 59.4 mmol) (1*R*,2*R*,5*S*)-2,6,6-trimethylbicyclo-[3.1.1]heptan-3-one [(1*R*,2*R*,5*S*)-(+)-isopinocampnone] (**4**) after column chromatography. $[\alpha]_D^{23} = +11.2$ (*c* 1.0, EtOH), 98% ee. The NMR data were identical to the data for the (−)-isomer. GC-MS *m/z*. See other enantiomer.

(−)-*Pinocampnone* (**5**). (1*S*,2*R*,5*R*)-2,6,6-Trimethylbicyclo[3.1.1]heptan-3-one. To a solution of (−)-isopinocampnone (**3**) (3.04 g, 20.0 mmol) dissolved in EtOH (10 mL) was added NaOEt in EtOH (21% w/w, 11 mL, 34 mmol), and the mixture stirred for 24 h at RT. Water (50 mL) and Et₂O (50 mL) were added after 24 h, when the 4:1 equilibrium ratio between (−)-pinocampnone (**5**) and (−)-isopinocampnone (**3**) had been established. The aqueous phase was extracted with Et₂O (2 × 50 mL), and the combined ether phases were washed with 15 mL of water to remove EtOH. After drying over MgSO₄, filtration, and evaporation, 15 mL of toluene was added before subsequent evaporation. The removal of water/EtOH by azeotropic distillation with toluene was repeated twice. The clear amber-colored residue was subjected to MPLC, yielding 2.9 g (19.1 mmol) of (1*S*,2*R*,5*R*)-(−)-pinocampnone (**5**) as a 95:5 mixture with (1*S*,2*S*,5*R*)-(−)-isopinocampnone (**3**). $[\alpha]_D^{23} = -20.2$ (*c* 1.0, EtOH); ¹H NMR (CDCl₃, 500 MHz) δ (in ppm) 2.67–2.59 (2H, m), 2.49 (2H, m), 2.14–2.07 (1H, m), 1.92 (1H, td, *J* = 6.1 and 2.1 Hz), 1.33 (3H, s, −CH₃), 1.16 (1H, d, *J* = 10.8 Hz), 1.10 (3H, d, *J* = 7.3 Hz, −CH₃), 0.89 (3H, s, −CH₃); ¹³C NMR (CDCl₃, 125 MHz) δ (in ppm) 215.7, 46.5, 44.4 (2C), 39.4, 38.2, 29.1, 26.4, 19.8, 15.1; GC-MS *m/z* 83 (100%), 55, 69, 41, 95, 81, 97, 67, 152 (M⁺), 53 (decreasing order of intensity).

(+)-*Pinocampnone* (**6**). In analogy with the (−)-antipode, (1*R*,2*R*,5*S*)-2,6,6-trimethylbicyclo[3.1.1]heptan-3-one [(1*R*,2*R*,5*S*)-(+)-isopinocampnone, **4**] (3.04 g, 20.0 mmol) was epimerized with NaOEt in EtOH. The yield of (1*R*,2*S*,5*S*)-2,6,6-trimethylbicyclo[3.1.1]heptan-3-one ((1*R*,2*S*,5*S*)-(+)-pinocampnone, **6**) as a 4:1 mixture with (1*R*,2*R*,5*S*)-(+)-isopinocampnone (**4**), after column chromatography, was 95% (2.89 g, 19.0 mmol). $[\alpha]_D^{23} = +22.7$ (*c* 1.0, EtOH); ¹H NMR (CDCl₃, 500 MHz) δ (in ppm) 2.69–2.58 (2H, m), 2.50–2.36 (2H, m), 2.11 (1H, td, *J* = 5.8 and 2.8 Hz), 1.92 (1H, td, *J* = 6.1 and 2.2 Hz), 1.33 (3H, s, −CH₃), 1.16 (1H, d, *J* = 10.8 Hz), 1.10 (3H, d, *J* = 7.3 Hz, −CH₃), 0.89 (3H, s, −CH₃); ¹³C NMR (CDCl₃, 125 MHz) δ (in ppm) 215.7, 46.5, 44.4 (2C), 39.4, 38.2, 29.1, 26.3, 19.8, 15.1; GC-MS *m/z* identical to the (−)-isomer **5** of pinocampnone (see above).

