



2-Substituted 4,5-dihydrothiazole-4-carboxylic acids are novel inhibitors of metallo- β -lactamases

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ABSTRACT

Bacterial resistance to β -lactam antibiotics caused by class B metallo- β -lactamases (MBL), especially for certain hospital-acquired, Gram-negative pathogens, poses a significant threat to public health. We report several 2-substituted 4,5-dihydrothiazole-4-carboxylic acids to be novel MBL inhibitors. Structure activity relationship (SAR) and molecular modeling studies were performed and implications for further inhibitor design are discussed.

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The development of β -lactam antibiotics over the past 60 years has led to the availability of drugs to treat a wide range of Gram-positive and Gram-negative bacterial infections. However, during the past few decades, populations of drug resistant bacteria have increased significantly.¹ One important mechanism bacteria use to resist β -lactam antibiotics is the production of β -lactamases, which can hydrolyze the 4-membered β -lactam ring and render the drugs inactive. There are two mechanistically distinct types of β -lactamases: serine β -lactamases and metallo- β -lactamases (MBLs).² The class A, C and D β -lactamases belong to the first type and use a serine –OH group as a nucleophile to attack and hydrolyze the amide bond in the β -lactam ring. In contrast, the class B β -lactamases are Zn²⁺ dependent metalloenzymes, with one or two Zn ions at the active site.^{3,4} MBLs are able to hydrolyze essentially all β -lactams, including imipenem, which have the widest antibacterial spectrum and are one of the very few effective drugs to treat some Gram-negative bacterial infections, such as *Pseudomonas aeruginosa*. The clinical significance of these enzymes has only recently been recognized, when plasmid-encoded, transferable MBLs were found to spread quickly among many species of Gram-negative bacteria around the world and confer resistance to imipenem and extended-spectrum cephalosporins.^{5–7}

There has been much interest in discovering and developing MBL inhibitors during the past decade.⁸ As representatively shown in Figure 1, a number of structurally diverse inhibitors have been

identified, such as thiol-containing compounds,^{9,10} trifluoromethyl ketones,¹¹ tetrazoles,¹² and succinates.¹³ However, none of these compounds have been used in clinical trials to sensitize β -lactam antibiotics due to chemical instability, a narrow spectrum of activity or side effects. For example, captopril (Fig. 1) was discovered to be very active against IMP-1 from *P. aeruginosa*, one of the most prevalent forms of transferable MBLs.^{5–7,14} It is also used in the clinic to treat hypertension as a potent inhibitor of angiotensin-converting enzyme. There is, therefore, a pressing need to find potent, drug-like inhibitors that have a new scaffold for further development. Here, we report the discovery and structure activity relationships (SAR) of 2-substituted 4,5-dihydrothiazole-4-carboxylic acids as a new class of MBL inhibitors.

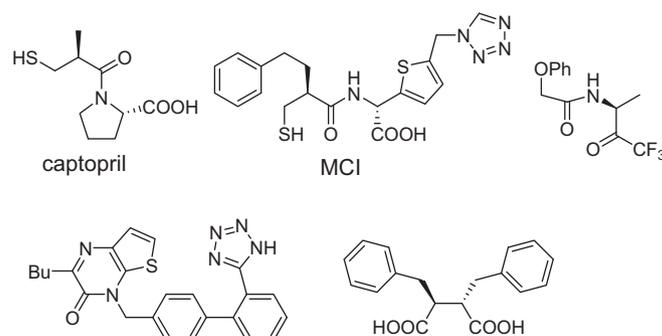
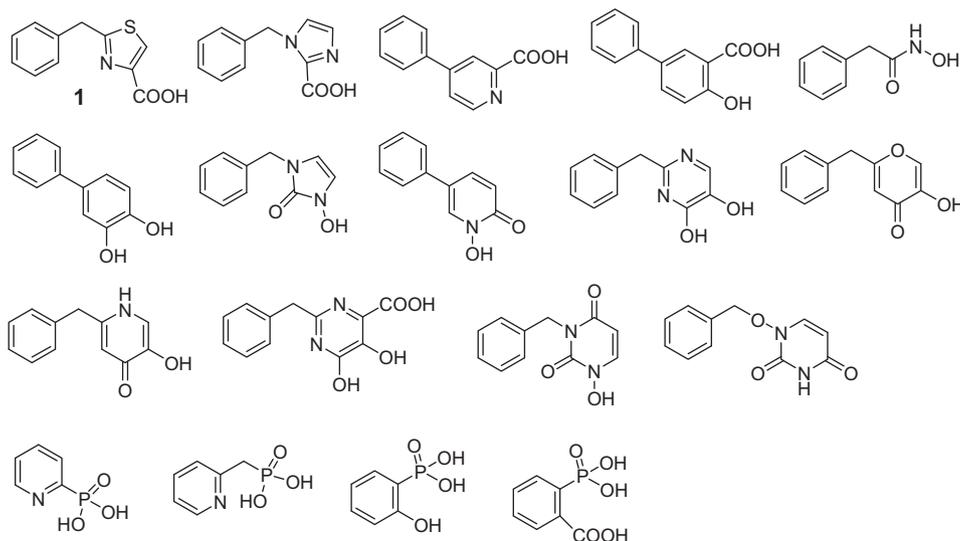


