

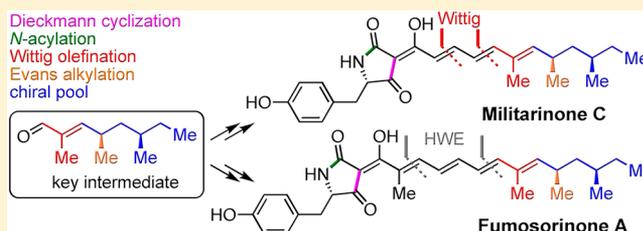
Synthesis of the Entomopathogenic Fungus Metabolites Militarinone C and Fumosorinone A

Sebastian Bruckner, Marie Weise, and Rainer Schobert*[✉]

Department of Chemistry, University Bayreuth, Universitaetsstrasse 30, 95440 Bayreuth, Germany

S Supporting Information

ABSTRACT: Militarinone C and fumosorinone A, 3-oligoenoyltetramic acids produced by insect pathogenic fungi, were synthesized for the first time. The pyrrolidine-2,4-dione ring was closed through a late-stage Dieckmann condensation of *N*-(β -ketoacyl) derivatives of tyrosine, obtained by its acylation with either thioesters or Meldrum's acid derivatives bearing the *all-trans*-polyene side chain. The latter was built up from (*S*)-citronellol via an Evans methylation and Wittig or HWE olefinations.



INTRODUCTION

In 2002 Hamburger et al. reported the isolation of militarinone A (**1**), a neurotrophic 2-pyridone alkaloid, from the entomogenous fungus *Paecilomyces militaris*.¹ Shortly after, they also identified two yellow tetramic acids, militarinone B (**2**) and militarinone C (**3**), as cometabolites (Figure 1).² It is

(PTP1B), a major negative regulator⁶ of the insulin signaling pathway. Such inhibitors are of interest as potential type II diabetes drugs since Klamann et al. had confirmed a higher sensitivity to insulin for mice deficient in PTP1B.⁷ Herein we report short syntheses that procure both compounds in quantities sufficient to study their conversion to 2-pyridones.

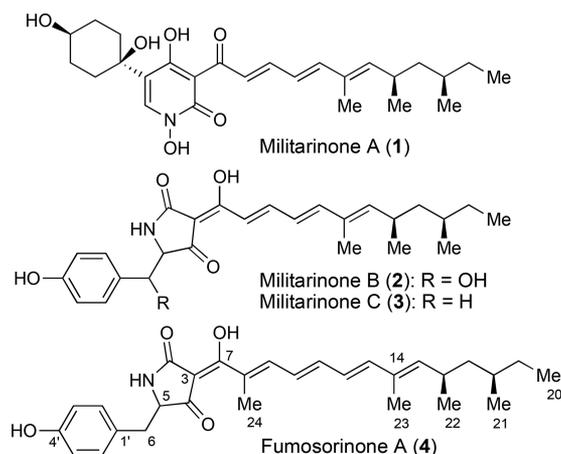


Figure 1. Structures of militarinones A–C (**1–3**) and fumosorinone A (**4**).

RESULTS AND DISCUSSION

The retrosynthetic approach is outlined in Scheme 1. Both target compounds **3** and **4** were finished by a Dieckmann cyclization⁸ of the respective functionalized β -ketoamide **5** or **6** followed by *N,O*-deprotection. These β -ketoamides were obtained by reaction of *N,O*-bisprotected methyl tyrosinates **7** with either Meldrum's acid derivative **8** or thioester **9** as *N*-acylating agents carrying the respective unsaturated side chain. The β -ketoesters **8** and **9** were accessible through consecutive Wittig or HWE olefinations of key aldehyde **10** which was built up from (*S*)-citronellol (**11**) using an Evans alkylation⁹ to introduce the second methyl group and an *E*-selective Wittig olefination to establish the trisubstituted double bond.

(*S*)-Citronellol (**11**) was first converted to imide **15** following a modified route by Nishida et al.¹⁰ (Scheme 2). It was quantitatively deoxygenated to alkene **13** in two steps via mesylation to give **12** which was reduced with LiAlH₄. Olefin **13** was subjected to a ruthenium-catalyzed oxidative cleavage according to a general procedure by Sharpless et al.¹¹ which afforded carboxylic acid **14**. This was converted to a mixed anhydride with pivaloyl chloride which was reacted with (*R*)-4-benzyloxazolidin-2-one to yield imide **15**. Its methylation at -78 °C gave the desired (*R,R*)-product **16** as a separable mixture of two diastereomers. Removal of the Evans auxiliary with LiBH₄/MeOH at 0 °C left enantiopure alcohol **17**. This was Swern oxidized to aldehyde **18** which was Wittig olefinated

not uncommon that fungi produce mixtures of tyrosine-derived tetramic acids and 2-pyridones, e.g., the family of torrubiellones, metabolites of the fungus *Torrubiella* sp. BCC 2165,³ or the (proto)tenellins, produced by the insect pathogenic fungus *Beauveria bassiana*. For the latter, Cox et al. established a radical oxidation–rearrangement conversion of the tetramic acid prototenellin D to the 2-pyridone tenellin.⁴ In 2017, Zhang et al.⁵ isolated fumosorinone A (**4**) from the entomogenous fungus *Isaria fumosorosea* and found it to inhibit (IC₅₀ 3.24 μ M) protein tyrosine phosphatase 1B

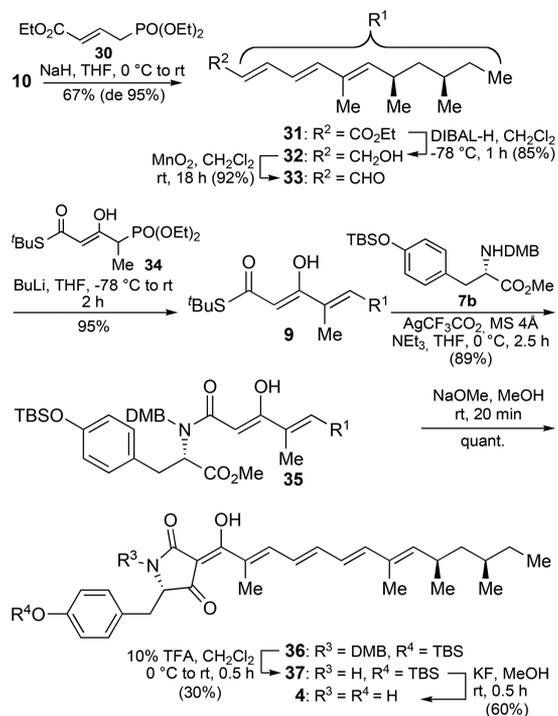
Received: June 18, 2018

Published: July 23, 2018

penicillinol A₂¹⁴ and (–)-hymenosein,¹⁵ our synthesis of militarinone C supports, yet does not prove, the absolute configuration proposed in the literature for the natural product. An unambiguous proof would, for instance, require a comparison of circular dichroism spectra¹⁵ of natural and synthetic samples (for an ECD spectrum of synthetic militarinone C (3) cf. Supporting Information).

For the synthesis of fumosorinone A (4), aldehyde 10 was submitted to a HWE olefination with phosphonate 30, followed by a DIBAL-H reduction of product ester 31 to alcohol 32 and its oxidation with MnO₂ to aldehyde 33, analogously to Dash et al.¹⁶ (Scheme 4). Another HWE

Scheme 4. Synthesis of Fumosorinone A (4)



olefination with phosphonate 34, according to Loscher et al.,¹⁷ afforded thioester 9 in excellent 95% yield as a 2:3 keto/enol mixture. It was used to acylate aminoester 7b in a surprisingly good yield of 89% according to Ley's silver(I)-mediated aminolysis protocol.¹⁸ The resulting β -ketoamide 35 was cyclized quantitatively under mild conditions to give doubly protected fumosorinone A 36. Due to its acid sensitivity, it had to be deprotected in two steps. Treatment with 10% TFA for only 30 min allowed the isolation of 30% TBS-protected fumosorinone A 37 aside of 30% recovered 36. Desilylation of all accumulated 37 with KF in methanol finally yielded fumosorinone A (4) in 60% after semipreparative HPLC. It proved identical to the natural isolate in terms of NMR spectra (cf. Supporting Information Table S2) and also specific optical rotations ($[\alpha]_{\text{D}}^{24}$ –229 (*c* 0.20, CH₃OH) for synthetic and $[\alpha]_{\text{D}}^{20}$ –207 (*c* 0.1, CH₃OH) as reported for natural 4).

CONCLUSIONS

In summary, fumosorinone A (4) and militarinone C (3) were each prepared in 18 steps and ca. 2% yield by N-acylating L-tyrosine esters with thioesters or 5-enoyl Meldrum's acids carrying the polyene side chains, followed by Dieckmann cyclization of the resulting β -ketoamides. The side chains were

built up from (S)-citronellol via an Evans methylation and consecutive Wittig or HWE olefinations. The agreement (good in the case of 4, reasonable for 3) between NMR spectra and optical rotations of our synthetic products and those reported for the natural isolates at least does not rule out the origin of the latter from L-tyrosine. Studies of the conversion of compounds 3 and 4 to the respective 2-pyridones by means of radical oxidants are already underway.

EXPERIMENTAL SECTION

General Remarks. IR spectra were recorded with an FT-IR spectrophotometer equipped with an ATR unit. ¹H NMR and ¹³C NMR spectra were obtained using a 500 MHz spectrometer. Chemical shifts are given in parts per million using the residual solvent peak as an internal standard 7.26 ppm (proton) and 77.16 ppm (carbon) for CDCl₃, 3.31 ppm (proton), and 47.60 ppm (carbon) for CD₃OD and 2.50 ppm (proton) and 39.51 ppm (carbon) for DMSO-*d*₆. Coupling constants (*J*) are quoted in hertz (Hz). Multiplicity abbreviation used: s singlet, d doublet, t triplet, q quartet, m multiplet, br broad. High resolution mass spectra were obtained with a UPLC/Orbitrap MS system in ESI mode. Optical rotations were measured at 589 nm (Na-D line). Photolysis was performed using a Pro Collect UV tester with 366 nm and 4 W.

