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Abstract: The reactions of 1-(2-haloethyl)pyrrole-2-carbaldehydes with (1*S*)-1-phenylglycinol or (1*S*)-valinol gave the corresponding fused tricyclic oxazolidines as single diastereomers from which 1,2-disubstituted 1,2,3,4-tetrahydropyrrole[1,2-*a*]pyrazines were obtained by addition of organometallic reagents. The diastereoselectivity was dependent on the nature of both the chiral auxiliary and the organometallic reagent. The best diastereoselectivity (dr \leq 98:2) was obtained by using Grignard reagents with the oxazolidine derived from (1*S*)-phenylglycinol. (+)-1-Methyl- and (+)-1-ethyl-1,2,3,4-tetrahydropyrrole[1,2-*a*]pyrazines were obtained by reductive removal of the N2-substituent.

Key words: asymmetric synthesis, diastereoselectivity, Grignard reagents, heterocycles, oxazolidines, tetrahydropyrrolopyrazines

Substituted 1,2,3,4-tetrahydropyrrolo[1,2-a]pyrazines, e.g. 1 (Scheme 1), are extremely useful because of their psychotropic,² antiamnesic, antihypoxic,¹ and antihypersensitive³ activities.⁴ Moreover, a stereochemically defined 3,5,5-trisubstituted 2,4-dioxo derivative is a potent inhibitor of aldose reductase.⁵ 1-Substituted and 1,2-disubstituted 1,2,3,4-tetrahydropyrrolo[1,2-a]pyrazines 1 have been synthesized by hydrogenation or reduction of 2-substituted 3,4-dihydropyrrolo[1,2-a]pyrazines $\mathbf{2}^{1,6-8}$ The dihydro compounds $\mathbf{2}$ are, in turn, prepared by reaction of 2-furylcarbaldehyde or 2-furyl ketones with ethylenediamine,^{9–11} or by addition of Grignard reagents to unsubstituted 2 ($R^1 = H$).¹² Furthermore, hydrogenation of the pyrrole ring under more-forcing conditions has been used to obtain octahydro derivatives which are useful intermediates for the synthesis of coronary dilators and neuroleptics.⁸ Dihydro compounds 2 have also been produced by phosphorus oxychloride-mediated condensation of N-[2-(pyrrol-1-yl)ethyl]carboxamides 3, which are obtained, in turn, from 2,5-dimethoxytetrahydrofuran.^{13,14} The N2-substituted derivatives 1 can be easily prepared from the N-unsubstituted precursors by routine alkylation/acylation reactions.^{2,15} An alternative route to 1 involves the reaction of 1-(2-aminoethyl)pyrroles 4 with two equivalents of formaldehyde and benzotriazole, followed by addition of a Grignard reagent that provided the R¹ substituent. Similarly, 5,6,9,10,11,11a-hexahydro-8H-pyrido[1,2*a*]pyrrolo[2,1-*c*]pyrazines 1 [$R^1R^2 = CH_2CH_2CH_2CH(R)$]

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Scheme 1 Retrosynthetic pathways for substituted 1,2,3,4-tetrahydropyrrolo[1,2-*a*]pyrazines **1**

have been synthesized by the benzotriazole method using glutaric dialdehyde.¹⁶

The same skeleton is present in compounds **5** and **6** (Figure 1). The former, which is a modulator for human metabotropic glutamate receptor 5 (mGluR5), is useful in the control and prevention of chronic neurological disorders.¹⁷ 7-(1,2,3,4-Tetrahydropyrrolo[1,2-*a*]pyrazin-7-yl)quinolones **6** have shown efficacy in vivo in a murine lethal systemic infection model; this efficacy is dependent on the position of the methyl group (R²) at C3 of the tetrahydropyrazine ring and its *S*-configuration. In this case, the 1,2,3,4-tetrahydropyrazine fragment was constructed stereoselectively starting from (4*R*)-hydroxy-L-proline, and the pyrrole ring was obtained by dehydrogenation of an intermediate pyrroline.¹⁸



Figure 1 Drugs containing substituted 1,2,3,4-tetrahydropyrro-lo[1,2-*a*]pyrazine fragments

substituted

ring-closure to the iminium ion 9 by subsequent reaction with an optically pure primary amine should occur in a single step by consecutive formation of two C-N bonds (b and c or c and b). The third and final advantage is that the enhanced reactivity of the iminium ion in comparison with an imine should permit the use of more-convenient

purification

by

column

derivative.

After

product was stored at 4 °C. On the other hand, when aldehyde 10 and 1,2-dichloroethane were stirred overnight in

a two-phase system of 50% aqueous sodium hydroxide

tive synthesis of 1-(pyrrol-2-yl)alkylamines,19 we decided to develop an efficient asymmetric route to the target compound 7 (Scheme 2). Two retrosynthetic pathways were envisaged, both involving the use of 2-pyrrolecar-Grignard reagents. baldehyde (10) as a convenient starting material an an op-We therefore began our investigation by looking for optitically pure primary amine as a chiral auxiliary. The two mal conditions for the conversion of aldehyde 10 into Nroutes differ in the sequence in which the three C-N bonds (2-haloethyl) derivatives **11a** and **11b** (Scheme 3). Treat-(a-c) and one C-C bond (d) are formed, and they therement with potassium hydroxide in dimethyl sulfoxide at fore involve 1-(2-pyrrolyl)alkylamines 8 or cyclic imini-0 °C for one hour followed by slow addition of 1,2-dibroum ions 9 as intermediates.²⁰ In both cases, the chiral moethane (20 equivalents)²² and stirring for twenty-four auxiliary, i.e. the nitrogen substituent R*, induces asymhours resulted in a good conversion to 11a; this was acmetry during the formation of the new stereocenter. Finalcompanied by small amounts of the corresponding N-vily, removal of the chiral auxiliary from 7 should lead to nyl the desired N-unsubstituted 1-substituted 1,2,3,4-tetrahychromatography, the desired 11a was isolated in 69% dropyrrolo[1,2-a]pyrazines in an enantiomerically pure or yield, but this compound decomposed slowly when stored enantiomerically enriched form. at room temperature. To suppress this decomposition, the

1,2,3,4-tetrahydropyrrolo[1,2-a]pyrazines,

Because of the potential value of configurationally pure 1-

and taking into account our experience in the stereoselec-



Scheme 2 Retrosynthetic pathways for chiral 1-substituted 1,2,3,4tetrahydropyrrolo[1,2-a]pyrazines 7

We have previously reported a highly diastereoselective synthesis of N-substituted 1-(2-pyrrolyl)alkylamines 8 from aldehyde 10 through formation of an imine with (1S)-phenylglycinol, followed by the addition of an organolithium reagent (paths c and d).^{19a} The products were converted into the corresponding primary amines by removal of the chiral auxiliary. To obtain the desired compounds 7 from 8, the formation of two C-N bonds would be required (paths a and b). However, in a preliminary attempt to perform this transformation, the insertion of the two-carbon tether between the two nitrogen atoms of 1-(pyrrole-2-yl)-3-butenamine 8 (R = allyl; $R^* = H$) by reaction with 1,2-dibromoethane by heating with sodium hydride in tetrahydrofuran failed. It is possible that this goal could be reached by an appropriate choice of the twocarbon 1,2-dielectrophilic reactant, e.g. chloroacetyl chloride, in a three-step sequence.²¹ However, we reasoned that the alternative route described in Scheme 2 (sequence: a, b, c, d) offers several advantages. The first is that the electron-withdrawing effect of the formyl function results in an increased acidity of the pyrrole N-H bond in aldehyde **10**, facilitating metalation of the pyrrole and the formation of the first C-N bond by reaction with a 1,2-dihaloethane (step *a*). The second advantage is that





Scheme 3 Diastereoselective synthesis of tricyclic oxazolidines 14 and 15 from 1H-pyrrole-2-carbaldehyde

