# Accepted Manuscript

A practical chromatography-free synthesis of a 5,6-dihydroimidazolo[1,5-*f*]pteridine derivative as a polo-like kinase-1 inhibitor

Kazuhisa Ishimoto, Keiichiro Nakaoka, Osamu Yabe, Atsuko Nishiguchi, Tomomi Ikemoto

PII: S0040-4020(18)30973-6

DOI: 10.1016/j.tet.2018.08.020

Reference: TET 29740

To appear in: *Tetrahedron* 

Received Date: 21 June 2018

Revised Date: 12 August 2018

Accepted Date: 14 August 2018

Please cite this article as: Ishimoto K, Nakaoka K, Yabe O, Nishiguchi A, Ikemoto T, A practical chromatography-free synthesis of a 5,6-dihydroimidazolo[1,5-*f*]pteridine derivative as a polo-like kinase-1 inhibitor, *Tetrahedron* (2018), doi: 10.1016/j.tet.2018.08.020.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



# **Graphical Abstract**

To create your abstract, type over the instructions in the template box below. Fonts or abstract dimensions should not be changed or altered.





# Tetrahedron journal homepage: www.elsevier.com



# A practical chromatography-free synthesis of a 5,6-dihydroimidazolo[1,5-*f*]pteridine derivative as a polo-like kinase-1 inhibitor

Kazuhisa Ishimoto \*, Keiichiro Nakaoka <sup>1</sup>, Osamu Yabe <sup>1</sup>, Atsuko Nishiguchi <sup>1</sup>, Tomomi Ikemoto <sup>1</sup>

Process Chemistry, Pharmaceutical Sciences, Takeda Pharmaceutical Company Limited, 17-85, Jusohonmachi 2-Chome, Yodogawa-ku, Osaka 532-8686, Japan

### ARTICLE INFO

Article history: Received Received in revised form Accepted Available online

Keywords:

5,6-dihydroimidazolo[1,5-f]pteridine polo-like kinase-1 inhibitor chromatography-free synthesis *trans* 1,4-cyclohexyldiamine piperazine ring formation amination

## ABSTRACT

A practical chromatography-free synthesis of a potent polo-like kinase-1 inhibitor possessing a unique 5,6-dihydroimidazolo[1,5-f]pteridine structure has been developed. We showed that key cyanoimidazole ring formation could be conducted at benign temperature and obtained a chiral 5,6-dihydroimidazolo[1,5-f]pteridine derivative in good yield without epimerization. An aniline derivative containing a *trans* 1,4-cyclohexyl diamine structure was prepared by a synthesis that makes use of defined stereocenters of commercially available trans-cyclohexane-1,4-diamine via selective piperazine ring formation from a primary diamine. A coupling reaction of the 3-chloro-5,6-dihydroimidazolo[1,5-f]pteridine derivative and the aniline derivative in the endgame was closely investigated, and good yields were achieved both by palladium-catalyzed amination and acid-promoted coupling under benign reaction conditions. As a result of these investigations, the polo-like kinase-1 inhibitor was successfully obtained in a practical way without concern for generation/separation of stereoisomers.

2009 Elsevier Ltd. All rights reserved.

## 1. Introduction <sup>i</sup>

Polo-like kinases (PLKs) were identified as one of the kinase families that play an essential role in various cell-cycle related processes.<sup>1</sup> Amongst five known PLKs (PLK1–5), PLK1 has attracted much attention as an important target for a medicine, and inhibitors of PLK1 have been developed and evaluated as anticancer drugs in clinical studies.<sup>2</sup> Compound **19** is a potent PLK1 inhibitor that has strong enzyme and cellular activities against PLK1, and also exhibits oral bioavailability.<sup>3</sup>

Compound **19** is characterized by its unique 5,6dihydroimidazolo[1,5-f]pteridine core structure, and medicinal chemists have synthesized **19** as described in Scheme 1 and Scheme 2.<sup>3</sup> Unnatural amino acid **1** was converted to methyl ester hydrochloric acid salt **2·HCl**, and reductive amination of **2·HCl** with acetone followed by an  $S_NAr$  reaction with 2,4-dichloro-5nitropyrimidine **4** gave **5** (Scheme 1). Reduction of the nitro group of **5** led to cyclization to give **6**. Activation of the amide group of **6** by phosphorylation was followed by addition of an anion derived from **7**, and the subsequent acid-promoted cyclization provided in one pot **8**, which has the 5,6dihydroimidazolo[1,5-*f*]pteridine structure. Aniline **15** was synthesized starting from commercially available ketone **9**. Amino ketone **10·HCl** prepared by deprotection of **9** was condensed with benzoic acid **11** to give ketone **12**. Imine formation of **12** with piperazine **13** followed by reduction using NaBH<sub>4</sub> provided *trans* **14** as the major product and *cis* isomer **14'** as the minor product, respectively. Hydrogenation of the mixture of **14** and **14'** provided aniline **15**.

For the synthesis of **19** from **8**, two routes have been reported (Scheme 2). In one route (Route A), compound **8** was reacted with aniline **15** in an acidic condition to afford **19**. In the other route (Route B), compound **19** was synthesized by sequential reactions: compound **8** was first reacted with aniline **16** using a palladium catalyst under a microwave condition, and then condensed with known amine **18**<sup>4</sup>.

Three key issues need to be overcome for scale-up of the medicinal chemistry syntheses. First, the cryogenic condition (-78 °C) used for the synthesis of **8** from **6** is difficult to perform in large-scale production. Second, construction of the *trans* diamine structure on the cyclohexyl ring of **15**/1**8** is a synthetic challenge. In the patent filed by the medicinal chemists, <sup>3a</sup> ratio of **14** to **14'** synthesized from **12** and **13** by reductive amination is not obviously mentioned. A mixture of **14** and **14'** was hydrogenated, and after simple workup, the obtained crude aniline **15** was used for the final coupling with **8** without further purification. Removal of the undesired cis isomer **19'** seemingly necessitated cumbersome workup procedures including column chromatography separation (Route A) or HPLC separation

<sup>&</sup>lt;sup>i</sup> \* Corresponding author.

E-mail address: kazuhisa.ishimoto@takeda.com (K. Ishimoto).

<sup>&</sup>lt;sup>1</sup> Present address: SPERA PHARMA, Inc., 17-85, Jusohonmachi 2-Chome, Yodogawa-ku, Osaka 532-0024, Japan.

E-mail address: tomomi.ikemoto@spera-pharma.co.jp (T. Ikemoto)



Scheme 1. Medicinal chemistry synthesis of 8 and 15.

Reagents and Conditions: (a) SOCl<sub>2</sub>, MeOH; (b) acetone, NaBH(OAc)<sub>3</sub>, NaOAc, AcOH, THF; (c) **4**, THF, rt; (d) 5% Pt/C, H<sub>2</sub> (75 psi), NH<sub>4</sub>VO<sub>3</sub>, P(OPh)<sub>3</sub>, 5 h, rt; (e) LHMDS, CIPO(OEt)<sub>2</sub>, THF, -78 °C; (f) **7**, LHMDS, THF, -78 °C; (g) AcOH, THF; (h) 4 M HCl in dioxane, rt, 6 h; (i) **11**, HATU, DIPEA, DMF, rt, 2.5 h; (j) **13**, cat. methanesulfonic acid, toluene, reflux with a Dean-Stark reflux condenser, 5 h; (k) NaBH<sub>4</sub>, EtOH, rt, overnight; (l) Pd/C, H<sub>2</sub> (balloon), MeOH/AcOH (5:1), rt, 8 h.



Scheme 2. Medicinal chemistry synthesis of 19 from 8.

Reagents and Conditions: (a) conc. HCl, 2-propanol, 95 °C, 2 days; (b) Pd(OAc)<sub>2</sub>, Xantphos, Cs<sub>2</sub>CO<sub>3</sub>, *N*,*N*-dimethylacetamide/dioxane, microwave, 160 °C, 15 min; (c) HATU, DIPEA, DMF, rt, 2 h.

(Route B).<sup>5</sup> Finally, the syntheses of **19** from **8** required long N reaction time or the use of microwave irradiation, and suffered from low yields. The acid-promoted coupling of **8** with **15** in 2-propanol using concentrated HCl required 2 days for the reaction, providing **19** in low yield (25%, Route A). The palladium-catalyzed amination of **8** with aniline **16** was conducted using the microwave system (160 °C, 15 min), and the subsequent coupling with **18** gave **19**, also in low yield (31%, Route B).

As a first step toward a future scale-up synthesis to supply 19 to preclinical/clinical studies, we initiated development of a practical column chromatography-free/HPLC separation-free synthesis of 19 that addresses these three key issues. Retrosynthetic analysis of 19 is shown in Scheme 3. Since it is known that a convergent synthesis is generally preferable from a standpoint of cost and efficiency,<sup>6</sup> we planned to conduct the coupling reaction of 8 with 15 in the last step, and to obtain 19 in a convergent manner. Compound 8 was expected to be synthesized from 2 and 4 as in the medicinal chemistry synthesis, though the synthesis was modified for scale-up, including revisit of reaction conditions for the preparation of 8 from 6. For the synthesis of 15, we adopted another synthetic route that makes use of defined *trans* stereocenters of commercially available trans diamine 22.7 Aniline 15 was envisioned to be synthesized from 20 and 21 via piperazine ring formation. Aniline 20 was anticipated to be synthesized from aniline 16 and 22, and 21 was envisaged to be prepared by chlorination of diol 23 synthesized by alkylation of diol 24 with 25.

#### 2. Results and Discussion

Lactam 6 was prepared from commercially available 2·HCl and 4 with a modified procedure (Scheme 4).<sup>8</sup> The reductive amination of 2·HCl with acetone using NaBH<sub>4</sub> instead of expensive NaBH(OAc)<sub>3</sub> as the reducing agent proceeded smoothly, and 3 was isolated as a hydrochloric acid salt in 81% yield. The subsequent  $S_NAr$  reaction of 4 with 3·HCl in toluene using NaHCO<sub>3</sub> as the base gave 5 in 73% yield.

