

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 14 (2004) 4539-4544

Evaluation of pyrrolidin-2-imines and 1,3-thiazolidin-2-imines as inhibitors of nitric oxide synthase $\stackrel{\diamond}{\sim}$

K. Shankaran,^{a,*} Karla L. Donnelly,^a Shrenik K. Shah,^a Ravindra N. Guthikonda,^a Malcolm MacCoss,^a John L. Humes,^b Stephen G. Pacholok,^b Stephan K. Grant,^c T. M. Kelly^c and K. K. Wong^c

^aDepartment of Medicinal Chemistry, Merck Research Laboratory, PO Box 2000, Rahway, NJ 07065, USA ^bDepartment of Immunology and Inflammations, Merck Research Laboratory, PO Box 2000, Rahway, NJ 07065, USA ^cDepartment of Enzymology, Merck Research Laboratory, PO Box 2000, Rahway, NJ 07065, USA

> Received 23 April 2004; revised 10 June 2004; accepted 10 June 2004 Available online 19 July 2004

Abstract—Syntheses and evaluation of pyrrolidin-2-imines and 1,3-thiazolidin-2-imines as inhibitors of nitric oxide synthase (NOS) are discussed. An extensive SAR was established for pyrrolidin-2-imines class of compounds. The amidines came out as the most potent inhibitors in addition to displaying selectivity.

© 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Nitric oxide synthases (NOS) are a group of oxidative enzymes, which produce nitric oxide (NO) from L-arginine.² These enzymes bring about a five electron oxidation of L-arginine to nitric oxide and L-citrulline as shown below.



To date three isoform of NOS enzymes have been discovered. Of these, the two constitutive versions namely, neuronal (nNOS or NOS1) and endothelial (eNOS or NOS3) isoform are involved in neurotransmission, and vasodilatation, respectively.³ The inducible form of nitric oxide synthase (iNOS or NOS2) is concerned with host defense, septic shock, and inflammatory diseases.⁴ Thus, selective inhibition of iNOS over the other isoforms may offer a new therapy for these diseases. Pursuit of NOS inhibitors has lead to amino acid analogs⁵ as well as nonamino acid based inhibitors including aminoguanidines,⁶ isothioureas,⁷ and amidines,⁸ benzoxazolones,⁹ and pyridines.¹⁰ A recent communication^{11a} disclosed that the analogs of 1,3-oxazolidin-2-imine derivatives (**2**) were shown to be inhibitors of the NOS enzymes prompted us to communicate our findings in these area.

We had previously reported that piperidin-2-imines (1) are potent inhibitors¹² of NOS enzymes displaying varying degree of selectivity. In parallel efforts we¹ and others studied pyrrolidin-2-imines as NOS inhibitors and toward this end Hagen and co-workers has already disclosed their results.^{11b} In addition to pyrrolidin-2-imines we also probed the related thiazolidin-2-imines and it was hoped that these classes (3) might provide selective iNOS inhibitors. The results of such a study are disclosed herein. While work related to the 1,3-thiazo-lidin-2-imines lead class was minimal in scope, a detailed structure-activity relationship was established for the pyrrolidin-2-imines.

Keywords: NOS; Pyrrolidin-2-imine; 1,3-Thiazolidin-2-imine. * See Ref. 1.

^{*} Corresponding author. Tel.: +1-732-594-3979; fax: +1-732-594-3007; e-mail: kothandaraman_shankaran@merck.com

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter © 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2004.06.033



2. Chemistry

Scheme 1 shows the preparation of mono- and disubstituted pyrrolidin-2-imines essentially in three steps. The first two steps consists of Michael addition of various nitroalkanes to the acrylates 4 followed by the catalytic hydrogenation of the intermediate nitroesters to give the lactams 5.13 The lactams 5 were transformed to the desired amidines following one of the several conditions shown below. Method A, which involved the conversion of 5 to the iminoether 6 followed by reflux with ammonium chloride in ethanol to furnish 8, as a mixture of stereoisomers was the first choice. In some incidences, the iminoether 6 was too volatile to be isolated and for these systems method B was preferred. This required the initial conversion of the amides 5 to the thioamides 7 by Lawesson's reagent.¹⁴ The thioamides 7 were then quaternized with excess iodomethane followed by reaction with methanolic ammonia to afford the amidines 8 as a hydroiodide salt. Alternatively, the thioamides 7 could be directly converted to the amidines 8 in a procedure utilizing mercuric chloride and gaseous ammonia as reported in the literature.¹⁵

The preparation of fused amidine 11 from the known¹⁶ iodolactam 9 is shown in Scheme 2. The reduction of 9 to 10 was efficiently accomplished under tinhydride conditions. The transformation of 10 to the amidine 11 was analogous to the transformation 5 to 8 (method B) shown in Scheme 1.