Both halo aldehydes were treated with a slight excess of (1S)-phenylglycinol or (S)-valinol in anhydrous dichloromethane in the presence of anhydrous magnesium sulfate as a dehydrating agent for two days. In all cases, the corresponding bicyclic iminium ions 12 (Scheme 3) were observed in the crude products isolated by filtration of the solid and evaporation of the solvent. The ¹H NMR spectra of 12 in CDCl₃ showed an adsorption for the $H-C=N^+$

proton at $\delta = 9.33$ ppm that was distinctly higher than that of the proton of the formyl group in **11a** (δ = 9.49 ppm). Minor amounts of starting material and another aldehyde, possibly 1-vinyl-2-pyrrolecarbaldehyde, were also detected. Moreover, an equilibrium was slowly established between the iminium 12 and the corresponding oxazolidine, and the chemical shift of the O-CH-N proton was observed at $\delta = 5.24$ ppm. In practice, the crude iminium bromide 12 precipitated from its dichloromethane solution upon addition of diethyl ether and was obtained in 70% yield. However, this result could not be always reproduced when the same procedure was repeated. On the other hand, treatment of the dichloromethane solution with saturated aqueous sodium bicarbonate and extraction of the organic phase gave the pure tricyclic oxazolidine 14 in 87% yield. This last procedure was therefore used to prepare the tricyclic compound 15a and 15b from the corresponding halides 11a and 11b via the intermediate iminium ion 13.24

The ¹H NMR spectra of the tricyclic structures **14** and **15** showed absorptions for the O–CH–N protons of the oxazolidine groups as singlets at $\delta = 5.58$ and 5.24, respectively. At room temperature, minor amounts (about 5%) of another diastereoisomer could be observed, with some difficulty, in the spectrum, but the relevant signals became narrower and more evident at lower temperatures. The structure of **14** in the solid state (Figure 2) was confirmed by means of single-crystal X-ray diffraction studies on crystals grown by slow evaporation of a tetrahydrofuran solution of **14**.²⁵ This structure was found to corresponds to that of the most stable of the possible diastereomers (in which the N atom is also a stereocenter), as calculated at the MM2 level.



Figure 2 ORTEP drawing of compound 14; thermal ellipsoids are at 30% probability

The tricyclic oxazolidines **14** and **15** were used as substrates for reactions with organometallic reagents.²⁶ Reactions of phenylglycinol-derived N-substituted oxazolidines with organometallic reagents have been previously used in the diastereoselective syntheses of secondary amines.^{27,28} In our hands, Grignard reagents proved to be the reagents of choice for the reactions of **14**, giving, in many cases, high yields and high diastereoselectivities of 1,2-disubstituted 1,2,3,4-tetrahydropyrrolo-[1,2-*a*]pyrazines **16** (Scheme 4 and Table 1).

The addition of methylmagnesium bromide in tetrahydrofuran at -78 °C was particularly effective in terms of both the yield and diastereoselectivity; the major diastereomer **16a** (dr 98:2) was obtained in a pure form and 95% yield after column chromatography (entry 1). The stereocontrol progressively decreased on increasing the length of the

Entry ^a	RM	Temp (°C)	Time (h)	dr ^b	Product(s) [Yield (%)] ^c
1	MeMgBr	-78	2	98:2	16a (95)
2	EtMgBr	-78	1	92:8	16b (78) + <i>epi</i> - 16b (3)
3	Me(CH ₂) ₅ MgBr	-78	1	93:7	16d (77) + <i>epi</i> - 16d (6)
4	CH ₂ =CHCH ₂ MgCl	-78	1	50:50	16e (41) + <i>epi</i> - 16e (35)
5	CH ₂ =CHMgCl/TiCl ₄	-78	2 ^d	-	_
6	CH ₂ =CHZnBr	-78 to 20	4	62:38	16e (47) + <i>epi</i> - 16e (24)
7	CH ₂ =CHZn(Et ₂)MgCl ^e	-781	1	76:24	16e (38) + <i>epi</i> - 16e (14)
8	BnMgCl	-78	1	65:35	16f (53) + <i>epi</i> - 16f (29)
9	PhMgBr	-78	1	>98:2	$[16g + epi-16g] (72)^{f}$
10	PhLi	-78 to 20	1	78:22	$[16g + epi-16g] (45)^{f}$

 Table 1
 Addition of Organometallic Reagents to the Tricyclic Oxazolidine 14

^a The reactions were performed by adding the organometallic reagents (4 equiv) to a soln of the imine in anhyd THF under N₂.

^b The diastereomeric ratios (dr) were determined by ¹H NMR analyses of the crude reaction product.

^c Yields refer to pure diastereoisomers isolated by column chromatography (SiO₂).

^d No reaction occurred.

^e The zincate was prepared by adding allylmagnesium chloride to Et₂Zn in THF and stirring for 1 h at 0 °C.

^f Pure diastereoisomers could not be obtained because epimerization occurred during chromatography on the SiO₂ column.



Scheme 4 Addition of organometallic reagents to the tricyclic oxazolidines 14 and 15

alkyl group in the Grignard reagents (entries 2 and 3). Allylmagnesium chloride showed no stereoselectivity (entry 4) and gave no reaction in the presence of titanium tetrachloride (entry 5). We therefore examined the reaction of allylzinc bromide; however, the diastereoselectivity increased only slightly (entry 6). Better stereocontrol (dr 76:24) was achieved by using a preformed mixed zincate (entry 7). Benzylmagnesium chloride gave only a moderate diastereoselectivity (dr 65:35; entry 8). Finally, phenylmagnesium bromide reacted with almost complete stereocontrol to give a high yield of the crude secondary amine with a better than 99:1 dr (entry 9), whereas phenyllithium reacted sluggishly even when the temperature was increased to 20 °C (entry 10). Unfortunately, attempts to purify the crude phenyl-substituted product 16g (entry 9) by chromatography on a silica-gel column resulted in epimerized product in all the eluted fractions. This can be explained by the acidity present in the silica inducing heterolytic cleavage of the benzydrylic C-N bond to form a carbenium ion that is stabilized by both adjacent aromatic rings.

Next, we examined the effectiveness of (S)-valinol as a chiral auxiliary by performing the same organometallic reactions on the oxazolidine 15 (Scheme 4 and Table 2). In all cases, we obtained unsatisfactory stereochemical outcomes, and the diastereomeric ratios ranged from 70:30 to 50:50 when Grignard reagents or butyllithium were used. Allyl(tributyl)stannane/tin tetrachloride and -boron trifluoride systems were totally unreactive, whereas the allyltitanium reagent formed in situ from the Grignard reagent and titanium tetraethoxide afforded 15e with a lower yield than that obtained with the Grignard reagent alone, and the dr remained unsatisfactory. Moreover, column chromatography did not provide adequate separation of the diastereoisomers of the products 17b,e-g. In particular, the phenyl-substituted product 17g, initially obtained with a dr of 65:35, was eluted with an inverted dr of 40:60.

The absolute stereochemistry of the main diastereoisomers could not be unambiguously demonstrated; however, an *R*-configuration can reasonably be postulated by analogy with all the previously reported outcomes of Grignard reactions performed on various N-substituted oxazolidines derived from (S)-phenylglycinol.²⁵ We assume that the same mechanism considered for those reactions also operates with our substrate 14, as shown in Scheme 5. The Lewis acidity of the Grignard reagent is a determinant for a successful reaction, which therefore proceeds by preliminary O-Mg coordination. This leads to the formation of the incipient carbenium ion 18 that undergoes attack of the R nucleophile from the side of the C-O bond being broken. In this way, adduct 20 is formed with an *R*-configuration at the newly formed stereocenter. The formation of a true carbenium ion 19 cannot be excluded and, in this case, a reduced diastereoselectivity might be expected owing to the possible rotation of the nitrogen substituent around the C*-N bond. Moreover, the

Entry ^a	RM	Temp (°C)	Time (h)	dr ^b	Product(s) [Yields (%)] ^c
1	MeMgBr	-78	2	70:30	17a (49) + <i>epi</i> - 17a (17)
2	EtMgBr	-78	1	65:35	$[17b + epi-17b] (68)^d$
3	BuLi	-78	1	70:30 ^e	17c ^f
4	CH ₂ =CHCH ₂ MgCl	-78	1	50:50	$[17e + epi-17e] (64)^d$
5	CH ₂ =CHCH ₂ MgCl/Ti(OEt) ₄	-78	1	54:46	$[17e + epi-17e] (50)^d$
6	CH ₂ =CHCH ₂ SnBu ₃ /SnCl ₄	-78 to 20	4	_f	_
7	CH ₂ =CHCH ₂ SnBu ₃ /BF ₃	-78 to 20	4	_f	_
8	BnMgCl	-78	1	50:50	$[17f + epi-17f] (76)^d$
9	PhMgBr	-78	1	65:35	$[17g + epi-17g] (78)^d$

Table 2Addition of Organometallic Reagents to the Tricyclic Oxazolidine 15

^a The reactions were performed by adding the organometallic reagents (4 equiv) to a soln of the imine in anhyd THF under argon.