For preparation of 6, reduction of the nitro group of 5 using inexpensive iron with acetic acid was examined to avoid the use of an expensive Pt catalyst<sup>9</sup>. When the reduction was conducted using acetic acid as the solvent, rapid and huge exotherm was observed. During the course of investigation, addition of a catalytic amount of AcOH was found to be sufficient for completion of the reaction. The use of a catalytic amount of AcOH decreased the reaction rate, which made the reaction proceed without rapid exotherm. Also, it is noteworthy that addition of a small amount of water was effective for improving fluidity of the reaction mixture, presumably because water helped acetic acid salt of 6 to dissolve into the reaction mixture. The reaction was carried out in the presence of 4.0 equiv of iron, 0.3 equiv of acetic acid, and water (50 w/w% to 5) in toluene at 75 °C for 3 h, leading to the reduction of the nitro group and the cyclization to provide 6 without notable exotherm. Upon completion of the reaction, 6 M HCl was added to the reaction mixture to dissolve iron. Following extraction with EtOAc, and washing with water, brine, and saturated aqueous NaHCO<sub>3</sub>, lactam 6 was crystallized from EtOH/H<sub>2</sub>O and isolated in 80% yield.

Compound  $7^{3a, 10}$  was prepared by the reaction of aminoacetonitrile hydrochloride 26·HCl with N.Ndimethylformamide dimethyl acetal in the presence of triethylamine in THF, and isolated in 67% yield by distillation (5.6 mmHg, 90 °C). During investigation into a workup procedure, we found that treatment with a pad of NH silica gel was effective for removing byproducts generated during the reaction. After the reaction was completed, the solvent was switched from THF to diisopropyl ether, and the mixture was passed through a pad of NH silica gel (50 w/w% to 26·HCl), which removed precipitated triethylamine hydrochloride salt and brown tarry byproducts. Following evaporation of the solvent, 7 was obtained in 64% yield. It was possible to use the obtained 7 for the next reaction with 6 without further purification, which allowed us to avoid time-consuming distillation.





Scheme 5. Precedent work and synthesis of 8 from 6.

Next, the preparation of **8** from **6** was investigated. There have been some precedents for this interesting imidazole ring formation reaction (Scheme 5). Rogers-Evans and co-workers reported cyanoimidazole ring formation from an iminochloride.<sup>10</sup> Addition of an anion of **28/7** generated by LHMDS to iminochloride **27** gave intermediates **29** and **29'**, and the subsequent acid-promoted isomerization of **29** to **29'** and cyclization provided **30**.<sup>11</sup> Fryer et al. reported that addition of an anion of methyl ester analogue of **28** to an iminophosphate gave a methoxycarbonyl imidazole.<sup>12</sup> The medicinal chemists used these chemistries in a very excellent manner and conducted the three reactions (phosphorylation, addition of **7**, and isomerization/cyclization) in one pot. In these works, addition of

the anions to the iminochloride/iminophosphate was conducted at -78 °C or at -30 °C, which largely restrict a manufacturing facility for scale-up. To confirm whether this reaction could be carried out under more benign conditions, we began by examining the reaction conditions closely (Table 1).

When we conducted the addition of the anion of **7** to **31** at -70 °C, **8** was obtained in 56% yield (entry 1). To our surprise, when the temperature for the addition of the anion of **7** was increased to -35 °C and -5 °C, the yield increased to 72% and 77%, respectively (entries 2, 3). Next, we conducted the phosphorylation of **6** at -5 °C. Under this condition, **8** was obtained in good yield (71%, entry 4) comparable to the yield of entry 3, where the phosphorylation was conducted at -70 °C.<sup>13</sup>

Under theses reaction conditions, phosphorylation of 6 did not M reach complete conversion, and 5-10% (HPLC area%) of 6 remained unreacted. On the basis of these results, we concluded **Table 1** Synthesis of 8 from 6.

A that both the phosphorylation and the addition of the anion of 7 could be performed at -5 °C. Fixing the temperatures for the phosphorylation and the addition of the anion of 7 at -5 °C, we



phosphorylation			addition of <b>7</b>						
entry	LHMDS (equiv)	ClPO(OEt) <sub>2</sub> (equiv)	temp (°C)	time (h)	7 (equiv)	LHMDS (equiv)	temp (°C)	yield <sup>a</sup> (%)	optical purity (% ee)
$1^b$	1.1	2	$-70 \rightarrow 0$	2.5	3	3	-70	56	7 -
2 <sup><i>b</i></sup>	1.1	2	$-70 \rightarrow 0$	2.5	3	3	-35	72	-
3 <sup><i>b</i></sup>	1.1	2	$-70 \rightarrow 0$	2.5	3	3	-5	77	-
4	1.1	2	-5	1	3	3	-5	71	99.8
5	1.1	2	-5	1	2	3	-5	74	99.8
6	1.1	1.5	-5	1	3	3	-5	74	99.6
7	1.1	1.5	-5	1	2	3	-5	75	97.3
8	1.1	1.5	-5	1	2	2.5	-5	60	99.8
9	1.1	1.5	-5	1	2	2	-5	44	-
10	1.1	1.2	-5	1	2	3	-5	56	<mark>98.5</mark>

<sup>a</sup>Isolated yield. <sup>b</sup>Compound 7 isolated by distillation was used for the reaction.

then tried to decrease the amounts of reagents. Whereas decrease of the amount of diethyl chlorophosphate from 2.0 equiv (entry 5) to 1.5 equiv did not affect the yield (75%, entry 7), decrease to 1.2 equiv caused large yield deterioration (56%, entry 10). Decrease of the amount of **7** from 3.0 equiv (entry 4) to 2.0 equiv was found to be acceptable (74%, entry 5). In contrast, decrease of the amount of LHMDS used for the deprotonation of **7** had crucial effects on the yields. When the amount of LHMDS for the deprotonation of **7** was decreased from 3.0 equiv (entry 7) to 2.5 equiv and 2.0 equiv, the yield decreased to 60% and 44%, respectively (entries 8, 9). Isolation of **8** was conducted by crystallization, and typically 3–5% (HPLC assay yield) of **8** was included in mother liquor.

After screening the reaction conditions, we analyzed optical purity of the obtained **8** and found that the optical purity of **8** prepared under the reaction conditions of entry 7 and entry 10 decreased to 97.3% ee and 98.5% ee, respectively. A plausible mechanism for the optical purity deterioration is described in Scheme 6. It is assumed that a strong base such as LHMDS could bring about deprotonation at the  $\gamma$  position of the cyano group of **32** at -5 °C to give an anion, and reprotonation of the anion would lead to the optical purity decrease. Under the reaction conditions of entry 7 and entry 10, the total amounts of diethyl chlorophosphate and **7** were less than the total amounts under the conditions of entries 4, 5, and 6. This situation would result in an increased amount of remaining active LHMDS in the reaction mixture and would cause the deprotonation of **32**, leading to the

optical purity decrease of **8**. We also examined the possibility of racemization of compound **6** by treating it with 1.1 equiv of LHMDS in THF at -5 °C for 5 h. A small portion of the reaction mixture was quenched with MeCN/20 mM aqueous KH<sub>2</sub>PO<sub>4</sub> (30:70) and analyzed by chiral HPLC, which showed that the optical purity of **6** (>99.7% ee) did not change during 5 h under this reaction condition. Considering these results, the reaction conditions of entry 5 were finally selected as the best reaction conditions.

Aniline 15 was prepared by the synthesis described in Scheme 7. Mono *N*-Boc protected *trans* diamine  $33^{14}$  was prepared from commercially available *trans*-cyclohexane-1,4-diamine 22 by treatment with Boc<sub>2</sub>O in MeCN. The obtained crude 33 containing di-Boc byproduct 34 (approximately 3:1) was used for condensation with 16 without purification. The condensation of 33 carried 16 with was out using 1-ethyl-3-(3dimethylaminopropyl)carbodiimide hydrochloride (EDCI·HCl) and 1-hydroxybenzotriazole monohydrate (HOBt·H<sub>2</sub>O) in MeCN, giving 35 exclusively without self-condensation of 16. Addition of water to the reaction mixture led to crystallization, which enabled isolation of 35 by filtration. The obtained 35 was dissolved in MeOH, and the Boc group was deprotected with 6 M HCl. After pH was adjusted to 7.8 with 8 M NaOH, the mixture was treated with activated carbon. Following evaporation of MeOH, filtration of the resulting aqueous slurry gave 20·HCl in 82% yield for 2 steps from 16. The di-Boc byproduct 34 was completely removed during isolation of 20·HCl.



Scheme 6. Plausible mechanism for optical purity decrease of 8.



Scheme 7. Synthesis of 15.

Compound 21 was synthesized via alkylation of 24 with (chloromethyl)cyclopropane followed by chlorination of the 24 hydroxyl groups. Diol was reacted with (chloromethyl)cyclopropane in the presence of NaI and K<sub>2</sub>CO<sub>3</sub> in refluxing EtOH. After evaporation of EtOH, 23 was extracted with EtOAc and washed with saturated brine, and the solvent was switch to toluene. The obtained toluene solution of 23 was treated with 2.0 equiv of SOCl<sub>2</sub> at 50 °C for 3 h. On completion of the reaction, the reaction mixture was neutralized with slow addition of 8 M NaOH. Following phase separation, filtering off insoluble matter with a pad of silica gel, and evaporation of the solvent, the obtained 21 (66% for 2 steps from 24, determined by <sup>1</sup>H NMR) was used for the piperazine ring formation without further purification.