Figure 1. Representative MBL inhibitors.

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High-throughput screening (HTS) has been widely used to find inhibitors of enzymes, including MBLs, in the pharmaceutical industry. However, due to the high costs, HTS is often not available in many academic institutes, where alternative methods have to be used. For metalloenzymes, a coordination chemistry based approach has been successfully applied, by us¹⁵ and other groups,^{16,17} to discover new inhibitors. In order to find novel MBL inhibitors, we synthesized or purchased the following compounds:



Each of these compounds features a different potential Zn²⁺-binding group. In addition, they also contain a hydrophobic group, such as a phenyl or benzyl, that could potentially extend to the lipophilic pocket of MBLs to enhance the binding. These compounds were

tested for their inhibitory activities against recombinant IMP-1,^{18,19} together with captopril as a positive control, which was found to be a good inhibitor with an IC₅₀ value of 5.0 μM (Table 1), or a K_i value of 2.4 μM. Another control is ethylenediaminetetraacetic acid (EDTA), a non-competitive, broad inhibitor of MBLs, which acts as a metal chelator depleting Zn²⁺ from the active site of the metalloenzyme. EDTA was found to inhibit the activity of IMP-1 with an IC₅₀ value of 27.9 μM. Compound **1**, 2-benzylthiazole-4-carboxylic acid was identified to be a novel IMP-1 inhibitor

having an IC₅₀ of 34.7 μM.

Since compound **1** potentially represents a new class of MBL inhibitors, we performed a medicinal chemistry study in an effort to find compounds with improved activity as well as structure activity relationships (SAR). First, compounds **2–7** (Fig. 2) were synthesized to examine the effects of related core structures as well as those of phenyl versus benzyl at the 2-position. The inhibitory activities of these compounds against IMP-1 are listed in Table 1. For the fully aromatic thiazole ring, **1** with a 2-benzyl is more

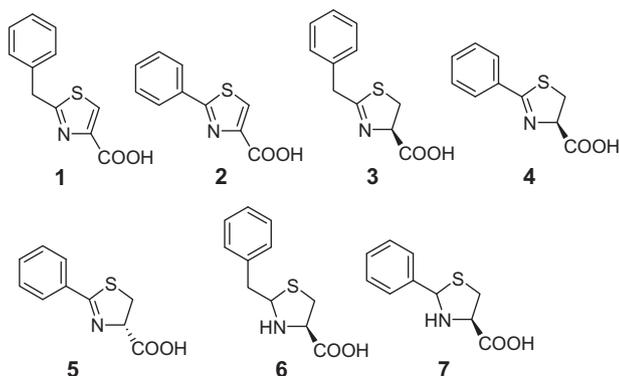


Figure 2. Structures of compounds 1–7.

Table 1
MBL inhibitory activities (IC₅₀ in μM) of 1–7

Compound	IC ₅₀ for IMP-1	IC ₅₀ for Bla2
Captopril	5.0	25.8
EDTA ^a	27.9	1060
1	34.7	>200
2	>200	>200
3	>200	>200
4	5.5	>200
5	>200	>200
6	>200	>200
7	>200	>200

^a with a 12-hour incubation.

