

Efficient Synthesis of Functionalized Unsymmetrical Dialkyl Trisulfanes

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Abstract: We have developed a convenient method for the synthesis of functionalized unsymmetrical dialkyl trisulfanes under mild conditions in very good yields. The designed method is based on the reaction of (5,5-dimethyl-2-thioxo-1,3,2-dioxaphosphorinan-2-yl)-disulfanyl derivatives with alkyl disulfanyl anions generated in situ from *S*-acetyl disulfanyl derivatives and sodium methoxide. The developed method allows for the preparation of unsymmetrical trisulfanes bearing additional hydroxyl, carboxyl, or amino functionalities on both sides of the trisulfane functionality.

Key words: unsymmetrical trisulfanes, thiols, hydrodisulfanes, *S*-acetyl alkyl disulfanes, phosphorodithioic acid

Our interest in the chemistry of diorganyl trisulfanes (trisulfides) arises from their diverse roles in living organisms and their occurrence in natural sources.^{1,2} In the literature, these compounds are often termed as organic trisulfides, but the IUPAC recommended nomenclature is trisulfanes.³ The name trisulfide should only be applied to ionic compounds, such as Na_2S_3 . Organic trisulfanes have been isolated from shiitake mushrooms,⁴ oil made from *erula asafetida*,⁵ durian fruit,⁶ garlic oil,^{7,8} and Hawaiian algae.⁹ It also should be mentioned that dialanyltrisulfane has been detected in acidic wool hydrolysates, but it is not clear whether the related amino acid ($\text{HOOCCH}(\text{NH}_2)\text{CH}_2\text{S}_3$) is a part of the wool structure or is formed from cysteine during hydrolysis.¹⁰ In the biochemical literature, dialanyltrisulfane is often incorrectly termed as ‘cysteine trisulfide’. A natural peptide containing a trisulfane group in place of a disulfane bridge has been isolated from genetically engineered *Escherichia coli* bacteria.¹¹ It is a derivative of the human growth hormone consisting of 191 amino acids in a single chain with a trisulfane bridge between cysteine (alanyl) residues 182 and 189.¹² A trisulfanyl functionality has also been observed in recombinant DNA-derived methionyl human growth hormone in the bridge between cysteine residues 53 and 165.¹³ Calicheamicin^{14,15} and esperamicins A_1 and A_2 ,^{16,17} members of the enediyne class of antibiotics, also contain the trisulfanyl functionality. These natural products are very potent antitumor antibiotics.

There are numerous reactions that can be used to prepare symmetrical organic trisulfanes. The most common methods include the reaction of thiols with sulfur dichloride,¹⁸ the coupling of alkyl halides with sodium trisulfide,¹⁹ and the reaction of thiols or disulfanes with sulfur.²⁰ Thio-

alkylation reactions of various thiosulfenate species can also produce trisulfanes. The most suitable substrates include Bunte salts,²¹ metal sulfides,²² and thiosulfonyl chloride.²³ The latter can also be used for the preparation of unsymmetrical trisulfanes. Other practical procedures involve the reduction of thiosulfonates and disulfonyl sulfides with phosphines,²⁴ sulfur insertion reactions into thiosulfonates, thiosulfonates,²⁵ and disulfanes,²⁶ alkoxide decomposition of sulfenylthiocarbonates,²⁷ and reactions of thiols with 1,1-thiobis(benzimidazole)²⁸ or diimidazolylsulfide.²⁹

Preparative methods that are efficient for the preparation of symmetrical trisulfanes are very often ineffective for the preparation of unsymmetrical compounds. Indeed, the synthesis of unsymmetrical trisulfanes is more complex. There are known procedures based on the coupling of chlorodisulfanes with *N*-arylamidodithiosulfites³⁰ or thiols^{31,32} or the sequential coupling of two thiols using sulfur dichloride.³³ Other procedures involve the desulfurization of unsymmetrical dialkanesulfonic thioanhydrides,²⁴ or the use of (often) unstable hydrodisulfanes.³⁴

Moderate yields and/or the formation of undesirable polysulfane side products are the major drawbacks of the methods presented above. The likelihood that pure trisulfanes can be separated from the reaction mixture is very poor. The best purification method is crystallization, but clearly this can only be applied to solid products. Moreover, the scopes of the presented methods are limited by the availability of reagents and the chemical reactivity of the additional functional groups.

