Tetrahedron Letters 56 (2015) 6593-6597

Contents lists available at ScienceDirect

Tetrahedron Letters

journal homepage: www.elsevier.com/locate/tetlet



Synthesis of various substituted 5-methyluridines (xm⁵U) and 2-thiouridines (xm⁵s²U) via nucleophilic substitution of 5-pivaloyloxymethyluridine/2-thiouridine

Karolina Bartosik, Grazyna Leszczynska*

Institute of Organic Chemistry, Lodz University of Technology, Zeromskiego 116, 90-924 Lodz, Poland

ARTICLE INFO

Article history: Received 2 September 2015 Revised 1 October 2015 Accepted 6 October 2015 Available online 8 October 2015

Keywords: Modified nucleosides 5-Methyl(-2-thio)uridines (xm⁵(s²)U) 5-Pivaloyloxymethyl(-2-thio)uridine (Pivom⁵(s²)U) Pivaloyloxyl group

ABSTRACT

5-Pivaloyloxymethyluridine and its 2-thio analogue have been utilized as convenient substrates for the synthesis of various 5-methyluridines (xm⁵U) and 5-methyl-2-thiouridines (xm⁵s²U). The pivaloyloxy group (OPiv) located at the pseudobenzylic position was effectively substituted with a series of nucleophiles: ammonia, primary and secondary amines including secondary cyclic amines, tetrabutylammonium salts of amino acids, an alkoxide and a thiolate.

© 2015 Elsevier Ltd. All rights reserved.

5-Methyluridines (xm⁵U) and their 2-thio analogues (xm⁵s²U) are a biologically important class of modified nucleosides.¹ In particular, 5-aminomethyluridine/2-thiouridine derivatives, for example, 5-methylaminomethyl- (mnm⁵U and mnm⁵s²U), 5-carboxymethylaminomethyl- (cmnm⁵U and cmnm⁵s²U) and 5-taurinomethyl-modified uridines ($\tau m^5 U$ and $\tau m^5 s^2 U$), are present at the first position of the anticodon (the wobble position) in several cytosolic and/or mitochondrial tRNAs that are responsible for precise recognition of the purine-ending codons.^{2,3} In some cases, the lack of xm⁵s²U wobble units totally inhibits the protein biosynthesis.^{4,5} For example, the absence of $\tau m^5 s^2 U_{34}$ in the sequence of human mitochondrial (mt) tRNA^{Lys} gives a defect of UUA and UUG-rich genes translation that results in mitochondrial encephalomyopathic disease MERRF.⁴ In DNA sequences, the presence of xm⁵-modified 2'-deoxyuridines (xm⁵dU) is limited to 5-hydroxymethyldU that is a well-characterized product of thymine oxidation and was recently shown to be an epigenetic base in mouse embryonic stem cells DNA.⁶

Site-specific incorporation of the native 5-methyluridines/ 2-thiouridines into oligoribonucleotide sequences offers a reliable tool for model studies on the structural requirements of tRNA interactions with partners in complex bioprocesses.^{3,7} Efficient and simple approaches to the synthesis of 5-methyluridine and 2-thiouridine derivatives are an important element of those studies. Moreover, native 5-methyluridines xm^5U/xm^5s^2U are frequently used as authentic standards to identify individual ribonucleosides in RNA hydrolyzates using liquid chromatography-mass spectrometry analysis.⁸ Some of them are important synthons for the preparation of more complex nucleosides, for example, mnm⁵s²U or cmnm⁵s²U can be considered as substrates for the synthesis of recently discovered 5-substituted *S*-geranylated 2-thiouridines.⁹

Unnatural C5-modified pyrimidine nucleosides have been screened for antiviral and antibacterial activities.¹⁰ Various 5-amine or 5-thiol containing pyrimidine nucleosides have found application as useful units for duplex structure stabilization as well as for the introduction of additional RNA functionality such as aptamers, biosensors or catalysts.¹¹ The protected forms of 5-hydroxymethyldU were applied in the transient chemical protection of DNA against cleavage by restriction endonucleases.¹²