Scheme 1. Reagents and conditions: (a) $R_3CH_2NO_2$, DBU, MeOH; (b) PtO₂, H₂, EtOH; (c) *Method A* (a) Me₃OBF₄, CH₂Cl₂, rt; (b) NH₄Cl, EtOH, reflux; *Method B* (a) Lawesson's reagent, toluene, 90 °C; (b) MeI, rt; (c) NH₃, MeOH; (d) NH₃, HgCl₂, THF, 60 °C.



Scheme 2. Reagents and conditions: (a) *n*-Bu₃SnH, AIBN, PhH reflux; (b) *Method B*, steps (a) and (d) (Scheme 1).



Scheme 3. Reagents and conditions: (a) Oxalyl chloride, DMSO, DCM, NEt₃; (b) Ph₃P=CHCOOMe, DCM, rt; (c) Pd/C, H₂, MeOH; (d) HCl and then K_2CO_3 ; (e) *Method B*, steps (a), (b), and (c) (Scheme 1).



Scheme 4. Reagents and conditions: (a) KOH, CS₂, EtOH reflux; (b) *Method A*, steps (a) and (b) (Scheme 1).



Scheme 5. Reagents and conditions: (a) $SOCl_2$, $CHCl_3$, reflux; (b) NEt_3 , CS_2 , CH_2Cl_2 ; (c) *Method B*, steps (b) and (c) (Scheme 1).

The fused amidine 15 was prepared from *N*-Boc protected aminol 12 as shown in Scheme 3. In a sequence of reaction that involved Swern oxidation of 12 followed by the Wittig–Horner reaction gave a mixtures of esters 13. Catalytic reduction of 13 followed by *N*-Boc removal led to a rapid cyclization affording 14. The amide 14 was transformed to 15 by the method B as in Scheme 1. Likewise, starting from the corresponding piperidinol and following the same procedure for 15 gave 16 (n = 2).

A general synthesis of 1,3-thiazolidin-2-imines is shown in Scheme 4. This involved the reaction of aminol 17 and carbon disulfide under basic conditions¹⁷ to afford 18. The thiazolidine-2-thiones 18 was transformed to 1,3-thiazolidin-2-imines 19 in two steps that involved initial reaction with Meerwein salt (to afford thioiminoether) that was followed by refluxing with ammonium chloride (Scheme 1, method A).

The bicyclic 1,3-thiozol-3-imine analogs 22 and 23 were accessed by a similar approach shown above for 19, but with a slight variation as shown in Scheme 5. A two step reaction that involved the transformation of aminol 20 to the intermediate chloro compound followed by reaction with CS_2 /triethylamine gave 21. The thione 21 was transformed to 22 by method B. Similarly, starting from 2-piperidinemethanol and following the aforesaid steps gave 23.

3. Results and discussion

The hydrochloride/hydroiodide salts of pyrrolidine-2imines and 1,3-thiazolidin-2-imines were evaluated for NOS activity by an assay protocol18 that used recombinant human version of all three isoform of NOS. The binding results for both the pyrrolidin-2-imines and 1,3-thiazolidin-2-imines classes are compared with the standard (L)-N-iminoethyl-lysine (L-NIL). The SAR studies on the pyrrolidin-2-imines analogs 8 centered on the placement of alkyl substituents at all the available positions of pyrrolidine core. The results from such a study, compared to the parent amidine (8a) are displayed in Table 1. The analog (8c) with the methyl group at C-4 of the pyrrolidine ring lead to significant improvement in the binding potency for the iNOS (20folds) and nNOS (12-folds) as compared to 8a. Compounds 8b and 8d bearing the methyl at C-3 and C-5 showed a modest 2- to 3-folds improvement in potency for iNOS, however these compounds (8b-d) displayed no selectivity. In probing other alkyl residues at C-4, it became clear that the amidine analog 8e (with ethyl at C-4) led to somewhat less potent compound compared to 8c. More importantly, 8e displayed better selectivity for iNOS (over eNOS) while the selectivity over nNOS was not as good. The gain in potency for 8e led to the preparation of other analogs 8f-h. The *n*-propyl analog 8f had lost significant activity compared to either 8c or 8e. This was also true for 8g but it still maintained good

selectivity for iNOS over eNOS. Finally, the compound with phenyl at C-4 (8h) was inactive.