Synthesis of the Pinocarvone Stereoisomers (Scheme 2). To a solution of (−)- β -pinene (**7**) (0.25 g, 1.8 mmol) in DCM (3 mL) was added SeO₂ (0.20 g, 1.8 mmol), and the mixture was refluxed for 2.5 h until GC-MS showed complete transformation. The solution was filtered through silica gel (in a Pasteur pipet), and the product washed out of the silica gel with additional aliquots of DCM. The product was concentrated under reduced pressure at 30 °C to obtain a mixture of pinocarvone and myrtenal. These two compounds were close on TLC and difficult to purify by column chromatography. To the concentrate, MeCN (2 mL), NaH₂PO₄ (70 mg) in Milli-Q-water (1 mL), and 35% H₂O₂ (0.2 mL) were added. The solution was stirred for approximately 1 h until the solution became clear. On an ice bath, NaClO₂ (0.32 g) in MQ-water (3 mL) was added dropwise, and the mixture was stirred overnight. One spatula of anhydrous Na₂SO₃ was added, and the mixture was extracted with DCM (3 × 5 mL). After removal of the solvents, the concentrate was purified on silica gel. The combined fractions were concentrated by rotatory low-vacuum evaporation to afford (+)-pinocarvone (**8**) (yield 60%, 165 mg, 1.1 mmol). The same experimental procedure was followed to produce (−)-pinocarvone (**9**) from (+)- β -pinene. The chemical purity of (+)-pinocarvone was 95%, and the optical purity was 97% ee.

(+)-*Pinocarvone* (**8**). $[\alpha]_D^{23} = +30.8$ (*c* 1.0, EtOAc) and +27.5 (*c* 1.0, DCM); ¹H NMR (CDCl₃, 500 MHz) δ (in ppm) 5.93 (1H, s), 4.98 (1H, s), 2.74 (1H, t, *J* = 5.9 Hz), 2.69–2.64 (1H, m), 2.63 (1H,

d, *J* = 2.6 Hz), 2.50 (1H, dd, *J* = 19.3 and 2.7 Hz), 2.18 (1H, dd, *J* = 5.8 and 3.0 Hz), 1.33 (3H, s, H-8), 1.27 (1H, d, *J* = 10.5 Hz), 0.78 (3H, s, H-9); ¹³C NMR (CDCl₃, 125 MHz) δ (in ppm) 200.1, 149.1, 117.5, 48.3, 42.6, 40.9, 38.6, 32.5, 26.1, 21.6; GC-MS *t_R* 9.06 *m/z* 81 (100%), 108, 53, 107, 135, 79, 41, 77, 150 (M⁺), 69, 91, 122 (intensity of decreasing order). NMR spectrometry data are in accordance with reported values.^{16,28}

(−)-*Pinocarvone* (**9**). $[\alpha]_D^{23} = -29.6$ (*c* 2.0, EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ (in ppm) 5.94 (1H, s, H-10 a), 4.99 (1H, s, H-10 b), 2.74 (1H, t, *J* = 5.9 Hz), 2.69–2.64 (1H, m), 2.63 (1H, d, *J* = 2.6 Hz), 2.52 (1H, dd, *J* = 19.3 and 2.7 Hz), 2.24–2.17 (1H, m), 1.33 (3H, s, H-8), 1.27 (1H, d, *J* = 10.5 Hz), 0.78 (3H, s, H-9); ¹³C NMR (CDCl₃, 125 MHz) δ (in ppm) 200.2, 149.1, 117.6, 48.3, 42.6, 40.41, 38.6, 32.5, 26.1, 21.6; GC-MS *t_R* 9.06 *m/z* 108 (100%), 81, 53, 107, 135, 79, 41, 150 (M⁺), 69, 91, 122 (intensity of decreasing order).