The most frequently used method for the synthesis of the title compounds involves nucleophilic substitution of 5-chloromethyl-2',3'-O-isopropylideneuridine/2-thiouridine (**1a/1b**, Fig. 1).^{13–17} The reactions of **1a/1b** with nucleophiles proceed under mild anhydrous conditions affording xm^5U/xm^5s^2U products (x = -CN, $-N_3$, $-NHCH_3$, $-NHCH_2COOH(R)$, $-NHCH_2CH_2SO_3H(R)$) in moderate yields.^{13–17} However, a significant decrease in yield has been observed in the synthesis of 2-thiouridine analogues.^{14–16} Additionally, the susceptibility of the chloromethyl group of **1a/1b** to hydrolysis renders some nucleophilic substitution





^{*} Corresponding author. Tel.: +48 42 631 31 50; fax: +48 42 636 55 30. *E-mail address:* grazyna.leszczynska@p.lodz.pl (G. Leszczynska).



Figure 1. Substrates utilized for the synthesis of 5-methyluridines xm^5U and 2-thiouridines xm^5s^2U reported in the literature.

reactions unsuccessful, for example, amination of **1a**/**1b** with aqueous NH₃.¹⁷ Alternative methods involve using *N*-methyl quaternary ammonium salts **2a**/**2b**, **3a** (Fig. 1),¹⁷⁻¹⁹ 5-hydroxymethy-luridine/2-thiouridine **4a**/**4b**,²⁰ 5-formyluridine/2-thiouridine **5a**/**5b**¹⁵ or 5-aminomethyluridine/2-thiouridine **6a**/**6b**²¹ derivatives as substrates. The reactions involving compounds **2a**/**2b**-**6a**/**6b** proceed with moderate to low yields and have been demonstrated only on a single target molecule. Notably, the treatment of 2-thiouridines **2b** or **4b** with a nucleophile resulted in partial desulfurization (s² \rightarrow o²).^{16,20}

In this work, we present an effective and convenient approach to the synthesis of various 5-methyluridines (xm⁵U) and 2-thiouridines (xm⁵s²U) utilizing the reaction of 5-pivalovloxymethyluridine 8a and its 2-thio analogue 8b and a set of different nucleophiles: ammonia, primary and secondary amines. amino acid salts, an alkoxide and a thiolate. Previous studies reported partial nucleophilic substitution of 5-acetoxymethyl-2'deoxyuridine/cytidine-modified DNAs with ammonia that involved a substitution of the acetoxy group (OAc) located at the pseudobenzylic position.²² However, ammonolysis of the acetate ester was predominant and regeneration of the parent pseudobenzyl alcohol was observed.²² Contrary to other ester based alcohol protecting groups, pivalate shows relative stability in the presence of base; thus we assumed that its ammonolysis would be significantly reduced or even eliminated under basic conditions. Additionally, the formation of a pivalate ester at the pseudobenzylic position could effectively promote an $S_{N}% \left(\boldsymbol{x}_{n}^{\prime}\right) = S_{N}\left(\boldsymbol{x}_{n}^{\prime}\right) + S_{N}\left(\boldsymbol{x}_{n}^{\prime}\right)$ nucleophilic attack occurring at the pseudobenzylic carbon with OPiv acting as a leaving group.