^b The diastereomeric ratios (dr) were determined by ¹H NMR analysis of the crude reaction product.

^c Yields refer to pure diastereoisomers isolated by column chromatography (SiO₂).

^d The diastereomers could not be separated by column chromatography.

^e No reaction occurred.

^f The reaction mixture contained mainly unreacted **15** and minor amount of the addition product **17c** (~10%).

^g The product was not isolated.

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lower diastereoselectivity of allylic reagents can be explained in terms of an unfavorable transition state for the γ -attack.



Scheme 5 Mechanism and stereochemical model for the organometallic addition to oxazolidine 14

Finally, we directed our efforts to the removal of the chiral auxiliary. Unfortunately, all efforts to achieve this goal by oxidative reactions of the amine **16** or amine **17**, including the use of periodic acid/methylamine and lead tetraacetate, were unsuccessful. Furthermore, hydrogenolysis of **16b** with ammonium formate and palladium/carbon gave a complex mixture of products. Finally, when **16a** or **16b** was subjected to 7 bar of hydrogen pressure in the presence of 10% palladium/carbon in methanol for two days, the desired secondary amines **21a** and **21b**, respectively, were obtained in about 80% yield together with 2-phenylethanol and trace amounts of the corresponding fully hydrogenated compounds **22** (Scheme 6).



Scheme 6 Removal of the chiral auxiliary from 1,2-disubstituted 1,2,3,4-tetrahydropyrrolo[1,2-*a*]pyrazines **16a,b**

In conclusion, we have developed the first asymmetric synthesis of 1-substituted 1,2,3,4-tetrahydropyrrolo[1,2-a]pyrazines by a four-step route starting from 1*H*-pyrrole-2-carbaldehyde. The key intermediate, a tricyclic oxazo-lopyrrolopyrazine, was formed efficiently and diastereo-selectively by condensation of a 1-(2-haloethyl)-2-pyrrolecarbaldehyde with (1*S*)-phenylglycinol. This intermediate reacted with Grignard reagents to give 1,2-disubstituted 1,2,3,4-tetrahydropyrrolo[1,2-a]pyrazines with variable levels of diastereoselectivity that decreased with increasing the length of the alkyl group in the Grignard reagent. The highest diastereomeric ratios were obtained with methylmagnesium bromide (98:2) and phenylmagnesium bromide (>98:2). In the latter case, the product could not be purified by chromatography on silica gel be-

cause considerable epimerization occurred. The chiral auxiliary was removed from 1-methyl- and 1-ethyl-substituted products by hydrogenolysis to give the corresponding unsubstituted cyclic amines with a presumed R-configuration at the C1 stereocenter.

Melting points are uncorrected. Optical rotations were measured on a digital polarimeter in a 1-dm cell, and $[\alpha]_D^{20}$ values are given in 10^{-1} deg cm³ g⁻¹. ¹H NMR spectra were recorded on Varian MR400 and Gemini 200 instruments for samples in CDCl₃ which was stored over Mg. The ¹H chemical shifts are reported in ppm relative to CHCl₃ ($\delta_H = 7.27$) and the *J*-values are given in Hz. IR spectra were recorded on a Nicolet FT-380 spectrometer, and IR assignments are reported in wave numbers (cm⁻¹). MS spectra were recorded at an ionizing voltage of 70 eV on a Hewlett-Packard 5975 spectrometer with GC injection from a HP-5 column (30 m; ID 0.25 mm). Molecular weights were determined on an Agilent Technologies MS 1100 instrument. Chromatographic separations were performed on columns of SiO₂ (Merck; 230–400 mesh) at medium pressure. All the organic, inorganic, and organometallic reagents and anhydrous solvents were purchased from Aldrich.

1-(2-Bromoethyl)-1H-pyrrole-2-carbaldehyde (11a)

KOH (5.88 g, 100 mmol) was added in one portion to a stirred soln of 1*H*-pyrrole-2-carbaldehyde (**10**; 1.00 g, 10 mmol) in anhyd DMSO (6 mL) at r.t. After 1 h, Br(CH₂)₂Br (18.1 mL, 0.2 mol) was slowly added at 0 °C and the resulting soln was stirred overnight at r.t. The reaction was quenched by addition of H₂O (10 mL), and the organic layer was extracted with EtOAc (3×20 mL). The collected organic layers were washed with brine (20 mL), dried (Na₂SO₄), and concentrated to give a crude product that was purified by flash column chromatography [SiO₂, cyclohexane–EtOAc (9:1)] to give a yellow oil; yield: 1.59 g (75%).

IR (neat): 3111, 2947, 2807, 2722, 1660, 1530, 1479, 1321, 1208, 1075, 822, 623, 607 cm⁻¹.

¹H NMR (200 MHz, CDCl₃): δ = 9.49 (s, 1 H), 7.04–6.99 (m, 2 H), 6.29–6.25 (m, 1 H), 4.80 (t, *J* = 5.8 Hz, *J* = 6.2 Hz, 2 H), 3.70 (t, *J* = 5.8 Hz, *J* = 6.2 Hz, 2 H).

¹³C NMR (50 MHz, CDCl₃): δ = 179.4, 150.9, 132.5, 125.6, 109.6, 50.6, 31.7.

MS (EI): *m*/*z* = 122 (100), 94 (36), 201 (14), 203 (12), 202 (3).

Anal. Calcd for C₇H₈BrNO (202.05): C, 41.61; H, 3.99; N, 6.93. Found: C, 41.50; H, 4.01; N, 6.91.

1-(2-Chloroethyl)-1*H*-pyrrole-2-carbaldehyde (11b)²³

1,2-Dichloroethane (9.7 mL, 0.12 mol), Bu_4NI (1.94 g, 5.3 mmol), and 50% aq NaOH (5 mL) were added to a soln of 1*H*-pyrrole-2carbaldehyde (**10**; 0.500 g, 5.3 mmol) in CH₂Cl₂ (2 mL), and the mixture was vigorously stirred overnight. H₂O (2 mL) was added and the organic layers were extracted with CH₂Cl₂ (2 × 30 mL) and washed with 1 M HCl (2 × 20 mL), sat. aq NaHCO₃ (2 × 20 mL), and brine (2 × 20 mL). After drying (Na₂SO₄), the organic layers were concentrated in vacuo to give an orange slurry that was purified by column chromatography [SiO₂, cyclohexane–EtOAc (9:1)] to give a yellow oil; yield: 0.76 g (92%).

IR (neat) 3111, 2959, 2809, 1663, 1655, 1479, 882, 766, 705, 678, 656, 608 $\rm cm^{-1}.$

¹H NMR (200 MHz, CDCl₃): δ = 9.51 (s, 1 H), 7.03–6.97 (m, 2 H), 6.23 (t, *J* = 2.6 Hz, 1 H), 4.57 (t, *J* = 5.8 Hz, *J* = 5.4 Hz, 2 H), 3.79 (t, *J* = 5.8 Hz, *J* = 5.4 Hz, 2 H).

¹³C NMR (50 MHz, CDCl₃): δ = 179.2, 132.7, 130.7, 125.4, 109.5, 50.5, 43.7.

