



Contents lists available at ScienceDirect

Chinese Chemical Letters

journal homepage: www.elsevier.com/locate/cclet

Original article

Design and concise synthesis of *gem*-difluoromethylenated analogue of 7-*epi*-castanospermineQ1 Xin-Yi Jiang^a, Xiu-Hua Xu^a, Feng-Ling Qing^{a,b,*}^a Key Laboratory of Organofluorine Chemistry, Shanghai Institute of Organic Chemistry, Chinese Academy of Science, Shanghai 200032, China^b College of Chemistry, Chemical Engineering and Biotechnology, Donghua University, Shanghai 201620, China

ARTICLE INFO

Article history:

Received 20 February 2014

Received in revised form 8 April 2014

Accepted 10 April 2014

Available online xxx

Keywords:

Indolizidine

Castanospermine

gem-Difluoromethylene

Cyclization

Dihydroxylation

ABSTRACT

A novel *gem*-difluoromethylenated castanospermine analogue **B** was designed and synthesized, starting from 3-bromo-3,3-difluoropropene and L-(–)-malic acid. The key steps involve substitution cyclization reaction and RCM reaction to construct the aza fused bicyclic framework.

© 2014 Feng-Ling Qing. Published by Elsevier B.V. on behalf of Chinese Chemical Society. All rights reserved.

1. Introduction

Polyhydroxylated indolizidine alkaloids are widely found in microorganisms, vertebrates, higher invertebrates, and plants [1]. Some of these aza fused bicyclic compounds, such as lentiginosine [2], swainsonine [3], and castanospermine [4], have been reported to exhibit good glycosidase inhibition activity, which makes them potential therapeutic agents (Fig. 1). Among them, castanospermine has attracted considerable attention. It is a strong inhibitor of α - and β -glucosidases [5] and has shown potential antitumor, antiviral, and immunomodulating activities [6]. Because of its unique biological activities, a lot of efforts have been devoted into the synthesis of castanospermine [7] as well as its analogues [8] to study their structure-activity relationship (SAR).

The major drawback for castanospermine in clinical applications is the low inhibition selectivity between α - and β -glucosidases. In 1997, Tyler and co-workers reported that 7-*epi*-castanospermine showed higher α/β glucosidase selectivity than castanospermine while retaining almost complete activity towards α -glucosidase (Scheme 1) [9]. To further improve the selectivity and the potency of the inhibition of glycosidases, it is highly desirable to develop new polyhydroxylated alkaloid analogues. In

2006, our group found that introduction of a *gem*-difluoromethylene group (CF₂) into 1-deoxymannonojirimycin (DMJ) could improve the inhibition selectivity of compound **A** [10]. As a part of our interest in the synthesis of fluorinated iminosugars [10,11], we designed a new *gem*-difluoromethylenated analogue **B** of 7-*epi*-castanospermine. The strongly electron-withdrawing *gem*-difluoromethylene group would reduce the pK_a value and might change the stereoconfiguration, thus affecting the inhibition activity and selectivity. In this paper, we present our results on the synthesis of this novel castanospermine analogue.

2. Experimental

2.1. (3*S*,4*R*)-Methyl 3-(benzyloxy)-5,5-difluoro-4-hydroxyhept-6-enoate (**3a**) and (4*S*,5*S*)-4-(benzyloxy)-5-(1,1-difluoroallyl)dihydrofuran-2(3*H*)-one (**3b**)

To a stirring solution of (*S*)-*O*-benzylmalic acid dimethyl ester **2** (0.90 g, 3.57 mmol) in CH₂Cl₂ (50 mL) was added magnesium bromide etherate (1.0 g, 4.0 mmol) at 0 °C. The solution was stirred for 1 h at 0 °C and cooled to –90 °C. To the solution was added a 1.5 mol/L solution of diisobutylaluminum hydride in toluene (3.0 mL, 4.5 mmol) via syringe pump over 90 min, then the reaction mixture was stirred at –90 °C for 2 h. Methanol (4 mL) was slowly added followed by saturated Rochelle salts (80 mL). The mixture was warmed to room temperature and stirred for 12 h. The layers were separated and the aqueous phase was

* Corresponding author at: Key Laboratory of Organofluorine Chemistry, Shanghai Institute of Organic Chemistry, Chinese Academy of Science, Shanghai 200032, China.

E-mail address: flq@mail.sioc.ac.cn (F.-L. Qing).

<http://dx.doi.org/10.1016/j.ccl.2014.04.018>

1001-8417/© 2014 Feng-Ling Qing. Published by Elsevier B.V. on behalf of Chinese Chemical Society. All rights reserved.

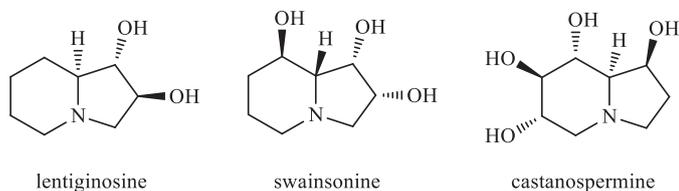


Fig. 1. Polyhydroxylated indolizidines.

54 extracted three times with methylene dichloride. The combined
55 organic phase was dried with Na₂SO₄ and concentrated. The
56 mixture was separated by flash column chromatography (petro-
57 leum ether/ethyl acetate = 8/1) to yield 0.61 g (77%) of a colourless
58 oil. The colourless oil was dissolved in DMF (10 mL). After that,
59 indium (474 mg, 4.12 mmol) and 3-bromo-3,3-difluoropropane
60 (420 μL, 4.20 mmol) was added. The reaction mixture was stirred
61 at room temperature for 24 h. The reaction mixture was then
62 quenched with 1 mol/L HCl and extracted with CH₂Cl₂. The
63 combined organic extract was washed with brine, dried over
64 anhydrous Na₂SO₄, and filtered. The solvent was removed *in vacuo*.
65 The residue was purified by silica gel column chromatography
66 (petroleum ether/ethyl acetate = 8/1) to give 255 mg (31% yield) of
67 compound **3a** as a clear oil and 362 mg (44% yield) of compound **3b**
68 as a clear oil.

69 **3a**: [α]_D²⁰ = -6.93 (c 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ
70 7.31–7.30 (m, 5H), 6.08–5.95 (m, 1H), 5.76–5.68 (m, 1H), 5.54–5.47
71 (m, 1H), 4.67–4.60 (m, 2H), 4.28–4.25 (m, 1H), 3.76–3.64 (m, 4H),
72 2.73–2.71 (m, 2H), 2.42 (br, 1H); ¹⁹F NMR (282 MHz, CDCl₃): δ
73 -107.0 (dt, 1F, J = 253.0 Hz, 9.45 Hz), -111.5 (dt, 1F, J = 252.4 Hz,
74 11.0 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 171.4, 137.4, 130.6 (t,
75 J = 25.7 Hz), 128.5, 128.1, 120.7 (t, J = 9.5 Hz), 119.4 (t,
76 J = 242.8 Hz), 74.1 (t, J = 29.6 Hz), 73.2, 72.9, 51.8, 37.3; IR (thin
77 film, cm⁻¹): 3500, 3033, 2953, 1738, 1439; MS (ESI): *m/z* 301.3
78 (M+H⁺), 318.3 (M+NH₄⁺); 323.2 (M+Na⁺); HRMS Calcd. for
79 C₁₅H₁₈O₄F₂Na: 323.1065; Found: 323.1073.