Chemicals. All reagents were purchased from commercial sources and were used without further purification. All anhydrous solvents were used as supplied, except tetrahydrofuran, diethyl ether, and dichloromethane which were freshly distilled according to standard procedures. Reactions were routinely carried out under an argon atmosphere unless stated otherwise. All glassware was flame-dried before use.

Chromatography. Analytical thin layer chromatography was carried out using Merck silica gel 60GF₂₅₄ precoated aluminum-backed plates and/or Merck 60 RP-18 F_{254S} foil plates. The compounds were visualized with UV light (254 nm and/or 360 nm) and/or ceric ammonium molybdate (CAM) and/or potassium permanganate and/or iodine on silica. Flash chromatography was performed at medium pressure using dry-packed Marchery-Nagel silica gel 60, pore size 40–63 μ m, with the eluent specified. Analytical HPLC measurements were performed on a Beckman System Gold Programmable Solvent Module 126 using a Phenomenex Kinetex C-18-HPLC column, length 250 \times 4.6 mm, pore size 100 \AA , particle size 5 μ m. Detection was by a Beckman Instruments Diode Array Detection Module 168. MPLC reversed phase chromatography was performed using a Büchi MPLC system with a "MN Polyproprep 100–50 C 18 end-capped" column, length 460 mm, diameter 49 mm. Detection was by a Büchi UV Photometer C-635. Semipreparative reversed phase HPLC was performed using an Amersham Biosciences ÄKTAbasic10 system with a Phenomenex Gemini-NX Su C18 110A, 250 \times 10.00 mm column. Detection was by an Amersham Biosciences ÄKTA UV-900 module.

Militarinone C (3). A solution of protected tetramic acid 28b (107 mg, 156 μ mol) in CH₂Cl₂ (50 mL) was cooled to 0 °C and treated dropwise with 20% trifluoroacetic acid in CH₂Cl₂ (50 mL). The resulting mixture was stirred at ambient temperature for 1 h, sat. aqueous phosphate buffer (pH 7, 50 mL) was added, and the phases were separated. The organic phase was washed with the same buffer (2 \times 50 mL) and aqueous KHSO₄ (5% wt, 50 mL) and then dried (Na₂SO₄) and concentrated in vacuo to give a mixture of O-TBS-protected tetramic acid 29 and militarinone C (3) as a yellow oil (93 mg) that was used in the next step without further purification.

The crude mixture of 29 and 3 was taken up in methanol p.a. (6 mL), a 10 M suspension of KF in methanol p.a. (624 μ L, 6.24 mmol) was added, and the mixture was stirred at ambient temperature for 1 h. A 1 M aqueous HCl (20 mL) solution and brine (50 mL) were added, and the mixture was extracted with EtOAc (2 \times 125 mL). The combined organic phases were dried (Na₂SO₄) and concentrated in vacuo to give a yellow oil which was purified by MPLC on an RP-18 column, eluting with 75% methanol in H₂O (with 0.1% formic acid) to 95% methanol in 10 min with a flow rate of 240 mL/min. The

product-containing fractions were collected, the methanol was removed in vacuo, and the aqueous phase was extracted with EtOAc (2 × 100 mL). The combined organic phases were washed with 1 M aqueous HCl (20 mL), dried (Na₂SO₄), and concentrated in vacuo to give militarinone C (3) as an orange-yellow solid foam (40.9 mg, 62% over two steps); [α]_D²⁴ −310 (c 0.30, MeOH) (lit.² [α]_D²⁴ −430.2 (c 0.17, MeOH)); IR ν_{\max} 3284, 2959, 2923, 1587, 1551, 1515, 1463, 1429, 1373, 1226, 1172, 1105, 1031, 1000, 895, 868, 822, 733, 626 cm^{−1}; for NMR data cf. Supporting Information Table S1; HRMS (ESI) *m/z* [M + H]⁺ calcd for C₂₆H₃₄NO₄⁺ 424.2482, found 424.2477.

Fumosorinone A (4). A solution of tetramic acid 36 (72 mg, 99 μ mol) in CH₂Cl₂ (36 mL) was cooled to 0 °C and treated dropwise with 20% trifluoroacetic acid in CH₂Cl₂ (36 mL), and the mixture was stirred at ambient temperature for 30 min. Saturated aqueous phosphate buffer (pH 7, 100 mL) was added at 0 °C, the phases were separated, and the organic phase was washed with the same buffer (2 × 100 mL) and 1 M aqueous HCl (50 mL), dried (Na₂SO₄), and concentrated in vacuo to afford an orange-yellow oil. It was purified by flash chromatography on RP-18 silica gel, eluting with 95% acetonitrile in H₂O to give O-TBS-protected tetramic acid 37 (16 mg, 30%) and residual starting material 36 (22 mg, 30%); R_f = 0.36 (8% MeOH in CH₂Cl₂). A solution of 37 (16 mg, 28 μ mol) in MeOH (1.8 mL) was treated with a 10 M suspension of potassium fluoride in MeOH (199 μ L, 1.99 mmol), and the mixture was stirred at ambient temperature for 30 min. Saturated aqueous NH₄Cl (20 mL) and 1 M aqueous HCl (10 mL) were added, and the mixture was extracted with EtOAc (2 × 50 mL). The combined organic phases were dried (Na₂SO₄) and concentrated in vacuo to leave an orange-red oil which was filtered over Sephadex LH-20 (MeOH) to afford an orange-red oil upon evaporation. The oil was further purified by semipreparative HPLC (AKTA system, flow rate: 5 mL/min on a Phenomenex Gemini-NX 5u C18 110A, 250 × 10.00 mm column, one column volume (CV) at 70% MeCN in H₂O (with 0.1% formic acid), then three CV at 90% MeCN, *t*_{ret} = 11.7–12.6 min, UV_{det} = 414 nm) to give fumosorinone A (4) as a bright orange-yellow oil (7.7 mg, 60%); [α]_D²⁴ −229 (c 0.20, MeOH) (lit.⁵ [α]_D²⁴ −207 (c 0.1, MeOH)); IR ν_{\max} 3310, 2960, 2925, 1650, 1591, 1516, 1442, 1261, 1171, 988, 812, 620 cm^{−1}; for NMR data cf. Supporting Information Table S2; HRMS (ESI) *m/z* [M + H]⁺ calcd for C₂₉H₃₈NO₄⁺ 464.2795, found 464.2785.

Methyl (S)-3-(4-((tert-Butyldimethylsilyloxy)phenyl)-2-((2-nitrobenzyl)amino)propanoate (7a). A solution of L-tyrosine methyl ester hydrochloride (1.16 g, 5.00 mmol) in 3% acetic acid in methanol (100 mL) was treated with o-nitrobenzaldehyde (1.51 g, 10.00 mmol) and MS 3 Å (100 mg), and the resulting mixture was stirred at ambient temperature for 1 h. NaBH₃CN (781 mg, 12.50 mmol) was added, and stirring was continued for 2 h. The molecular sieves were filtered off, and the reaction mixture was quenched with sat. aqueous NaHCO₃ (300 mL). Ethyl acetate (300 mL) was added, the phases were separated, and the organic phase was washed with brine (200 mL), dried (Na₂SO₄), and concentrated in vacuo to give a yellowish oil that was adsorbed on silica gel (wt ratio oil/silica 1:10) and purified by flash chromatography (silica gel, 1% MeOH in CH₂Cl₂ ⇒ 1.5% MeOH ⇒ 2% MeOH) to give oNb-L-Tyr-OMe as a yellow oil (1.145 g, 69%); R_f = 0.30 (4% MeOH in CH₂Cl₂); [α]_D²⁴ +33.6 (c 1.00, CHCl₃); IR ν_{\max} 3324, 2953, 1732, 1613, 1596, 1578, 1516, 1444, 1344, 1206, 1173, 1107, 991, 829, 789, 731, 702, 666, 556 cm^{−1}; ¹H NMR (CDCl₃, 500 MHz) δ 7.90 (dd, *J* = 1.1, 8.1 Hz, 1H), 7.49 (ddd, *J* = 1.1, 7.2, 7.3 Hz, 1H), 7.46 (dd, *J* = 1.5, 7.3 Hz, 1H), 7.36 (ddd, *J* = 1.5, 7.2, 8.1 Hz, 1H), 6.96 (d, *J* = 8.5 Hz, 2H), 6.69 (d, *J* = 8.5 Hz, 2H), 4.08 (d, *J* = 15.0 Hz, 1H), 3.93 (d, *J* = 15.0 Hz, 1H), 3.64 (s, 3H), 3.47 (dd, *J* = 6.1, 7.3 Hz, 1H), 2.91 (dd, *J* = 6.1, 13.4 Hz, 1H), 2.86 (dd, *J* = 7.3, 13.4 Hz, 1H); ¹³C NMR (CDCl₃, 125 MHz) δ 175.0, 155.0, 149.0, 134.9, 133.3, 131.2, 130.4, 128.5, 128.2, 124.9, 115.5, 62.6, 52.0, 49.2, 38.8; HRMS (ESI) *m/z* [M + H]⁺ calcd for C₁₇H₁₉N₂O₅⁺ 331.1288, found 331.1284.