We have previously demonstrated the preparation of functionalized unsymmetrical molecules such as dialkyl disulfanes, alkyl-aryl disulfanes,³⁵ ‘bioresistant’ disulfanes,³⁶ unsymmetrical disulfanes of L-cysteine and L-cystine,³⁷ and diaryl disulfanes³⁸ based on the readily available 5,5-dimethyl-2-thioxo-1,3,2-dioxaphosphorinane-2-disulfanyl derivatives **1**. These disulfanyl derivatives **1** of phosphorodithioic acid were also convenient for the preparation of α -sulfenylated carbonyl compounds³⁹ and symmetrical trisulfanes.⁴⁰

The limitations of our previous method for the preparation of unsymmetrical trisulfanes⁴¹ have encouraged us to develop a new synthetic strategy for the preparation of unsymmetrical trisulfanes bearing additional functionalities on either side of the trisulfane. The idea is based on the reaction of electrophilic disulfanyl derivatives of phospho-

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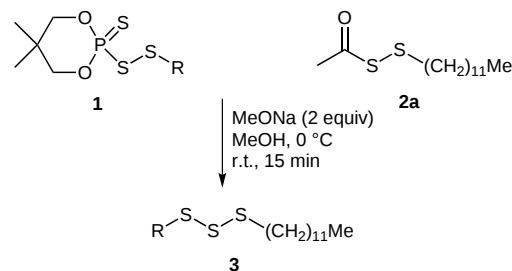
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rodithioic acid **1** with a nucleophilic dodecylsulfanyl anion generated in situ from *S*-acetyl dodecylsulfane (**2a**) and sodium methoxide (Table 1).

Table 1 Formation of Unsymmetrical Trisulfanes **3** from *S*-Acetyl Dodecylsulfane (**2a**)

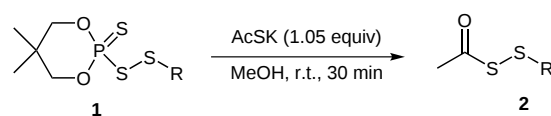


Entry	R	3	Yield (%)
1	(CH ₂) ₁₁ OH	3a	99
2	(CH ₂) ₁₀ CO ₂ H	3b	99
3	(CH ₂) ₁₁ NHBoc	3c	96
4	(CH ₂) ₁₀ CO ₂ Me	3d	75
5	Ph	3e	63

It is known that the preparation of polysulfanes is frequently hampered by the formation of their homologues with a variable number of sulfur atoms.³² The purification of such a mixture is, at best, difficult and, in most cases, impossible due to the similar properties of the products. We were able to avoid the formation of side products by using a small excess of the disulfanyl derivatives **1** (1.05 equiv). In this case, the potentially unstable dodecylsulfanyl anion was consumed very quickly and sulfur extrusion or exchange with product **3** was not observed.

The *S*-acetyl alkylsulfanes **2** are readily available from disulfanyl derivatives **1** and potassium thioacetate (Table 2).

Table 2 Preparation of *S*-Acetyl Alkylsulfanes **2**



Entry	R	2	Yield (%)
1	(CH ₂) ₁₁ Me	2a	99
2	(CH ₂) ₁₁ OH	2b	80
3	(CH ₂) ₁₀ CO ₂ H	2c	87
4	(CH ₂) ₁₁ NHBoc	2d	91

We selected disulfanyl derivatives **1** and *S*-acetyl alkylsulfanes **2** bearing hydroxy, carboxy, methyl ester, *tert*-butoxycarbonylamino, and phenyl functionalities to examine the scope and limitations of this method for the preparation of functionalized unsymmetrical trisulfanes **3**. All of the above alkyl derivatives afforded, after purification, unsymmetrical trisulfanes **3** in very good yield (Table 3).