80 **3b**: [α]_D²⁰ = -15.17 (c 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃):
81 δ 7.40–7.30 (m, 5H), 6.04–5.87 (m, 1H), 5.80–5.74 (m, 1H), 5.63–
82 5.60 (m, 1H), 4.65–4.51 (m, 3H), 4.46–4.44 (m, 1H), 2.87–2.78 (m,
83 1H), 2.65–2.59 (m, 1H); ¹⁹F NMR (282 MHz, CDCl₃): δ -109.8 (ddd,
84 1F, J = 258.6 Hz, 12.7 Hz, 4.5 Hz), -113.0 (ddd, 1F, J = 258.9 Hz,
85 18.6 Hz, 10.6 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 174.2, 136.7, 128.8
86 (t, J = 25.8 Hz), 128.7, 128.3, 127.9, 122.9 (t, J = 9.5 Hz), 117.6 (t,
87 J = 242.7 Hz), 84.3 (t, J = 31.4 Hz), 73.4, 71.4, 34.9; IR (thin film,
88 cm⁻¹): 3033, 2932, 1691, 1455, 1208; MS (ESI): *m/z* 286.2
89 (M+NH₄⁺); HRMS Calcd. for C₁₄H₁₄O₃F₂Na: 291.0803; Found:
90 291.0811.

91 2.2. (3*S*,4*S*)-3-(Benzyloxy)-5,5-difluorohept-6-ene-1,4-diol (**4**)

92 To a suspension of LiAlH₄ (91.2 mg, 2.4 mmol) in dry THF (5 mL)
93 was added a solution of **3b** (0.36 g, 1.2 mmol) in dry THF at 0 °C.
94 The mixture was stirred for 3 h at 0 °C and quenched by careful
95 addition of water and then extracted with CH₂Cl₂. The combined
96 organic layer was washed with brine, dried over anhydrous

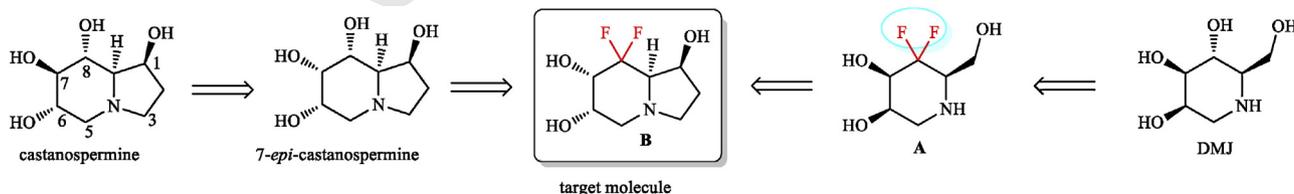
Na₂SO₄, and concentrated *in vacuo*. The residue was quickly
97 purified by silica gel column chromatography (petroleum ether/
98 ethyl acetate = 2/1) to afford compound **4** (0.28 g, 86%) as a clear
99 oil. [α]_D²⁰ = -17.01 (c 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ
100 7.35–7.25 (m, 5H), 6.10–5.93 (m, 1H), 5.73–5.68 (m, 1H), 5.50–5.47
101 (m, 1H), 4.52 (dd, 2H, J = 17.4 Hz, 11.4 Hz), 4.07–3.98 (m, 1H), 3.90–
102 3.63 (m, 3H), 2.96 (br, 2H), 2.02–1.92 (m, 2H); ¹⁹F NMR (282 MHz,
103 CDCl₃): δ -107.7 (dt, 1F, J = 251.5 Hz, 10.3 Hz), -109.3 (dt, 1F,
104 J = 251.8 Hz, 12.0 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 137.6, 130.9 (t,
105 J = 25.4 Hz), 128.6, 128.1, 128.0, 120.4 (t, J = 9.5 Hz), 119.7 (t,
106 J = 243.1 Hz), 76.3, 73.8 (t, J = 27.9 Hz), 71.8, 58.9, 31.7; IR (thin
107 film, cm⁻¹): 3392, 3033, 2928, 1664, 1056; MS (ESI): *m/z* 295.2
108 (M+Na⁺); HRMS Calcd. for C₁₄H₁₈O₃F₂Na: 295.1116; Found:
109 295.1120.

111 2.3. (2*R*,3*S*)-1-Allyl-3-(benzyloxy)-2-(1,1-difluoroallyl)pyrrolidine (**5**)

112
113 To a solution of compound **4** (1.0 g, 3.68 mmol) in anhydrous
114 CH₂Cl₂ (12 mL), and NEt₃ (3.4 mL, 23.8 mmol), MsCl (1.3 mL,
115 16.7 mmol) was added slowly at 0 °C. The reaction mixture was
116 then warmed to room temperature and stirred overnight. The
117 reaction was quenched with water. The resulting mixture was
118 extracted with CH₂Cl₂. The combined organic layer was washed
119 with brine, dried over anhydrous Na₂SO₄, and filtered. The solvent
120 was removed *in vacuo*. The residue was dissolved in allylamine
121 (6 mL). Then the reaction mixture was heated to 145 °C in the
122 sealed tub for 10 h. The allylamine was removed *in vacuo* and then
123 the residue was purified by silica gel column chromatography
124 (petroleum ether/ethyl acetate = 15/1) to give 870 mg (81% yield,
125 two steps) of compound **5** as a clear oil. [α]_D²⁰ = -16.86 (c 1.10,
126 CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.34–7.25 (m, 5H), 6.35–6.17
127 (m, 1H), 5.93–5.79 (m, 1H), 5.68–5.62 (m, 1H), 5.42–5.38 (m, 1H),
128 5.19–5.09 (m, 2H), 4.53 (s, 2H), 4.12 (q, 1H, J = 6.3 Hz), 3.57–3.51
129 (m, 1H), 3.18–3.00 (m, 3H), 3.17 (q, 1H, J = 8.7 Hz), 1.99–1.92 (m,
130 2H); ¹⁹F NMR (282 MHz, CDCl₃): δ -96.4 (d, 1F, J = 255.2 Hz), -97.3
131 (dt, 1F, J = 254.9 Hz, 13.25 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 138.2,
132 134.8, 133.0 (t, J = 24.5 Hz), 128.3, 127.7, 127.6, 121.6 (t,
133 J = 239.9 Hz), 119.3 (t, J = 9.65 Hz), 117.4, 79.1 (d, J = 7.0 Hz),
134 72.1, 68.7 (dd, J = 30.3 Hz, 25.6 Hz), 58.2, 50.0 (d, J = 6 Hz), 30.4; IR
135 (thin film, cm⁻¹): 3066, 2924, 2359, 1419, 1354, 1121, 738; MS
136 (ESI): *m/z* 298.4 (M+H⁺); HRMS Calcd. for C₁₇H₂₂O₂F₂N: 294.1664;
137 Found: 294.1657.