A solution of oNb-L-Tyr-OMe (610 mg, 1.85 mmol) in CH₂Cl₂ p.a. (19 mL) was cooled to 0 °C and treated with imidazole (378 mg, 5.55 mmol) and TBSCl (613 mg, 4.07 mmol). The resulting mixture was

stirred for 19 h while reaching room temperature. The mixture was filtered, the filtrate was taken up in CH₂Cl₂ (50 mL), the organic phase was washed with sat. aqueous NH₄Cl (100 mL), and the aqueous phase was extracted with CH₂Cl₂ (50 mL). The combined organic layers were washed with brine (150 mL), dried (Na₂SO₄), and concentrated in vacuo to give a yellow oil that was purified by flash chromatography (silica gel, 12% ethyl acetate in hexane) to afford 7a as a yellow oil (746 mg, 91%); R_f = 0.74 (hexane/EtOAc 1:1); [α]_D²⁴ +26.8 (c 1.00, CHCl₃); IR ν_{\max} 2954, 2931, 2858, 1737, 1609, 1580, 1527, 1510, 1471, 1444, 1346, 1255, 1200, 1170, 1131, 1105, 1007, 914, 840, 782, 729, 691, 668 cm^{−1}; ¹H NMR (CDCl₃, 500 MHz) δ 7.91 (d, *J* = 8.2 Hz, 1H), 7.45–7.54 (m, 2H), 7.34–7.41 (m, 1H), 7.00 (d, *J* = 8.5 Hz, 2H), 6.74 (d, *J* = 8.5 Hz, 2H), 4.09 (d, *J* = 15.0 Hz, 1H), 3.92 (d, *J* = 15.0 Hz, 1H), 3.63 (s, 3H), 3.45 (dd, *J* = 6.4, 7.3 Hz, 1H), 2.91 (dd, *J* = 6.4, 13.7 Hz, 1H), 2.86 (dd, *J* = 7.3, 13.7 Hz, 1H), 2.07 (br. s, 1H), 0.97 (s, 9H), 0.18 (s, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 174.9, 154.6, 149.1, 135.3, 133.2, 130.9, 130.3, 129.8, 128.0, 124.8, 120.1, 62.7, 51.9, 49.1, 39.1, 25.8, 18.3, −4.3; HRMS (ESI) *m/z* [M + H]⁺ calcd for C₂₃H₃₃N₂O₅Si⁺ 445.2153, found 445.2137.

Methyl (S)-3-(4-((tert-Butyldimethylsilyloxy)phenyl)-2-((2,4-dimethoxybenzyl)amino)propanoate (7b). According to a modified literature procedure,¹⁹ a suspension of L-tyrosine methyl ester hydrochloride (580 mg, 2.50 mmol) in CH₂Cl₂ (12 mL) was treated with imidazole (510 mg, 15.00 mmol) and TBSCl (452 mg, 6.00 mmol). The resulting mixture was stirred at room temperature for 19 h. Saturated aqueous NaHCO₃ (50 mL) was added, and the jellylike mixture was extracted with CH₂Cl₂ (3 × 75 mL). The combined organic phases were washed with H₂O (50 mL), dried (MgSO₄), and concentrated in vacuo to give an oil that was purified by flash chromatography (silica gel, 90% ethyl acetate in hexane) to afford L-Tyr(OTBS)-OMe as a clear oil (479 mg, 77%); R_f = 0.24 (hexane/EtOAc 1:4); [α]_D²⁴ +10.0 (c 1.00, CHCl₃); IR ν_{\max} 2954, 2931, 2893, 2858, 1739, 1609, 1509, 1472, 1464, 1438, 1252, 1195, 1169, 1109, 1102, 1008, 911, 837, 802, 779, 688 cm^{−1}; ¹H NMR (CDCl₃, 500 MHz) δ 7.03 (d, *J* = 8.5 Hz, 2H), 6.77 (d, *J* = 8.5 Hz, 2H), 3.70 (s, 3H), 3.68 (dd, *J* = 5.2, 7.6 Hz, 1H), 3.00 (dd, *J* = 5.2, 13.7 Hz, 1H), 2.80 (dd, *J* = 7.6, 13.7 Hz, 1H), 1.45 (br. s., 2H), 0.97 (s, 9H), 0.18 (s, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 175.7, 154.7, 130.3, 129.9, 120.3, 56.1, 52.1, 40.5, 25.8, 18.3, −4.3.

A solution of L-Tyr(OTBS)-OMe (881 mg, 2.85 mmol) in 3% acetic acid in methanol (10 mL) was treated with 2,4-dimethoxybenzaldehyde (450 mg, 2.71 mmol), and the mixture was stirred at room temperature for 30 min. NaBH(OAc)₃ (804 mg, 3.79 mmol) was added, stirring continued for 1.5 h, and the reaction mixture was quenched with sat. aqueous NaHCO₃ (50 mL). The mixture was extracted with ethyl acetate (3 × 75 mL), and the combined organic phases were washed with brine (75 mL), dried (Na₂SO₄), and concentrated in vacuo to give an oil that was purified by flash chromatography (silica gel, 15% EtOAc with 0.5% NEt₃ in hexane ⇒ 30% EtOAc with 0.5% NEt₃) to leave 7b as a clear oil (870 mg, 71%); R_f = 0.68 (hexane/EtOAc 1:1); [α]_D²⁴ +1.98 (c 1.00, CHCl₃); IR ν_{\max} 2952, 2931, 2858, 1735, 1611, 1589, 1508, 1463, 1438, 1418, 1278, 1250, 1207, 1156, 1132, 1104, 1036, 911, 835, 797, 779, 688, 634 cm^{−1}; ¹H NMR (CDCl₃, 500 MHz) δ 7.02 (d, *J* = 8.9 Hz, 1H), 6.99 (d, *J* = 8.2 Hz, 2H), 6.73 (d, *J* = 8.2 Hz, 2H), 6.36–6.40 (m, 2H), 3.78 (s, 3H), 3.68 (s, 3H), 3.58 (s, 3H), 3.45 (t, *J* = 7.2 Hz, 1H), 2.89 (dd, *J* = 6.7, 13.4 Hz, 1H), 2.85 (dd, *J* = 7.6, 13.4 Hz, 1H), 1.87–2.05 (m, 1H), 0.97 (s, 9H), 0.17 (s, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 175.2, 160.2, 158.7, 154.5, 130.5, 130.21, 130.17, 120.2, 120.1, 103.7, 98.5, 62.4, 55.5, 55.3, 51.7, 47.3, 39.1, 25.8, 18.3, −4.3.

S-(tert-Butyl) (2Z,4E,6E,8E,10E,12R,14R)-3-Hydroxy-4,10,12,14-tetramethylhexadeca-2,4,6,8,10-pentaenethioate (9). Following a general literature protocol,¹⁷ thioester 9 (236 mg, 95%) was prepared from phosphonate 34 (289 mg, 889 μ mol) and aldehyde 33 (140 mg, 635 μ mol) as an orange-yellow oil and as a 2:3 keto/enol mixture; R_f = 0.86 (10% EtOAc in hexane); [α]_D²⁴ −43.2 (c 0.50, CHCl₃); IR ν_{\max} 2960, 2923, 2871, 1688, 1651, 1614, 1586, 1456, 1376, 1364, 1311, 1250, 1163, 1100, 1062, 986, 907, 859, 797, 769, 652 cm^{−1}; ¹H NMR (CDCl₃, 500 MHz) δ 12.96 (s, 1H), 7.15 (d, *J* = 11.0 Hz, 1H),

7.11 (d, $J = 11.0$ Hz, 1H), 6.22–6.75 (m, 8H), 5.57 (s, 1H, HCCOS enol), 5.41 (d, $J = 9.8$ Hz, 1H), 5.36 (d, $J = 9.8$ Hz, 1H), 3.88 (s, 2H, H₂CCOS keto), 2.56–2.69 (m, 2H), 1.92 (s, 3H), 1.86 (d, $J = 0.9$ Hz, 3H), 1.81 (d, $J = 0.9$ Hz, 3H), 1.80 (d, $J = 0.9$ Hz, 3H), 1.53 (s, 9H), 1.47 (s, 9H), 1.21–1.35 (m, 6H), 1.05–1.17 (m, 4H), 0.96 (d, $J = 6.4$ Hz, 3H), 0.95 (d, $J = 6.4$ Hz, 3H), 0.80–0.88 (m, 12H); ¹³C NMR (CDCl₃, 125 MHz) δ 196.2, 193.6, 193.3, 169.6, 144.3, 143.2, 142.4, 141.8, 141.5, 139.9, 134.7, 132.6, 127.3, 127.2, 127.0, 126.4, 126.0, 98.1, 54.1, 49.0, 48.5, 45.0, 44.9, 32.5, 32.4, 30.84, 30.76, 30.35, 30.27, 30.25, 29.8, 21.63, 21.56, 19.2, 12.6, 12.4, 11.8, 11.5; HRMS (ESI) m/z [M + Na]⁺ calcd for C₂₄H₃₈O₂Na⁺ 413.2485, found 413.2482.

(4*R*,6*R*,*E*)-2,4,6-Trimethyloct-2-enal (10). A solution of alcohol 21 (1.60 g, 9.37 mmol) in CH₂Cl₂ p.a. (94 mL) was treated with MnO₂ (29.48 g, 327.95 mmol), and the mixture was stirred at ambient temperature for 18 h and then filtered over Celite. The filtrate was concentrated in vacuo to give a clear liquid that was purified by flash chromatography (silica gel, 10% ethyl acetate in hexane) to afford aldehyde 10 (1.31 g, 83%) as a clear oil; $R_f = 0.38$ (6% ethyl acetate in hexane); $[\alpha]_D^{24} -36.3$ (c 1.00, CHCl₃) (lit.²⁰ $[\alpha]_D^{20} -43$ (c 0.75, CHCl₃)); IR ν_{\max} 2961, 2928, 2875, 2707, 1688, 1644, 1457, 1405, 1379, 1313, 1243, 1201, 1128, 1051, 1015, 874, 827, 805, 675 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 9.38 (s, 1H), 6.21 (dd, $J = 1.2, 10.1$ Hz, 1H), 2.75–2.86 (m, 1H), 1.72–1.78 (m, 3H), 1.10–1.39 (m, 5H), 1.03 (d, $J = 6.7$ Hz, 3H), 0.81–0.87 (m, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 195.8, 161.0, 138.0, 44.1, 32.5, 31.4, 30.1, 20.5, 19.2, 11.4, 9.5.