An observation (from Table 1) that the amidine **8e** was not only potent (iNOS) but also displayed modest selectivity encouraged us to prepare analogs of **8e** with additional substituents on pyrrolidine core to improve its profile further. The results of binding data from such a study compared with **8e** are shown in Table 2.

The placement of alkyl substituents at C-3 and C-4 leading to **8i** resulted in an analog that was less potent for iNOS but surprisingly was more potent for nNOS. This was also reflected to some extent in **8j**, whose selectivity for iNOS over eNOS was very good. Despite slight loss of potency, the selectivity displayed by **8j** (eNOS/iNOS) was the best we have seen for these analogs. The C-3/C-5 disubstituted compound **8k** had lost significant binding for the iNOS as a result this substitution pattern was not further pursued. The C-4/C-5 substituted compounds were explored in greater depth (**8l–p**). The data for **8l,m**, and **8o** clearly indicates that small alkyl substituents at C-4 or C-5 were the most preferred as it lead to potent analogs (compared to **8e**) in addition to maintaining good selectivity. The amidines

Table 1. Inhibition of NOS by monosubstituted pyrrolidin-2-imines $(IC_{50} = \mu M)^a$

Entry	Structure ^b	iNOS	eNOS	Selectivity eNOS/iNOS	nNOS	Selectivity nNOS/iNOS
_	l-NIL	2.1	7.9	3.7	17.5	8.3
8a		4.1	4.6	1.1	9.6	2.3
8b	NH H	1.5	1.8	1.2	0.9	0.6
8c	NH H	0.2	2.6	13	0.8	0.3
8d	NH H	1.3	1.9	1.4	3.1	2.3
8e	NH H	0.5	14.5	27	2.6	4.8
8f	N NH	38.7	53	1.4	53	1.4
8g	NH H	3.5	87.8	25	10.6	3
8h	Ph NH H	>50	>50	ND	>50	ND

ND = not determined.

 a IC₅₀s are obtained from 10 point titration using sigma plot, where each individual point is an average of duplicate determination at that concentration.

^bThe pyrrolidin-2-imines were synthesized according to Scheme 1 and the IC₅₀s are reported for racemic mixture.

Table	2.	Inhibition	of NOS	by	disubstituted	pyrrolidin-2-imines	$(IC_{50} = \mu M)^a$
-------	----	------------	--------	----	---------------	---------------------	-----------------------

Entry	Structure ^b	iNOS	eNOS	Selectivity eNOS/iNOS	nNOS	Selectivity nNOS/iNOS
8e	NH H	0.5	14.5	27	2.6	4.8
8i	NH H	1.2	8.7	7.2	0.61	0.5
8j	NH H	0.93	60.6	65	0.61	0.65
8k	NH H	23	5.5	0.24	2.4	0.1
81	NH H	0.3	0.6	17	0.13	3.8
8m	NH H	0.02	0.52	26	0.08	4.0
8n	N NH	2	49	24	4.8	2.4
80	NH H	0.16	6.3	38	0.14	0.8
8p	NH H	13.1	104	8	27	2
8q	NH H	10.2	227	22	17.3	1.7

 a IC₅₀s are obtained from 10 point titration using sigma plot, where each individual point is an average of duplicate determination at that concentration.

^b The pyrrolidin-2-imines were synthesized according to Scheme 1 and the IC₅₀s are reported for the diastereomeric and racemic mixture.

8n,p, and **8q** with *n*-propyl group at both C-5 and C-4 led to significant loss in iNOS activity. This result was akin to the one seen for **8f** (Table 1) as discussed previously. It is also worth noticing that both **8n** and **8q** still retained good selectivity for iNOS over eNOS despite loss of potency.

While we had achieved significant potency going from **8e** (Table 1) to **8m** (Table 2) the selectivity remained unchanged. We felt that perhaps the preparation of rigid analogs might overcome the mediocre selectivity (over nNOS) seen for **8m**. The results of binding for the select fused amidines are displayed in Table 3. In general the fused amidines displayed binding activity that was drastically less for the all three isoforms and more so for the iNOS compared to **8m**. In a dramatic reversal, all fused analogs consistently maintained better binding potency for the nNOS over iNOS. Thus, the amidine analog **11** (fused version of **8m**) was practically inactive and fared much worse than we had hoped. Similar

behavior was noticed for 15 and 16. The bicyclic analogs 17 and 18 were considerably more potent for nNOS while the potency for the iNOS was unimpressive.