To prepare 5-pivaloyloxymethyluridine (8a) and its 2-thio analogue **8b**, 5-hydroxymethyl-2',3'-O-isopropylideneuridine/2thiouridine²³ (**4a/4b**) was converted into **8a/8b** by selective pivaloylation of the 5-hydroxymethyl group followed by removal of the 2',3'-isopropylidene protecting group (Scheme 1). Different reactivities of the 5'-hydroxyl and 5-hydroxymethyl groups under acidic conditions were previously reported by Sowers and Beardsley for selective acetylation of 5-hydroxymethyl-2'deoxyuridine.²⁴ Using a similar procedure, compound **4a** was treated with pivalic acid (PivOH) in the presence of trifluoroacetic acid at 120 °C for 10 h. Although the desired regioisomer 7a was formed, the following isolation and purification proved difficult for separation of the pure product from the pivalic acid sideproducts. Multiple attempts to purify 7a resulted in a significant decrease in yield (<30%). A more effective method of 5-hydroxvmethyl acylation involved a strategy previously reported for the regioselective pivalovlation of carbohydrates.²⁵ Compounds 4a/4b were treated with 1.1 equiv of pivaloyl chloride (PivCl) in pyridine (Scheme 1) under mild conditions (0 °C, 7 h). Although the low temperature significantly increased the selectivity of pivaloylation, a mixture of unreacted substrate 4a/4b, 5-pivalate-7a/7b and 5,5'-bispivalate esters in a 1:2:1 ratio was obtained. To achieve the complete conversion of substrates 4a/4b, an additional 0.9 equiv of PivCl was added and the reaction continued for 10 h. Mono 5-pivalate component 7a/7b was separated by column chromatography and treated with 50% aq acetic acid for effective removal of the 2',3'-acetonide. After purification, 5-pivaloyloxymethyluridine (8a, Pivom⁵U) and its 2-thio analogue (8b, Pivom⁵s²U) were obtained in ca. 50% total yield in each case.

Nucleosides **8a/8b** were then reacted with a wide range of structurally diverse nucleophilic reagents: ammonia, primary and secondary amines including cyclic amines (piperidine, morpholine), tetrabutylammonium salts of amino acids (glycine and taurine), an alkoxide and a thiolate (ESI, pg. S6–S14). The reaction conditions were optimized in terms of solvent (EtOH/MeOH, water, DMF and neat conditions), reaction time and the excess of nucle-ophilic reagent (Table 1). The products of nucleophilic substitution **9a–17a/9b–17b** were purified by preparative RP HPLC and their structures unambiguously confirmed by ¹H and ¹³C NMR as well as mass spectrometry (ESI, pg. S32–S49).

In general, these reactions were carried out at elevated temperature (50–60 °C) except for the nucleophilic substitution with methylamine (entry 4) which proceeded at room temperature. Esters **8a/8b** rapidly reacted with ammonia and methylamine (1–4 h) under anhydrous conditions using 8 M NH₃/EtOH (entry 1) and 8 M MeNH₂/EtOH (entries 3 and 4) to give products **9a/9b** and **10a/10b**, respectively, in good yields of 86–90%. Interestingly, changing from anhydrous conditions to an aqueous solution of ammonia or methylamine (entries 2 and 5) resulted in only a slight decrease in yield which was attributed to aminolysis of the pivalate ester and regeneration of 5-hydroxymethyluridine/2-thiouridine (hm⁵U/hm⁵s²U). On the other hand, effective nucleophilic



Scheme 1. Synthetic route to 5-pivaloyloxymethyluridine/2-thiouridine (8a/8b). Reagents and conditions: (i) PivCl ($1.1 \rightarrow 2$ equiv), py, 17 h, 0 °C; (ii) 50% aq AcOH, 1.5 h, 85 °C.

Table 1

Nucleophilic substitution of 5-pivaloyloxymethyluridine (8a) and 5-pivaloyloxymethyl-2-thiouridine (8b)