(*S*)-3,7-Dimethyloct-6-en-1-yl Methanesulfonate (12). According to a literature procedure,²¹ compound 12²² (4.67 g, 100%) was prepared from (*S*)-citronellol (11) (3.63 mL, 20.00 mmol), MsCl (1.6 mL, 21.00 mmol), and NEt₃ (2.9 mL, 21.00 mmol) as a yellow oil; $R_f = 0.55$ (hexane/EtOAc 5:1); $[\alpha]_D^{24} -2.11$ (c 1.00, CHCl₃); IR ν_{\max} 2964, 2914, 2859, 1456, 1378, 1351, 1333, 1171, 1036, 973, 938, 889, 818, 796, 730 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 5.04–5.11 (m, 1H), 4.21–4.32 (m, 2H), 3.00 (s, 3H), 1.90–2.07 (m, 2H), 1.75–1.85 (m, 1H), 1.68 (s, 3H), 1.60 (s, 3H), 1.50–1.65 (m, 2H), 1.30–1.40 (m, 1H), 1.15–1.25 (m, 1H), 0.93 (d, $J = 6.7$ Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 131.8, 124.4, 68.7, 37.5, 36.9, 36.0, 29.1, 25.9, 25.4, 19.3, 17.8.

(*R*)-2,6-Dimethyloct-2-ene (13). According to a literature protocol,²³ compound 13 (6.13 g, 100%) was prepared from 12 (10.30 g, 43.95 mmol) and LiAlH₄ (3.34 g, 88.00 mmol) as a clear oil; $R_f = 0.89$ (hexane); $[\alpha]_D^{24} -6.2$ (c 1.00, CHCl₃) (lit.²⁴ $[\alpha]_D^{25} -7.55$ (neat)); IR ν_{\max} 2963, 2915, 2875, 2853, 1456, 1377, 833 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 5.08–5.14 (m, 1H), 1.89–2.04 (m, 2H), 1.69 (s, 3H), 1.61 (s, 3H), 1.28–1.39 (m, 3H), 1.08–1.18 (m, 2H), 0.86 (d, $J = 6.3$ Hz, 3H), 0.86 (t, $J = 7.3$ Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 131.1, 125.3, 36.9, 34.2, 29.6, 25.9, 25.8, 19.3, 17.8, 11.5.

(*R*)-4-Methylhexanoic Acid (14). According to a modified literature protocol,¹¹ a solution of alkene 13 (6.15 g, 43.65 mmol) in CH₂Cl₂ p.a. (130 mL) was treated with acetonitrile p.a. (130 mL), H₂O (170 mL), NaO₄ (37.35 g, 174.60 mmol), and RuCl₃ × H₂O (181 mg, 873 μmol, 2 mol %), and the mixture was stirred at ambient temperature for 18 h. Na₂S₂O₃ (3 g) was added, and the mixture was stirred for 15 min. The solids were filtered off, and the filtrate was concentrated in vacuo. A 1 M aqueous NaOH (100 mL) solution was added, and the resulting mixture was washed with diethyl ether (2 × 200 mL). The aqueous phase was acidified with 1 M aqueous HCl (175 mL), and the brown mixture was extracted with diethyl ether (3 × 200 mL). The combined organic phases were washed with aqueous Na₂S₂O₃ (20% wt, 2 × 150 mL) and brine (150 mL), dried (Na₂SO₄), and concentrated in vacuo to give acid 14 (3.66 g, 64%) as a brownish oil; $R_f = 0.61$ (hexane/EtOAc 4:1); $[\alpha]_D^{24} -10.9$ (c 1.00, CHCl₃) (lit.²⁵ $[\alpha]_D^{10} -10.1$ (c 1.00, CHCl₃)); IR ν_{\max} 3042, 2962, 2931, 2876, 2654, 1705, 1463, 1413, 1380, 1281, 1250, 1216, 936 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 11.45 (br. s, 1H), 2.27–2.44 (m, 2H), 1.63–1.74 (m, 1H), 1.40–1.50 (m, 1H), 1.30–1.40 (m, 2H), 1.12–1.22 (m, 1H), 0.88 (d, $J = 6.4$ Hz, 3H), 0.88 (t, $J = 7.3$ Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 180.6, 34.1, 32.0, 31.3, 29.2, 18.9, 11.4.

(*R*)-4-Benzyl-3-((*R*)-4-methylhexanoyl)oxazolidin-2-one (15). According to a literature procedure,⁹ compound 15 (526 mg, 88%) was prepared from 14 (268 mg, 2.06 mmol), pivaloyl chloride (0.27 mL, 2.16 mmol), and (*R*)-benzyloxazolidin-2-one (383 mg, 2.16 mmol) as a colorless solid; $R_f = 0.64$ (hexane/EtOAc 4:1); $[\alpha]_D^{24} -56.9$ (c 1.00, CHCl₃) (lit.²⁶ $[\alpha]_D^{27} -57.5$ (c 1.00, CHCl₃)); mp 36–37 °C; IR ν_{\max} 2961, 2925, 2874, 1777, 1698, 1455, 1384, 1351, 1324, 1280, 1210, 1194, 1138, 1097, 1052, 1015, 761, 741, 700, 628, 595, 564 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 7.31–7.36 (m, 2H), 7.27–7.30 (m, 1H), 7.19–7.23 (m, 2H), 4.64–4.71 (m, 1H), 4.20 (dd, $J = 7.6, 9.0$ Hz, 1H), 4.16 (dd, $J = 3.0, 9.0$ Hz, 1H), 3.30 (dd, $J = 3.3, 13.3$ Hz, 1H), 2.99 (ddd, $J = 5.4, 10.0, 16.7$ Hz, 1H), 2.89 (ddd, $J = 5.7, 9.8, 16.7$ Hz, 1H), 2.76 (dd, $J = 9.6, 13.3$ Hz, 1H), 1.66–1.76 (m, 1H), 1.47–1.55 (m, 1H), 1.35–1.47 (m, 2H), 1.15–1.24 (m, 1H), 0.92 (d, $J = 6.4$ Hz, 3H), 0.90 (t, $J = 7.2$ Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 173.9, 153.6, 135.5, 129.6, 129.1, 127.5, 66.3, 55.3, 38.1, 34.1, 33.5, 31.0, 29.4, 19.1, 11.5.

(*R*)-4-Benzyl-3-((2*R*,4*R*)-2,4-dimethylhexanoyl)oxazolidin-2-one (16). According to a literature procedure,⁹ compound 16 (5.63 g, 90%) was prepared from 15 (5.98 g, 20.65 mmol) as a colorless solid of mp 32–33 °C; $R_f = 0.54$ (hexane/EtOAc 7:1); $[\alpha]_D^{24} -66.6$ (c 1.00, CHCl₃) (lit.²⁷ $[\alpha]_D^{27} -68.6$ (c 4.13, CHCl₃)); IR ν_{\max} 2962, 2929, 2875, 1775, 1695, 1455, 1384, 1349, 1290, 1239, 1207, 1098, 1075, 1052, 1015, 971, 917, 761, 739, 701, 625, 593 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 7.30–7.36 (m, 2H), 7.26–7.30 (m, 1H), 7.19–7.24 (m, 2H), 4.65–4.71 (m, 1H), 4.20 (dd, $J = 7.4, 9.0$ Hz, 1H), 4.17 (dd, $J = 2.8, 9.0$ Hz, 1H), 3.82–3.92 (m, 1H), 3.26 (dd, $J = 3.1, 13.3$ Hz, 1H), 2.76 (dd, $J = 9.6, 13.3$ Hz, 1H), 1.85 (ddd, $J = 5.5, 8.6, 13.2$ Hz, 1H), 1.29–1.43 (m, 2H), 1.22 (d, $J = 6.8$ Hz, 3H), 1.09–1.20 (m, 2H), 0.86 (d, $J = 6.4$ Hz, 3H), 0.88 (t, $J = 7.2$ Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 177.6, 153.2, 135.5, 129.6, 129.1, 127.5, 66.1, 55.5, 40.6, 38.0, 35.5, 32.4, 29.6, 19.5, 18.5, 11.4.

(2*R*,4*R*)-2,4-Dimethylhexan-1-ol (17). According to a literature procedure,²⁸ alcohol 17 (2.11 g, 95%) was prepared from 16 (5.19 g, 17.10 mmol) and LiBH₄ (4 M in THF, 4.9 mL, 19.67 mmol) as a clear oil; $R_f = 0.33$ (hexane/EtOAc 5:1); $[\alpha]_D^{24} +4.7$ (c 1.00, CHCl₃) (lit. $[\alpha]_D +3.7$ (c 1.67, CHCl₃)); IR ν_{\max} 3342, 2959, 2915, 2875, 1462, 1378, 1028, 986, 612 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 3.48–3.56 (m, 1H), 3.34–3.42 (m, 1H), 1.66–1.76 (m, 1H), 1.40–1.48 (m, 1H), 1.35–1.40 (m, 1H), 1.24–1.35 (m, 2H), 1.03–1.13 (m, 1H), 0.92 (d, $J = 6.7$ Hz, 3H), 0.90 (s, 1H), 0.87 (d, $J = 6.5$ Hz, 3H), 0.86 (t, $J = 7.3$ Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 68.6, 40.7, 33.3, 31.7, 29.2, 19.9, 17.4, 11.3.

Triphenyl(1-ethoxycarbonyl)ethyl)phosphorane (19). According to literature procedure,²⁹ ylide 19³⁰ (10.13 g, 50.00 mmol, 64%) was prepared as a beige solid from 2-bromoethyl propanoate (6.5 mL, 50.00 mmol) and PPh₃ (13.12 g, 50.00 mmol); ¹H NMR (CDCl₃, 500 MHz) δ 7.44–7.63 (m, 15H), 3.56–4.00 (m, 2H), 1.61 (d, $J = 13.7$ Hz, 3H), 0.22–0.92 (m, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 133.8, 133.7, 131.7, 128.6, 128.5, 57.6, 12.8; IR ν_{\max} 3055, 2979, 2927, 1626, 1589, 1571, 1484, 1435, 1381, 1365, 1310, 1298, 1185, 1158, 1087, 1071, 1029, 998, 950, 861, 773, 760, 747, 714, 692, 618, 576 cm⁻¹.