While we had modest success in the pyrrolidin-2-imines lead class, the results of the binding assay for the 1,3-thiazolidin-2-imines analogs as shown below (Table 4) were far from satisfactory and stand in sharp contrast with the results of pyrrolidin-2-imines discussed above.

To start with, the lead compound **19a** was equipotent in all three isoform, and showed lack of selectivity. Partial attempts to improve the potency and selectivity specifically for the iNOS enzyme suggests that this lead class was quite sensitive to the substituents on the thiazole ring as exemplified by the analogs **19b,c**, and **19d**. All three analogs had lost significant activity in binding assay compared to **19a**. Notice also that the fused analog **22** and **23** were less impressive and share the trait similar to **15** and **16** (Table 3). But for the fused analogs,

Table 3. Inhibition of NOS by fused pyrrolidin-2-imines $(IC_{50} = \mu M)^a$

Entry	Structure ^b	iNOS	eNOS	Selectivity eNOS/iNOS	nNOS	Selectivity nNOS/iNOS
8m	NH H	0.02	0.52	26	0.08	4.0
11	NH H	>50	>50	ND	9.9	ND
15	NH NH	>50	33	ND	9.6	ND
16	NH NH	>50	>50	ND	>50	ND
17	NH H	3.1	4.7	1.5	0.31	0.1
18	NH H	1.2	24	20	0.66	0.55

ND = not determined.

 ${}^{a}IC_{50}s$ are obtained from 10 point titration using sigma plot, where each individual point is an average of duplicate determination at that concentration.

^b The amidine 11 was synthesized according to Scheme 2, 15 and 16 were prepared according to Scheme 3 and 17 and 18 by Scheme 1. The IC₅₀ s are reported for the racemic mixture.

Table	4.	Inhibition	of	NOS b	οv	disubstituted	1.	3-thiazol	idin	-2-i	mines	$(IC_{50} =$	μM) ^a
-------	----	------------	----	-------	----	---------------	----	-----------	------	------	-------	--------------	------------------

Entry	Structure ^b	iNOS	eNOS	Selectivity eNOS/iNOS	nNOS	Selectivity nNOS/iNOS	
	l-NIL	2.1	7.9	3.7	17.5	8.3	
19a	S NH H	0.9	1.1	1.2	1.8	2	
19b	S NH H	>50	>50	ND	>50	ND	
19c	NH NH	5.2	>50	ND	13.2	2.5	
19d	Ph NH	>50	>50	ND	>50	ND	
22		>50	>50	ND	14	ND	
23		>50	>50	ND	>50	ND	

ND = not determined.

 a IC₅₀s are obtained from 10 point titration using sigma plot, where each individual point is an average of duplicate determination at that concentration.

^b The 1,3-thiazolidin-2-imines were synthesized according to Schemes 4 and 5 and the IC₅₀s are reported for the racemic mixture.

the pyrrolidin-2-imines, which were investigated in parallel with 1,3-thiazolidin-2-imines had a more

encouraging outcome. For these reasons 1,3-thiazolidin-2-imine as a lead class was not pursued further. In conclusion, the synthesis and evaluation of pyrrolidin-2-imines and 1,3-thiazolidin-2-imines as iNOS inhibitor have been demonstrated. Among the pyrrolidin-2-imines, **81** and **8m** displayed superb potency for iNOS and was the best with in this class. Our limited efforts in the 1,3-thiazolidin-2-imines suggest that they were far less significant both in potency and selectivity compared to the pyrrolidin-2-imines or for that matter 1,3-oxazolidin-2-imines class of compounds that was reported recently.^{11a}

In the final analysis, it has been our experience that, in general, the pyrrolidin-2-imines lead class was less potent and considerably less selective compared to the piperidin-2-imines as iNOS inhibitors.¹²