There have been many examples of piperazine ring formation from a primary amine by a reaction with bis(2-chloroethyl)amine derivatives.<sup>15</sup> However, preparation of a piperazine ring from an unsymmetrical primary diamine by reacting one of the primary amino groups selectively with bis(2-chloroethyl)amine derivatives without protection seems to be difficult. To the best of our knowledge, there has been only one report (patent) that includes this selective piperazine ring formation from an unsymmetrical primary diamine, and the reported yield was quite low (12%).<sup>16</sup> We thought that protection of the aromatic amino group of **20-HCl** was not indispensable for the reaction with **21**, considering difference of nucleophilicity between an aromatic amino group and an alkyl amino group.

For the synthesis of 15 from 20·HCl by the selective piperazine ring formation, we screened several bases (NaHCO<sub>3</sub>, K<sub>2</sub>CO<sub>3</sub>, NaOAc, N,N-diisopropylethylamine) and solvents (EtOH, THF, acetone, MeCN) using 1.2 equiv of 21 at 50 °C for 3 h. The reaction was rather slow, and in addition, generation of many byproducts was observed, which resulted in low conversion of the reaction. Among the screened solvents, EtOH was found to be the best solvent; the conversion exceeded 50% only when EtOH was used as the solvent. As for the bases,  $K_2CO_3$  and  $N,N_2$ diisopropylethylamine gave good results. The best result was achieved when K<sub>2</sub>CO<sub>3</sub> and EtOH were used for the reaction with NaI as an additive. The  $S_N 2$  reaction of 20·HCl with 21 using 1.0 equiv of NaI and 3.0 equiv of K<sub>2</sub>CO<sub>3</sub> in EtOH mainly occurred not at the aromatic amino group, but at the alkyl amino group on the cyclohexyl ring, and provided 15 as expected. During the reaction, generation of several byproducts was observed, and HPLC assay yield was found to be moderate (67%). Dialkylated product 36 (the structure was suggested by LC-MS analysis) was identified as the most notable byproduct for this reaction, and even under the best reaction condition, approximately 17% (HPLC area%) of 36 was observed. After workup, 15 was once isolated as a hydrochloric acid salt, and then dissolved in water and treated with activated carbon. The activated carbon was

filtered off, and pH was adjusted to 9.0 with 25% aqueous NH<sub>3</sub>. M Filtration of the resulting slurry afforded pure **15** (**36**: 0.4 HPLC area%) in 37% isolated yield with 17% of **15** in the mother liquor.

Having obtained **8** and **15** in hand, the last coupling reaction of **8** with **15** was examined. Other than the patent filed by the medicinal chemists<sup>3a</sup>, there has been only one report (patent) that describes a coupling reaction of 3-halo-5,6-dihydroimidazo[1,5f]pteridine with an amine, and the reported yield was only moderate (70 mg scale reaction, 52%).<sup>17</sup> As described previously, the acid-promoted coupling reaction required long reaction time (>24 h) and the palladium-catalyzed amination needed high reaction temperature (microwave, 160 °C), and in addition, the yields were quite low; therefore, more benign reaction conditions that afford the coupling product in good yield are highly demanded for scale-up.

Table 2 Synthesis of 19 by palladium-catalyzed amination.

A First, we tried to conduct the coupling reaction using a base; however, the reaction did not proceed well in a basic condition. When *N*,*N*-diisopropylethylamine was used for the reaction in *N*,*N*-dimethylacetamide (DMAC) at 100 °C, the reaction did not proceed at all. Additionally, when  $Cs_2CO_3$  or *t*-BuOK was used for the reaction in DMAC at 100 °C, decomposition of **8** was mainly observed with a small amount of **19**. Thus, we turned our attention to the palladium-catalyzed amination<sup>18</sup> (Table 2) and the acid-promoted coupling reaction (Table 3).

Screening of palladium catalysts and ligands using *t*-BuOH as the solvent showed that the combination of  $Pd(OAc)_2$  and Xantphos was good for the reaction (Table 2, entries 1–4). When the reaction was conducted in *t*-BuOH using 10 mol% of  $Pd(OAc)_2$  and 10 mol% of Xantphos in the presence of 2.0 equiv of  $K_2CO_3$ , the reaction reached full conversion in 1 h. HPLC assay yield was 86%, and after workup, **19** was isolated in 61%



<sup>a</sup>HPLC assay yield. <sup>b</sup>Isolated yield in parentheses. <sup>c</sup>1.1 equiv of 8 was used. <sup>d</sup>Not detected.

yield (entry 2). Several kinds of other solvents were screened for the reaction. While the use of THF as the solvent decreased the yield (57%, entry 5), *s*-BuOH was found to be a good solvent for the reaction. HPLC assay yield reached 90%, and **19** was isolated in 78% yield (entry 7). Although we tried to decrease the amounts of  $Pd(OAc)_2$  and Xantphos to 1 mol%, the reaction completely stalled (entry 8). This was probably because **8** and/or **15** contained a small amount of impurities that deactivated the palladium catalyst.

In preliminary experiments of the acid prompted coupling reaction of **8** with **15**, several kinds of acids (concentrated HCl, MsOH, TsOH·H<sub>2</sub>O, H<sub>2</sub>SO<sub>4</sub>, H<sub>3</sub>PO<sub>4</sub>, 6 M HCl, 48% aqueous HBr, AcOH) and solvents (alcohols, DMF, DMAC, *N*-methylpyrrolidone, toluene, dioxane, *n*-BuOAc) were screened. Among the screened solvents, alcohols gave relatively good conversion. Whereas the reaction in 2-propanol (bp: 82 °C) with

concentrated HCl was very slow and reached as little as 6% conversion after 8 h at reflux condition (Table 3, entry 1), it was found that the use of MsOH and TsOH·H<sub>2</sub>O largely promoted the conversion. When MsOH and TsOH·H2O were used for the reaction, the conversion reached 44% and 58% after 8 h, respectively (entries 2, 3). To achieve faster conversion, alcohols that have a higher boiling point than 2-propanol were used for the reaction. When the reaction was conducted in primary alcohol i-BuOH (bp: 108 °C), a large amount of byproduct was formed by addition of i-BuOH to 8 (byproduct 1), and a small amount of hydrolyzed form of 8 (byproduct 2) was observed, which resulted in moderate conversion (48%, entry 4). Although tertiary alcohol *t*-amylalcohol (bp: 102 °C) did not give byproduct 1, the reaction in t-amylalcohol was not so fast and conversion after 8 h was also moderate (59%, entry 5). Fortunately, secondary alcohol sbutanol (bp: 100 °C) was found to be a good solvent for this reaction. The reaction in s-butanol with TsOH·H<sub>2</sub>O reached up to

#### Tetrahedron

81% conversion after 8 h (entry 6). Furthermore, increase of the amount of 8 to 2.0 equiv and 2.4 equiv led to higher conversions (entries 7, 8). We also confirmed that the reaction did not proceed in *s*-BuOH without acid (entry 9).

In the initial screening of acids, concentrated HCl was not found to be effective for the reaction (entry 1, 10). During the investigation, we happened to find that the reaction employing concentrated HCl as the acid was largely accelerated by addition of a small amount of water. When the reaction was carried out in *s*-BuOH/H<sub>2</sub>O (95:5) using 3.0 equiv of concentrated HCl, the reaction speed largely increased and became comparable to that of the reaction using TsOH·H<sub>2</sub>O (entry 11, 12). We presume that addition of a small amount of water helps hydrochloric acid salt of **15** formed by addition of concentrated HCl to dissolve in the solvent, which led to acceleration of the reaction. Finally, the reaction conditions of entry 12 were selected as the best **Table 3** Synthesis of **19** by acid-promoted coupling reaction. conditions, taking into account that complete removal of 3.0 equiv of TsOH was found not to be easy.

Isolation of pure **19** required elaborated workup procedures. On completion of the reaction, *s*-butanol was evaporated, and the product was extracted with 1 M HCl and washed with EtOAc. After pH of the aqueous extract was adjusted to approximately 9 with 8 M NaOH, the product was extracted with EtOAc and washed with 5% aqueous NaHCO<sub>3</sub>. Following solvent switch to 2-propanol, **19** was crystallized from 2-propanol/*n*-heptane (1:3). The obtained crude product was dissolved in MeOH and treated with activated carbon (Darco<sup>®</sup> G-60) at 50 °C, which was very effective for removing red color of the crude product. The activated carbon was filtered off, and solvent was switched to 2-propanol, and crystallization from 2-propanol gave pure **19** in 71% yield as an off-white solid (entry 12). The obtained **19** was



onteri	8 acid		colvent	temp	time	conversion	yield <sup>a</sup>		
entry	(equiv)	(equiv)	sorvent	(°C)	(h)	(%)	(%)		
$1^b$	0.83	c.HCl (3.0)	2-propanol <mark>(bp: 82 °C)</mark>	reflux	8	6	-		
2	1.5	MsOH (3.0)	2-propanol	reflux	8	44	-		
3	1.5	TsOH (3.0)	2-propanol	reflux	8	58	-		
4	1.5	TsOH (3.0)	<i>i</i> -BuOH <mark>(bp: 108 °C)</mark>	reflux	8	48	-		
5	1.5	TsOH (3.0)	<i>t</i> -amylalcohol ( <mark>bp: 102 °C)</mark>	reflux	8	59	-		
6	1.5	TsOH (3.0)	<i>s</i> -BuOH <mark>(bp: 100 °C)</mark>	reflux	8	81	-		
7	2.0	TsOH (3.0)	s-BuOH	reflux	10	89	85 (56) <sup>c</sup>		
8	2.4	TsOH (3.0)	s-BuOH	reflux	13	88	88 (60) <sup>c</sup>		
<mark>9</mark>	<mark>1.5</mark>		s-BuOH	<mark>reflux</mark>	<mark>8</mark>	N.D. <sup>d</sup>	-		
<mark>10</mark>	2.0	c.HCl (3.0)	s-BuOH	reflux	8	26	-		
<mark>11</mark>	2.0	c.HCl (3.0)	s-BuOH/H <sub>2</sub> O 95:5	reflux	8	79	-		
<mark>12</mark>	2.4	c.HCl (3.0)	s-BuOH/H <sub>2</sub> O 95:5	reflux	11	87	86 (71) <sup>c</sup>		

<sup>a</sup>HPLC assay yield. <sup>b</sup>Reagents were used based on the amount of 8 (15: 1.2 equiv, c.HCl: 3.0 equiv). <sup>c</sup>Isolated yield in parentheses. <sup>d</sup>Not detected.

analyzed by chiral SFC, which showed that the obtained **19** did not contain three other potential stereoisomers ((S)-enantiomer, (R)-cis isomer, (S)-cis isomer).