Ethyl (4*R*,6*R*,*E*)-2,4,6-Trimethyloct-2-enoate (20). According to a modified literature procedure,¹² a cooled (–78 °C) solution of oxalyl chloride (2.2 mL, 25.87 mmol) in CH₂Cl₂ (27 mL) was treated with a solution of dimethyl sulfoxide (2.9 mL, 40.43 mmol) in CH₂Cl₂ (13.5 mL), and the mixture was stirred at –78 °C for 30 min. A solution of alcohol 17 (2.11 g, 16.17 mmol) in CH₂Cl₂ (13.5 mL) was added, and stirring was continued at –78 °C for 30 min. NEt₃ (11.2 mL, 80.85 mmol) was added, and the mixture was stirred at –78 °C for 1 h before it was warmed to ambient temperature and stirred for 1 h. Then 1 M aqueous HCl (100 mL) was added, and the mixture was extracted with *n*-pentane (3 × 100 mL). The combined organic phases were washed with H₂O (100 mL), dried (Na₂SO₄), and concentrated in vacuo to give aldehyde 18 as a pale-yellow liquid that was used in the next step without purification.

A solution of aldehyde 18 (2.07 g, 16.17 mmol) in CH₂Cl₂ p.a. (35 mL) was treated with Ph₃P=C(CH₃)CO₂Et (19) (8.79 g, 24.26 mmol), and the mixture was stirred at ambient temperature for 19 h.

The solvent was removed in vacuo to give a brownish oil that was purified by flash chromatography (silica gel, 2% diethyl ether in hexane) to give ester **20** (2.41 g, 70% over two steps) as an 98:2 mixture of *E/Z* isomers as a pale yellow liquid; $R_f = 0.34$ (2% diethyl ether in hexane); IR ν_{\max} 2961, 2929, 2875, 1710, 1650, 1462, 1367, 1311, 1271, 1249, 1217, 1152, 1099, 1035, 991, 750 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 6.50 (d, $J = 9.8$ Hz, 1H), 4.18 (q, $J = 7.2$ Hz, 2H), 2.56–2.64 (m, 1H), 1.84 (s, 3H), 1.29 (t, $J = 7.2$ Hz, 3H), 1.20–1.34 (m, 3H), 1.08–1.18 (m, 2H), 0.98 (d, $J = 6.4$ Hz, 3H), 0.80–0.87 (m, 6H); $^{13}\text{C NMR}$ (CDCl_3 , 125 MHz) δ 168.7, 148.4, 126.3, 60.5, 44.3, 32.4, 31.0, 30.2, 20.7, 19.1, 14.4, 12.6, 11.4.

(4*R*,6*R*,*E*)-2,4,6-Trimethyloct-2-en-1-ol (**21**).¹⁶ According to a modified literature procedure,¹² a cooled (-78 °C) solution of ester **20** (2.29 g, 10.79 mmol) in CH_2Cl_2 (36 mL) was treated with DIBAL-H (1 M in hexane, 27 mL, 26.98 mmol) using a syringe pump (2 mL/min). The mixture was stirred at -78 °C for 1 h, treated with aqueous citric acid (20% wt, 150 mL), stirred at room temperature for 20 min, and then extracted with ethyl acetate (3 \times 150 mL). The combined organic phases were washed with aqueous citric acid (20% wt, 100 mL) and brine (100 mL), dried (Na_2SO_4), and concentrated in vacuo to give a clear oil which was purified by flash chromatography (silica gel, 15% ethyl acetate in hexane) to afford alcohol **21** (1.66 g, 90%) as a clear oil; $R_f = 0.31$ (16% ethyl acetate in hexane); $[\alpha]_{\text{D}}^{24} -27.9$ (c 1.00, CHCl_3); IR ν_{\max} 3315, 2959, 2923, 2871, 1457, 1378, 1010, 850, 618 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 5.12 (dd, $J = 1.2, 9.5$ Hz, 1H), 3.99 (d, $J = 5.8$ Hz, 2H), 2.43–2.55 (m, 1H), 1.68 (d, $J = 1.2$ Hz, 3H), 1.23–1.31 (m, 3H), 1.01–1.16 (m, 2H), 0.92 (d, $J = 6.7$ Hz, 3H), 0.80–0.86 (m, 6H); $^{13}\text{C NMR}$ (CDCl_3 , 125 MHz) δ 133.25, 133.16, 69.3, 44.9, 32.2, 30.2, 29.7, 21.8, 19.3, 14.0, 11.4.

Ethyl (2*E*,4*E*,6*R*,8*R*)-4,6,8-Trimethyldeca-2,4-dienoate (**23**). According to a literature procedure,¹² compound **23** (990 mg, 68%) was prepared from **10** (1.03 g, 6.09 mmol) and $\text{Ph}_3\text{PCHCO}_2\text{Et}$ (**22**) (4.24 g, 12.18 mmol) as a colorless oil; $R_f = 0.32$ (6% ethyl acetate in hexane); $[\alpha]_{\text{D}}^{24} -66.6$ (c 1.00, CHCl_3) (lit.¹² $[\alpha]_{\text{D}}^{22.5} -41.8$ (c 1.00, CHCl_3)); IR ν_{\max} 2961, 2926, 2874, 1713, 1623, 1461, 1393, 1366, 1289, 1260, 1239, 1162, 1131, 1096, 1033, 982, 846 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 7.31 (d, $J = 15.6$ Hz, 1H), 5.78 (d, $J = 15.6$ Hz, 1H), 5.63 (d, $J = 9.8$ Hz, 1H), 4.20 (q, $J = 7.2$ Hz, 2H), 2.58–2.70 (m, 1H), 1.76–1.80 (m, 3H), 1.29 (t, $J = 7.2$ Hz, 3H), 1.19–1.35 (m, 3H), 1.07–1.17 (m, 2H), 0.96 (d, $J = 6.7$ Hz, 3H), 0.84 (t, $J = 7.3$ Hz, 3H), 0.81 (d, $J = 6.4$ Hz, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 125 MHz) δ 167.8, 150.1, 148.9, 131.3, 115.6, 60.3, 44.6, 32.5, 31.0, 30.3, 21.2, 19.2, 14.5, 12.4, 11.4.

(2*E*,4*E*,6*R*,8*R*)-4,6,8-Trimethyldeca-2,4-dien-1-ol (**24**). Analogously to **21**, alcohol **24** (383 mg, 98%) was prepared as a colorless oil from ester **23** (477 mg, 2.00 mmol) and DIBAL-H (1 M in hexane, 5.0 mL, 5.00 mmol); $R_f = 0.15$ (16% ethyl acetate in hexane); $[\alpha]_{\text{D}}^{24} -40.3$ (c 1.00, CHCl_3) (lit.¹² $[\alpha]_{\text{D}}^{22.5} -37.1$ (c 1.00, CHCl_3)); IR ν_{\max} 3313, 2960, 2923, 2871, 1651, 1457, 1377, 1097, 1024, 995, 964, 872, 772 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 6.25 (qd, $J = 0.8, 15.6$ Hz, 1H), 5.71 (td, $J = 6.1, 15.6$ Hz, 1H), 5.23 (d, $J = 9.8$ Hz, 1H), 4.20 (dd, $J = 0.8, 6.1$ Hz, 2H), 2.53–2.65 (m, 1H), 1.76 (d, $J = 1.2$ Hz, 3H), 1.36 (br. s, 1H), 1.20–1.32 (m, 3H), 1.03–1.18 (m, 2H), 0.93 (d, $J = 6.7$ Hz, 3H), 0.84 (t, $J = 7.3$ Hz, 3H), 0.81 (d, $J = 6.41$ Hz, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 125 MHz) δ 140.7, 137.4, 131.3, 125.1, 64.2, 45.0, 32.4, 30.4, 30.3, 21.7, 19.2, 12.8, 11.5.

(2*E*,4*E*,6*R*,8*R*)-4,6,8-Trimethyldeca-2,4-dienal (**25**). Analogously to **10**, aldehyde **25** (363 mg, 96%) was prepared as a colorless oil from alcohol **24** (383 mg, 1.95 mmol) and MnO_2 (3.39 g, 39.02 mmol); $R_f = 0.55$ (16% ethyl acetate in hexane); $[\alpha]_{\text{D}}^{24} -47.3$ (c 1.00, CHCl_3) (lit.¹² $[\alpha]_{\text{D}}^{22.5} -61.4$ (c 1.00, CHCl_3)); IR ν_{\max} 2961, 2925, 2873, 2722, 1680, 1623, 1605, 1456, 1379, 1315, 1127, 1010, 969, 822, 595 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 9.55 (d, $J = 7.9$ Hz, 1H), 7.11 (d, $J = 15.6$ Hz, 1H), 6.09 (dd, $J = 7.9, 15.6$ Hz, 1H), 5.76 (d, $J = 9.8$ Hz, 1H), 2.62–2.74 (m, 1H), 1.83 (d, $J = 1.2$ Hz, 3H), 1.19–1.39 (m, 3H), 1.10–1.19 (m, 2H), 1.00 (d, $J = 6.7$ Hz, 3H), 0.85 (t, $J = 7.3$ Hz, 3H), 0.83 (d, $J = 6.4$ Hz, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 125 MHz) δ 194.4, 158.4, 151.3, 132.0, 126.9, 44.5, 32.5, 31.3, 30.2, 21.1, 19.2, 12.6, 11.4.

2,2-Dimethyl-5-(triphenylphosphoranylidene)acetyl-1,3-dioxan-4,6-dione (**26**). According to a literature procedure,¹³ ylide **26** (2.31 g, 56%) was prepared as a colorless solid from Meldrum's acid (1.32 g, 9.18 mmol) and ketenylidetriphenylphosphorane (2.78 g, 9.18 mmol); IR ν_{\max} 3062, 2984, 1685, 1626, 1587, 1573, 1548, 1516, 1375, 1314, 1272, 1258, 1204, 1175, 1158, 1104, 1055, 1029, 996, 985, 935, 86, 798, 783, 757, 747, 723, 716, 692, 659, 651, 579 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 13.53 (dd, $J = 0.8, 2.9$ Hz, 1H), 7.60–7.69 (m, 9H), 7.49–7.55 (m, 6H), 5.76 (dd, $J = 2.9, 21.7$ Hz, 1H), 1.70 (s, 6H); $^{13}\text{C NMR}$ (CDCl_3 , 125 MHz) δ 175.9, 133.3 (d, $J = 10.9$ Hz), 133.0 (d, $J = 2.7$ Hz), 129.3 (d, $J = 11.8$ Hz), 124.9 (d, $J = 91.7$ Hz), 102.4, 57.0 (d, $J = 108.1$ Hz), 26.4.