MS (EI): *m*/*z* = 122 (100), 94 (50), 157 (34), 53 (19), 108 (18), 80 (14).

Anal. Calcd for C_7H_8 ClNO (157.6): C, 53.35; H, 5.12; N, 8.89. Found: C, 53.19; H, 5.14; N, 8.86.

2-[(1*S*)-2-Hydroxy-1-phenylethyl]-3,4-dihydropyrrolo[1,2*a*]pyrazin-2-ium bromide (12)

A mixture of carbaldehyde **11a** (0.203 g, 1.0 mmol), (1*S*)-phenylglycinol (0.138 g, 1.2 mmol), and MgSO₄ (0.5 g) in anhyd CH₂Cl₂ (4 mL) was protected from light and magnetically stirred for 2 d under an inert atmosphere. The mixture was then filtered through a small pad of Celite that was washed with CH₂Cl₂. The filtered soln was concentrated under reduced pressure to a final volume of 1 mL, and Et₂O (3 mL) was slowly added to give a brown precipitate; yield: 0.228 g (71%).

IR (neat): 3381, 2962, 2924, 2848, 1646, 1629, 1491, 1377, 1103, 1067 cm⁻¹.

¹H NMR (400 MHz, CDCl₃): δ = 9.33 (s, 1 H), 7.39–7.26 (m, 7 H), 6.44–6.36 (m, 1 H), 5.56–5.49 (m, 1 H), 5.39 (br s, 1 H), 4.60–4.20 (m, 3 H), 4.10–3.93 (m, 2 H), 3.80–3.60 (m, 1 H).

¹³C NMR (100 MHz, CDCl₃): δ = 151.8, 134.5, 132.7, 129.5, 127.9, 127.3, 122.6, 115.4, 72.5, 60.1, 45.9, 43.4.

MS (ES): $m/z = 241.1 [M - HBr + H]^+ (100)$.

(3*S*,10*bR*)-3-Phenyl-2,3,5,6-tetrahydro-10*bH*-[1,3]oxazolo[3,2*a*]pyrrolo[2,1-*c*]pyrazine (14); Typical Procedure

A mixture of carbaldehyde **11a** (1.515 g, 7.5 mmol), (1*S*)-phenylglycinol (1.233 g, 9.0 mmol), and MgSO₄ (5 g) in anhyd CH₂Cl₂ (15 mL) was protected from light and stirred for 2 d under an inert atmosphere. The mixture was then filtered through a small pad of Celite that was washed with CH₂Cl₂. The collected organic layers were washed with sat. aq NaHCO₃ (20 mL) and brine (20 mL), and the organic phase was concentrated under reduced pressure to give a white solid; yield: 1.531 g (85%); mp 85.1–85.5 °C; $[\alpha]_D^{20}$ +8.5 (*c* 1.0, CHCl₃).

IR (KBr): 3101, 3027, 2847, 1600, 1449, 1212, 1190, 873, 798, 757, 698 cm⁻¹.

¹H NMR (400 MHz, CDCl₃): δ = 7.49–7.34 (m, 5 H), 6.63 (dd, J = 1.6 Hz, J = 2.8 Hz, 1 H), 6.28 (d, J = 2.2 Hz, 1 H), 6.20 (t, J = 3.0 Hz, 1 H), 5.53 (s, 1 H), 4.50 (t, J = 7.9 Hz, 1 H), 4.30 (t, J = 6.8 Hz, 1 H), 4.14–4.03 (m, 2 H), 3.76 (dd, J = 6.6 Hz, J = 8.5 Hz, 1 H), 3.37–3.27 (m, 1 H), 3.16–3.07 (m, 1 H).

 ^{13}C NMR(100 MHz, CDCl₃): δ = 141.4, 134.9, 128.7, 127.3, 126.5, 119.5, 108.7, 107.6, 86.6, 71.2, 67.8, 47.4, 44.1.

MS (EI): *m/z* = 239 (100), 210 (20), 106 (15).

Anal. Calcd for $C_{15}H_{16}N_2O$ (240.30): C, 74.97; H, 6.71; N, 11.66. Found: C, 74.78; H, 6.74; N, 11.62.

(3*S*,10*bR*)-3-Isopropyl-2,3,5,6-tetrahydro-10*bH*-[1,3]oxazo-lo[3,2-*a*]pyrrolo[2,1-*c*]pyrazine (15)

This was prepared by the same procedure as **14**, starting from **11a** (1.515 g, 7.5 mmol).

Yellow oil; yield: 1.443 g (92%). [α]_D²⁰ –13.0 (*c* 1.2, CHCl₃).

IR (neat): 3101, 2957, 2868, 1663, 1494, 1469, 1299, 1195, 1037, 861, 769, 609 cm⁻¹.

¹H NMR (400 MHz, CDCl₃): $\delta = 6.59$ (s, 1 H), 6.28–6.24 (m, 1 H), 6.19 (t, J = 2.9 Hz, 1 H), 5.24 (s, 1 H), 4.14 (t, J = 8.0 Hz, 1 H), 4.06 (ddd, J = 3.5 Hz, J = 12.0 Hz, 1 H), 3.98–3.88 (m, 1 H), 3.50 (dd, J = 6.0 Hz, J = 8.1 Hz, 1 H), 3.21 (dt, J = 3.4 Hz, J = 11.5 Hz, 1 H), 2.86–2.78 (m, 1 H), 2.74–2.64 (m, 1 H), 1.74–1.64 (m, 1 H) 1.07 (d, J = 6.4 Hz, 3 H), 0.88 (d, J = 6.4 Hz, 3 H).

MS (EI): *m/z* = 205 (100), 161 (90), 176 (13).

Anal. Calcd for $C_{12}H_{18}N_2O$ (206.28): C, 69.87; H, 8.80; N, 13.58. Found: C, 69.60; H, 8. 83; N, 13.54.

(2*S*)-2-[(1*R*)-1-Methyl-3,4-dihydropyrrolo[1,2-*a*]pyrazin-2(1*H*)-yl]-2-phenylethanol (16a); Typical Procedure

A 3.0 M soln of MeMgCl in Et₂O (1.3 mL, 4.0 mmol) was added to a magnetically stirred soln of tricycle **14** (0.240 g, 1.0 mmol) in anhyd THF (15 mL) at -78 °C, and the mixture was stirred until no starting material remained (TLC). The reaction then was quenched by adding sat. aq NaHCO₃ (10 mL), and the organic material was extracted with Et₂O (3 × 10 mL). The collected ethereal layers were dried (Na₂SO₄) and concentrated to give an oil. The diastereomeric ratio was determined by ¹H NMR analysis of a sample soln in CDCl₃. Flash column chromatography [SiO₂, cyclohexane–EtOAc (9:1]) gave a yellow oil; yield: 0.253 g (99%): [α]_D²⁰ –21.4 (*c* 1.3, CHCl₃).

IR (neat): 3415, 3101, 3050, 2970, 2928, 1601, 1492, 1453, 1266, 1186, 1056, 735, 609 cm⁻¹.

¹H NMR (400 MHz, CDCl₃): δ = 7.45–7.29 (m, 5 H), 6.50 (t, J = 1.7 Hz, 1 H), 6.15 (t, J = 3.0 Hz, 1 H), 5.85–5.80 (m, 1 H), 4.30 (q, J = 6.8 Hz, J = 13.0 Hz, 1 H), 4.02–3.90 (m, 3 H), 3.82–3.75 (m, 1 H), 3.43 (t, J = 6.7 Hz, 1 H), 3.8–3.21 (m, 1 H), 3.10–3.02 (m, 1 H), 1.39 (d, J = 6.6 Hz, 3 H).

¹³C NMR (100 MHz, CDCl₃): δ = 140.4, 132.1, 128.6, 128.4, 128, 3, 117.9, 107.8, 102.9, 65.4, 63.2, 50.8, 43.38, 41.3, 19.2.

MS (ES): $m/z = 257.2 [M + H]^+ (100)$.

Anal. Calcd for $C_{16}H_{20}N_2O$ (256.34): C, 74.97; H, 7.86; N, 10.93. Found: C, 74.75; H, 7.88; N, 10.90.