#### 3. Conclusions

A practical chromatography-free synthesis of **19** was successfully developed. We studied cyanoimidazole ring formation closely and showed that the cryogenic condition was not imperative for the reaction provided that appropriate care was taken for the potential epimerization of **8**. This result will allow us to conduct this reaction on a large scale using a normal manufacturing facility. An alternative synthesis of **15** utilizing the advantage of the defined stereochemistry of commercially available *trans* diamine **22** was also developed. The piperazine ring was successfully constructed from primary diamine **20-HCl** without protection. Although the piperazine ring formation requires some improvements to achieve higher yield, this

synthesis provided 15 in a simple way without concern for generation of stereoisomers. The last coupling reaction of 3chloro-5,6-dihydroimidazolo[1,5-f]pteridine 8 with aniline 15 was scrutinized to find feasible reaction conditions and to improve the low yields of the medicinal chemistry synthesis. Consequently, much higher yields were achieved both by the palladium-catalyzed amination and the acid-promoted coupling under benign reaction conditions. Although the acid-promoted coupling seems to be preferable for scale-up from the viewpoint of cost at this point, future development of the palladiumcatalyzed amination might enable further decrease of the amount of a palladium catalyst and render the palladium-catalyzed amination a more cost-effective procedure. The obtained 19 did not include stereoisomers, and column chromatography/HPLC separation was not required. These findings will lead to a successful future scale-up synthesis and development of a potent anticancer drug candidate 19, and at the same time, will

#### 4. Experimental section

#### 4.1 General

All materials were purchased from commercial suppliers and used without any additional purification. NH silica gel (CHROMATOREX) was purchased from Fuji Silysia Chemical Ltd. Silica gel (Silica Gel 60, spherical) was purchased from Nacalai Tesque, Inc. Celite<sup>®</sup> (No. 500) and DARCO<sup>®</sup> (G-60) were purchased from Wako Pure Chemical Industries, Ltd. Melting points were determined on a Stanford Research Systems OptiMelt MPA 100, and are uncorrected unless otherwise noted. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a BRUKER AVANCE 600 spectrometer or a BRUKER AVANCE 500 spectrometer with tetramethylsilane as an internal standard. Chemical shifts are shown in ppm. HPLC analysis of the compounds and reaction monitoring was carried out on a Shimadzu LC-2010C<sub>HT</sub>. High-resolution mass spectrometry (HRMS) data was obtained on a Shimadzu Prominence UFLC system with a Thermofisher LTQ Orbitrap Discovery. IR spectra were recorded on a Thermo Electron FT-IR Nicolet 4700 (ATR). Data of elemental analyses, data of melting points for 35 and 20·HCl (determined by differential scanning calorimetry: DSC), data of HRMS for 7, 23 and 21, and IR spectra of 7, 35, 20·HCl, 23, 21 and 19 were obtained by Sumika Chemical Analysis Service, Ltd.

#### 4.2 HPLC, Chiral HPLC, chiral SFC, and GC conditions

HPLC conditions. (A) Inertsil ODS-3 column, 5  $\mu$ m, 150 mm  $\times$  4.6 mm i.d.; UV detector at 254 nm; isocratic elution with MeCN/50 mM aqueous KH<sub>2</sub>PO<sub>4</sub> (50:50) at 1.0 mL/min flow rate; column temperature: 25 °C. Retention times: **6** (4.0 min), **31** (10.2 min).

(B) YMC A-313 column, 5  $\mu$ m, 250 mm × 6.0 mm i.d.; UV detector at 254 nm; isocratic elution with MeCN/10 mM aqueous AcONH<sub>4</sub> (40:60) at 1.0 mL/min flow rate; column temperature: 25 °C. Retention times: **15** (8.3 min), **20·HCl** (5.4 min), **36** (41 min).

Chiral HPLC conditions. (A) CHIRALCEL OJ-RH, 5  $\mu$ m, 150 mm × 4.6 mm i.d.; UV detector at 254 nm; isocratic elution with MeCN/20 mM aqueous KH<sub>2</sub>PO<sub>4</sub> (40:60) at 1.0 mL/min flow rate; column temperature: 25 °C. Retention times: **5** (11.8 min), *ent*-**5** (15.1 min).

(B) CHIRALCEL OJ-RH, 5  $\mu$ m, 150 mm × 4.6 mm i.d.; UV detector at 254 nm; isocratic elution with MeCN/20 mM aqueous KH<sub>2</sub>PO<sub>4</sub> (30:70) at 1.0 mL/min flow rate; column temperature: 25 °C. Retention times: **6** (8.2 min), *ent*-**6** (5.7 min).

(C) CHIRALCEL OD-RH, 5  $\mu$ m, 150 mm × 4.6 mm i.d.; UV detector at 254 nm; isocratic elution with MeOH at 0.7 mL/min flow rate; column temperature: 25 °C. Retention times: **8** (5.0 min), *ent-***8** (6.5 min).

Chiral SFC condition. CHIRALPAK AD-H, 5  $\mu$ m, 250 mm × 4.6 mm i.d.; UV detector at 220 nm; isocratic elution with CO<sub>2</sub>/EtOH/diethylamine (700:300:3) at 3.0 mL/min flow rate; pressure: 100 bar; column temperature: 35 °C. Retention times: **19** (20.3 min), *ent*-**19** (13.3 min), cis isomer of **19** (9.3 min), cis isomer of *ent*-**19** (7.7 min).

GC condition. SUPELCO SPB<sup>®</sup>-5, Capillary GC Column, 30 m  $\times$  0.53 mm i.d., 5  $\mu m$  film; FID detector; He carrier gas (approximately 6 mL/min); oven heating 90 °C for 5 min,

# 4.3 Methyl (2R)-2-(Propan-2-ylamino)butanoate Hydrochloride (1:1) (**3·HCl**)

To a mixture of methyl (2R)-2-aminobutanoate hydrochloride (1:1) (2·HCl) (14.7 g, 95.7 mmol) in THF (59 mL) were added acetic acid (23.0 g, 383 mmol) and acetone (7.23 g, 124 mmol), and the mixture was stirred at 20-30 °C for 1 h under N<sub>2</sub> atmosphere. To the mixture was added sodium borohydride (3.62 g, 95.7 mmol) portionwise at 10 °C, maintaining the temperature below 20 °C. After being stirred at 20-30 °C for 2 h, the mixture was cooled to 0-10 °C, and H<sub>2</sub>O (44 mL) was slowly added. To the mixture was added 8 M NaOH (54 mL), and pH was adjusted to 9.0-9.5, maintaining the temperature below 30 °C. Toluene (74 mL) was added, and the layers were separated. The organic layer was filtered and insoluble matter was washed with toluene (7 mL). To the combined filtrate were added seed crystals of 3·HCl (15.0 mg) at 0–10 °C. To the mixture was added 4 M HCl in EtOAc (35.9 mL, 144 mmol) dropwise, and the mixture was stirred at 0-10 °C for 1 h. The resulting solids were collected by filtration and washed with EtOAc (29 mL) and dried in vacuo at 50 °C to give 3·HCl (15.1 g, 81%) as a white solid. Mp 155-157 °C; <sup>1</sup>H NMR (600 MHz, DMSO- $d_6$ )  $\delta$  0.92 (t, J = 7.6 Hz, 3H), 1.28 (dd, J = 6.4, 4.2 Hz, 6H), 1.89 (dquin, J = 14.4, 7.5 Hz, 1H), 1.98–2.08 (m, 1H), 3.28 (br s, 1H), 3.78 (s, 3H), 4.04 (br s, 1H), 9.18 (br s, 1H), 9.87 (br s, 1H); <sup>13</sup>C NMR (151 MHz, DMSO-d<sub>6</sub>) δ 9.2, 18.1, 19.1, 22.6, 49.0, 52.8, 57.3, 169.3; IR (ATR) 2969, 2650, 2504, 1736, 1561, 1470, 1436, 1389, 1314, 1264, 1219, 1165, 1117, 1011, 959, 924, 852, 798, 753, 653, 535, 455 cm<sup>-1</sup>; Anal. Calcd for C<sub>8</sub>H<sub>18</sub>ClNO<sub>2</sub>: C, 49.10; H, 9.27; Cl, 18.12; N, 7.16; O, 16.35. Found: C, 49.08; H, 9.03; N, 7.35.