138 2.4. (1*S*,8*aR*)-1-(Benzyloxy)-8,8-difluoro-1,2,3,5,8,8*a*- 139 hexahydroindolizine (**6**)

140 To a solution of compound **5** (520 mg, 1.77 mmol) in anhydrous
141 toluene (50 mL) was added Grubbs' II catalyst (70 mg,
142 0.0824 mmol). The reaction mixture was then warmed to 80 °C
143 and stirred for 10 h. After the solvent was evaporated, the crude
144 product was purified by flash silica gel column chromatography
145 (petroleum ether/ethyl acetate = 6/1) to give compound **6** (375 mg,
146 80%) as a clear oil. [α]_D²⁰ = 98.66 (c 1.15, CHCl₃); ¹H NMR
147 (300 MHz, CDCl₃): δ 7.40–7.23 (m, 5H), 6.15–6.11 (m, 1H), 5.92–
148 5.86 (m, 1H), 4.66 (dd, 2H, J = 37.5 Hz, 12.3 Hz), 4.39–4.33 (m, 1H),



Scheme 1. Design of target molecule B.

3.61–3.55 (m, 1H), 3.28 (t, 1H, $J = 8.6$ Hz), 3.00–2.93 (m, 1H), 2.73–2.64 (m, 1H), 2.36 (q, 1H, $J = 8.4$ Hz), 2.26–2.15 (m, 1H), 2.07–1.95 (m, 1H); ^{19}F NMR (282 MHz, CDCl_3): δ –98.3 (d, 1F, $J = 270.4$ Hz), –100.5 (d, 1F, $J = 269.3$ Hz); ^{13}C NMR (100 MHz, CDCl_3): δ 138.7, 133.8 (t, $J = 9.8$ Hz), 128.3, 127.4, 127.3, 123.7 (dd, $J = 30.3$ Hz, 26.4 Hz), 117.7 (dd, $J = 244.8$ Hz, 229.7 Hz), 78.2, 72.2, 65.9 (d, $J = 20.7$ Hz), 65.6 (d, $J = 21.3$ Hz), 51.6, 31.2; IR (thin film, cm^{-1}): 3033, 2987, 1620, 1237, 987; MS (ESI): m/z 266.3 ($\text{M}+\text{H}^+$); HRMS Calcd. for $\text{C}_{15}\text{H}_{17}\text{OF}_2\text{NNa}$: 288.1170; Found: 288.1164.

2.5. (3*S*,4*S*)-3-(Benzyloxy)-5,5-difluoro-4-hydroxyhept-6-enyl acetate (7)

To a solution of compound **4** (1.032 g, 3.79 mmol) in vinyl acetate (15 mL) was added lipase AK (0.52 g). The solution was stirred at room temperature. After 24 h, the solution was filtered and volatiles were removed by reduced pressure. The crude product was chromatographed (petroleum ether/ethyl acetate = 8/1) to afford **7** (1.037 g, 87%) as a clear oil. $[\alpha]_{\text{D}}^{20} = -35.71$ (c 1.00, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.35–7.25 (m, 5H), 6.05–5.92 (m, 1H), 5.69 (d, 1H, $J = 17.2$ Hz), 5.49 (d, 1H, $J = 11.2$ Hz), 4.51 (dd, 2H, $J = 34.4$ Hz, 11.2 Hz), 4.23–4.12 (m, 2H), 4.03 (td, 1H, $J = 11.2$ Hz, 3.2 Hz), 3.77–3.74 (m, 1H), 2.04–1.91 (m, 5H); ^{19}F NMR (282 MHz, CDCl_3): δ –107.5 (dt, 1F, $J = 251.5$ Hz, 11.3 Hz), –109.4 (dt, 1F, $J = 251.3$ Hz, 11.3 Hz); ^{13}C NMR (100 MHz, CDCl_3): δ 171.1, 137.4, 130.5 (t, $J = 25.3$ Hz), 128.5, 128.1, 128.0, 120.6 (t, $J = 9.7$ Hz), 119.4 (dd, $J = 241.2$ Hz, 243.3 Hz), 74.6 (t, $J = 1.5$ Hz), 73.6 (t, $J = 28.3$ Hz), 72.0, 61.1, 28.7, 20.9; IR (thin film, cm^{-1}): 3466, 3032, 2904, 1736, 819; MS (ESI): m/z 315.0 ($\text{M}+\text{H}^+$); HRMS Calcd. for $\text{C}_{16}\text{H}_{20}\text{O}_4\text{F}_2\text{NNa}$: 337.1222; Found: 337.1223.

2.6. (3*S*,4*R*)-4-Azido-3-(benzyloxy)-5,5-difluorohept-6-enyl acetate (8)

Compound **7** (2.62 g, 8.34 mmol) was dissolved in dry CH_2Cl_2 (50 mL). After that, DMAP (2.03 g, 16.68 mmol) was added. The resulting mixture was cooled to -35°C . Then, F_2O (2.10 mL, 12.52 mmol) was added dropwise to the solution with stirring. After that, the reaction mixture was stirred for about 3 h at 0°C . Water and NaHCO_3 solution were added successively after the mixture was warmed to room temperature. Then the mixture was extracted with CH_2Cl_2 , dried over anhydrous Na_2SO_4 . The mixture was separated by flash column (petroleum ether/ethyl acetate = 8/1) to yield a colourless oil. The colourless oil was dissolved in DMF (20 mL). Then, sodium azide (2.7 g, 41.7 mmol) was added carefully with stirring at 0°C in an ice bath. The reaction mixture was stirred overnight at room temperature. Water was added to quench the reaction. The aqueous phase was extracted with CH_2Cl_2 . The combined organic layer was washed with brine, dried over anhydrous Na_2SO_4 , and concentrated *in vacuo*. The residue was quickly purified by silica gel column chromatography (petroleum ether/ethyl acetate = 8/1) to afford compound **8** (1.95 g, 69% yield) as a clear oil. $[\alpha]_{\text{D}}^{20} = -45.97$ (c 1.00, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ 7.35–7.25 (m, 5H), 6.08–5.95 (m, 1H), 5.78 (dt, 1H, $J = 17.4$ Hz, 2.1 Hz), 5.49 (d, 1H, $J = 11.1$ Hz), 4.59 (dd, 2H, $J = 39.3$ Hz, 11.1 Hz), 4.24–4.08 (m, 2H), 3.92–3.86 (m, 1H), 3.58–3.49 (m, 1H), 2.03–1.97 (m, 5H); ^{19}F NMR (282 MHz, CDCl_3): δ –97.98 (d, 1F, $J = 253.2$ Hz), –104.70 (d, 1F, $J = 251.3$ Hz); ^{13}C NMR (100 MHz, CDCl_3): δ 170.3, 136.9, 129.8 (t, $J = 25.1$ Hz), 128.0, 127.6, 127.5, 120.7 (t, $J = 9.6$ Hz), 119.0 (dd, $J = 244.7$ Hz, 243.6 Hz), 73.8 (d, $J = 2.7$ Hz), 72.8, 66.7 (t, $J = 28.3$ Hz), 60.1, 31.0, 20.4; IR (thin film, cm^{-1}): 3032, 2962, 2114, 1740, 1240, 989; MS (ESI): m/z 357.1 ($\text{M}+\text{NH}_4^+$); HRMS Calcd. for $\text{C}_{16}\text{H}_{19}\text{O}_3\text{F}_2\text{N}_3\text{Na}$: 362.1287; Found: 362.1287.