(2*S*)-2-[(1*R*)-1-Ethyl-3,4-dihydropyrrolo[1,2-*a*]pyrazin-2(1*H*)-yl]-2-phenylethanol (16b)

Yellowish oil; yield: 0.210 g (78%); $[\alpha]_D^{20}$ –10.5 (*c* 0.5, CHCl₃).

IR (neat): 3411, 3109, 3046, 2974, 2921, 1605, 1457, 1174, 1050, 736 $\rm cm^{-1}$

¹H NMR (400 MHz, CDCl₃): δ = 7.40–7.29 (m, 5 H), 6.52 (t, J = 1.8 Hz, 1 H), 6.12 (t, J = 3.2 Hz, 1 H), 5.79–6.75 (m, 1 H), 4.04 (dt, J = 4.6 Hz, J = 11.8 Hz, 1 H), 3.96–3.88 (m, 3 H), 3.82–3.73 (m, 2 H), 3.46–3.36 (m, 1 H), 3.22–3.14 (m, 1 H), 1.86–1.74 (m, 1 H), 1.71–1.58 (m, 1 H), 0.92 (t, J = 7.4 Hz, 3 H).

¹³C NMR (100 MHz, CDCl₃): δ = 140.5, 130.0, 128.5, 127.7, 118.3, 107.4, 104.0, 65.2, 63.6, 56.4, 41.6, 40.9, 26.8, 11.0.

MS (ES): $m/z = 271.2 [M + H]^+ (100)$.

Anal. Calcd for $C_{17}H_{22}N_2O$ (270.37): C, 75.52; H, 8.20; N, 10.36. Found: C, 75.68; H, 8.21; N, 10.32.

epi-16b

Yellowish oil; yield: 0.008 g (3%); $[\alpha]_D^{20}$ +6.8 (*c* 1.2, CHCl₃).

IR (neat): 3414, 3107, 3054, 2978, 2923, 1611, 1497, 1451, 1263, 1051, 736 $\rm cm^{-1}.$

¹H NMR (400 MHz, CDCl₃): δ = 7.38–7.24 (m, 5 H), 6.47 (t, J = 1.8 Hz, 1 H), 6.12 (t, J = 2.9 Hz, 1 H), 5.85–5.81 (m, 1 H), 4.20 (dd, J = 5.2 Hz, J = 9.6 Hz, 1 H), 4.07 (dd, J = 10.4 Hz, 1 H), 3.98 (t, J = 4.2 Hz, 1 H), 3.96–3.84 (m, 2 H), 3.76 (dd, J = 4.9 Hz, J = 10.8 Hz, 1 H), 3.28–3.20(m, 1 H), 3.02 (br s, 1 H), 2.54–2.45 (m, 1 H), 2.21–2.14 (m, 1 H), 1.98–1.88 (m, 1 H), 0.92 (t, J = 7.3 Hz, 3 H).

 ^{13}C NMR (100 MHz, CDCl₃): δ = 135.3, 129.9, 129.0, 128.4, 118.1, 107.9, 103.5, 62.4, 60.7, 55.9, 44.3, 41.7, 25.6, 8.3.

MS (ES): $m/z = 271.2 [M + H]^+ (100)$.

(2S)-2-[(1R)-1-Hexyl-3,4-dihydropyrrolo[1,2-*a*]pyrazin-2(1H)-yl]-2-phenylethanol (16d)

Light-brown oil; yield: 0.251 g (77%); $[\alpha]_D^{20}$ –5.9 (*c* 0.6, CHCl₃).

¹H NMR (400 MHz, CDCl₃): δ = 7.39–7.28 (m, 5 H), 6.52 (t, J = 2.2 Hz, 1 H), 6.11 (t, J = 3.2 Hz, 1 H), 5.78–5.76 (m, 1 H), 4.05 (dt, J = 4.7 Hz, J = 11.7 Hz, 1 H), 3.97–3.91 (m, 3 H), 3.80–3.74 (m, 1 H), 3.50–3.38 (m, 1 H), 3.21–3.16 (m, 1 H), 1.85–1.71 (m, 1 H), 1.70–1.55 (m, 1 H), 1.41–1.18 (m, 10 H), 0.90 (t, J = 6.7 Hz, 3 H).

¹³C NMR (100 MHz, CDCl₃): δ = 140.5, 128.6, 128.4, 127.8, 118.3, 107.5, 104.0, 65.3, 63.8, 54.9, 41.4, 40.9, 34.1, 31.8, 29.2, 26.4, 22.6, 14.0.

MS (ES): $m/z = 327.1 [M + H]^+ (100)$.

Anal. Calcd for $C_{21}H_{30}N_2O$ (326.48): C, 77.26; H, 9.26; N, 8.58. Found: C, 77.56; H, 9.29; N, 8.55.

epi-16d

Yellow oil; yield: 0.020 g (6%); $[\alpha]_D^{20}$ +5.4 (*c* 0.9, CHCl₃).

¹H NMR (400 MHz, CDCl₃): δ = 7.41–7.28 (m, 5 H), 6.60 (t, *J* = 2.2 Hz, 1 H), 6.20 (t, *J* = 2.5 Hz, 1 H), 5.80–5.75 (m, 1 H), 4.36–4.26 (m, 1 H), 4.25–4.16 (m, 2 H), 3.72–3.66 (m, 1 H), 3.37–3.27 (m, 1 H), 3.15–3.08 (m, 1 H), 2.04–1.98 (m, 2 H), 1.40–1.18 (m, 10 H), 0.90 (t, *J* = 6.8 Hz, 3 H).

 ^{13}C NMR (100 MHz, CDCl₃): δ = 139.4, 131.3, 128.6, 127.8, 127.5, 118.6, 107.9, 104.9, 72.7 63.0, 42.2, 31.8, 30.1, 29.6, 29.4, 26.8, 23.6, 22.6, 14.1.

MS (ES): $m/z = 327.1 [M + H]^+ (100)$.

(2S)-2-[(1R)-1-Allyl-3,4-dihydropyrrolo[1,2-*a*]pyrazin-2(1*H*)yl]-2-phenylethanol (16e)

Light-brown oil; yield: 0.107 g (38%); $[a]_D^{20}$ +18.3 (*c* 0.9, CHCl₃). IR (neat): 3416, 3068, 2924, 2852, 1639, 1569, 1492, 1333, 1290,

 $1069, 1029, 914, 702 \text{ cm}^{-1}$.

¹H NMR (400 MHz, CDCl₃): δ = 7.39–7.26 (m, 5 H), 6.52 (t, *J* = 1.7 Hz, 1 H), 6.11 (t, *J* = 2.9 Hz, 1 H), 5.88–5.80 (m, 1 H), 5.79–5.76 (m, 1 H), 5.09–4.99 (m, 2 H), 4.06 (t, *J* = 6.7 Hz, 1 H), 4.03–3.93 (m, 2 H), 3.92–3.87 (m, 2 H), 3.79–3.72 (m, 1 H), 3.46–3.38 (m, 1 H), 3.20–3.13 (m, 1 H), 2.63–2.52 (m, 1 H), 2.47–2.38 (m, 1 H).

¹³C NMR (100 MHz, CDCl₃): δ = 140.2, 136.1, 129.5, 128.5, 128.4, 127.8, 118.4, 116.8, 107.5, 103.9, 65.7, 63.4, 54.6, 41.7, 41.6, 38.8.

MS (ES): $m/z = 283.2 [M + H]^+ (100), 305.2 [M + Na]^+ (15).$

Anal. Calcd for $C_{18}H_{22}N_2O$ (282.38): C, 76.56; H, 7.85; N, 9.92. Found: C, 76.58; H, 7.87; N, 9.90.

epi-16e

 $\overline{\text{Yellowish oil}}$; yield: 0.039 g (14%); [α]_D²⁰ –13.4 (*c* 0.8, CHCl₃).

IR (neat): 3415, 3078, 2975, 2926, 2849, 2793, 1639, 1496, 1485, 1339, 1219, 1119, 1002, 849, 704 cm⁻¹.