References and notes

- The part of results of the work described herein was presented at the 214th ACS National Meeting, Las Vegas, NV, 1997 (*Abstract* #037).
- Several excellent reviews have appeared on this subject, see: (a) Kerwin, J. F.; Lancaster, J. R.; Feldman, P. L. J. Med. Chem. 1995, 38, 4343; (b) Marletta, M. A. Cell 1994, 78, 927; (c) Feldman, P. L.; Griffith, O. W.; Stuehr, D. J. Chem. Eng. News 1993, 26; (d) Snyder, S. H.; Bredt, D. S. Sci. Am. 1992; (May), 68.
- 3. Moncada, S.; Higgs, A. N. Engl. J. Med. 1993, 329, 2002.
- 4. White, K. A.; Marletta, M. A. Biochemistry 1992, 31, 6627.
- (a) Cochran, F. R.; Selph, J.; Sherman, P. Med. Chem. Res. 1996, 16, 547; (b) Nussler, A. K.; Billiar, T. A. J. Leukoc. Biol. 1993, 54, 171.
- (a) Corbett, J. A.; Tilton, R. G.; Chang, K.; Hasan, K. S.; Ido, Y.; Wang, J. L.; Sweetland, M. A.; Lancaster, J. R.; Willamson, J. R.; McDaniel, M. L. *Diabetes* 1992, *41*, 552; (b) Joly, G. A.; Ayres, M.; Chelly, F.; Kilbourn, R. G. *Biochem. Biophys. Res. Commun.* 1994, *199*, 147.
- (a) Nakane, M.; Klinghofer, V.; Kuk, J. E.; Donnelly, J. L.; Budzik, G. P.; Pollock, J. S.; Basha, F.; Carter, G. E. *Mol. Pharmacol.* **1995**, *47*, 831; (b) Garvey, E. P.; Oplinger, J. A.; Tanoury, G. J.; Sherman, P. A.; Fowler,

M.; Marshall, S.; Harmon, M. F.; Paith, J. E.; Fufine, E. S. J. Biol. Chem. **1994**, 269, 26669.

- Moore, W. M.; Webber, K. R.; Fok, K. F.; Jerome, G. M.; Conner, J. R.; Manning, P. T.; Wyatt, P. S.; Misko, T. P.; Tjoeng, F. S.; Misko, T. P.; Currie, M. G. *J. Med. Chem.* **1996**, *39*, 3886.
- Shankaran, K.; Donnelly, K. L.; Shah, S. K.; Humes, J. L.; Pacholok, S.; Green, B. G.; Grant, S. K.; MacCoss, M. Bioorg. Med. Chem. Lett. 1997, 7, 2887.
- Hagmann, W. K.; Caldwell, C. G.; Chen, P.; Durette, P.; Esser, C. K.; Lanza, T. J.; Kopka, I. E.; Guthikonda, R.; Shah, S. K.; Green, B. G.; Humes, J. L.; Kelly, T. M.; Luell, S.; Meurer, R.; Moore, V.; Pacholok, S. G.; Pavia, T.; Williams, H. R.; Wong, K. K. *Bioorg. Med. Chem. Lett.* 2000, 10, 1975.
- (a) Ueda, S.; Terauchi, H.; Yano, A.; Ido, M.; Matsumoto, M.; Kawasaki, M. *Bioorg. Med. Chem. Lett.* 2004, 14, 313; (b) Hagen, T. J.; Bergmanis, A. A.; Kramer, S. W.; Fok, K. F.; Schmelzer, A. E.; Pitzele, B. S.; Swenton, L.; Jerome, G. M.; Kornmeier, C. M.; Moore, W. M.; Branson, L. F.; Conner, J. R.; Manning, P. T.; Currie, M. G.; Hallinan, E. A. J. Med. Chem. 1998, 41, 3675.
- 12. Guthikonda, R. N. *Abstract #90*, National Medicinal Chemistry Symposium, Ann Arbor, MI, 1996.
- 13. Moffett, R. B. Org. Synth. 1963, 4, 357.
- 14. Scheibye, S.; Pedersen, B. S.; Lawesson, S.-O. Bull. Soc. Chim. Belg. 1978, 87, 229.
- 15. Foloppe, M. P.; Rault, S.; Robba, M. Tetrahedron Lett. **1992**, *33*, 2803.
- 16. Knapp, S.; Gibson, F. S. Org. Synth. 1991, 70, 101.
- Nagao, Y.; Hagiwara, Y.; Kumagai, T.; Ochiai, M.; Inoue, T.; Hasimoto, K.; Fujita, E. J. Org. Chem. 1986, 51, 2391.
- 18. The NOS inhibitory activity of the compound was determined by comparing the conversion of 3 H-L-arginine to 3 H-citrulline in the presence of inhibitor with control. The assay mixture containing 1 μ M of 3 H-L-arginine, cofactors and the inhibitor or aqueous DMSO (control) was incubated for 30 min at room temperature. The reaction was quenched by adding a slurry of Dowex 50W-K8 resin to complex and remove the unreacted substrate. The concentration of the 3 H-L-citrulline product in the supernatant fluid was determined on a scintillation counter. Under the assay conditions the production of L-citrulline was linear with time for the duration of the experiment.