(5*S*,3*Z*)-5-(4-((*tert*-Butyldimethylsilyloxy)benzyl)-3-((2*E*,4*E*,6*E*,8*R*,10*R*)-1-hydroxy-6,8,10-trimethyldeca-2,4,6-trien-1-ylidene)-1-(2-nitrobenzyl)pyrrolidine-2,4-dione (**28a**) and (5*S*,3*Z*)-5-(4-((*tert*-Butyldimethylsilyloxy)benzyl)-1-(2,4-dimethoxybenzyl)-3-((2*E*,4*E*,6*E*,8*R*,10*R*)-1-hydroxy-6,8,10-trimethyldeca-2,4,6-trien-1-ylidene)pyrrolidine-2,4-dione (**28b**). (A) A suspension of ylide **26** (790 mg, 1.77 mmol) and KO^tBu (199 mg, 1.77 mmol) in THF (20 mL) was treated with a solution of aldehyde **25** (344 mg, 1.77 mmol) in THF (15 mL), and the resulting mixture was heated at reflux for 22 h. It was concentrated in vacuo, and the remainder was taken up in CH_2Cl_2 (150 mL) and sat. aqueous NaHCO_3 (100 mL). The phases were separated, and the organic one was washed with sat. aqueous NaHCO_3 (2 \times 150 mL) and 1 M aqueous HCl (100 mL). The organic phase was dried (Na_2SO_4) and concentrated in vacuo to give an inseparable mixture of 42% of Meldrum's acid derivative **8**, 42% PPh_3O , and 16% residual ylide **26**. It was taken up in acetonitrile p.a. (30 mL), and the resulting solution was split in two 15 mL portions which were used in the next step without further purification.

(B) The first 15 mL portion was treated with *o*Nb-L-Tyr(OTBS)-OME **7a** (326 mg, 734 μmol , 1.00 equiv), the resulting mixture was heated at reflux for 1 h, and all volatiles were removed in vacuo to leave an orange oil that was purified by flash chromatography (silica gel, 30% EtOAc in hexane, $R_f = 0.67$) to give β -ketoamide **27a** as a yellow oil (161 mg, 30% over two steps) that was used in the next step without further purification.

The second 15 mL portion was reacted analogously with DMB-L-Tyr(OTBS)-OME **7b** (381 mg, 829 μmol , 1.13 equiv) to give β -ketoamide **27b** as a yellow oil (120 mg, 25% over two steps) that was also used in the next step without further purification; $R_f = 0.57$ (30% EtOAc in hexane). (C) A solution of β -ketoamide **27a** (161 mg, 228 μmol) in methanol p.a. (23 mL) was treated with sodium methoxide (62 mg, 1.140 mmol), and the resulting mixture was stirred at ambient temperature for 15 min. A 1 M aqueous HCl (15 mL) solution and brine (10 mL) were added, and the mixture was extracted with EtOAc (2 \times 100 mL). The combined organic phases were dried (Na_2SO_4) and concentrated in vacuo to afford tetramic acid **28a** as a foamy orange yellow solid (152 mg, quant); $R_f = 0.15$ (30% EtOAc in hexane); $[\alpha]_{\text{D}}^{24} -415$ (c 0.50, CHCl_3); IR ν_{\max} 2957, 2927, 2857, 1698, 1645, 1626, 1592, 1556, 1525, 1510, 1443, 1353, 1338, 1305, 1259, 1173, 1120, 1001, 920, 875, 857, 839, 809, 781, 749, 727, 688, 608, 571 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 8.02 (dd, $J = 1.2, 8.2$ Hz, 1H), 7.58 (ddd, $J = 1.2, 7.6, 7.6$ Hz, 1H), 7.53 (dd, $J = 11.3, 15.0$ Hz, 1H), 7.44 (ddd, $J = 1.2, 7.6, 8.2$ Hz, 1H), 7.31 (dd, $J = 1.2, 7.6$ Hz, 1H), 7.17 (d, $J = 15.0$ Hz, 1H), 6.87 (d, $J = 8.5$ Hz, 2H), 6.69 (d, $J = 15.3$ Hz, 1H), 6.64 (d, $J = 8.5$ Hz, 2H), 6.43 (dd, $J = 11.3, 15.3$ Hz, 1H), 5.56 (d, $J = 9.8$ Hz, 1H), 5.22 (d, $J = 17.1$ Hz, 1H), 4.72 (d, $J = 17.1$ Hz, 1H), 4.00 (dd, $J = 4.3, 5.2$ Hz, 1H), 3.14 (dd, $J = 4.3, 14.7$ Hz, 1H), 3.04 (dd, $J = 5.2, 14.7$ Hz, 1H), 2.60–2.71 (m, 1H), 1.82 (s, 3H), 1.20–1.36 (m, 3H), 1.08–1.17 (m, 2H), 0.97 (d, $J = 6.4$ Hz, 3H), 0.94 (s, 9H), 0.81–0.87 (m, 6H), 0.14 (s, 6H); $^{13}\text{C NMR}$ (CDCl_3 , 125 MHz) δ 193.7, 174.6, 173.9, 154.8, 149.5, 148.3, 147.9, 146.6, 133.9, 132.9, 132.1, 130.3, 129.4, 128.6, 127.6, 125.4, 125.2, 120.2, 119.8, 99.7, 66.0, 44.7, 41.1, 35.0, 32.5, 31.1, 30.2, 25.8, 21.3, 19.2, 18.3, 12.5, 11.4, –4.4; HRMS (ESI) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{39}\text{H}_{53}\text{N}_2\text{O}_6\text{Si}^+$ 673.3667, found 673.3651.

Analogously, tetramic acid **28b** (113 mg, quant) was obtained as a solid yellow foam from β -ketoamide **27b** (120 mg, 167 μmol) and sodium methoxide (45 mg, 835 μmol); $R_f = 0.41$ (30% EtOAc in

hexane); $[\alpha]_D^{24}$ -412 (c 0.50, CHCl_3); IR ν_{max} 2958, 2928, 2858, 1697, 1609, 1593, 1558, 1509, 1451, 1361, 1260, 1209, 1158, 1131, 1108, 1035, 1000, 915, 838, 782, 686, 610 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 7.44 (dd, $J = 11.3, 15.0$ Hz, 1H), 7.06–7.12 (m, 2H), 6.96 (d, $J = 8.5$ Hz, 2H), 6.69 (d, $J = 8.5$ Hz, 2H), 6.64 (d, $J = 15.3$ Hz, 1H), 6.41–6.45 (m, 2H), 6.39 (dd, $J = 11.3, 15.3$ Hz, 1H), 5.51 (d, $J = 9.8$ Hz, 1H), 4.99 (d, $J = 14.6$ Hz, 1H), 4.19 (d, $J = 14.6$ Hz, 1H), 3.83–3.87 (m, 1H), 3.76–3.82 (m, 6H), 3.06–3.14 (m, 2H), 2.59–2.73 (m, 1H), 1.82 (s, 3H), 1.21–1.34 (m, 3H), 1.08–1.16 (m, 2H), 0.96 (d, $J = 6.7$ Hz, 3H), 0.94 (s, 9H), 0.80–0.86 (m, 6H), 0.14 (s, 6H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 194.8, 173.8, 173.1, 160.9, 158.7, 154.6, 148.5, 147.2, 145.3, 132.9, 131.5, 130.7, 128.1, 125.3, 120.3, 120.1, 116.4, 104.4, 100.6, 98.5, 65.1, 55.51, 55.48, 44.7, 38.4, 34.4, 32.5, 31.0, 30.2, 25.8, 21.4, 19.2, 18.3, 12.5, 11.4, -4.3 ; HRMS (ESI) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{41}\text{H}_{58}\text{NO}_6\text{Si}^+$ 688.4028, found 688.4031.

Ethyl (2E,4E,6E,8R,10R)-6,8,10-Trimethyldodeca-2,4,6-trienoate (31). According to a literature procedure,¹⁶ ester **31** (248 mg, 67%) was prepared as a colorless oil from aldehyde **10** (235 mg, 1.394 mmol) and phosphonate **30** (1.12 g, 4.46 mmol); $R_f = 0.40$ (6% EtOAc in hexane); $[\alpha]_D^{24}$ -51.7 (c 1.00, CHCl_3) (lit.¹⁶ $[\alpha]_D^{19}$ -41.8 (c 1.00, CHCl_3)); IR ν_{max} 2960, 2925, 2872, 1710, 1613, 1457, 1392, 1367, 1329, 1304, 1256, 1234, 1201, 1178, 1136, 1096, 1039, 996, 861, 718 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 7.34 (dd, $J = 11.0, 15.3$ Hz, 1H), 6.56 (d, $J = 15.3$ Hz, 1H), 6.22 (dd, $J = 11.0, 15.3$ Hz, 1H), 5.85 (d, $J = 15.3$ Hz, 1H), 5.44 (d, $J = 9.8$ Hz, 1H), 4.20 (q, $J = 7.0$ Hz, 2H), 2.55–2.69 (m, 1H), 1.80 (d, $J = 1.2$ Hz, 3H), 1.29 (t, $J = 7.0$ Hz, 3H), 1.20–1.33 (m, 3H), 1.07–1.17 (m, 2H), 0.96 (d, $J = 6.7$ Hz, 3H), 0.80–0.87 (m, 6H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 167.5, 146.4, 145.6, 145.5, 132.3, 123.8, 119.6, 60.3, 44.8, 32.4, 30.8, 30.2, 21.5, 19.2, 14.5, 12.6, 11.4.