Entry	-Nu	Nu system	Time (h)	Product	Yield ^b (%)	Product	Yield ^b (%)
1	-NH ₂	8 M NH ₃ /EtOH	4	9a	87	9b	86
2	-NH ₂	30% aq NH ₃	2	9a	80	9b	80
3	-NHMe	8 M MeNH ₂ /EtOH	1	10a	90	10b	90
4	-NHMe	8 M MeNH ₂ /EtOH	2	10a	89	10b	87
5	-NHMe	40% aq MeNH ₂	1	10a	85	10b	84
6	-NEt ₂	$Et_2NH/H_2O 4/1 v/v$	20	11a	70	11b	70
7	-NC ₄ H ₈ O	Morpholine/H ₂ O 4/1 v/v	20	12a	84	12b	80
8	$-NC_5H_{10}$	Piperidine/EtOH 1/1 v/v	20	13a	85	13b	80
9	$-NC_5H_{10}$	Piperidine/H ₂ O 4/1 v/v	20	13a	80	13b	75
10	-NHCH ₂ COOH	0.8 M NH ₂ CH ₂ COO ⁻ NBu ₄ ⁺ /EtOH	20	14a	77	14b	75
11	-NH(CH ₂) ₂ SO ₃ H	0.8 M NH ₂ (CH ₂) ₂ SO ₃ NBu ⁺ /EtOH	20	15a	75	15b	75
12	-OMe	0.1 M K ₂ CO ₃ /MeOH	1	16a	72	16b	70
13	–SEt	0.5 M EtSNa/EtOH	1	17a	70	17b	70

^a rt for synthesis of **10a**, **10b** (entry 4), 50 °C for synthesis of **11a**, **11b** (entry 6).

^b Isolated yields.



Scheme 2. Synthetic route to 5'-O-pivaloyl-5-pivaloyloxymethyluridine/2-thiouridine (19a/19b). Reagents and conditions: (i) PivCl (3 equiv), py, 2 h, rt; (ii) 50% aq AcOH, 2 h, 85 °C.

substitution of 8a/8b with a secondary aliphatic amine and morpholine (entries 6 and 7) required aqueous conditions for complete substrate conversion. For piperidine, both anhydrous and aqueous solutions provided products 13a/13b in good yield (entries 8 and 9). Contrary to ammonia and methylamine, the reactions of 8a/8b with secondary amines required prolonged reaction times. Due to the use of aqueous conditions, the formation of minor amounts of hm⁵U/hm⁵s²U (1–5%) was also observed. Attempts to substitute the 5-OPiv group of 8a/8b with amino acids were performed with glycine and taurine which are the fragments of native, wobble 5-substituted uridines and 2-thiouridines. Prior to the reaction, both amino acids were converted into their corresponding tetrabutylammonium salts, providing good solubility in EtOH as well as enhanced nucleophilicity compared to their zwitterionic forms. The reactions of 8a/8b with 0.8 M solutions of the tetrabutylammonium salts of glycine or taurine in anhydrous EtOH (entries 10 and 11) proceeded at 60 °C for 20 h to give 5-carboxymethylaminomethyluridine/2-thiouridine 14a/14b and 5-taurinomethyluridine/2-thiouridine 15a/15b, all in ca. 75% yields.

In addition to nitrogen nucleophiles, alkoxide and thiolate nucleophilic reagents were utilized for 5-OPiv substitution. The reactions of **8a/8b** with 0.1 M K₂CO₃/MeOH (entry 12) and 0.5 M EtSNa/EtOH (entry 13) proceeded rapidly under anhydrous conditions yielding 5-methoxymethyluridine/2-thiouridine (**16a/16b**)

and 5-ethylthiomethyluridine/2-thiouridine (**17a/17b**), respectively, along with small amounts of hm^5U/hm^5s^2U (<5%). For the preparation of **16a/16b**, 10% DBU/MeOH and 0.5 M MeONa/MeOH were also tested, however methanolysis of the pivalate ester was predominant.

To investigate the potential of the new method, we carried out the nucleophilic substitution of the 5-OPiv group in a moiety containing an additional pivalate ester at the aliphatic 5'-hydroxyl position. The conditions previously screened for 5-OPiv substitution (Table 1) were used to investigate the reactivity of the 5'-pivaloyl group. For the synthesis of bispivalate esters **19a/19b** 5-hydroxymethyl-2',3'-O-isopropylideneuridine/2-thiouridine (**4a/4b**) were reacted with 3 equiv of PivCl (rt, 2 h) and the cis diol functional group was selectively deprotected by treatment with 50% aq acetic acid to give **19a/19b** in 70% overall yield (Scheme 2) (ESI).