4.4 Methyl (2R)-2-[(2-Chloro-5-nitropyrimidin-4-yl)(propan-2-yl)amino]butanoate (5)

To a mixture of 2,4-dichloro-5-nitropyrimidine (4) (50.0 g, 258 mmol) and 3·HCl (55.5 g, 284 mmol) in toluene (500 mL) was added NaHCO3 (54.1 g, 644 mmol), and the mixture was stirred at 85-95 °C for 3 h under N2 atmosphere. After cooling to room temperature, the mixture was filtered and insoluble matter was washed with toluene (50 mL). To the combined filtrate was added H<sub>2</sub>O (250 mL), and the layers were separated. To the organic layer were added EtOAc (250 mL) and 1 M HCl (250 mL), and the layers were separated. The organic layer was concentrated in vacuo until the weight of the mixture became approximately 100 g. To this mixture was added 2-propanol (250 mL), and the mixture was concentrated in vacuo again until the weight of the mixture became approximately 100 g. To the resulting mixture were added 2-propanol (200 mL) and seed crystals of 5 (50.0 mg), and the mixture was stirred at 20-30 °C for 1 h. The mixture was cooled to 0-10 °C and stirred for 1 h, and then filtrated. Wet solids were washed with 2-propanol/H<sub>2</sub>O (1:1, 100 mL) and dried in vacuo at 50 °C to give 5 (59.7 g, 73%) as a yellow solid. Mp 89–91 °C; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 1.07 (t, J = 7.6 Hz, 3H), 1.31 (d, J = 6.4 Hz, 3H), 1.36 (d, J = 6.4 Hz, 3H), 1.86–2.04 (m, 1H), 2.37–2.56 (m, 1H), 3.55 (dt, J =13.1, 6.5 Hz, 1H), 3.75 (s, 3H), 3.78 (t, J = 6.8 Hz, 1H), 8.63 (s, 1H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 12.0, 19.5, 21.6, 23.2, 52.5, 53.7, 58.8, 131.0, 153.5, 156.5, 159.4, 170.8; IR (ATR) 1739, 1572, 1543, 1513, 1466, 1439, 1374, 1345, 1311, 1215, 1197, 1180, 1165, 1084, 993, 912, 866, 767, 559, 445 cm<sup>-1</sup>; HRMS (ESI):  $[M+H]^+$  calcd for  $C_{12}H_{18}CIN_4O_4$ , 317.1011; found, 317.1008. Optical purity: 99.9% ee (chiral HPLC condition A).

*4.5* (7*R*)-2-*Chloro-7-ethyl-8-(propan-2-yl)-7,8-dihydropteridin-6(5H)-one* (**6**)

To a mixture of 5 (20.0 g, 63.1 mmol) and reduced iron  $\mathcal{M}$ (14.1 g, 253 mmol) in toluene (100 mL) were added AcOH (1.14 g, 18.9 mmol) and H<sub>2</sub>O (10 mL) in this order, and the mixture was stirred at 70-80 °C for 3 h under N<sub>2</sub> atmosphere. To the mixture was added 6 M HCl (100 mL) with vigorous stirring, and the mixture was stirred at 70-80 °C for 1 h. After cooling to room temperature, the mixture was filtered and insoluble matter was washed with H<sub>2</sub>O (40 mL) and EtOAc (40 mL). To the combined filtrate were added EtOAc (160 mL) and H<sub>2</sub>O (60 mL), and the layers were separated. The aqueous layer was extracted with EtOAc (2  $\times$  200 mL), and the combined organic layer was washed with 10% aqueous NaCl (2  $\times$  100 mL) and saturated aqueous NaHCO<sub>3</sub> (200 mL). The organic layer was concentrated in vacuo until the weight of the mixture became approximately 60 g. EtOH (100 mL) was added and the mixture was concentrated in vacuo until the weight of the mixture became approximately 60 g. EtOH (100 mL) was added and the mixture was concentrated in vacuo again until the weight of the mixture became approximately 60 g. To the resulting mixture was added EtOH (20 mL), and H<sub>2</sub>O (90 mL) was added dropwise. The mixture was cooled to 0-10 °C and stirred for 1 h, and then filtrated. Wet solids were washed with EtOH/H2O (1:2, 40 mL) and dried in vacuo at 50 °C to give 6 (12.7 g, 80%) as a white solid. Mp 200–201 °C; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 0.94 (t, J = 7.6 Hz, 3H), 1.37 (d, J = 6.8 Hz, 3H), 1.41 (d, J = 6.8 Hz, 3H), 1.81 (dt, J = 14.5, 7.5 Hz, 1H), 1.99 (ddd, J = 14.5, 7.5, 3.0 Hz, 1H), 4.28 (dd, J = 7.4, 3.2 Hz, 1H), 4.59 (spt, J = 6.8 Hz, 1H), 7.71 (s, 1H), 9.80 (br s, 1H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 8.6, 19.8, 20.9, 27.7, 49.2, 58.6, 117.8, 139.1, 151.9, 154.4, 165.8; IR (ATR) 3229, 2964, 1692, 1655, 1604, 1476, 1410, 1365, 1269, 1237, 1194, 1155, 1127, 1088, 1002, 932, 773, 688, 558, 411, 401 cm<sup>-1</sup>; HRMS (ESI):  $[M+H]^+$  calcd for  $C_{11}H_{16}ClN_4O$ , 255.1007; found, 255.1007. Optical purity: 99.4% ee (chiral HPLC condition B).

#### 4.6 N'-(Cyanomethyl)-N,N-dimethylimidoformamide (7)

To a mixture of aminoacetonitrile hydrochloride (1:1) (26·HCl) (10.0 g, 108 mmol) in THF (100 mL) were added N,Ndimethylformamide dimethyl acetal (16.7 g, 141 mmol) and triethylamine (16.4 g, 162 mmol), and the mixture was stirred at 45-55 °C for 5 h. After cooling to room temperature, the mixture was concentrated in vacuo until the weight of the mixture became approximately 40 g. Diisopropyl ether (50 mL) was added and the mixture was concentrated in vacuo again until the weight of the mixture became approximately 40 g. To the resulting mixture was added diisopropyl ether (100 mL), and the mixture was passed through a pad of NH silica gel (100-200 mesh, 5.00 g), followed by washing with IPE ( $2 \times 100$  mL). The combined filtrate was concentrated in vacuo to give 7 (8.75 g, 88.1 w/w%, 64% GC assay yield) as a pale yellow oil. The obtained 7 was used for the next reaction without further purification. A small portion of the product was purified by distillation (5.6 mmHg, 90 °C) to obtain a sample for analysis. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 2.90 (br s, 6H), 4.18 (s, 2H), 7.44 (s, 1H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 34.6 (br s), 40.0 (br s), 41.5, 118.1, 157.5; IR (ATR) 2917, 1639, 1489, 1432, 1415, 1377, 1326, 1260, 1233, 1114, 1066, 1042, 1017, 984, 970, 903, 865 cm<sup>-1</sup>; HRMS (ESI):  $[M+H]^+$  calcd for C<sub>5</sub>H<sub>10</sub>N<sub>3</sub>, 112.0869; found, 112.0875.

#### 4.7 (6R)-3-Chloro-6-ethyl-5-(propan-2-yl)-5,6dihydroimidazo[1,5-f]pteridine-7-carbonitrile (8)

To a mixture of **6** (5.00 g, 19.6 mmol) in THF (50 mL) was added LHMDS (1.1 M THF solution, 19.6 mL, 21.6 mmol) dropwise at -5 °C, and the mixture was stirred at -5 °C for 0.5 h under N<sub>2</sub> atmosphere. To the mixture was added diethyl

chlorophosphate (6.76 g, 39.2 mmol) dropwise at -5 °C, and the mixture was stirred at -5 °C for 0.5 h. To the mixture was added 7 (4.96 g, 88.1 wt%, 39.3 mmol), and then LHMDS (1.1 M THF solution, 53.5 mL, 58.9 mmol) was added dropwise at -5 °C, and the mixture was stirred at -5 °C for 1 h. To the mixture was added AcOH (30 mL) dropwise, maintaining the temperature below 20 °C, and the mixture was stirred at 40-50 °C for 3 h. After cooling to room temperature, H<sub>2</sub>O (50 mL), EtOAc (50 mL) and THF (50 mL) were added, and the layers were separated. The organic layer was washed with 1 M NaOH ( $3 \times 50$  mL) and H<sub>2</sub>O (50 mL), and concentrated in vacuo until the weight of the mixture became approximately 30 g. EtOAc (50 mL) was added, and the mixture was concentrated in vacuo until the weight of the mixture became approximately 25 g. EtOAc (50 mL) was added and the mixture was concentrated in vacuo again until the weight of the mixture became approximately 25 g. To the mixture was added diisopropyl ether (75 mL) dropwise at 40-50 °C, and the mixture was stirred at 40-50 °C for 1 h. The mixture was gradually cooled to 0–10 °C and stirred for 1 h, and then filtrated. Wet solids were washed with diisopropyl ether (25 mL) and dissolved in THF (150 mL). To the mixture was added activated carbon (500 mg) at 40-50 °C, and the mixture was stirred at 40-50 °C for 1 h. Activated carbon was filtered off and washed with THF (10 mL). The combined filtrate was concentrated in vacuo until the weight of the mixture became approximately 25 g. EtOH (50 mL) was added and the mixture was concentrated in vacuo again until the weight of the solution became approximately 25 g. To the resulting mixture was added EtOH (20 mL), and then  $H_2O$  (50 mL) was added dropwise at 20–30 °C. The mixture was stirred at 20-30 °C for 1 h and then filtrated. Wet solids were washed with H<sub>2</sub>O (25 mL) and dried in vacuo at 50 °C to give 8 (4.40 g, 74%) as a brown solid. Mp 252–253 °C (decomp); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  0.84 (t, *J* = 7.4 Hz, 3H), 1.44 (d, J = 6.8 Hz, 3H), 1.48 (d, J = 6.8 Hz, 3H), 1.84 (dquin, J = 14.6, 7.5 Hz, 1H), 1.97 (ddd, J = 14.4, 7.6, 3.4 Hz, 1H), 4.68 (quin, J = 6.9 Hz, 1H), 5.11 (dd, J = 8.3, 3.4 Hz, 1H), 8.02 (s, 1H), 8.36 (s, 1H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 8.6, 20.1, 20.8, 30.6, 50.6, 51.6, 109.9, 113.4, 114.3, 131.6, 133.1, 141.2, 152.8, 158.2; IR (ATR) 3106, 2970, 2938, 2223, 1605, 1536, 1492, 1419, 1402, 1377, 1362, 1341, 1331, 1236, 1142, 1114, 1081, 962, 755, 651, 586, 507, 463 cm<sup>-1</sup>; HRMS (ESI): [M+H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>16</sub>ClN<sub>6</sub>, 303.1119; found, 303.1118. Optical purity: 99.8% ee (chiral HPLC condition C).