Table 2
Structures and activities of compounds 8–17

	R-	IC ₅₀ for IMP-1	IC ₅₀ for Bla2
8	4-Cl-phenyl	>200	>200
9	4-Br-phenyl	>200	>200
10	3-NH ₂ -phenyl	77.0	4.9
11	3-acetamido-phenyl	>200	>200
12	3-BOCNH-phenyl	>200	74.1
13	2-OH-phenyl	>200	>200
14		>200	>200
15		>200	>200
16		>200	>200
17		>200	>200

active than **2** with a phenyl ($IC_{50} > 200 \mu M$). However, for the partially saturated 4,5-dihydrothiazole ring, compound **3** with a 2-benzyl group is inactive, while compound **4**, (*R*)-2-phenyl-4,5-dihydrothiazole-4-carboxylic acid, was found to be a good IMP-1 inhibitor with an IC_{50} value of $5.5 \mu M$ (or a K_i of $3.3 \mu M$), $\sim 6\times$ more active than **1**. However, its enantiomer **5**, (*S*)-2-phenyl-4,5-dihydrothiazole-4-carboxylic acid, was found to be inactive. In addition, none of compounds **6** and **7** having a thiazolidine ring possess inhibitory activity against IMP-1.

We next synthesized compounds **8–13** shown in Table 2 to investigate the SAR for a substituent on the 2-phenyl ring of compound **4**. Compounds **8** and **9** with a 2-(4-chlorophenyl) and 2-(4-bromophenyl), respectively, were found to completely lose the inhibitory activity. Compound **10** with a 3-NH₂ substituent was observed to have a weak inhibitory activity with an IC_{50} of $77.0 \mu M$. Its derivatives **11** and **12** containing an additional acetyl and *tert*-butoxycarbonyl (BOC) group, respectively, are inactive. Compound **13** with a 2-OH substituent on the phenyl ring has also no activity. Compounds **14–17** (Table 2) were synthesized to find the effects of

replacing 2-phenyl of **4** with an lipophilic amide or carbamate group. These groups were designed to mimic the sidechain of penicillin, one of the substrates of IMP-1. However, as can be seen in Table 2, none of these compounds show activity against IMP-1.

Molecular modeling (docking) was used to rationalize the above observed SARs. The crystal structure¹⁰ (PDB code: 1DD6) of IMP-1 in complex with the thiol inhibitor MCI (Fig. 1) was chosen to be the docking template, which was prepared, according to our previous method,^{15,20,21} by removing only the inhibitor using the program Glide²² in Schrödinger (version 2010).²³ The two Zn²⁺ ions were treated as an integrated part of the protein. Compound **4** was built, energy-minimized using OPLS_2005 force field in Schrödinger, and docked into the prepared IMP-1 structure using Glide. Figure 3A shows the 20 docking structures of **4** with the lowest energies, which can be basically classified into two binding conformations A and B. Conformation A represents 7 tightly clustered docking structures with a lower average energy, and there are 13 docking poses in conformation B, as representatively shown in Figure 3B and C. The common feature of conformations A and B is that one O atom of the carboxylate group binds to both Zn²⁺ ions, behaving similarly as the S atom of MCI. The other O atom of **4** in conformation A also interacts with the tightly bound Zn1 (coordinated by three imidazoles of His77, 79 and 139), while that of B binds to the Zn2 (which is bound to IMP-1 via coordination by Asp81, Cys158 and His197 with less affinity). None of the N atoms of **4** in conformations A and B are predicted to bind to Zn²⁺, with the distances being $>3 \text{ \AA}$. It is of interest that the binding of the 2-phenyl group of **4** in conformations A and B is different. The phenyl of A is located almost in the same position as the phenyl of MCI, and the mainly hydrophobic pocket is tightly surrounded by Val25, Val31, Glu23 (the hydrophobic carbon skeleton), Ser21, Pro32, Phe51 and His197 (Fig. 3b). The phenyl group of conformation B is situated in a much larger and longer channel that holds the two aromatic rings of MCI (Fig. 3c). Our SAR results suggest the conformation A better mimics the real binding structure of compound **4** to IMP-1, because the pocket shown in Fig. 3b could only accommodate a phenyl group favorably. Any additional substituent, even as small as a -Cl or -OH, could result in an unfavored steric and/or hydrophobic interaction. In addition, our SAR and docking studies could provide useful implications for future inhibitor design. For example, derivatives of compound **4** that possess an additional group that extends to the other binding site of IMP-1 could be more active.