Synthesis of 4-Thujanol Isomers (Scheme 3). **Synthesis of Sabinenediol (11).** To a solution of (−)-sabinene (**10**) 86% ee (1 g, 7.4 mmol) in THF (3 mL) was added KMnO₄ (2.3 g, 14.6 mmol) in water (4 mL) over a period of 2.5 h. The mixture was stirred for another hour before the precipitate was filtered off. The filtrate was extracted with EtOAc (2 × 50 mL), and the combined organic layers were washed with brine and dried over Na₂SO₄. The solution was concentrated by rotatory evaporation to yield 875 mg (5.1 mmol) of crude sabinene diol (**11**) (70% yield). ¹H NMR (CDCl₃, 500 MHz) δ (in ppm) 3.55 (2H, app t, *J* = 11.6 Hz), 2.45 (1H, br s, −OH), 2.26 (1H, br s, −OH), 1.95–1.89 (1H, m), 1.67–1.61 (1H, m), 1.54 (1H, dd, 14.1 and 8.6 Hz), 1.45 (1H, heptet, *J* = 6.9 Hz), 1.25–1.18 (1H, m), 1.11 (1H, ddd, *J* = 8.5, 3.6, and 1.4 Hz), 0.98 (3H, d, *J* = 6.8 Hz), 0.89 (3H, d, *J* = 6.8 Hz), 0.41 (1H, dd, *J* = 8.3 and 5.3 Hz), 0.24 (1H, dd, *J* = 5.2 and 3.5 Hz); ¹³C NMR (CDCl₃, 125 MHz) δ (in ppm) 83.5, 68.1, 34.5, 32.5, 32.3, 30.4, 25.5, 20.2, 20.1, 12.9.

Synthesis of Sabina Ketone (12). To a stirred solution of sabinenediol diastereomers (**11**) (800 mg, 4.7 mmol) in THF/H₂O (1:1, 5 mL) was added NaIO₄ (3.7 g, 17.3 mmol) in five portions during 30 min at RT. After 4 h, the mixture was diluted with water (20 mL) and extracted with EtOAc (2 × 30 mL). The combined organic layers were dried over Na₂SO₄ and concentrated *in vacuo* to obtain a crude mixture, which was purified on silica gel column chromatography using 10% EtOAc in *n*-hexane as eluent. The yield was 650 mg (4.7 mmol) of sabina ketone (**12**, 99% yield, 86% ee, chemical purity 99%). $[\alpha]_D^{23} = +24.4$ (*c* 1.0, DCM); ¹H NMR (CDCl₃, 400 MHz) δ (in ppm) 2.14–2.06 (2H, m), 1.97–1.94 (2H, m), 1.63 (1H, dd, *J* = 8.8 and 2.8 Hz), 1.56 (1H, app quin, *J* = 6.8 Hz), 1.17 (1H, dd, *J* = 9.2 and 4.8 Hz), 1.06 (1H, dd, *J* = 4.8 and 3.2 Hz), 0.98 (3H, d, *J* = 6.8 Hz), 0.93 (3H, d, *J* = 6.8 Hz); ¹³C NMR (CDCl₃, 100 MHz) δ (in ppm) 214.9, 39.5, 33.7, 33.2, 32.2, 23.6, 19.5, 19.3, 19.1; GC-MS *t_R* 24.00 (5.9%), 24.42 (94.1%) (Figure S29, Supporting Information), *m/z* 81 (100%), 96, 95, 67, 55, 123, 41, 138 (M⁺), 109, 110 (decreasing order of intensity).