#### 4.8 tert-Butyl (trans-4-Aminocyclohexyl)carbamate $(33)^{14}$

To a mixture of *trans*-cyclohexane-1,4-diamine (**22**) (150 g, 1.31 mol) in MeCN (2.7 L) was added (Boc)<sub>2</sub>O (143 g, 657 mmol) in MeCN (300 mL) maintaining the temperature below 30 °C, and the mixture was stirred at 20–30 °C for 3 h. The resulting slurry was concentrated *in vacuo*. To the residue was added saturated aqueous NaHCO<sub>3</sub> (3 L), and the mixture was stirred at 20–30 °C for 1 h, and then filtrated. Wet solids were washed with H<sub>2</sub>O (400 mL) and dried *in vacuo* at 50 °C to give **33** (**33**:**34** = approximately 3:1, 129 g, 46%) as a white solid. The obtained **33** was used for the next reaction without further purification.

#### 4.9 4-Amino-N-(trans-4-aminocyclohexyl)-2-fluoro-5methoxybenzamide Hydrochloride (1:1) (20·HCl)

To a mixture of 4-amino-2-fluoro-5-methoxybenzoic acid **16** (65.8 g, 355 mmol), **33** (129 g, 604 mmol), *N*,*N*-diisopropylethylamine (91.2 mL, 533 mmol) and 1-hydroxybenzotriazole monohydrate (65.2 g, 426 mmol) in MeCN (1.32 L) was added 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (81.7 g, 426

mmol), and the mixture was stirred at 40 °C for 2 h To-the mixture was added H<sub>2</sub>O (1.32 L) dropwise, and the mixture was stirred at 20-30 °C for 1 h, and then filtrated. Wet solids were washed with H<sub>2</sub>O (526 mL) and dissolved in MeOH (1.32 L). To the mixture was added 6 M HCl (526 mL), and the mixture was stirred at 40 °C for 3 h. After cooling to 5 °C, 8 M NaOH (324 mL) was added dropwise and pH of the mixture was adjusted to 7.8, maintaining the temperature below 20 °C. To the mixture was added activated carbon (6.60 g), and the mixture was stirred at 20-30 °C for 0.5 h. Activated carbon was filtered off and washed with MeOH (132 mL). The combined filtrate was concentrated in vacuo to remove MeOH. The resulting slurry was stirred at 20-30 °C for 0.5 h, and then filtrated. Wet solids were washed with acetone (132 mL) and dried in vacuo at 50 °C to give 20·HCl (92.0 g, 82% for 2 steps) as a brown solid. Mp 309 °C (decomp.) (DSC); <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ )  $\delta$ 1.29–1.53 (m, 4H), 1.84–1.92 (m, 2H), 2.01 (br d, J = 10.4 Hz, 2H), 2.92 (ddd, J = 11.4, 7.5, 4.1 Hz, 1H), 3.60–3.72 (m, 1H), 3.77 (s, 3H), 5.61 (s, 2H), 6.40 (d, J = 13.2 Hz, 1H), 7.03 (d, J = 6.9 Hz, 1H), 7.32–7.54 (m, 1H), 8.30 (br s, 3H);  $^{13}\mathrm{C}$  NMR (126 MHz, DMSO-d<sub>6</sub>) δ 29.6 (2C), 30.4 (2C), 47.6, 49.1, 56.3, 99.8  $(^{2}J_{CF} = 29.0 \text{ Hz}), 108.3 (^{2}J_{CF} = 13.9 \text{ Hz}), 111.4 (^{3}J_{CF} = 5.0 \text{ Hz}),$ 142.4, 142.9 ( ${}^{3}J_{CF} = 12.6$  Hz), 156.0 ( ${}^{1}J_{CF} = 239.4$  Hz), 163.3  $({}^{3}J_{CF} = 2.5 \text{ Hz}); \text{ IR (ATR) } 3428, 3310, 2926, 1622, 1607, 1541,$ 1510, 1319, 1256, 1217, 1169, 1022, 874, 853, 768, 729 cm<sup>-1</sup>; HRMS (ESI):  $[M+H]^+$  calcd for  $C_{14}H_{21}FN_3O_2$  (free form), 282.1612; found, 282.1610. A sample of tert-butyl {trans-4-[(4amino-2-fluoro-5-methoxybenzoyl)amino]cyclohexyl}carbamate (35) for analysis was prepared by the reaction of 20·HCl with 1.0 equiv of Boc<sub>2</sub>O in EtOAc/5% aqueous NaHCO<sub>3</sub>. Data for 35: Mp 248 °C (DSC); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.22–1.38 (m, 4H), 1.44 (s, 9H), 2.01–2.08 (m, 2H), 2.08–2.16 (m, 2H), 3.45 (br s, 1H), 3.88 (s, 3H), 3.90–3.99 (m, 1H), 4.26 (s, 2H), 4.43 (br d, J =6.3 Hz, 1H), 6.36 (d, J = 13.2 Hz, 1H), 6.50 (br dd, J = 15.0, 7.7 Hz, 1H), 7.44 (d, J = 6.9 Hz, 1H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$ 28.4 (3C), 31.8 (2C), 32.1 (2C), 48.0, 49.0, 56.0, 79.2, 100.7  $({}^{2}J_{CF} = 30.2 \text{ Hz}), 108.9 ({}^{2}J_{CF} = 11.3 \text{ Hz}), 111.8 ({}^{3}J_{CF} = 3.8 \text{ Hz}), 141.1 ({}^{3}J_{CF} = 12.6 \text{ Hz}), 143.2, 155.3, 156.4 ({}^{1}J_{CF} = 239.4 \text{ Hz}),$ 163.1 ( ${}^{3}J_{CF} = 2.5 \text{ Hz}$ ); IR (ATR) 3497, 3468, 3317, 2941, 1686, 1620, 1508, 1450, 1366, 1315, 1275, 1261, 1213, 1157, 1072, 1020, 766, 727, 652 cm<sup>-1</sup>; HRMS (ESI): [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>29</sub>FN<sub>3</sub>O<sub>4</sub>, 382.2137; found, 382.2131.

#### 4.10 2-Chloro-N-(2-chloroethyl)-N-(cyclopropylmethyl)ethanamine (21)

To a mixture of 2,2'-iminodiethanol 24 (400 g, 3.80 mol) in EtOH (4 L) were added NaI (684 g, 4.57 mol), (chloromethyl)cyclopropane (413 g, 4.57 mol), and K<sub>2</sub>CO<sub>3</sub> (789 g, 5.71 mol), and the mixture was stirred under reflux for 5.5 h. After cooling to room temperature, the resulting solids were filtered off and washed with EtOAc. The combined filtrate was concentrated in vacuo. To the residue were added EtOAc (8 L) and saturated brine (1.6 L), and the layers were separated. The aqueous layer was extracted with EtOAc (2  $\times$  4 L), and to the combined organic layer was added Na<sub>2</sub>SO<sub>4</sub> (400 g, 2.82 mol), and the mixture was stirred at room temperature for 10 min. The mixture was filtrated and insoluble matter was washed with EtOAc. The combined filtrate was concentrated in vacuo to give 2,2'-[(cyclopropylmethyl)imino]diethanol 23 (472 g, pale yellow oil). The obtained 23 was used for the next reaction without further purification. To a solution of 23 (300 g, 1.88 mol) in toluene (3 L) was added thionyl chloride (448 g, 3.77 mol) dropwise maintaining the temperature below 10 °C, and the mixture was stirred at 50 °C for 3 h. After cooling to room temperature, the mixture was neutralized by slow addition of 8 M NaOH (1.2 L), and the layers were separated. To the organic

layer were added Na<sub>2</sub>SO<sub>4</sub> (500 g, 3.52 mol) and activated carbon (30 g), and the mixture was stirred at room temperature for 30 min. The mixture was filtrated and insoluble matter was washed with toluene, and the combined filtrate was concentrated in vacuo. To the residue was added toluene (300 mL), and the mixture was passed through a pad of silica gel, and insoluble matter was washed with toluene (200 mL). The combined filtrate was concentrated in vacuo to give 21 (343 g, 91.6 wt% determined by <sup>1</sup>H NMR using diphenylmethane as an internal standard, 66% for two steps from 24, pale brown oil). The obtained 21 was used for the next reaction without further purification. A small portion of the obtained 21 was purified by distillation (4 mmHg, 88 °C) to obtain a sample for analysis. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ; 0.06-0.19 (m, 2H), 0.47-0.59 (m, 2H), 0.85 (ttt, J = 8.1, 6.5, 5.0Hz, 1H), 2.49 (d, J = 6.6 Hz, 2H), 2.96 (t, J = 7.3 Hz, 4H), 3.54 (t, J = 7.3 Hz, 4H); <sup>13</sup>C NMR (126 MHz,CDCl<sub>3</sub>)  $\delta$  3.9 (2C), 9.2, 42.0 (2C), 56.5 (2C), 59.8; IR (ATR) 3079, 3001, 2960, 2817, 1462, 1447, 1299, 1251, 1102, 1051, 1019, 936, 828, 802, 765, 726, 661 cm<sup>-1</sup>; HRMS (ESI): [M+H]<sup>+</sup> calcd for C<sub>8</sub>H<sub>16</sub>Cl<sub>2</sub>N, 196.0654; found, 196.0630. A small portion of the obtained 23 was purified by distillation (3 mmHg, 134 °C) to obtain a sample for analysis. Data for 23: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ; 0.08-0.16 (m, 2H), 0.45–0.60 (m, 2H), 0.89 (ttt, J = 8.1, 6.6, 5.0 Hz, 1H), 2.45 (d, J = 6.6 Hz, 2H), 2.73–2.79 (m, 4H), 3.57–3.71 (m, 4H);  $^{13}$ C NMR (126 MHz,CDCl<sub>3</sub>)  $\delta$  4.0 (2C), 8.8, 56.0 (2C), 59.6, 59.6 (2C); IR (ATR) 3347, 2939, 2876, 2820, 1033, 888, 873, 828 cm<sup>-1</sup>; HRMS (ESI):  $[M+H]^+$  calcd for C<sub>8</sub>H<sub>18</sub>NO<sub>2</sub>, 160.1332; found, 160.1335.