2.7. (3*S*,4*R*)-4-Azido-3-(benzyloxy)-5,5-difluorohept-6-en-1-ol (9)

Compound **8** (0.87 g, 4.06 mmol) was dissolved in MeOH (15 mL). After that, KOH (0.12 g) was added. The resulting mixture was stirred for about 1 h. Water was added and the aqueous phase was extracted with CH_2Cl_2 . Then, the combined organic layers were washed with brine. After the resultant solution was dried over anhydrous Na_2SO_4 and filtered, the solvent was removed *in vacuo*. The residue was purified by flash silica gel column chromatography (petroleum ether/ethyl acetate = 6/1) to give **9** (713 mg, 96%) as a clear oil. $[\alpha]_{\text{D}}^{20} = -49.66$ (c 1.00, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.37–7.27 (m, 5H), 6.09–5.96 (m, 1H), 5.77 (d, 1H, $J = 17.2$ Hz), 5.54 (d, 1H, $J = 11.1$ Hz), 4.62 (dd, 2H, $J = 36.8$ Hz, 26.0 Hz), 4.02–3.98 (m, 1H), 3.73 (t, 2H, $J = 5.6$ Hz), 3.62–3.56 (m, 1H), 1.92 (q, 2H, $J = 6.0$ Hz), 1.73 (br, 1H); ^{19}F NMR (282 MHz, CDCl_3): δ –98.6 (dt, 1F, $J = 253.5$ Hz, 9.9 Hz), –105.24 (dt, 1F, $J = 251.5$ Hz, 11.8 Hz); ^{13}C NMR (100 MHz, CDCl_3): δ 137.5, 130.3 (t, $J = 25.3$ Hz), 128.5, 128.0, 127.9, 121.1 (t, $J = 9.7$ Hz), 119.5 (t, $J = 24.2$ Hz), 75.0 (d, $J = 3.7$ Hz), 73.2, 67.3 (t, $J = 28.3$ Hz), 59.1, 34.8 (d, $J = 1.5$ Hz); IR (thin film, cm^{-1}): 3387, 2929, 2883, 2113, 989; MS (ESI): m/z 268.0 ($[\text{M}-\text{N}_2-\text{H}]^+$); HRMS Calcd. for $\text{C}_{14}\text{H}_{16}\text{O}_2\text{F}_2\text{N}$: 268.1149; Found: 268.1152.

2.8. 1-((2*R*,3*S*)-3-(Benzyloxy)-2-(1,1-difluoroallyl)pyrrolidin-1-yl)prop-2-en-1-one (10)

A solution of compound **9** (3.897 g, 13.12 mmol) in dry CH_2Cl_2 (30 mL) was cooled to 0°C . Et_3N (3.975 g, 39.36 mmol), DMAP (80 mg, 0.656 mmol), and MsCl (5 mL, 65.6 mmol) were added. The mixture was stirred at room temperature for 12 h and then quenched with water. The two layers were separated and the aqueous layer was extracted with CH_2Cl_2 . The combined organic layer was dried over Na_2SO_4 and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 6/1) to give methanesulfonate (4.543 g, 12.08 mmol) as a clear oil. To a solution of methanesulfonate in THF (50 mL) was added Ph_3P (4.75 g, 18.12 mmol) and water (4 mL). The reaction mixture was warmed to 80°C and stirred for 4 h and then the reaction mixture was monitored by TLC. When the starting material was consumed, 10% NaOH (aq., 15 mL) was added and the reaction mixture was stirred for 12 h at room temperature. The reaction mixture extracted with ethyl acetate. The combined organic layer was washed with water and dried over Na_2SO_4 . The residue was dissolved in CH_2Cl_2 (20 mL). Then, K_2CO_3 (3.33 g, 24.16 mmol) and acryloyl chloride (2.18 g, 24.16 mmol) was added. The mixture was stirred at room temperature for 12 h and then quenched with water. The two layers were separated, and the aqueous layer was extracted with CH_2Cl_2 . The combined organic layer was dried over Na_2SO_4 and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 10/1) to give compound **10** (2.54 g, 63%) as a clear oil. $[\alpha]_{\text{D}}^{20} = -56.83$ (c 2.00, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.34–7.26 (m, 5H), 6.57–6.51 (m, 0.5H), 6.41–6.37 (m, 1.5H), 6.23–6.07 (m, 1H), 5.74–5.66 (m, 2H), 5.44 (dd, 1H, $J = 21.2$, 11.2 Hz), 4.95–4.87 (m, 0.5H), 4.70–4.63 (m, 1H), 4.55–4.49 (m, 1H), 4.45–4.38 (m, 0.5H), 4.25–4.08 (m, 1H), 3.74–3.64 (m, 1H), 3.57–3.46 (m, 1H), 2.37–2.12 (m, 2H); ^{19}F NMR (376 MHz, CDCl_3): δ –96.2 (dd, 0.48F, $J = 252.3$ Hz, 10.9 Hz), –102.7 (dt, 0.52F, $J = 249.3$ Hz, 13.9 Hz), –102.8 (dt, 0.48F, $J = 250.8$ Hz, 10.9 Hz), –103.4 (dt, 0.52F, $J = 249.3$ Hz, 12.4 Hz); ^{13}C NMR (100 MHz, CDCl_3): δ 166.1, 137.5, 137.3, 132.4 (t, $J = 24.6$ Hz), 131.9 (t, $J = 24.6$ Hz), 129.1, 128.5, 128.4, 128.2, 127.9, 127.8, 127.7, 127.6, 127.5, 120.7 (t, $J = 11.1$ Hz), 119.5 (t, $J = 9.6$ Hz), 118.3 (t, $J = 245.3$ Hz), 117.4 (t, $J = 244.9$ Hz), 77.8, 76.5, 72.7, 61.4 (t, $J = 23.1$ Hz), 58.1 (t, $J = 26.8$ Hz), 44.0, 43.1, 29.5, 27.5; IR (thin film, cm^{-1}): 3030, 2896, 1655, 1614, 1421; MS

274 (ESI): m/z 308 (M+H⁺); HRMS Calcd. for C₁₇H₁₉O₂F₂NNa: 327
275 330.1276; Found: 330.1270.