Compounds **19a/19b** were reacted with the same series of nucleophiles (Table 2). In general, the 5'-O-pivaloyl group proved to be stable under the employed conditions, affording compounds **20a–25a/20b–25b**. Exceptions included Et_2NH/H_2O (4/1 v/v), 0.1 M K₂CO₃/MeOH and 0.5 M EtSNa/EtOH solutions (entries 3, 8 and 9), in which nucleophilic substitution at the pseudobenzylic carbon was accompanied by deprotection of the 5'-hydroxyl functional group of the sugar moiety. Products **11a/11b**, **16a/16b** and **17a/17b** were afforded in 60–70% yields. In these cases, detectable

Table 2

Nucleophilic substitution of 5'-O-pivaloyl-5-pivaloyloxymethyluridine (19a) and 5'-O-pivaloyl-5-pivaloyloxymethyl-2-thiouridine (19b)



Entry	-Nu	Nu system	Time (h)	Product	Yield ^c (%)	Product	Yield ^c (%)
1	-NH ₂	8 M NH ₃ /EtOH	4	9a	80 ^d	9b	77 ^d
2	-NHMe	8 M MeNH ₂ /EtOH	1	10a	90 ^d	10b	88 ^d
3	-NEt ₂	Et ₂ NH/H ₂ O 4/1 v/v	20	11a	67	11b	65
4	-NC ₄ H ₈ O	Morpholine/H ₂ O 4/1 v/v	20	12a	85 ^d	12b	80 ^d
5	$-NC_5H_{10}$	Piperidine/EtOH 1/1 v/v	20	13a	70 ^d	13b	70 ^d
6	-NHCH ₂ COOH	0.8 M NH ₂ CH ₂ COO ⁻ NBu ₄ /EtOH	20	14a	75 ^d	14b	72 ^d
7	-NH(CH ₂) ₂ SO ₃ H	0.8 M NH ₂ (CH ₂) ₂ SO ₃ ⁻ NBu ⁺ ₄ /EtOH	20	15a	85 ^d	15b	75 ^d
8	-OMe	0.1 M K ₂ CO ₃ /MeOH	1	16a	70	16b	60
9	–SEt	0.5 M EtSNa/EtOH	1	17a	70	17b	70

^a The numbering of the intermediate compounds has been assigned as follows: **20a/20b** (-NH₂), **21a/21b** (-NHMe), **22a/22b** (-NC₄H₈O), **23a/23b** (-NC₅H₁₀), **24a/24b** (-NHCH₂COOH), **25a/25b** (-NHCH₂CO₄H₈O), **23a/25b** (-NHCH₂CO₄

^b 50 °C for the synthesis of **11a**, **11b** (entry 3).

^c Isolated yields.

^d Yield over 2 steps.

amounts of hm^5U/hm^5s^2U (<5%) were also formed. The crude compounds **20a–25a/20b–25b** were then deprotected using 40% aq MeNH₂ (60 °C, 1 h) to give the products of 5'-pivalate ester aminolysis. Compounds **9a–17a/9b–17b** were isolated and purified by preparative RP HPLC with overall yields of 60–90% (ESI). The yields obtained were comparable with those for substitution of Pivom⁵U/Pivom⁵s²U **8a/8b** (Table 1) and both methods can be used interchangeably.

In conclusion, the nucleophilic substitution of 5-pivaloyloxymethyluridine/2-thiouridine (**8a/8b**) was found to be a simple and efficient approach for the preparation of various 5-methyluridines and 5-methyl-2-thiouridines (xm^5U and xm^5s^2U), including natural tRNA components (mnm^5U , mnm^5s^2U cmnm⁵U, cmnm⁵s²U, τm^5U , τm^5s^2U). The remarkable stability of **8a/8b** under acidic and mild basic conditions indicates the possibility for their transformation into suitable phosphoramidites and incorporation into RNA sequences. Pivom⁵U/Pivom⁵s²U-containing oligoribonucleotides can serve as a valuable tool for the synthesis of variously xm^5U/xm^5s^2 U-modified oligomers by post-synthetic nucleophilic substitution of the pivaloyloxy group. The chemical synthesis of RNA oligomers containing Pivom⁵U/Pivom⁵s²U is currently underway.