¹H NMR (400 MHz, $CDCl_3$): $\delta = 7.38-7.23$ (m, 5 H), 6.47 (t, J = 1.6 Hz, 1 H), 6.12 (t, J = 3.3 Hz, 1 H), 5.89–5.86 (m, 1 H), 5.80–5.67 (m, 1 H), 5.22 (s, 1 H), 5.14 (t, J = 11.6 Hz, 1 H), 4.22 (dd, J = 4.8 Hz, J = 9.7 Hz, 1 H), 4.13 (t, J = 4.1 Hz, 1 H), 4.05 (t, J = 10.6 Hz, 1 H), 3.94–3.87 (m, 2 H), 3.73 (dd, J = 4.8 Hz, J = 10.9 Hz, 1 H), 3.26–3.18 (m, 1 H), 2.94–2.85 (m, 1 H), 2.76–2.68 (m, 1 H), 2.55–2.47 (m, 1 H), 1.57 (br s, 1 H).

¹³C NMR (100 MHz, CDCl₃): δ = 136.0, 134.6, 128.9, 128.4, 128.2, 118.2, 117.7, 108.0, 103.8, 62.7, 60.6, 54.7, 44.2, 41.5, 38.3.

MS (ES): $m/z = 283.2 [M + H]^+ (100), 305.1 [M + Na]^+ (23).$

(2S)-2-[(1R)-1-Benzyl-3,4-dihydropyrrolo[1,2-*a*]pyrazin-2(1*H*)-yl]-2-phenylethanol (16f)

Yellowish oil; yield: 0.176 g (53%); $[\alpha]_D^{20}$ –86.2 (*c* 1.0, CHCl₃).

¹H NMR (400 MHz, CDCl₃): δ = 7.34–7.03 (m, 10 H), 6.48 (dd, J = 1.7 Hz, J = 2.6 Hz, 1 H), 6.06 (t, J = 3.1 Hz, 1 H), 5.48 (dd, J = 1.4 Hz, J = 3.4 Hz, 1 H), 4.14 (t, J = 7.4 Hz, 1 H), 4.10–3.98 (m, 1 H), 3.90 (t, J = 5.1 Hz, 1 H), 3.79–3.70 (m, 2 H), 3.67–3.58 (m, 1 H), 3.36–3.24 (m, 2 H), 3.13 (dd, J = 7.9 Hz, J = 13.2 Hz, 1 H), 2.83 (dd, J = 6.3 Hz, J = 13.2 Hz, 1 H).

¹³C NMR (100 MHz, CDCl₃): δ = 140.0, 139.2, 131.2, 129.6, 129.1, 128.5, 128.4, 128.2, 128.1, 127.7, 127.6, 127.5, 126.2, 118.3, 107.4, 104.2, 66.1, 63.8, 56.3, 41.5, 41.3, 40.9.

MS (ES): $m/z = 333.2 [M + H]^+ (100)$.

Anal. Calcd for $C_{22}H_{24}N_2O$ (332.44): C, 79.48; H, 7.28; N, 8.43. Found: C, 79.28; H, 7.31; N, 8.41.

epi-16f

Yellowish oil; yield: 0.096 g (29%); $[\alpha]_D^{20}$ +14.3 (*c* 1.6, CHCl₃).

¹H NMR (400 MHz, CDCl₃): δ = 7.32–7.12 (m, 10 H), 6.46 (m, 1 H), 6.12 (t, *J* = 3.1 Hz, 1 H), 5.77 (m, 1 H), 4.38 (t, *J* = 6.4 Hz, 1 H), 4.04 (dd, *J* = 4.9 Hz, *J* = 7.1 Hz, 1 H), 3.75 (dd, *J* = 7.7 Hz, *J* = 11.0 Hz, 1 H), 3.71–3.54 (m, 3 H), 3.30–3.21 (m, 2 H), 3.10 (dd, *J* = 5.8 Hz, *J* = 13.2 Hz, 1 H), 2.75 (ddd, *J* = 4.2 Hz, *J* = 5.8 Hz, *J* = 13.2 Hz, 1 H).

 ^{13}C NMR (100 MHz, CDCl₃): δ = 138.8, 137.2, 129.5, 128.7, 128.4, 128.3, 128.2, 128.0, 126.5, 118.2, 104.2, 64.4, 61.7, 56.7, 42.5, 41.9, 40.4.

MS (ES): $m/z = 333.1 [M + H]^+ (100)$.

(2*S*)-2-Phenyl-2-[(1*R*/1*S*)-1-phenyl-3,4-dihydropyrrolo[1,2*a*]pyrazin-2(1*H*)-yl]ethanol (16g + *epi*-16g) Oil; yield: 0.141 g (45%); dr 60:40.

Representative signals for **16g** in the ¹H NMR spectrum (400 MHz, CDCl₃) of the mixture were observed at $\delta = 6.61$ (dd, J = 2.0 Hz, J = 2.8 Hz, 1 H), 6.15 (dd, J = 2.8 Hz, J = 3.6 Hz, 1 H), 5.59 (m, 1 H), 5.06 (s, 1 H), 3.29 (m, 1 H), 3.21 (m, 1 H).

MS (ES): $m/z = 319.1 [M + H]^+ (100)$.

Representative ¹H NMR signals for *epi*-**16g**: $\delta = 6.51$ (t, J = 1.6 Hz, 1 H), 6.03 (t, J = 3.2 Hz, 1 H), 5.27 (m, 1 H), 4.79 (s, 1 H), 4.22 (dd, J = 4.6 Hz, J = 10.6 Hz, 1 H), 4.22 (ddd, J = 1.9 Hz, J = 3.0 Hz, J = 12.1 Hz, 1 H), 2.59 (ddd, J = 3.7 Hz, J = 12.1 Hz, J = 12.2 Hz, 1 H).

MS (ES): $m/z = 319.1 [M + H]^+ (100)$.

Anal. Calcd for $C_{21}H_{22}N_2O$ (318.41): C, 79.21; H, 6.96; N, 8.80. Found: C, 79.02; H, 6.97; N, 8.78.

(2*S*)-3-Methyl-2-[(1*R*)-1-methyl-3,4-dihydropyrrolo[1,2*a*]pyrazin-2(1*H*)-yl]butan-1-ol (17a)

Yellowish oil; yield: 0.033 g (15%); $[\alpha]_D^{20}$ +23.4 (*c* 1.0, CHCl₃).

IR (neat): 3361, 2968, 2934, 2853, 1466, 1449, 1367, 1315, 1114, 1080, 1069, 1019, 886, 733, 715 cm⁻¹.

¹H NMR (400 MHz, CDCl₃): $\delta = 6.56-6.53$ (m, 1 H), 6.18 (t, J = 3.3 Hz, 1 H), 5.91–5.89 (m, 1 H), 4.34 (q, J = 6.2 Hz, J = 12.2 Hz, 1 H), 4.00–3.92 (m, 2 H), 3.62 (dd, J = 5 Hz, J = 10.2 Hz, 1 H), 3.31 (t, J = 10.6 Hz, 1 H), 3.19–3.12 (m, 1 H), 3.10–3.00 (m, 1 H), 2.99–2.90 (m, 1 H), 2.00–1.90 (m, 1 H), 1.47 (d, J = 6.4 Hz, 3 H), 1.08 (d, J = 6.4 Hz, 3 H), 0.90 (d, J = 6.9 Hz, 3 H).

 ^{13}C NMR (100 MHz, CDCl₃): δ = 132.5, 118.4, 108.0, 103.2, 63.1, 58.8, 53.0, 45.7, 41.4, 27.9, 22.6, 20.9, 19.9.

MS (EI): *m/z* = 207 (100), 121 (53), 191 (35), 222 (1).

MS (ES): $m/z = 223.1 [M + H]^+ (100)$.

Anal. Calcd for C₁₃H₂₂N₂O (222.33): C, 70.23; H, 9.97; N, 12.60. Found: C, 70.19; H, 10.00; N, 12.58.

epi-(17a)

Yellow oil; yield: 0.011 g (5%); $[α]_D^{20}$ –15.2 (*c* 1.0, CHCl₃).