276 2.9. (1*S*,8*aR*)-1-(Benzyloxy)-8,8-difluoro-2,3,8,8*a*-
277 tetrahydroindolizin-5(1*H*)-one (**11**)

278 Compound **10** (643 mg, 0.31 mmol) and titanium isopropoxide
279 (180 mg, 0.643 mmol) in dry toluene (20 mL) was refluxed for 3 h
280 under an argon atmosphere. Then Grubbs' II catalyst dissolved in
281 toluene (5 mL) was added dropwise to the mixture. The reaction
282 mixture was stirred at reflux for 10 h. The reaction mixture was
283 cooled to room temperature and concentrated under reduced
284 pressure. The residue was purified by silica gel column chroma-
285 tography (petroleum ether/ethyl acetate = 8/1) to give compound
286 **11** (450 mg, 78% yield) as a yellow oil. [α]_D²⁰ = -716.00 (c 0.51,
287 CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.33–7.24 (m, 5H), 6.52–6.47
288 (m, 1H), 6.20 (d, 1H, J = 7.8 Hz), 4.63 (dd, 2H, J = 19.5 Hz, 9.0 Hz),
289 4.48 (d, 1H, J = 1.8 Hz), 3.97–3.88 (m, 1H), 3.77–3.65 (m, 2H), 2.21–
290 2.15 (m, 1H), 1.93–1.84 (m, 1H); ¹⁹F NMR (376 MHz, CDCl₃): δ
291 -101.4 (dd, 1F, J = 254.9 Hz, 25.6 Hz), -106.4 (dt, 1F, J = 273.7 Hz,
292 8.3 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 161.0 (d, J = 3.3 Hz), 131.7,
293 133.7 (dd, J = 32.6 Hz, 24.3 Hz), 131.0 (dd, J = 11.3 Hz, 8.3 Hz),
294 128.4, 127.8, 127.5, 116.2 (dd, J = 251.3 Hz, 231.5 Hz), 77.85, 72.2
295 (d, J = 1.5 Hz), 62.8 (dd, J = 35.7 Hz, 24.3 Hz), 43.0, 29.9; IR (thin
296 film, cm⁻¹): 2952, 1675, 1616, 1445, 1206; MS (ESI): m/z 302.0
297 (M+Na⁺), 280.0 (M+H⁺); HRMS Calcd. for C₁₅H₁₆O₂F₂N: 280.1144;
298 Found: 280.1154.

299 2.10. (1*S*,6*R*,7*S*,8*aR*)-1-(Benzyloxy)-8,8-difluoro-6,7-
300 dihydroxyhexahydroindolizin-5(1*H*)-one (**12**)

301 To a solution of compound **11** (450 mg, 1.61 mmol) in acetone
302 (5 mL) was added NMNO (438 mg, 3.23 mmol), followed by
303 addition of water (10 mL) at room temperature with stirring.
304 Then a catalytic amount of OsO₄ (5 mol %) solution in water (4%
305 solution) was added. After the reaction mixture was stirred at room
306 temperature for 48 h, it was quenched with saturated NaHSO₃
307 solution and extracted with ethyl acetate. The combined organic
308 layer was washed with brine, dried over anhydrous Na₂SO₄, and
309 filtered. The solvent was removed *in vacuo*. The residue was
310 purified by silica gel column chromatography (petroleum ether/
311 ethyl acetate = 2/1) to give 402 mg (80% yield) of compound **12** as a
312 yellow oil. [α]_D²⁰ = 93.56 (c 1.00, CHCl₃); ¹H NMR (400 MHz,
313 CDCl₃): δ 7.37–7.27 (m, 5H), 4.63 (dd, 2H, J = 14.7 Hz, 9.0 Hz), 4.40–
314 4.32 (m, 2H), 4.32 (s, 1H), 4.15 (d, 1H, J = 19.5 Hz), 3.69–3.65 (m,
315 2H), 2.21–2.15 (m, 1H), 1.97–1.92 (m, 1H); ¹⁹F NMR (376 MHz,
316 CDCl₃): δ -113.6 (dd, 1F, J = 256.0 Hz, 8.3 Hz), -117.7 (dd, 1F,
317 J = 256.0 Hz, 25.6 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 169.0, 137.7,
318 128.4, 127.8, 127.5, 118.6 (t, J = 248.6 Hz), 76.9, 71.9 (d, J = 1.5 Hz),
319 70.3 (dd, J = 32.8, 21.6 Hz), 69.1 (d, J = 8.2 Hz), 59.9 (dd, J = 37.2,
320 19.4 Hz), 43.4, 29.6; IR (thin film, cm⁻¹): 3372, 2896, 1644, 1117,
321 1068; MS (ESI): m/z 314.0 (M+H⁺); HRMS Calcd. for C₁₅H₁₈O₄F₂N:
322 314.1198; Found: 314.1212.

323 2.11. (1*S*,6*S*,7*S*,8*aR*)-1-(Benzyloxy)-8,8-difluoro-octahydroindolizine-
324 6,7-diol (**13**)

325 To a stirred, cooled (0 °C, ice bath) solution of **12** (185 mg,
326 0.585 mmol) in THF (3 mL) was added borane dimethylsulfide

complex (2.0 mol/L in THF, 3.0 mL, 6.0 mmol). After 30 min, the
327 cold bath was removed, and the reaction was heated to reflux and
328 then stirred for 20 h. The reaction was then quenched with MeOH
329 (1 mL) and concentrated under reduced pressure. The residue was
330 dissolved in MeOH (5 mL). Then, the reaction was heated to reflux
331 and stirred for 12 h. The mixture was concentrated under reduced
332 pressure and the residue purified by column chromatography on
333 silica gel (MeOH/CH₂Cl₂ = 1/50) to afford **13** (147 mg, 85%) as
334 a white solid. [α]_D²⁰ = 18.06 (c 1.00, CD₃OD); ¹H NMR (400 MHz,
335 CD₃OD): δ 7.34–7.20 (m, 5H), 4.85 (s, 2H), 4.53 (dd, 2H, J = 11.1 Hz,
336 9.3 Hz), 4.28–4.25 (m, 1H), 3.93–3.84 (m, 2H), 3.14–3.10 (m, 1H),
337 2.93–2.90 (m, 1H), 2.66 (dd, 1H, J = 19.2 Hz, 3.3 Hz), 2.33 (t, 1H,
338 J = 8.4 Hz), 2.26–2.15 (m, 2H), 1.93–1.89 (m, 1H); ¹⁹F NMR
339 (376 MHz, CD₃OD): δ -114.5 (d, 1F, J = 253.5 Hz), -117.1 (dd,
340 1F, J = 254.5 Hz, 26.0 Hz); ¹³C NMR (100 MHz, CD₃OD): δ 138.5,
341 127.6, 127.2, 127.0, 120.0 (t, J = 243.8 Hz), 76.9, 71.3, 70.9 (dd,
342 J = 32.6 Hz, 20.5 Hz), 66.6 (d, J = 6.8 Hz), 63.2 (dd, J = 30.4 Hz,
343 19.0 Hz), 51.9, 51.3, 30.8; IR (thin film, cm⁻¹): 3531, 2952, 1116,
344 1049, 740; MS (ESI): m/z 300.0 (M+H⁺); HRMS Calcd. for
345 C₁₅H₁₉O₃F₂NNa: 322.1225; Found: 322.1212.