Acknowledgments

This work was supported by grant 1306/B/H03/2011/40 from the National Science Centre (for G.L.) and grant W-3/FMN/19G/2014 from Young Scientists' Fund at the Faculty of Chemistry, Technical University of Lodz (for K.W.), Poland.

Supplementary data

Supplementary data (general information, detailed synthesis and characterization of the reported compounds) associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.tetlet.2015.10.023.

References and notes

- (a) Machnicka, M. A.; Milanowska, K.; Osman Oglou, O.; Purta, E.; Kurkowska, M.; Olchowik, A.; Januszewski, W.; Kalinowski, S.; Dunin-Horkawicz, S.; Rother, K. M.; Helm, M.; Bujnicki, J. M.; Grosjean, H. *Nucleic Acids Res.* **2013**, *41*, D262; (b) Cantara, W. A.; Crain, P. F.; Rozenski, J.; McCloskey, J. A.; Harris, K. A.; Zhang, X.; Vendeix, F. A.; Fabris, D.; Agris, P. F. *Nucleic Acids Res.* **2011**, *39*, D195.
- (a) Rodriguez-Hernandez, A.; Spears, J. L.; Gaston, K. W.; Limbach, P. A.; Hou, Y. M.; Kaiser, R.; Agris, P. F.; Perona, J. J. *J. Mol. Biol.* **2013**, 425, 3888; (b) Takai, K.; Yokoyama, S. *Nucleic Acids Res.* **2003**, *31*, 6383; (c) Takai, K. *Nucleic Acids Symp. Ser.* **2005**, 49, 317; (d) Armengod, M. E.; Meseguer, S.; Villarroya, M.; Prado, S.; Moukadiri, I.; Ruiz-Partida, R.; Garzon, M. J.; Navarro-Gonzalez, C.; Martinez-Zamora, A. *RNA Biol.* **2015**, *11*, 1495.
- 3. Kurata, S.; Weixlbaumer, A.; Ohtsuki, T.; Shimazaki, T.; Wada, T.; Kirino, Y.; Watanabe, K.; Ramakrishnan, V.; Suzuki, T. J. Biol. Chem. 2008, 283, 18801.
- 4. Suzuki, T.; Nagao, A.; Suzuki, T. WIREs RNA 2011, 2, 376.
- 5. Wang, X.; Yan, Q.; Guam, M.-X. J. Mol. Biol. 2010, 395, 1038.
- Pfaffeneder, T.; Spada, F.; Wagner, M.; Brandmayr, C.; Laube, S. K.; Eisen, D.; Truss, M.; Steinbacher, J.; Hackner, B.; Kotljarova, O.; Schuermann, D.; Michalakis, S.; Kosmatchev, O.; Schiesser, S.; Steigenberger, B.; Raddaoui, N.; Kashiwazaki, G.; Müller, U.; Spruijt, C. G.; Vermeulen, M.; Leonhardt, H.; Schär, P.; Müller, M.; Carell, T. Nat. Chem. Biol. 2014, 10, 574.
- Murphy, F. V., IV; Ramakrishnan, V.; Malkiewicz, A.; Agris, P. F. Nat. Struct. Mol. Biol. 2001, 11, 1186.
- (a) Erwin, V. C.; Songe-Møller, L.; Leihne, V.; Lien, G.; Leszczynska, G.; Malkiewicz, A.; Krokan, H.; Kirpekar, F.; Klungland, A.; Falnes, P. Nat. Commun. 2011, 2, 1; (b) Pomerantz, S. C.; McCloskey, J. A. Methods Enzymol. 1990, 193, 796.
- Dumelin, C. E.; Chen, Y.; Leconte, A. M.; Chen, Y. G.; Liu, D. R. Nat. Chem. Biol. 2012, 8, 913.
- (a) Abdel-Rahman, A. A.-H.; Wada, T. Z. Naturforsch. 2009, 64c, 163; (b) Johar, M.; Manning, T.; Kunimoto, D.; Kumar, R. Bioorg. Med. Chem. 2005, 13, 6663; (c) Rai, D.; Johar, M.; Manning, T.; Agrawal, B.; Kunimoto, D.; Kumar, R. J. Med. Chem. 2005, 48, 7012; (d) Kore, A. R.; Charles, I. Curr. Org. Chem. 2012, 16, 1996; (e) Ashida, N.; Watanabe, Y.; Miura, S.; Kano, F.; Sakata, S.; Yamaguchi, T.; Suzutani, T.; Machida, H. Antiviral Res. 1997, 35, 167; (f) Brulikova, L.; Hlavac, J. Beilstein J. Org. Chem. 2011, 7, 678.
- (a) Bittker, J. A.; Philips, K. J.; Liu, D. R. Curr. Opin. Chem. Biol. 2002, 6, 367; (b) Verma, S.; Jager, S.; Thum, O.; Famulok, M. Chem. Rec. 2003, 3, 51; (c) Hashimoto, H.; Nelson, M. G.; Switzer, Ch. J. Am. Chem. Soc. 1993, 115, 7128.
- 12. Vanikova, Z.; Hocek, M. Angew. Chem., Int. Ed. 2014, 53, 6734.
- 13. Ikeda, K.; Tanaka, S.; Mizuno, Y. Chem. Pharm. Bull. 1975, 23, 2958.
- 14. Malkiewicz, A.; Sochacka, E. *Tetrahedron Lett.* **1983**, *24*, 5359. 15. Leszczynska, G.; Leonczak, P.; Dziergowska, A.; Malkiewicz, A. Nucleosides
- Leszczyńska, G.; Leonczak, P.; Dziergowska, A.; Malkiewicz, A. Nucleosidi Nucleotides Nucleic Acids 2013, 32, 599.
- Ogata, T.; Shumazaki, T.; Umemoto, T.; Kurata, S.; Ohtsuki, T.; Suzuki, T.; Wada, T. J. Org. Chem. 2009, 74, 2585.