As shown in Table 3, the same unsymmetrical trisulfane **3** can be obtained in two different ways. For example, **3b** can be prepared from **1c** and **2a** or from **1a** and **2b**. Both approaches gave product **3b** in very good yield (Table 3). However, the reaction of disulfanyl derivative **1f** (R¹ = Ph) with *S*-acetyl alkylsulfanes **2** was successful only for compound **2a**. Unfortunately, the other *S*-acetyl alkylsulfanes **2b–d** provided products **3i**, **3l**, and **3n** that were contaminated with symmetrical and unsymmetrical di- and tetrasulfanes, respectively. Although di-, tri-, and tetrasulfanes could not be separated by column chromatography, the presence of these impurities can be easily confirmed by ¹H NMR spectroscopy. The chemical shift of the methylene group (triplet in CDCl₃) adjacent to the sulfur for di-, tri-, and tetrasulfanes are 2.66, 2.87, and 3.05, respectively. The versatility of the method is limited

Table 3 Synthesis of Unsymmetrical Trisulfanes **3**^a

	R ¹ (CH ₂) ₁₁ Me 1a	R ¹ (CH ₂) ₁₁ OH 1b	R ¹ (CH ₂) ₁₀ CO ₂ H 1c	R ¹ (CH ₂) ₁₁ NHBoc 1d	R ¹ (CH ₂) ₁₀ CO ₂ Me 1e	R ¹ Ph 1f
R ² (CH ₂) ₁₁ Me 2a	–	3a (99)	3b (99)	3c (96)	3d (75)	3e (63)
R ² (CH ₂) ₁₁ OH 2b	3a (71)	–	3f (72)	3g (78)	3h (60)	3i (0)
R ² (CH ₂) ₁₀ CO ₂ H 2c	3b (85)	3f (70)	–	3j (73)	3k (62)	3l (0)
R ² (CH ₂) ₁₁ NHBoc 2d	3c (87)	3g (76)	3j (74)	–	3m (77)	3n (0)

^a The yields (%) of the isolated product are reported in parentheses.

by very fast sulfur extrusions and exchange reactions in the case of aryl alkyl trisulfanes **3i**, **3l**, and **3n**. It seems that aromatic groups can promote these side reactions because the arylthiolate anion is a very good leaving group.

In conclusion, a convenient method for the preparation of unsymmetrical dialkyl trisulfanes **3**, bearing hydroxy, carboxy, methyl ester, or *tert*-butoxycarbonylamino groups on either or both sides of the trisulfane has been developed. Reactions of **1** with a variety of **2** in the presence of sodium methoxide in methanol at room temperature were generally complete within 30 minutes and gave exclusively unsymmetrical dialkyl trisulfanes **3** in very good yields after purification. Because the reactions of the *S*-acetyl alkyl disulfanes **2** proceeded with a small excess of **1** under mild reaction conditions and in a short time, thiol–trisulfane exchange and sulfur extrusion did not occur during the reaction. The simplicity and very good yields of this method make it one of the most attractive approaches for the preparation of functionalized unsymmetrical dialkyl trisulfanes.

Typical Procedure for Trisulfane Preparation

A solution of NaOMe (2.0 mmol) in dry MeOH (2 mL) was added to a solution of **1** (1.05 mmol) and **2** (1.0 mmol) in dry MeOH (20 mL) at 0 °C under N₂ atmosphere. Then, the ice bath was removed, and the mixture was stirred for 30 min at r.t. After evaporation of the solvent under reduced pressure, the residue was purified by column chromatography. The addition of small amount of AcOH prior to purification on a silica gel column inhibited sulfur extrusion during separation. The yields are reported in Table 3.

Representative Analytical Data

11-(11-Hydroxyundec-1-yl-trisulfanyl)undecanoic Acid (**3f**)

Chromatography (CH₂Cl₂–EtOAc, 10:1); yield: 0.326 g (72%); white solid; mp 49–52 °C. IR (KBr): ν = 3345 (w, OH), 2918 (s), 2848 (m, CH), 1710 (m, C=O), 1472 (m, SCH₂), 1036 (m, OCH₂), 716 (m, CH₂) cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ = 1.25–1.48 (m, 28 H, CH₂), 1.53–1.77 (m, 6 H, CH₂), 2.34 [t, J = 7.3 Hz, 2 H, C(O)CH₂], 2.87 (t, J = 7.3 Hz, 4 H, SCH₂), 3.18 (br s, 2 H, OH, COOH), 3.64 (t, J = 6.5 Hz, 2 H, OCH₂). ¹³C NMR (50 MHz, CDCl₃): δ = 178.5, 62.9, 38.9, 33.9, 32.7, 29.7, 29.6, 29.5, 29.4, 29.3, 29.1, 29.0, 28.9, 28.8, 28.6, 28.5, 28.2, 25.7, 24.7; signals: 22 expected; 19 observed. ESI-HRMS: m/z [M + Na]⁺ calcd for C₂₂H₄₄NaO₃S₃: 475.2350; found: 475.2344.