Synthesis of Sabinene Hydrates (4-Thujanols 13–16). To a solution of sabina ketone **12** (200 mg, 1.4 mmol) in anhydrous Et₂O (5 mL) was carefully added MeLi (1.8 mL, 2.8 mmol, 2.0 equiv, 1.6 M in Et₂O) at −78 °C. The mixture was stirred at the same temperature for 1 h and another 1 h at RT. The reaction was quenched by the addition of aqueous NH₄Cl (20 mL). The mixture was extracted with Et₂O (2 × 30 mL), before the organic phase was dried over Na₂SO₄, filtered, and concentrated *in vacuo* to afford a crude mixture (~91% conversion) of four sabinene hydrates (4-thujanols). The diastereomeric ratio was 90% *cis*-isomers [(−)-*cis*-(1*S*,4*S*)-4-thujanol (**13** plus its enantiomer **16**)] and 10% *trans*-isomers [(−)-*trans*-(1*S*,4*R*)-4-thujanol (**14** plus **15**)]. The *cis*- and *trans*-thujanol diastereomers could be separated by column chromatography on silica gel. Isolated yield of the *cis*-diastereomers was 150 mg (0.97 mmol). GC-MS: The retention times of the *trans*-isomers were (19.86, 20.02) and (21.08, 21.26) for the *cis*-isomers (Figure S29, Supporting Information).

(−)-*cis*-(1*S*,4*R*)-4-Thujanol (**13**). $[\alpha]_D^{23} = -40$ (*c* 0.5, DCM) (91% ee); ¹H NMR (CDCl₃, 400 MHz) δ (in ppm) 1.69 (1H, br s, OH), 1.61–1.49 (3H, m), 1.29 (3H, s), 1.31–1.28 (2H, m), 1.01 (1H, dd, *J* = 8.0 and 3.6 Hz), 0.89 (3H, d, *J* = 6.8 Hz), 0.84 (3H, d, *J* = 6.8 Hz),

0.62 (1H, dd, $J = 5.2$ and 3.6 Hz), 0.28 (1H, dd, $J = 8.0$ and 5.2 Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ (in ppm) 79.2, 36.0, 33.5, 33.1, 32.5, 27.9, 25.4, 19.5, 19.4, 11.1; GC-MS m/z 71 (100%), 111, 93, 81, 43, 139, 121, 79, 69, 55, 136, 154 (M^+) (decreasing order of intensity).

(+)-*trans*-(1*S*,4*S*)-4-Thujanol (**15**) (from Sigma-Aldrich, Schnelldorf, Germany). $[\alpha]_{\text{D}}^{23} = +29.8$ (c 0.5, DCM) (99% ee); ^1H NMR (CDCl_3 , 400 MHz) δ (in ppm) 1.88–1.80 (1H, m), 1.58 (1H, dd, $J = 12.0$ and 8.0 Hz), 1.52 (1H, dd, $J = 14.0$ and 8.4 Hz), 1.48–1.39 (2H, m), 1.29 (3H, s), 1.26 (1H, ddd, $J = 16.0, 5.6$ and 0.3 Hz), 1.07–1.03 (1H, ddd, $J = 8.4, 3.6$, and 1.2 Hz), 0.96 (3H, d, $J = 6.8$ Hz), 0.88 (3H, d, $J = 6.8$ Hz), 0.38 (1H, ddd, $J = 8.0, 5.2$, and 0.8 Hz), 0.20 (1H, dd, $J = 5.2$ and 3.6 Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ (in ppm) 80.5, 36.6, 34.6, 34.4, 32.2, 25.9, 24.9, 20.0, 19.9, 13.3; GC-MS m/z 71 (100%), 43, 93, 111, 81, 121, 139, 69, 55, 79, 136, 154 (M^+) (decreasing order of intensity).

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jnatprod.0c00669>.

GC chromatograms, ^{13}C NMR and ^1H NMR spectra (PDF)

GC chromatograms, ^{13}C NMR and ^1H NMR spectra (PDF)

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Notes

The authors declare no competing financial interest.

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