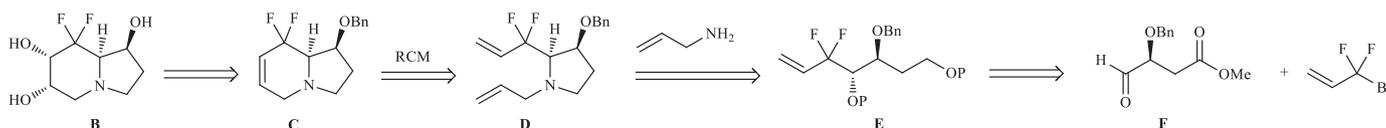
2.12. 7-*epi*-8,8-Difluorocastanospermine (**B**) 347

HCOOH (1.045 mL) was added to a mixture of compound **13** 348
349 (25 mg, 0.084 mmol) and 10% Pd/C (334 mg) in MeOH (5 mL)
350 under an argon atmosphere. The suspension was stirred for 4 h and
351 then filtered through a short pad of celite. The filtrate was
352 concentrated under reduced pressure and the residue was
353 dissolved in water and passed through a column of ion-exchange
354 resin (Dowex 1 × 8, OH-form) eluting with MeOH. The eluent was
355 concentrated under reduced pressure to give 7-*epi*-8,8-difluoro-
356 castanospermine (**B**) (16 mg, 92%) as a colourless solid.
357 [α]_D²⁰ = 20.52 (c 0.50, CD₃OD); ¹H NMR (400 MHz, CD₃OD): δ
358 4.47 (s, 1H), 3.91–3.89 (m, 1H), 3.82 (s, 1H), 3.13 (t, 1H, J = 3.3 Hz),
359 2.93–2.91 (m, 1H), 2.51 (d, 1H, J = 12.0 Hz), 2.33–2.22 (m, 2H), 2.15
360 (q, 1H, J = 8.8 Hz), 1.77–1.69 (m, 1H); ¹⁹F NMR (376 MHz, CD₃OD):
361 δ -112.6 (d, 1F, J = 251.9 Hz), -113.8 (dd, 1F, J = 250.3 Hz, 22.9 Hz);
362 ¹³C NMR (100 MHz, CD₃OD): δ 120.2 (t, J = 247.5 Hz), 70.8 (dd,
363 J = 32.6 Hz, 21.3 Hz), 69.3, 66.9 (d, J = 6.8 Hz), 63.8 (dd, J = 28.8 Hz,
364 19.0 Hz), 51.9, 51.1, 33.1; IR (thin film, cm⁻¹): 3401, 3321, 2938,
365 1079, 1057; MS (ESI): m/z 209.9 (M+H⁺); HRMS Calcd. for
366 C₈H₁₄O₃F₂N: 210.0936; Found: 210.0946.

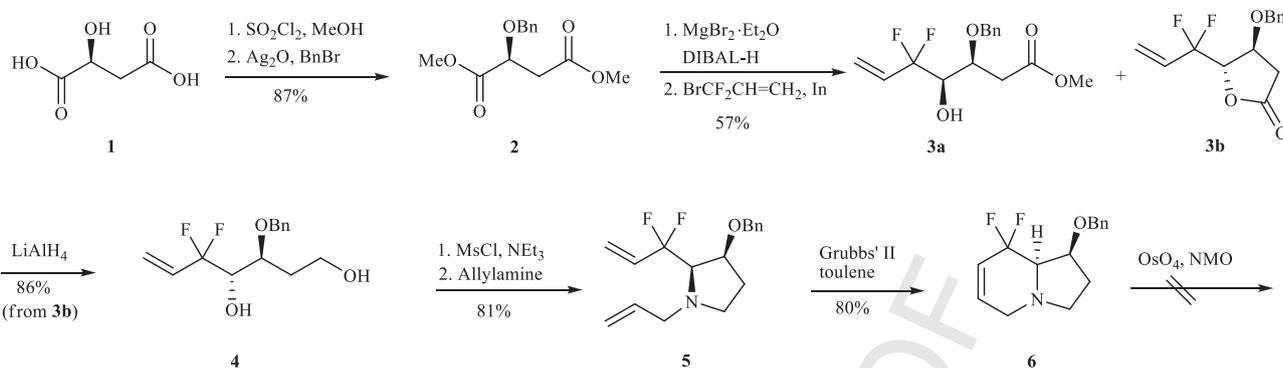
3. Results and discussion 367

368 The retrosynthetic analysis of target molecule **B** is shown in
369 Scheme 2. Compound **B** could be prepared by substrate-controlled
370 *cis*-dihydroxylation of cycloalkene **C**. The six-member ring in
371 compound **C** could be constructed by ring-closing metathesis
372 (RCM) reaction from diene **D**, in which the pyrrolidine ring is easily
373 accessible by intermolecular cyclization of allylamine and
374 compound **E**. Compound **E** is expected to be obtained by coupling
375 of 3-bromo-3,3-difluoropropene and aldehyde **F**.

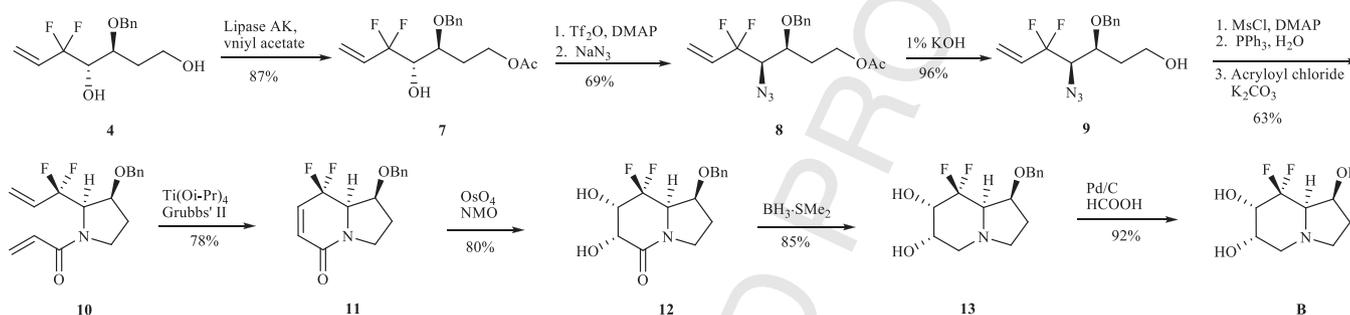
376 Our initial route towards compound **B** commenced from the
377 cheap and commercially available L-(–)-malic acid **1** (Scheme 3).
378 Esterification of compound **1** with SO₂Cl₂/MeOH and then
379 protection of the hydroxyl group under the condition of Ag₂O/
380 BnBr gave compound **2** in high yield. Lewis acid mediated selective
381 reduction of compound **2** produced the desired aldehyde [12],



Scheme 2. Retrosynthetic analysis of target molecule **B**.



Scheme 3. Initial synthetic route.



Scheme 4. Modified synthetic route.

which reacted with 3-bromo-3,3-difluoropropene in the presence of indium affording two diastereoisomers **3a** and **3b**. Interestingly, compound **3b** was obtained in the form of lactone. It was converted into diol **4** in high yield by reduction with LiAlH₄. Reaction of both of the hydroxyl groups in diol **4** with MsCl and subsequent intramolecular cyclization with allylamine afforded the RCM reaction precursor **5**. The RCM reaction of compound **5** with Grubbs' II catalyst proceeded smoothly to afford alkene **6** in 80% yield. To our disappointment, despite trying many different reaction systems [13]. The failure of the dihydroxylation reaction might be ascribed to the coordination of nitrogen atom to the catalyst OsO₄ [14].