#### 4.11 4-Amino-N-{trans-4-[4-(cyclopropylmethyl)piperazin-1yl]cyclohexyl}-2-fluoro-5-methoxybenzamide (15)

To a mixture of **20·HCl** (10.0 g, 31.5 mmol), K<sub>2</sub>CO<sub>3</sub> (13.1 g, 94.8 mmol) and NaI (4.72 g, 31.5 mmol) in EtOH (50 mL) was added 21 (7.40 g, 37.7 mmol), and the mixture was stirred at 20-30 °C for 15 h. To the mixture were added EtOAc (50 mL), THF (50 mL) and H<sub>2</sub>O (100 mL), and the layers were separated. To the organic layer were added 10% aqueous NaCl (90 mL) and 12 M HCl (13 mL) maintaining the temperature below 30 °C, and the layers were separated. The aqueous layer was concentrated in vacuo to remove EtOH and THF. To the resulting aqueous layer was added activated carbon (1.00 g), and the mixture was stirred at 20-30 °C for 0.5 h. Activated carbon was filtered off, and washed with H<sub>2</sub>O (20 mL). To the combined filtrate were added EtOAc (50 mL) and THF (50 mL). To the mixture was added 25% aqueous NH<sub>3</sub> (18 mL) dropwise maintaining the temperature below 30 °C, and the layers were separated. The organic layer was washed with saturated aqueous NaCl (50 mL), dried over MgSO<sub>4</sub>, and concentrated in vacuo. To the residue was added EtOH (60 mL), and 15% HCl in EtOH (40 mL) was added dropwise at 20-30 °C. The resulting slurry was stirred at 20-30 °C for 1 h, and 5-10 °C for 1 h, and then filtrated. Wet solids were washed with EtOH (15 mL), and dissolved in H<sub>2</sub>O (65 mL). To the mixture was added activated carbon (600 mg), and the mixture was stirred at 20-30 °C for 0.5 h. Activated carbon was filtered off, and washed with H<sub>2</sub>O (10 mL). To the combined filtrate was added 25% aqueous NH3 (3.1 mL), and pH was adjusted to 9.0. The resulting slurry was stirred at 20-30 °C for 1 h and then filtrated. Wet solids were washed with H<sub>2</sub>O (13 mL) and dried in vacuo at 50 °C to give 15 (4.73 g, 37%) as a white solid. Mp 184–185 °C; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 0.10 (q, J = 4.9 Hz, 2H), 0.45-0.57 (m, 2H), 0.82-0.92 (m, 1H), 1.20-1.31 (m, 2H), 1.43 (qd, J = 12.5, 3.0 Hz, 2H), 1.97 (br d, J = 12.5 Hz, 2H), 2.16 (br d, J = 11.3 Hz, 2H), 2.25 (d, J = 6.8 Hz, 2H), 2.27-2.32 (m, 1H), 2.64 (br s, 8H), 3.87 (s, 3H), 3.89-3.95 (m, 1H), 4.27 (s, 2H), 6.36 (d, J = 13.2 Hz, 1H), 6.50 (br dd, J = 14.9, 7.7 Hz, 1H), 7.45 (d, J = 7.2 Hz, 1H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$ 

3.9 (2C), 8.3, 27.3 (2C), 32.3 (2C), 48.7, 49.0 (2C), 53.7 (2C), M 56.0, 62.7, 63.8, 100.6 ( ${}^{2}J_{CF} = 31.7$  Hz), 108.9 ( ${}^{2}J_{CF} = 12.1$  Hz), 111.7 ( ${}^{3}J_{CF} = 3.0$  Hz), 141.0 ( ${}^{3}J_{CF} = 13.6$  Hz), 143.1, 156.4 ( ${}^{1}J_{CF} = 238.6$  Hz), 163.0 ( ${}^{3}J_{CF} = 4.5$  Hz); IR (ATR) 3481, 3346, 2937, 2809, 1738, 1619, 1512, 1451, 1309, 1236, 1214, 1160, 1005, 871, 836, 797, 766, 729, 584, 523 cm<sup>-1</sup>; HRMS (ESI): [M+H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>34</sub>FN<sub>4</sub>O<sub>2</sub>, 405.2660; found, 405.2659.

4.12 Synthesis of 4-{[(6R)-7-Cyano-6-ethyl-5-(propan-2-yl)-5,6dihydroimidazo[1,5-f]pteridin-3-yl]amino}-N-{trans-4-[4-(cyclopropylmethyl)piperazin-1-yl]cyclohexyl}-2-fluoro-5methoxybenzamide (**19**) by coupling of **8** with **15** using a palladium catalyst

To a mixture of 15 (1.00 g, 2.47 mmol), 8 (823 mg, 2.72 mmol) and potassium carbonate (683 mg, 4.94 mmol) in sbutanol (10 mL) were added Xantphos (143 mg, 0.247 mmol) and palladium(II) acetate (55.5 mg, 0.247 mmol), and the mixture was stirred at 85 °C for 1 h. After cooling to room temperature, H<sub>2</sub>O (15 mL) and 6 M HCl (3 mL) were added, and the mixture was filtered through a pad of Celite<sup>®</sup> (No. 500, 2.00 g). To the filtrate was added EtOAc (10 mL), and the layers were separated. To the aqueous layer was added 8 M NaOH (1.5 mL) and pH was adjusted to 9.5. The resulting aqueous layer was extracted with EtOAc/THF (3:2, 25 mL). The organic layer was washed with 25% aqueous NH<sub>3</sub> (10 mL) and saturated aqueous NaCl (10 mL). The organic layer was filtrated to remove insoluble matter, and the filtrate was concentrated in vacuo. To the residue was added 2-propanol (10 mL), and the mixture was concentrated in vacuo. To the residue was added 2-propanol (5 mL), and the mixture was stirred at 20-30 °C for 1 h, and then filtrated. Wet solids were washed with 2-propanol (2 mL) and H<sub>2</sub>O (2 mL), and dried in vacuo at 50 °C to give 19 (1.30 g, 78%) as a pale yellow solid. Mp 240-242 °C; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 0.07–0.14 (m, 2H), 0.47–0.55 (m, 2H), 0.81–0.92 (m, 4H), 1.21–1.35 (m, 2H), 1.44–1.50 (m, 5H), 1.54 (d, *J* = 6.8 Hz, 3H), 1.76–1.87 (m, 1H), 1.93 (dtd, J = 14.2, 7.3, 3.4 Hz, 1H), 1.99 (br d, J = 12.5 Hz, 2H), 2.15–2.22 (m, 2H), 2.26 (d, J = 6.8 Hz, 2H), 2.28–2.34 (m, 1H), 2.65 (br s, 8H), 3.90–3.96 (m, 1H), 3.97 (s, 3H), 4.79 (spt, J = 6.8 Hz, 1H), 5.08 (dd, J = 8.3, 3.4 Hz, 1H), 6.61 (br dd, *J* = 14.9, 7.7 Hz, 1H), 7.58 (d, *J* = 6.8 Hz, 1H), 7.86 (s, 1H), 7.96 (s, 1H), 8.30–8.37 (m, 1H), 8.43 (d, J = 15.1 Hz, 1H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 3.9 (2C), 8.3, 8.8, 20.2, 21.4, 27.3 (2C), 30.6, 32.3 (2C), 48.8, 49.1 (2C), 49.1, 50.6, 53.7 (2C), 56.3, 62.7, 63.8, 104.9 ( ${}^{2}J_{CF} = 36.2 \text{ Hz}$ ), 109.2, 109.8, 111.1  $({}^{3}J_{CF} = 4.5 \text{ Hz}), 112.2 ({}^{2}J_{CF} = 13.6 \text{ Hz}), 114.0, 130.8, 133.0,$ 133.2 ( ${}^{3}J_{CF} = 13.6$  Hz), 141.2, 143.7, 152.4, 155.6 ( ${}^{1}J_{CF} = 238.6$ Hz), 157.2, 162.5 ( ${}^{3}J_{CF} = 4.5$  Hz); IR (nujol) 3406, 2227, 1637, 1623, 1607, 1550, 1521, 1501, 1490, 1447, 1313, 1275, 1252, 1204, 1176, 1154, 1065, 1022, 1007, 964, 875, 780 cm<sup>-1</sup>; HRMS (ESI):  $[M+H]^+$  calcd for  $C_{36}H_{48}FN_{10}O_2$ , 671.3940; found, 671.3943. Optical purity: >99.9% ee (chiral SFC condition).