- 17. Seio, K.; Wada, T.; Sakamoto, K.; Yokoyama, S.; Sekine, M. J. Org. Chem. 1998, 63, 1429.
- 18. Reese, C. B.; Sanghvi, Y. S. J. Chem. Soc., Chem. Commun. 1984, 62.
- Sekine, M.; Peshakova, L. S.; Hata, T.; Yokoyama, S.; Miyazawal, T. J. Org. Chem. 19. **1987**, *52*, 5061.
- Wada, T.; Shimazaki, T.; Nakagawa, S.; Ohtsuki, T.; Kurata, S.; Suzuki, T.; Watanabe, K.; Saigo, K. *Nucleic Acids Symp. Ser.* 2002, *2*, 11.
 (a) Ogata, T.; Wada, T. *Nucleic Acids Symp. Ser.* 2006, *50*; (b) Leszczynska, G.; Leonczak, P.; Wozniak, K.; Malkiewicz, A. *RNA* 2014, *20*, 938.
- 22. (a) http://www.glenresearch.com/GlenReports/GR23-27.html.; (b) Dai, Q.; Song, Ch.-X.; Pan, T.; He, Ch. J. Org. Chem. 2011, 76, 4182; (c) de Kort, M.; de Visser, P. C.; Kurzeck, J.; Meeuwenoord, N. J.; van der Marel, G. A.; Rüger, W.; van Boom, J. H. Eur. J. Org. Chem. 2001, 2001, 2075.
- Scheit, K. H. Chem. Ber. 1966, 99, 3884.
 Sowers, L. C.; Beardsley, G. P. J. Org. Chem. 1993, 58, 1664.
 Jiang, L.; Chan, T.-H. J. Org. Chem. 1998, 63, 6035.