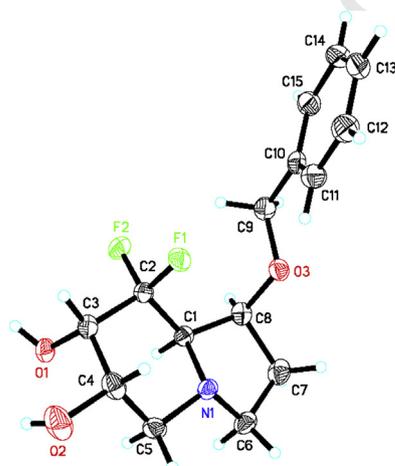
To reduce the coordination ability of nitrogen to osmium, we decided to connect an electron-withdrawing group to the nitrogen atom. The modified synthetic route is shown in Scheme 4. Selective

protection of the primary hydroxyl group in compound **4** with vinyl acetate and *Pseudomonas* (AK) [15] gave the secondary alcohol **7** in 87% yield. Reaction of alcohol **7** with Tf₂O in presence of DMAP afforded the corresponding triflate, which then reacted with NaN₃ to give azide **8**. Cleavage of the O-Ac group in a methanolic solution of 1% KOH afforded the desired alcohol **9**. Mesylation of the hydroxyl group in compound **9** gave the corresponding methanesulfonate. Reduction of azide group with triphenylphosphine and subsequent intramolecular substitution cyclization afforded the pyrrolidine intermediate, which then was directly treated with acryloyl chloride to give diene **10** in 63% overall yield. Considering the high electron-deficient properties of diene **10**, we performed the ring-closing metathesis (RCM) reaction under the reaction conditions developed by our group [16], with Grubbs' II catalyst and co-catalyst Ti(*i*-Pr)₄, affording the desired α,β -unsaturated lactam **11** in 78% yield. Dihydroxylation of lactam **11** catalyzed by OsO₄ proceeded well giving diol **12** as a single isomer. The high diastereoselectivity can be explained by the steric hindrance of benzyl ether. Subsequent reduction of lactam **12** with borane dimethyl sulfide complex gave indolizidine **13** in 85% yield. The absolute configuration of **13** was confirmed by single-crystal X-ray diffraction analysis (Fig. 2). Finally, removal of the benzyl group, via hydrogenolysis in the presence of 10% Pd/C, provided the target molecule 7-*epi*-8,8-difluorocastanospermine **B**.

The synthesized 7-*epi*-8,8-difluorocastanospermine **B** was evaluated for its inhibitory activities against α -glucosidase from baker's yeast and β -glucosidase from almonds. Unfortunately no significant inhibitory activity was observed.

4. Conclusion

In conclusion, we have designed and prepared a novel *gem*-difluoromethylenated castanospermine analogue **B**. Intramolecular cyclization reaction was applied to construct pyrrolidine ring, while RCM reaction was used to achieve the desired bicyclic

Fig. 2. X-ray crystallographic structures of compound **13**.

432 framework. Comparing the two synthetic routes, it was found that
433 amide **11** showed much better reactivity than amine **6** in the
434 dihydroxylation reaction. Thus, the introduction of an electron-
435 withdrawing group to the nitrogen atom was the highlight of the
436 modified route. The synthesis of other difluoromethylenated
437 castanospermine isomers as well as evaluation of their biological
438 activity are currently on progress.

439 Acknowledgments

440 Q3 We thank the National Natural Science Foundation of China
441 (Nos. 21272036, 21332010) and the National Basic Research
442 Program of China (No. 2012CB21600) for funding this work.

443 References

- 444 [1] (a) J.P. Michael, Indolizidine and quinolizidine alkaloids, *Nat. Prod. Rep.* 24 (2007)
445 191-222;
446 (b) J.P. Michael, Indolizidine and quinolizidine alkaloids, *Nat. Prod. Rep.* 25 (2008)
447 139-165.
- 448 [2] (a) I. Pastuszak, R.J. Molyneux, L.F. James, A.D. Elbein, Lentiginosine, a dihydroxy-
449 yndolizidine alkaloid that inhibits amyloglucosidase, *Biochemistry* 29 (1990)
450 1886-1891;
451 (b) A. Brandi, S. Cicchi, F.M. Cordero, et al., Assignment of the absolute configura-
452 tion of natural lentiginosine by synthesis and enzymic assays of optically pure
453 (+) and (-)-enantiomers, *J. Org. Chem.* 60 (1995) 6806-6812.
- 454 [3] (a) M.J. Schneider, F.S. Ungemach, H.P. Broquist, T.M. Harris, (1S,2R,8R,8aR)-1,2,8-
455 trihydroxyoctahydroindolizine (swainsonine), an α -mannosidase inhibitor from
456 *Rhizoctonia leguminicola*, *Tetrahedron* 39 (1983) 29-32;
457 (b) G.P. Kaushal, T. Szumilo, I. Pastuszak, A.D. Elbein, Purification to homogeneity
458 and properties of mannosidase II from mung bean seedlings, *Biochemistry* 29
459 (1990) 2168-2176.
- 460 [4] (a) L.D. Hohenschutz, E.A. Bell, P.J. Jewess, et al., Castanospermine, A 1,6,7,8-
461 tetrahydroxyoctahydroindolizine alkaloid, from seeds of *Castanospermum austral*,
462 *Phytochemistry* 20 (1981) 811-814;
463 (b) R.J. Nash, L.E. Fellows, J.V. Dring, et al., Castanospermine in *Alexa* species,
464 *Phytochemistry* 27 (1988) 1403-1404.
- 465 [5] (a) R. Saul, J.P. Chambers, R.J. Molyneux, A.D. Elbein, Castanospermine, a tetra-
466 hydroxylated alkaloid that inhibits β -glucosidase and β -glucocerebrosidase,
467 *Arch. Biochem. Biophys.* 211 (1983) 593-597;
468 (b) G. Trugnan, M. Rousset, A. Zweibaum, Castanospermine: a potent inhibitor of
469 sucrose from the human enterocyte-like cell line Caco-2, *FEBS Lett.* 195 (1986)
470 28-32;
471 (c) B.C. Campbell, R.J. Molyneux, K.C. Jones, Differential inhibition by casta-
472 nospermine of various insect disaccharidases, *J. Chem. Ecol.* 13 (1987)
473 1759-1770;
474 (d) A.M. Scofield, J.T. Rossiter, P. Witham, et al., Inhibition of thioglucosidase-
475 catalysed glucosinolate hydrolysis by castanospermine and related alkaloids,
476 *Phytochemistry* 29 (1990) 107-109;
477 (e) A.P. Valaitis, D.F. Bowers, Purification and properties of the soluble midgut
478 trehalase from the gypsy moth, *Lymantria dispar*, *Insect Biochem. Mol. Biol.* 23
479 (1993) 599-606.
- 480 [6] (a) H. Nojima, I. Kimura, F.J. Chen, et al., Antihyperglycemic effects of N-contain-
481 ing sugars from *Xanthocercis zambesiaca*, *Morus bombycis*, *Aglaonema treubii*, and
482 *Castanospermum australe* in Streptozotocin-Diabetic Mice, *J. Nat. Prod.* 61 (1998)
483 397-400;
484 (b) R. Pili, J. Chang, R.A. Partis, R.A. et, et al., The α -glucosidase I inhibitor
485 castanospermine alters endothelial cell glycosylation, prevents angiogenesis,
486 and inhibits tumor growth, *Cancer Res.* 55 (1995) 2920-2926;
487 (c) S. Walter, K. Fassbender, E. Gulbins, et al., Glycosylation processing inhibition by
488 castanospermine prevents experimental autoimmune encephalomyelitis by inter-
489 ference with IL-2 receptor signal transduction, *J. Neuroimmunol.* 132 (2002) 1-10;
490 (d) E. De Clercq, Current lead natural products for the chemotherapy of human
491 immunodeficiency virus (HIV) infection, *Med. Res. Rev.* 20 (2000) 323-349;
492 (e) P.M. Grochowicz, A.D. Hibberd, Y.C. Smart, et al., Castanospermine, an