4.13 Synthesis of 4-{[(6R)-7-Cyano-6-ethyl-5-(propan-2-yl)-5,6dihydroimidazo[1,5-f]pteridin-3-yl]amino}-N-{trans-4-[4-(cyclopropylmethyl)piperazin-1-yl]cyclohexyl}-2-fluoro-5methoxybenzamide (**19**) by coupling of **8** with **15** in an acidic condition

To a mixture of **15** (3.00 g, 7.42 mmol) and **8** (5.39 g, 17.8 mmol) in *s*-BuOH/H<sub>2</sub>O (95:5, 30 mL) was added concentrated hydrochloric acid (2.25 g, 22.3 mmol), and the mixture was stirred at reflux for 11 h. After cooling to room temperature, the mixture was concentrated *in vacuo*. To the residue were added EtOAc (45 mL) and 1 M HCl (60 mL), and the layers were separated. The organic layer was extracted with 1 M HCl (30 mL). To the combined aqueous layer was added 8 M NaOH (30 mL) and pH was adjusted to 9. The resulting aqueous layer was

extracted with EtOAc (60 mL). The organic layer was washed with 5% aqueous NaHCO<sub>3</sub> (3 × 60 mL) and concentrated *in vacuo*. To the residue was added 2-propanol (30 mL), and the mixture was stirred at 40–50 °C for 1 h. The mixture was gradually cooled to 0–10 °C and stirred for 1 h, and then filtrated. Wet solids were washed with 2-propanol/*n*-heptane (1:3, 9 mL) and dissolved in MeOH (60 mL). To the mixture was added DARCO<sup>®</sup> (G-60, 600 mg), and the mixture was stirred at 40– 50 °C for 1.5 h. DARCO<sup>®</sup> was filtered off, and the filtrate was concentrated *in vacuo*. To the residue was added 2-propanol (15 mL), and the mixture was stirred at 40–50 °C for 1 h. The mixture was gradually cooled to 0–10 °C and stirred for 1 h, and then filtrated. Wet solids were washed with *n*-heptane (9 mL) and dried *in vacuo* at 50 °C to give **19** (3.54 g, 71%) as a pale gray solid.

#### Acknowledgments

We are grateful to Mr. Keiichi Satou for LC/MS analysis, and Ms. Rie Kasai for chiral SFC analysis, and Dr. Makoto Kanematsu for his advice on the manuscript.

#### Appendix A. Supplementary Data

Supplementary data related to this article can be found at

#### **References and notes**

- For recent reviews, see: (a) Lens, S. M. A.; Voest, E. E.; Medema, R. H. *Nat. Rev. Cancer* **2010**, *10*, 825–841. (b) Wurzenberger, C.; Gerlich, D.W. *Nat. Rev. Mol. Cell Biol.* **2011**, *12*, 469–482. (c) Bruinsma, W.; Raaijmakers, J. A.; Medema, R. H. *Trends Biochem. Sci.* **2012**, *37*, 534–542. (d) Zitouni, S.; Nabais, C.; Jana, S. C.; Guerrero, A.; Bettencourt-Dias, M. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 433–452. (e) Archambault, V.; Lépine, G.; Kachaner, D. *Oncogene* **2015**, *34*, 4799–4807.
- For recent reviews, see: (a) Strebhardt, K. Nat. Rev. Drug Discov.
  2010, 9, 643–660. (b) Christoph, D. C.; Schuler, M. Expert Rev. Anticancer Ther. 2011, 11, 1115–1130. (c) McInnes, C.; Wyatt, M. D. Drug Discov. Today 2011, 16, 619–625. (d) Craig, S. N.; Wyatt, M. D.; McInnes, C. Expert Opin. Drug Discov. 2014, 9, 773–789. (e) Palmisiano, N. D.; Kasner, M. T. Am. J. Hematol. 2015, 90, 1071–1076. (f) Berg, A.; Berg, T. Chembiochem 2016, 17, 650– 656. (f) Talati, C.; Griffiths, E. A.; Wetzler, M.; Wang, E. S. Crit. Rev. Oncol. Hematol. 2016, 98, 200–210.
- (a) Cao, S. X.; Ichikawa, T.; Kiryanov, A. A.; Mcbride, C.; Natala, S. R.; Kaldor, S. W.; Stafford, J. A. PCT Int. Appl. WO 2010025073, 2010. (b) Kiryanov, A.; Natala, S.; Jones, B.; McBride, C.; Feher, V.; Lam, B.; Liu, Y.; Honda, K.; Uchiyama, N.; Kawamoto, T.; Hikichi, Y.; Zhang, L.; Hosfield, D.; Skene, R.; Zou, H.; Stafford, J.; Cao, X.; Ichikawa, T. *Bioorg. Med. Chem. Lett.* 2017, *27*, 1311–1315.
- (a) Hoffmann, M.; Grauert, M.; Brandl, T.; Breitfelder, S.; Eickmeier, C.; Steegmaier, M.; Schnapp, G.; Baum, A.; Quant, J. J.; Solca, F.; Colbatzky, F. PCT Int. Appl. WO 2004076454, 2004.
   (b) Linz, G.; Kraemer, G. F.; Gutschera, L.; Asche, G. PCT Int. Appl. WO 2006018220, 2006. (c) Schnaubelt, J.; Herter, R. PCT Int. Appl. WO 2012049153, 2012.
- 5. The ratio of 19 to 19' of a representative lot prepared by palladium-catalyzed amination of 8 with 15 was found to be 93:7 before HPLC separation. Our initial target for the amount of 19' was set as not more than 0.50%.
- Anderson, N. G. Practical Process Research & Development A Guide for Organic Chemists, 2nd ed.; Academic Press: New York, 2012.
- Leopoldo, M.; Lacivita, E.; Colabufo, N. A.; Berardi, F.; Perrone, R. J. Pharm. Pharmacol. 2006, 58, 209–218.
- For other syntheses of 6, see: (a) Duran, A.; Linz, G. PCT Int. Appl. WO 2006058876, 2006. (b) Grauert, M.; Linz, G.; Schmid, R.; Sieger, P. PCT Int. Appl. WO 2007090844, 2007. (c) Linz, G.; Sieger, P.; Schmid, R.; Goepper, S. PCT Int. Appl. WO 2009019205, 2009.

- Hoffmann, M.; Grauert, M.; Brandl, T.; Breitfelder, S.; Eickmeier, MANUSCRIPT C.; Steegmaier, M.; Schnapp, G.; Baum, A.; Quant, J. J.; Solca, F.; Colbatzky, F. U.S. Pat. Appl. US20040176380, 2004.
- 10. Rogers-Evans, M.; Spurr, P.; Hennig, M. *Tetrahedron Lett.* **2003**, *44*, 2425–2428.
- 11. Reference 10 reported that it was possible to isolate intermediate **29a** after non-aqueous workup.
- Fryer, R. I.; Kudzma, L. V.; Gu, Z.-Q.; Lin, K.-Y.; Rafalko, P. W. J. Org. Chem. 1991, 56, 3715–3719.
- 13. We checked stability of **31** by extending the reaction time for phosphorylation. Compound **6** was treated with 1.2 equiv of LHMDS and 1.5 equiv of diethyl chlorophosphate in THF at -5 °C for 3 h, and the reaction mixture was monitored by HPLC every hour. HPLC area% of **31** at each hour was as follows: 80.1 area% after 1 h, 80.2 area% after 2 h, and 81.6 area % after 3 h. These results indicate that **31** was stable up to 3 h at -5 °C.
- (a) Liu, B.; Liu, G.; Xin, Z.; Serby, M. D.; Zhao, H.; Schaefer, V. G.; Falls, H. D.; Kaszubska, W.; Collins, C. A.; Sham, H. L. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 5223–5226. (b) Wang, N.; Xiang, J.; Ma, Z.; Quan, J.; Chen, J.; Yang, Z. J. Comb. Chem. **2008**, *10*, 825–834. (c) Caron, K.; Lachapelle, V.; Keillor, J. W. Org. Biomol. Chem. **2011**, *9*, 185–197. (d) Bailey, S.; Barber, C. G.; Glossop, P. A.; Middleton, D. S. PCT Int. Appl. WO 2005009966, 2005. (e) Lin, S.; Yang, Z. PCT Int. Appl. WO 2010022159, 2010.
- For examples of piperazine ring formation, see: (a) Lazar, S.; Soukri, M.; Leger, J. M.; Jarry, C.; Akssira, M.; Chirita, R.; Grig-Alexa, I. C.; Finaru, A.; Guillaumet, G. *Tetrahedron* 2004, 60, 6461–6473. (b) Mutulis, F.; Yahorava, S.; Mutule, I.; Yahorau, A.; Liepinsh, E.; Kopantshuk, S.; Veiksina, S.; Tars, K.; Belyakov, S.; Mishnev, A.; Rinken, A.; Wikberg, J. E. S. *J. Med. Chem.* 2004, 47, 4613–4626. (c) Féau, C.; Klein, E.; Dosche, C.; Kerth, P.; Lebeau, L. *Org. Biomol. Chem.* 2009, 7, 5259–5270. (d) Andrews, M.; Brown, A.; Chiva, J.-Y.; Fradet, D.; Gordon, D.; Lansdell, M.; MacKenny, M. *Bioorg. Med. Chem. Lett.* 2009, *19*, 2329–2332. (e) Zheng, Y.-Y.; Xie, P.; Xing, L.-X.; Wang, J.-Y.; Li, J.-Q. Org. Process Res. Dev. 2012, *16*, 1921–1926.
- Bartkovitz, D. J.; Chu, X.-J.; Ding, Q.; Jiang, N.; Lovey, A. J.; Moliterni, J. A.; Mullin, J. G. Jr.; Vu, B. T.; Wovkulich, P. M. PCT Int. Appl. WO 2004069139, 2004
- 17. Charrier, J. D.; Kay, D.; Knegtel, R. PCT Int. Appl. WO 2008076392, 2008.
- (a) Wolfe, J. P.; Wagaw, S.; Marcoux, J.-F.; Buchwald, S. L. Acc. Chem. Res. 1998, 31, 805–818. (b) Hartwig, J. F. Acc. Chem. Res. 1998, 31, 852–860. (c) Hartwig, J. F. Angew. Chem. Int. Ed. 1998, 37, 2046–2067. (d) Yang, B. H.; Buchwald, S. L. J. Organomet. Chem. 1999, 576, 125–146.