N-(*tert*-Butoxycarbonyl)-11-(11-hydroxyundec-1-yl-trisulfanyl)-1-undecylamine (**3g**)

Chromatography (CH₂Cl₂–EtOAc, 10:1); yield: 0.420 g (78%); colorless oil. IR (KBr): ν = 3381 (m, NH), 3345 (w, OH), 2920 (s), 2850 (s, CH), 1685 (s, C=O), 1637 (m, NH), 1170 (m, CN), 720 (w, CH₂) cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ = 1.20–1.45 (m, 33 H, CH₂, OH), 1.46 (s, 9 H, *t*-Bu), 1.65–1.77 (m, 4 H, SCCH₂), 2.88 (t, J = 7.3 Hz, 4 H, SCH₂), 3.00–3.20 (m, 2 H, NCH₂), 3.64 (t, J = 6.5 Hz, 2 H, OCH₂), 4.48 (br s, 1 H, NH). ¹³C NMR (50 MHz, CDCl₃): δ = 155.8, 78.8, 62.9, 40.6, 39.1, 38.8, 31.8, 30.2, 30.1, 29.7, 29.5, 29.4, 29.3, 29.2, 29.1, 29.0, 28.8, 28.5, 28.4, 26.8, 22.7; signals: 25 expected; 21 observed. ESI-HRMS: m/z [M + Na]⁺ calcd for C₂₇H₅₅NNaO₃S₃: 560.3242; found: 560.3245.

11-(11-Hydroxyundec-1-yl-trisulfanyl)undecanoic Acid Methyl Ester (**3h**)

Chromatography (PE–CH₂Cl₂ = 2:1); yield: 0.280 g (60%); yellow solid; mp 26–28 °C. IR (KBr): ν = 3378 (w, OH), 2922 (s), 2852 (m, CH), 1743 (m, C=O), 1472 (m, SCH₂), 1036 (m, OCH₂), 720

(m, CH₂) cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ = 1.02–1.77 (m, 35 H, CH₂, OH), 2.22–2.34 (m, 2 H, CH₂), 2.87 (t, J = 7.3 Hz, 4 H, SCH₂), 3.64 (t, J = 6.5 Hz, 2 H, OCH₂), 3.67 (s, 3 H, COOCH₃). ¹³C NMR (50 MHz, CDCl₃): δ = 174.7, 63.0, 51.4, 39.4, 39.2, 38.8, 34.4, 34.0, 32.7, 29.7, 29.5, 29.4, 29.3, 29.2, 29.1, 29.0, 28.8, 28.6, 28.5, 28.4, 25.7, 24.9; signals: 23 expected, 22 observed. ESI-HRMS: m/z [M + H]⁺ calcd for C₂₃H₄₇O₃S₃: 467.2687; found: 467.2691.

11-[11-(*N*-*tert*-Butoxycarbonylamino)undec-1-yl-trisulfanyl]undecanoic Acid (**3j**)

Chromatography (CH₂Cl₂–EtOAc = 10:1); yield: 0.403 g (73%), white solid, mp 54–56 °C. IR (KBr): ν = 3381 (m, NH), 2920 (s), 2850 (s, CH), 1895 (m, C=O), 1684 (s, C=O), 1639 (m, NH), 1172 (m, CN), 730 (w, CH₂) cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ = 1.20–1.44 (m, 30 H, CH₂), 1.46 (s, 9 H, *t*-Bu), 1.66–1.76 (m, 4 H, SCCH₂), 2.36 (t, J = 7.3 Hz, 2 H, CH₂COO), 2.88 (t, J = 7.3 Hz, 4 H, SCH₂), 3.00–3.20 (m, 2 H, NCH₂), 4.48 (br s, 1 H, NH). ¹³C NMR (50 MHz, CDCl₃): δ = 179.9, 156.0, 78.9, 40.7, 39.2, 38.9, 34.0, 31.9, 29.6, 29.6, 29.5, 29.3, 29.2, 29.1, 29.0, 28.8, 28.7, 28.5, 24.6, 22.7; signals: 25 expected, 20 observed. ESI-HRMS: m/z [M + H]⁺ calcd for C₂₇H₅₄NO₄S₃: 552.3215; found: 552.3220.

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