(2E,4E,6E,8R,10R)-6,8,10-Trimethyldodeca-2,4,6-trien-1-ol (32). Analogously to **21**, alcohol **32** (179 mg, 85%) was prepared as a colorless cloudy oil from ester **31** (226 mg, 948 μmol) and DIBAL-H (1 M in hexane, 2.4 mL, 2.40 mmol) in 179 mg (85%); $R_f = 0.19$ (16% EtOAc in hexane); $[\alpha]_D^{24}$ -37.5 (c 1.00, CHCl_3) (lit.¹⁶ $[\alpha]_D^{20}$ -28.8 (c 1.00, CHCl_3)); IR ν_{max} 3306, 3024, 2959, 2922, 2870, 1625, 1455, 1376, 1309, 1087, 982, 842 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 6.10–6.33 (m, 3H), 5.82 (td, $J = 6.1, 15.2$ Hz, 1H), 5.25 (d, $J = 9.5$ Hz, 1H), 4.19 (d, $J = 6.1$ Hz, 2H), 2.54–2.65 (m, 1H), 1.77 (d, $J = 1.2$ Hz, 3H), 1.22–1.31 (m, 3H), 1.04–1.16 (m, 2H), 0.94 (d, $J = 6.7$ Hz, 3H), 0.79–0.86 (m, 6H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 141.3, 138.9, 132.7, 132.1, 130.6, 125.3, 63.8, 45.0, 32.4, 30.5, 30.3, 21.7, 19.2, 12.7, 11.5.

(2E,4E,6E,8R,10R)-6,8,10-Trimethyldodeca-2,4,6-trienal (33). Analogously to **10**, aldehyde **33** (150 mg, 92%) was prepared as a yellowish oil from alcohol **32** (165 mg, 742 μmol) and MnO_2 (1.29 g, 14.84 mmol); $R_f = 0.68$ (16% EtOAc in hexane); $[\alpha]_D^{24}$ -49.4 (c 0.50, CHCl_3); IR ν_{max} 2960, 2925, 2874, 2730, 1676, 1603, 1457, 1377, 1315, 1159, 1129, 1114, 1009, 984, 858, 647 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 9.55 (d, $J = 7.9$ Hz, 1H), 7.15 (dd, $J = 11.0, 15.3$ Hz, 1H), 6.68 (d, $J = 15.3$ Hz, 1H), 6.36 (dd, $J = 11.0, 15.3$ Hz, 1H), 6.16 (dd, $J = 8.1, 15.1$ Hz, 1H), 5.55 (d, $J = 9.8$ Hz, 1H), 2.60–2.72 (m, 1H), 1.83 (d, $J = 1.2$ Hz, 3H), 1.21–1.35 (m, 3H), 1.09–1.17 (m, 2H), 0.98 (d, $J = 6.7$ Hz, 3H), 0.80–0.88 (m, 6H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 193.9, 153.5, 148.6, 147.5, 132.4, 130.5, 124.0, 44.7, 32.5, 31.0, 30.2, 21.4, 19.2, 12.6, 11.4; HRMS (ESI) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{15}\text{H}_{25}\text{O}^+$ 221.1900, found 221.1901.

S-tert-Butyl-4-(diethoxyphosphoryl)-3-oxopentanothioate (34). According to a literature procedure,¹⁷ phosphonate **34**³¹ (4.32 g, 59%) was prepared from bromopropionyl bromide (2.3 mL, 22.30 mmol) and Meldrum's acid (2.900 g, 20.20 mmol) as a pale orange oil and as a keto/enol mixture (5.6:1); IR ν_{max} 2966, 1723, 1674, 1614, 1478, 1456, 1398, 1365, 1314, 1250, 1163, 1016, 959, 836, 789, 688, 644, 591 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 13.01–13.06 (m, 1H), 5.42–5.47 (m, 1H), 4.06–4.16 (m, 4H), 4.03 (d, $J = 15.3$ Hz, 1H), 3.72 (d, $J = 15.3$ Hz, 1H), 3.46 (qd, $J = 7.0, 26.2$ Hz, 1H), 2.67 (qd, $J = 7.3, 23.5$ Hz, 1H), 1.41–1.50 (m, 9H), 1.35 (d, $J = 7.0$ Hz, 1H), 1.29–1.33 (m, 6H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 198.3 (d, $J = 4.5$ Hz), 192.9, 171.6 (d, $J = 6.4$ Hz), 101.0 (d, $J = 7.3$ Hz), 62.9

(dd, $J = 18.2, 7.3$ Hz), 58.3, 49.2, 48.5, 47.2 (d, $J = 126.3$ Hz), 38.9 (d, $J = 135.3$ Hz), 30.2, 29.7, 16.5 (d, $J = 8.2$ Hz), 12.7 (d, $J = 5.4$ Hz), 10.7 (d, $J = 6.4$ Hz).

(5S,3Z)-5-(4-((tert-Butyldimethylsilyloxy)benzyl)-1-(2,4-dimethoxybenzyl)-3-((2E,4E,6E,8E,10R,12R)-1-hydroxy-2,8,10,12-tetramethyltetradeca-2,4,6,8-tetraen-1-ylidene)pyrrolidine-2,4-dione (36). A mixture of thioester **9** (100 mg, 256 μmol), THF (3.2 mL), and 4 Å molecular sieves (powdered, 40 mg) was cooled to 0 °C and treated with a solution of DMB-L-Tyr(OTBS)-OMe **7b** (130 mg, 282 μmol), NEt_3 (0.14 mL, 1.024 mmol), and silver trifluoroacetate (113 mg, 512 μmol) in THF (3.2 mL). The mixture was stirred at 0 °C under exclusion of light for 2.5 h, diluted with diethyl ether (100 mL), and filtered over Celite. The filtrate was washed with sat. aqueous NH_4Cl (75 mL) and brine (75 mL), dried (Na_2SO_4), and concentrated in vacuo to give an orange oil. It was purified by flash chromatography (silica gel, 50% EtOAc in hexane; $R_f = 0.76$) to give β -ketoamide **35** as a yellow oil (173 mg, 89%) that was used in the next step without further purification. A solution of β -ketoamide **35** (159 mg, 209 μmol) in methanol (21 mL) was treated with sodium methoxide (56 mg, 1.05 mmol), and the mixture was stirred at ambient temperature for 20 min. Saturated aqueous NH_4Cl (40 mL) and 1 M aqueous HCl (30 mL) were added, the mixture was extracted with EtOAc (100 mL), and the organic phase was dried (Na_2SO_4) and concentrated in vacuo to give bisprotected fumosorinone **A** (**36**) as a red gum (149 mg, quant); $R_f = 0.56$ (50% EtOAc in hexane); $[\alpha]_D^{24}$ -367.1 (c 0.75, CHCl_3); IR ν_{max} 2958, 2928, 2859, 1652, 1611, 1590, 1544, 1508, 1452, 1389, 1361, 1255, 1209, 1171, 1158, 1119, 1034, 988, 913, 837, 807, 781, 725, 687, 640, 605, 578 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 7.74 (d, $J = 11.6$ Hz, 1H), 7.09 (d, $J = 7.9$ Hz, 1H), 6.99 (d, $J = 8.5$ Hz, 2H), 6.68–6.75 (m, 3H), 6.57 (dd, $J = 11.6, 14.3$ Hz, 1H), 6.41–6.49 (m, 3H), 6.31 (dd, $J = 10.7, 15.0$ Hz, 1H), 5.40 (d, $J = 9.5$ Hz, 1H), 4.99 (d, $J = 14.6$ Hz, 1H), 4.24 (d, $J = 14.6$ Hz, 1H), 3.82–3.86 (m, 1H), 3.78–3.82 (m, 6H), 3.15 (dd, $J = 4.6, 14.3$ Hz, 1H), 3.10 (dd, $J = 4.0, 14.3$ Hz, 1H), 2.56–2.70 (m, 1H), 1.95 (s, 3H), 1.81 (s, 3H), 1.22–1.35 (m, 3H), 1.06–1.17 (m, 2H), 0.97 (d, $J = 6.4$ Hz, 3H), 0.95 (br. s., 9H), 0.85 (dd, $J = 7.0, 7.6$ Hz, 3H), 0.82 (d, $J = 6.4$ Hz, 3H), 0.14 (s, 6H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 192.6, 182.7, 175.9, 160.9, 158.7, 154.6, 144.1, 143.2, 142.9, 142.6, 132.7, 131.5, 130.8, 128.21, 128.18, 126.9, 126.3, 120.1, 116.4, 104.4, 99.4, 98.5, 64.5, 55.51, 55.49, 44.9, 38.7, 34.5, 32.4, 30.8, 30.2, 25.8, 21.5, 19.2, 18.3, 12.64, 12.61, 11.4, -4.32 , -4.34 ; HRMS (ESI) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{44}\text{H}_{62}\text{NO}_6\text{Si}^+$ 728.4341, found 728.4340.

■ ASSOCIATED CONTENT

📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.8b01530.

NMR spectra of all compounds, HPLC chromatograms of target compounds **3** and **4**, ECD spectrum of compound **3** (PDF)

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: Rainer.Schobert@uni-bayreuth.de. Fax: +49 (0)921 552671.

ORCID

Rainer Schobert: 0000-0002-8413-4342

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

We thank Prof. Dr. Birte Höcker and Sooruban Shanmugaratnam (all University of Bayreuth) for assistance with measuring ECD spectra.