IR (neat) 3407, 2954, 2921, 1581, 1450, 1348, 1066, 1029, 731, 702 cm^{-1} .

¹H NMR (400 MHz, CDCl₃): $\delta = 6.53-6.50$ (m, 1 H), 6.15 (t, J = 3.0 Hz, 1 H), 5.88–5.84 (m, 1 H), 4.21 (q, J = 6.4 Hz, J = 12.7Hz, 1 H), 4.00–3.86 (m, 2 H), 3.81 (dd, J = 3.7 Hz, J = 11.2 Hz, 1 H), 3.69 (dd, J = 7.1 Hz, J = 11.1 Hz, 1 H), 3.24–314 (m, 1 H), 2.68-2.60 (m, 1 H), 2.03-190 (m, 1 H), 2.00-1.90 (m, 1 H), 1.41 (d, J = 6.2 Hz, 3 H), 1.02 (d, J = 6.8 Hz, 3 H), 0.98 (d, J = 6.8 Hz, 3 H).

¹³C NMR (100 MHz, CDCl₃): δ = 133.2 118.0, 107.8, 102.8, 66.2, 60.2, 52.2, 45.7, 44.9, 27.6, 21.7, 20.9, 20.1.

MS (EI): *m/z* = 207 (100), 121 (51), 191 (19), 222 (1).

MS (ES): $m/z = 223.1 [M + H]^+ (100)$.

(2S)-2-[(1R/1S)-1-Ethyl-3,4-dihydropyrrolo[1,2-a]pyrazin-2(1H)-yl]-3-methylbutan-1-ol (17b + *epi*-17b)

Yellow oil; yield: 0.160 g (68%); dr 65:35.

¹H NMR (400 MHz, CDCl₃): δ (representative signals for **17b**) = 4.23 (t, J = 4.6 Hz, 1 H), 3.64 (dd, J = 5.2 Hz, J = 10.4 Hz, 1 H), 3.35 (t, *J* = 10.4 Hz, 1 H), 3.04 (dt, *J* = 3.2 Hz, *J* = 11.7 Hz, 1 H), 2.77 (m, 1 H); δ (representative signals for *epi*-**17b**) = 4.08 (t, *J* = 4.7 Hz, 1 H), 3.82 (dd, *J* = 3.5 Hz, *J* = 11.4 Hz, 1 H), 3.69 (dd, J = 6.1 Hz, J = 6.8 Hz, 1 H), 3.20 (dt, J = 4.0 Hz, J = 12.3 Hz, 1 H), 2.50 (m, 1 H).

Anal. Calcd for C₁₄N₂₄N₂ (236.25): C, 71.14; H, 10.23; N, 11.85. Found: C, 71.11; H, 10.25; N, 11.84.

(2S)-2-[(1R/1S)-1-Allyl-3,4-dihydropyrrolo[1,2-a]pyrazin-2(1H)-yl]-3-methylbutan-1-ol (17e + epi-17e) Yellowish oil; yield: 0.158 g (64%); dr 50:50.

¹H NMR (400 MHz, CDCl₃): δ (representative signals for 17e) = 6.57-6.55 (m, 1 H), 5.95-5.88 (m, 1 H), 5.62-5.49 (m, 1 H), 4.39 (t, J = 4.2 Hz, 1 H), 4.23 (t, J = 5.1 Hz, 1 H), 3.36 (t, J = 10.4 Hz, 1 H), 3.09–2.93 (m, 2 H), 1.05 (d, J = 6.8 Hz, 3 H), 0.90 (d, J = 6.8 Hz, 3 H); δ (representative signals for *epi*-17e) = 6.54–6.50 (m, 1 H), 5.87–5.84 (m, 1 H), 5.83–5.71 (m, 1 H), 3.28 (dt, J = 4.6 Hz, J = 12.8 Hz, 1 H), 2.85–2.77 (m, 1 H), 0.87 (t, J = 7.2 Hz, 3 H).

Anal. Calcd for C₁₅H₂₄N₂ (248.36): C, 72.54; H, 9.74; N, 11.28. Found: C, 72.28; H, 9.76; N, 11.24.

(2S)-2-[(1R/1S)-1-Benzyl-3,4-dihydropyrrolo[1,2-a]pyrazin-2(1H)-yl]-3-methylbutan-1-ol (17f + epi-17f)

Light-red oil; yield: 0.226 g (76%); dr 50:50.

¹H NMR (400 MHz, CDCl₃): δ (representative signals for 17f) = 7.33–7.21 (m, 4 H), 6.95 (m, 1 H), 6.54 (m, 1 H), 6.11 (dd, J = 2.8 Hz, J = 3.6 Hz, 1 H), 5.64 (ddd, J = 0.7 Hz, J = 1.6 Hz, J = 3.5 Hz, 1 H), 4.54 (dd, J = 4.8 Hz, J = 6.1 Hz, 1 H), 3.86 (dt, J = 3.9 Hz, *J* = 12.1 Hz, 1 H), 3.64 (dd, *J* = 5.1 Hz, *J* = 10.7 Hz, 1 H), 3.54 (m, 1 H), 3.33 (t, J = 10.7 Hz, 1 H), 3.17 (dd, J = 4.6 Hz, J = 13.1 Hz, 1 H), 3.00–2.87 (m, 3 H), 2.82 (ddd, J = 5.1 Hz, J = 7.9 Hz, J = 13.1 Hz, 1 H), 1.87 (m, 1 H), 0.97 (d, J = 6.7 Hz, 3 H), 0.87 (d, J = 6.7Hz, 3 H).

¹³C NMR (100 MHz, CDCl₃): δ (representative signals for **17f**) = 138.3, 129.7, 128.5, 128.1, 126.3, 118.3, 107.4, 104.5, 66.8, 50.1, 44.9, 43.8, 43.0, 40.6, 28.4, 22.0, 19.7.

MS (ES): $m/z = 299.2 [M + H]^+ (100)$.

¹H NMR (400 MHz, CDCl₃): δ (representative signals for *epi*-17f) = 4.34 (t, J = 6.7 Hz, 1 H), 3.93 (ddd, J = 4.3 Hz, J = 8.3 Hz,

J = 12.3 Hz, 1 H), 3.78 (ddd, J = 4.3 Hz, J = 5.3 Hz, J = 12.3 Hz, 1 H), 3.44 (dd, J = 4.3 Hz, J = 8.3 Hz, J = 13.1 Hz, 1 H), 2.65 (t, J = 7.7 Hz, 1 H), 2.58 (ddd, J = 3.8 Hz, J = 6.3 Hz, J = 10.2 Hz, 1 H), 1.80 (m, 1 H), 0.89 (d, J = 6.8 Hz, 3 H), 0.85 (d, J = 6.8 Hz, 3 H).

MS (ES): $m/z = 299.2 [M + H]^+ (100)$.

Anal. Calcd for $C_{19}H_{26}N_2$ (298.42): C, 76.47; H, 8.78; N, 9.39. Found: C, 76.45; H, 8.78; N, 9.37.

(2S)-3-Methyl-2-[(1R/1S)-1-phenyl-3,4-dihydropyrrolo[1,2a]pyrazin-2(1H)-yl]butan-1-ol (17g + epi-17g) Light-red oil; yield: 0.221 g (78%); dr 40:60.

¹H NMR (400 MHz, CDCl₃): δ (representative signals for 17g) = 7.36–7.28 (m, 5 H), 6.58 (m, 1 H), 6.08 (dd, J = 2.8 Hz, *J* = 3.5 Hz, 1 H), 5.32 (m, 1 H), 5.19 (s, 1 H), 4.15 (ddd, *J* = 4.0 Hz, *J* = 11.3 Hz, *J* = 11.4 Hz, 1 H), 4.08 (ddd, *J* = 2.1 Hz, *J* = 3.9 Hz, J = 11.4 Hz, 1 H), 3.42 (dd, J = 5.2 Hz, J = 10.7 Hz, 1 H), 3.35 (t, *J* = 10.7 Hz, 1 H), 3.29 (ddd, *J* = 1.9 Hz, *J* = 4.0 Hz, *J* = 12.1 Hz, 1 H), 3.16 (m, 1 H), 2.61 (ddd, J = 5.4 Hz, J = 6.4 Hz, J = 11.6 Hz, 1 H), 2.04 (m, 1 H), 1.05 (d, J = 6.4 Hz, 3 H), 0.83 (d, J = 6.4 Hz, 3 H).