- oligosaccharide processing inhibitor, reduces membrane expression of adhe-
sion molecules and prolongs heart allograft survival in rats, *Transpl. Immunol.* 4
(1996) 275-285.
- [7] (a) T. Machan, A.S. Davis, B. Liawruangrath, S.G. Pyne, Synthesis of castanosper-
mine, *Tetrahedron* 64 (2008) 2725-2732;
(b) T. Jensen, M. Mikkelsen, A. Lauritsen, et al., A concise synthesis of castanosper-
mine by the use of a transannular cyclization, *J. Org. Chem.* 74 (2009)
8886-8889;
(c) J. Ceccon, G. Danoun, A.E. Greene, J.F. Poisson, Asymmetric synthesis of (+)-
castanospermine through enol ether metathesis-hydroboration/oxidation, *Org.*
Biomol. Chem. 7 (2009) 2029-2031;
(d) G. Liu, T.J. Wu, Y.P. Ruan, P.Q. Huang, Asymmetric total syntheses of (+)-
castanospermine, (+)-7-deoxy-6-epi-castanospermine, and (+)-1-epi-castanosper-
mine, *Chem. Eur. J.* 16 (2010) 5755-5768;
(e) E.G. Bowen, D.J. Wardrop, Diastereoselective nitrenium ion-mediated cyclo-
functionalization: total synthesis of (+)-castanospermine, *Org. Lett.* 12 (2010)
5330-5333.
- [8] (a) J. Louvel, C. Botuha, F. Chemla, et al., Asymmetric total synthesis of (+)-6-epi-
castanospermine by the stereoselective formation of a syn, anti acetylenic 2-
amino-1,3-diol stereotriad, *Eur. J. Org. Chem.* (2010) 2921-2926;
(b) N.B. Kalamkar, V.G. Puranik, D.D. Dhavale, Synthesis of C1- and C8a-epimers
of (+)-castanospermine from d-glucose derived γ,δ -epoxyazide: intramolecular
5-endo epoxide opening approach, *Tetrahedron* 67 (2011) 2773-2778;
(c) P.R. Sultane, A.R. Mohite, R.G. Bhat, Total synthesis of 1-deoxy-7,8-di-epi-
castanospermine and formal synthesis of pumiliotoxin-251D, *Tetrahedron* 53
(2012) 5856-5858;
(d) H. Yun, J. Kim, J. Sim, et al., Asymmetric syntheses of 1-deoxy-6,8-di-epi-
castanospermine and 1-deoxy-6-epi-castanospermine, *J. Org. Chem.* 77 (2012)
5389-5393;
(e) A.T. Serafidou, E.G. Yioti, J.K. Gallos, A protection-free synthetic access to (\pm)-
1-deoxy-6-epi-castanospermine and (\pm)-1-deoxy-6,8-di-epi-castanospermine,
Eur. J. Org. Chem. (2013) 939-943.
- [9] R.H. Furneaux, G.J. Gainsford, J.M. Mason, et al., The chemistry of castanosper-
mine, part V: synthetic modifications at C-1 and C-7, *Tetrahedron* 53 (1997)
245-268.
- [10] R.W. Wang, X.L. Qiu, M. Bols, O.C. Fernando, F.L. Qing, Synthesis and biological
evaluation of glycosidase inhibitors: gem-difluoromethylenated nojirimycin ana-
logues, *J. Med. Chem.* 49 (2006) 2989-2997.
- [11] (a) R.W. Wang, F.L. Qing, Highly stereocontrolled synthesis of gem-difluoro-
methylenated azasugars: D- and L-1,4,6-trideoxy-4,4-difluoronojirimycin, *Org.*
Lett. 7 (2005) 2189-2192;
(b) R.J. Li, M. Bols, C. Rousseau, et al., Synthesis and biological evaluation of potent
glycosidase inhibitors: 4-deoxy-4,4-difluoroisofagomine and analogues, *Tetrahe-*
dron 65 (2009) 3717-3727;
(c) R.W. Wang, J. Xu, O. Lopez, M. Bols, F.L. Qing, Difluoromethylenated poly-
hydroxylated pyrrolidines: facile synthesis, crystal structure and biological eval-
uation, *Future Med. Chem.* 1 (2009) 991-997;
(d) Y. Yang, F. Zheng, M. Bols, L.G. Marinescu, F.L. Qing, Synthesis of monofluori-
nated isofagomine analogues and evaluation as glycosidase inhibitors, *J. Fluorine*
Chem. 132 (2011) 838-845.
- [12] G.E. Keck, M.B. Andrus, D.R. Romer, A useful new enantiomerically pure synthon
from malic acid: chelation controlled activation as a route to regioselectivity, *J.*
Org. Chem. 56 (1991) 417-420.
- [13] (a) H.C. Kolb, M.S. VanNieuwenhze, K.B. Sharpless, Catalytic asymmetric dihy-
droxylation, *Chem. Rev.* 94 (1994) 2483-2547;
(b) Z.X. Jiang, Y.Y. Qin, F.L. Qing, Asymmetric synthesis of both enantiomers of
anti-4,4,4-trifluorothreonine and 2-amino-4,4,4-trifluorobutanoic acid, *J. Org.*
Chem. 68 (2003) 7544-7547.
- [14] S.G. Hentges, K.B. Sharpless, Asymmetric induction in the reaction of osmium
tetroxide with olefins, *J. Am. Chem. Soc.* 102 (1980) 4263-4265.
- [15] K. Burgess, L.D. Jennings, Enantioselective esterifications of unsaturated alcohols
mediated by a lipase prepared from *Pseudomonas* sp., *J. Am. Chem. Soc.* 113 (1991)
6129-6139.
- [16] (a) Z.W. You, Y.Y. Wu, F.L. Qing, Synthesis of gem-difluoromethylenated
massoialactone by ring-closing metathesis, *Tetrahedron Lett.* 45 (2004)
9479-9481;
(b) Z.W. You, X. Zhang, F.L. Qing, Stereocontrolled synthesis of gem-difluoro-
methylenated goniodiols and goniothalamin epoxides based on ring-closing
metathesis, *Synthesis* (2006) 2535-2542.

493
494
495
496
497
498
499
500
501
502
503
504
505
506
507
508
509
510
511
512
513
514
515
516
517
518
519
520
521
522
523
524
525
526
527
528
529
530
531
532
533
534
535
536
537
538
539
540
541
542
543
544
545
546
547
548
549
550
551
552
553
554
555
556
557
558
559
560
561