REFERENCES

- (1) Schmidt, K.; Günther, W.; Stoyanova, S.; Schubert, B.; Li, Z.; Hamburger, M. Militarinone A, a Neurotrophic Pyridone Alkaloid from *Paecilomyces militaris*. *Org. Lett.* **2002**, *4*, 197–199.
- (2) Schmidt, K.; Riese, U.; Li, Z.; Hamburger, M. Novel Tetramic Acids and Pyridone Alkaloids, Militarinones B, C, and D, from the Insect Pathogenic Fungus *Paecilomyces militaris*. *J. Nat. Prod.* **2003**, *66*, 378–383.
- (3) (a) Isaka, M.; Chinthanom, P.; Supothina, S.; Tobwor, P.; Hywel-Jones, N. L. Pyridone and Tetramic Acid Alkaloids from the Spider Pathogenic Fungus *Torrubiella* sp. BCC 2165. *J. Nat. Prod.* **2010**, *73*, 2057–2060. (b) Bruckner, S.; Bilitewski, U.; Schobert, R. Synthesis and Antibacterial Activity of Four Stereoisomers of the Spider-Pathogenic Fungus Metabolite *Torrubiellone* D. *Org. Lett.* **2016**, *18*, 1136–1139.
- (4) (a) Cox, R. J.; O'Hagan, D. Synthesis of Isotopically Labeled 3-Amino-2-phenylpropionic Acid and its Role as a Precursor in the Biosynthesis of Tenellin and Tropic Acid. *J. Chem. Soc., Perkin Trans. 1* **1991**, *1*, 2537–2540. (b) Halo, L. M.; Heneghan, M. N.; Yakasai, A. A.; Song, Z.; Williams, K.; Bailey, A. M.; Cox, R. J.; Lazarus, C. M.; Simpson, T. J. Late Stage Oxidations during the Biosynthesis of the 2-Pyridone Tenellin in the Entomopathogenic Fungus *Beauveria bassiana*. *J. Am. Chem. Soc.* **2008**, *130*, 17988–17996. (c) Wasil, Z.; Pahirulzaman, K. A. K.; Butts, C.; Simpson, T. J.; Lazarus, C. M.; Cox, R. J. One Pathway, Many Compounds: Heterologous Expression of a Fungal Biosynthetic Pathway Reveals its Intrinsic Potential for Diversity. *Chem. Sci.* **2013**, *4*, 3845–3856.
- (5) Zhang, J.; Meng, L.-L.; Wei, J.-J.; Fan, P.; Liu, S.-S.; Yuan, W.-Y.; Zhao, Y.-X.; Luo, D.-Q. PTP1B Inhibitors from the Entomogenous Fungi *Isaria fumosorosea*. *Molecules* **2017**, *22*, 2058–2512.
- (6) van Huijsduijnen, R. H.; Sauer, W. H. B.; Bombrun, A.; Swinnen, D. Prospects for Inhibitors of Protein Tyrosine Phosphatase 1B as Antidiabetic Drugs. *J. Med. Chem.* **2004**, *47*, 4142–4146.
- (7) Klamann, L. D.; Boss, O.; Peroni, O. D.; Kim, J. K.; Martino, J. L.; Zabolotny, J. M.; Moghal, N.; Lubkin, M.; Kim, Y.-B.; Sharpe, A. H.; Stricker-Krongrad, A.; Shulman, G. I.; Neel, B. G.; Kahn, B. B. Increased Energy Expenditure, Decreased Adiposity, and Tissue-Specific Insulin Sensitivity in Protein-Tyrosine Phosphatase 1B-Deficient Mice. *Mol. Cell. Biol.* **2000**, *20*, 5479–5489.
- (8) Bruckner, S.; Haase, R. G.; Schobert, R. A Synthetic Route to β -Hydroxytyrosine-Derived Tetramic Acids: Total Synthesis of the Fungal Metabolite F-14329. *Chem. - Eur. J.* **2017**, *23*, 5692–5695.
- (9) Haase, R. G.; Schobert, R. Synthesis of the Bioherbicidal Fungal Metabolite Macrocidin A. *Org. Lett.* **2016**, *18*, 6352–6355.
- (10) Nishida, A.; Fuwa, M.; Fujikawa, Y.; Nakahata, E.; Furuno, A.; Nagagawa, M. First Total Synthesis of Martefragin A, a Potent Inhibitor of Lipid Peroxidation Isolated from Sea Alga. *Tetrahedron Lett.* **1998**, *39*, 5983–5986.
- (11) Carlsen, P. H. J.; Katsuki, T.; Martin, V. S.; Sharpless, K. B. A Greatly Improved Procedure for Ruthenium Tetroxide Catalyzed Oxidations of Organic Compounds. *J. Org. Chem.* **1981**, *46*, 3936–3938.
- (12) Ding, F.; Leow, M. L.; Ma, J.; William, R.; Liao, H.; Liu, X.-W. Collective Synthesis of 4-Hydroxy-2-pyridone Alkaloids and Their Antiproliferation Activities. *Chem. - Asian J.* **2014**, *9*, 2548–2554.
- (13) Lovmo, K.; Dütz, S.; Harras, M.; Haase, R. G.; Milius, W.; Schobert, R. A Short Synthesis of 3-Enoyltetramic Acids Employing a New Acyl Ylide Conjugate of Meldrum's Acid. *Tetrahedron Lett.* **2017**, *58*, 4796–4798.
- (14) Sengoku, T.; Nagae, Y.; Ujihara, Y.; Takahashi, M.; Yoda, H. A Synthetic Approach to Diverse 3-Acyltetramic Acids via O- to C-Acyl Rearrangement and Application to the Total Synthesis of Penicillenol Series. *J. Org. Chem.* **2012**, *77*, 4391–4401.
- (15) Kauh, U.; Andernach, L.; Weck, S.; Sandjo, L. P.; Jacob, S.; Thines, E.; Opatz, T. Total Synthesis of (–)-Hymenoseitin. *J. Org. Chem.* **2016**, *81*, 215–228.
- (16) Dash, U.; Sengupta, S.; Sim, T. A Concise and Efficient Total Synthesis of Militarinone D. *Eur. J. Org. Chem.* **2015**, *2015*, 3963–3970.
- (17) Loscher, S.; Schobert, R. Total Synthesis and Absolute Configuration of Epicoccamide D, a Naturally Occurring Mannosylated 3-Acyltetramic Acid. *Chem. - Eur. J.* **2013**, *19*, 10619–10624.
- (18) Ley, S. V.; Smith, S. C.; Woodward, P. R. Further Reactions of t-Butyl 3-Oxobutanthioate and t-Butyl 4-Diethyl-phosphono-3-oxobutanthioate: Carbonyl Coupling Reactions, Amination, Use in the Preparation of 3-Acyltetramic Acids and Application to the Total Synthesis of Fuligorubin A. *Tetrahedron* **1992**, *48*, 1145–1174.
- (19) Riache, N.; Bailly, C.; Deville, A.; Dubost, A.; Nay, B. Total Synthesis of Tyrosine-Derived Tetramic Acid Pigments from a Slime Mould. *Eur. J. Org. Chem.* **2010**, *2010*, 5402–5408.
- (20) Jessen, H. J.; Schumacher, A.; Shaw, T.; Pfaltz, A.; Gademann, K. A Unified Approach for the Stereoselective Total Synthesis of Pyridone Alkaloids and their Neurotogenic Activity. *Angew. Chem., Int. Ed.* **2011**, *50*, 4222–4226.
- (21) Caporali, S.; Chiappe, C.; Ghilardi, T.; Iuliano, A.; Longhi, G.; Margari, P.; Pomelli, C. S. Arrangements of Enantiopure and Racemic Ionic Liquids at the Liquid/Air Interface: the Role of Chirality on Self-assembly and Layering. *RSC Adv.* **2016**, *6*, 8053–8060.
- (22) Dooley, D. J.; Taylor, C. P., Jr.; Thorpe, A. J.; Wustrow, D. J. Preparation of Pregabalin Derivatives for the Treatment of Fibromyalgia and Other Disorders. *PCT Int. Appl.* WO 2004054566 A1, July 1, 2004.
- (23) Li, G.; Yang, X.; Zhai, H. Total Synthesis of (–)-5,6-Dihydrocineromycin B. *J. Org. Chem.* **2009**, *74*, 1356–1359.
- (24) Rossi, R.; Salvadori, P. A. Synthesis of Both Enantiomers of 6-Methyl-3-octanone, a Component of the Alarm Pheromone of Ants in the Genus *Crematogaster*. *Synthesis* **1979**, *1979*, 209–219.
- (25) Song, S.; Zhu, S.-F.; Yu, Y.-B.; Zhou, Q.-L. Carboxy-Directed Asymmetric Hydrogenation of 1,1-Diarylethenes and 1,1-Dialkylethenes. *Angew. Chem., Int. Ed.* **2013**, *52*, 1556–1559.
- (26) Mori, M.; Nakagawa, M.; Nishida, A.; Fuwa, M.; Saito, H.; Matsunaga, T.; Takahashi, S.; Hasegawa, C. Preparation of Stereoisomeric 3-(4-Carboxyoxazol-5-yl)indole Compounds by Oxidative Cyclization of N-acyl-L-tryptophan. *PCT Int. Appl.* WO 9912923 A1, March 18, 1999.
- (27) Tokuyama, H.; Yamada, K.; Fujiwara, H.; Sakata, J.; Okano, K.; Sappan, M.; Isaka, M. Structural Determination of (–)-SCH 64874 and Hirsutellomycin by Semisynthesis. *J. Org. Chem.* **2017**, *82*, 353–371.
- (28) Organ, M.; Bilokin, Y.; Bratovanov, S. Approach toward the Total Synthesis of Orevactaene. 2. Convergent and Stereoselective Synthesis of the C18–C31 Domain of Orevactaene. Evidence for the Relative Configuration of the Side Chain. *J. Org. Chem.* **2002**, *67*, 5176–5183.
- (29) Isler, O.; Gutmann, H.; Montavon, M.; Rüegg, R.; Ryser, G.; Zeller, P. Synthesen in der Carotinoid-Reihe. 10. Mitteilung. Anwendung der Wittig-Reaktion zur Synthese von Estern des Bixins und Crocetinins. *Helv. Chim. Acta* **1957**, *40*, 1242–1249.
- (30) Lang, R. W.; Hansen, H.-J. Eine einfache Allencarbonsäureester-Synthese mittels der Wittig-Reaktion. *Helv. Chim. Acta* **1980**, *63*, 438–455.
- (31) Burke, L. T.; Dixon, D. J.; Ley, S. V.; Rodriguez, F. Total Synthesis of the Fusarium Toxin Equisetin. *Org. Biomol. Chem.* **2005**, *3*, 274–280.