¹³C NMR (100 MHz, CDCl₃): δ = 141.8, 131.8, 129.4, 128.6, 128.0, 118.2, 108.3, 105.7, 63.7, 62.5, 58.5, 45.7, 42.0, 26.9, 22.9, 19.6.

¹H NMR (400 MHz, CDCl₃): δ (representative signals for epi-17g) = 6.55 (m, 1 H), 6.05 (dd, J = 2.8 Hz, J = 3.6 Hz, 1 H), 5.30 (m, 1 H), 5.10 (s, 1 H), 3.94 (dd, J = 2.9 Hz, J = 11.7 Hz, 1 H), 3.72 (dd, J = 5.7 Hz, J = 11.7 Hz, 1 H), 2.26 (m, 1 H), 0.93 (d, J = 6.6 Hz),3 H), 0.80 (d, J = 6.6 Hz, 3 H).

Anal. Calcd for $C_{18}H_{24}N_2O$ (284.40): C, 76.02; H, 8.51; N, 9.85. Found: C, 76.16; H, 8.50; N, 9.83.

(1R)-1-Methyl-1,2,3,4-tetrahydropyrrolo[1,2-a]pyrazine (21a); **Typical Procedure**

10% Pd/C (0.048 g) was added to a soln of 16a (0.476 g, 1.9 mmol) in MeOH (10 mL) inside an autoclave and the mixture was kept under H₂ (7 bar) for 2 d. The catalyst was filtered off on a small pad of Celite and the organic soln was concentrated under vacuum. The oily residue was purified by column chromatography [SiO₂, CH₂Cl₂–MeOH (95:5)] to give an orange oil; yield: 0.205 g (81%); $[\alpha]_{D}^{20}$ +8.3 (*c* 0.7, CHCl₃).

¹H NMR (400 MHz, CDCl₃): $\delta = 6.54$ (t, J = 2.4 Hz, 1 H), 6.15 (dd, J = 2.8 Hz, J = 3.6 Hz, 1 H), 5.90–5.80 (m, 1 H), 4.06 (q, J = 6.8 Hz, 1 H), 3.95-3.90 (m, 2 H), 3.38-3.31 (m, 1 H), 3.24-3.15 (m, 1 H), 1.06 (d, J = 6.8 Hz, 3 H).

¹³C NMR (100 MHz, CDCl₃): δ = 132.9, 119.0, 107.7, 102.3, 49.5, 45.4, 43.4, 31.0.

MS (ES): $m/z = 136.1 [M + H]^+$.

Anal. Calcd for C₈H₁₂N₂ (136.19): C, 70.55; H, 8.88; N, 20.57. Found: C, 70.28; H, 8.91; N, 20.50.

(1*R*)-1-Ethyl-(1,2,3,4-tetrahydropyrrolo[1,2-*a*]pyrazine (21b)

This was prepared from 16b (0.156 g, 0.6 mmol) as described above.

Orange oil; yield: 0.156 g (79%); $[\alpha]_D^{20}$ +11.8 (*c* 0.7, CHCl₃).

¹H NMR (400 MHz, CDCl₃): $\delta = 6.55$ (t, J = 2.1 Hz, 1 H), 6.15 (t, *J* = 3.4 Hz, 1 H), 5.90–5.88 (m, 1 H), 3.95–3.87 (m, 3 H), 3.38–3.35 (m, 1 H), 3.20–3.12 (m, 1 H), 2.03–1.94 (m, 1 H), 1.74–1.62 (m, 1 H), 1.06 (t, J = 7.4 Hz, 3 H).

¹³C NMR (100 MHz, CDCl₃): δ = 131.5, 118.8, 107.5, 102.3, 55.0, 45.3, 42.9, 28.2, 10.2.

MS (ES): $m/z = 151.1 [M + H]^+ (100)$.

Anal. Calcd for $C_9H_{14}N_2$ (150.22): C, 71.96; H, 9.39; N, 18.65. Found: C, 71.63; H, 9.43; N, 18.58.

X-ray Crystallographic Study of Oxazolopyrrolopyrazine 14

The X-ray intensity data for **14** were measured on a Bruker SMART Apex II diffractometer equipped with a CCD area detector and a graphite monochromated Mo-K α radiation source ($\lambda = 0.71073$ Å). Cell dimensions and the orientation matrix were initially determined from a least-squares refinement on reflections measured in three sets of 20 exposures, collected in three different ω regions, and eventually refined against all data. For all crystals, a full sphere of reciprocal space was scanned by 0.3° ω steps. The software SMART²⁹ was used for collecting frames of data, indexing reflections, and determining lattice parameters. The collected frames were then processed for integration by SAINT²⁹ software, and an empirical absorption correction was applied with SADABS.³⁰ The structure was solved by direct methods (SIR 97)³¹ and subsequent Fourier syntheses, and refined by full-matrix least-squares calculations on *F*² (SHELXTL),³² attributing anisotropic thermal parame-

 Table 3
 Crystal Data and Structure Refinement for 14

Compound	14			
Formula	$C_{15}H_{16}N_2 O$			
М	240.30			
Т, К	296(2)			
Crystal symmetry	monoclinic			
Space group	$P2_1$			
<i>a</i> (Å)	8.2411(4)			
<i>b</i> (Å)	7.3720(4)			
<i>c</i> (Å)	11.1066(6)			
α (°)	90			
β (°)	110.220(3)			
γ (°)	90			
V (Å ³)	633.18(6)			
Ζ	2			
$D_c (Mg m^{-3})$	1.260			
μ (Mo-K α) (mm ⁻¹)	0.080			
F(000)	256			
Crystal size (mm)	$0.30\times0.28\times0.25$			
θ limits (°)	1.95–26.99			
Reflections collected	9380			
Unique obs. reflections $[F_o > 4\sigma(F_o)]$	2761 [$R(int) = 0.0529$]			
Goodness-of-fit-on F ²	1.114			
$R_1(F)^a, wR_2(F^2)^b$	0.0436, 0.1008			
Largest diff. peak and hole, e. (Å-3)	0.323 and -0.217			

^a $R_1 = \Sigma ||F_0| - |F_c|| / \Sigma |F_0|.$

^b $wR_2 = [\Sigma w(F_o^2 - F_c^2)^2 / \Sigma w(F_o^2)^2]^{1/2}$ where $w = 1/[\sigma^2(F_o^2) + (aP)^2 + bP]$ where $P = (F_o^2 + F_c^2)/3$.

ters to the nonhydrogen atoms. All hydrogen atoms were located in the Fourier map. The aromatic and methylene hydrogen atoms were placed in the calculated positions, refined with isotropic thermal parameters $U(H) = 1.2 \ Ueq(C)$, and allowed to ride on their carrier carbons, whereas the methine H atoms were located in the Fourier map and refined isotropically $[U(H) = 1.2 \ U_{eq}(C)]$. Crystal data and details of the data collection for **14** are reported in Table 3.

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- (25) Crystallographic data for compound 14 have been deposited with the Cambridge Crystallographic Data Centre as supplementary publications CCDC 792774; copies can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK [fax: +44 (1223)336033 or email: deposit@ccdc.cam.ac.uk].
- (26) It is noteworthy that tricyclic tetrahydrooxazolopyrazolopyridine structures with three carbon stereocenters have been prepared as single diastereomers from 2-pyrrolecarboxaldehyde, (*S*)- or (*R*)-α-amino acid esters, and (+)- or (–)-norephedrine. The configuration of the newly formed stereocenter in the oxazolidine ring depends only on the

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