Finally, we tested the inhibitory activities of newly synthesized compounds against another MBL, *Bacillus anthracis* Bla2,²⁴ in order

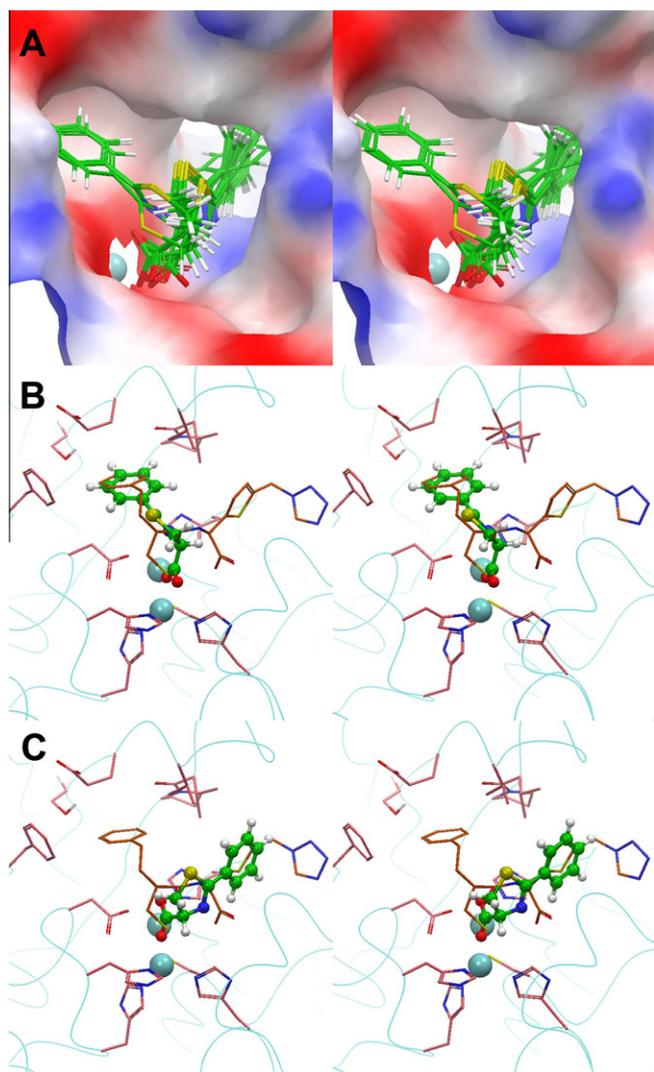
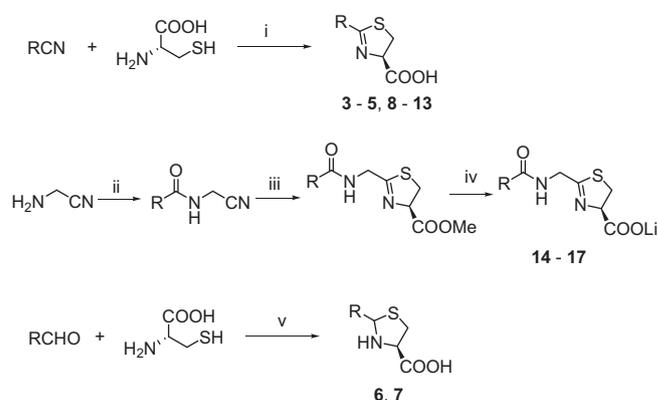


Figure 3. (A) Stereoview of the 20 lowest-energy docking structures of compound **4** in the active site of the crystal structure of IMP-1:MCI complex, with the protein shown as an electrostatic potential surface and Zn²⁺ as a light blue sphere; (B) The lowest-energy docking structure of conformation A of **4** (the ball and stick model in green) in the active site, superimposed with the crystal structure of MCI (in orange); (C) The lowest-energy docking structure of conformation B of **4** superimposed with the structure of MCI.



Scheme 1. General synthesis for compounds **3–17**. Reagents and conditions: (i) NaHCO₃, phosphate buffer (pH 6), MeOH, 66 °C, 72 h; (ii) RCOCl or (BOC)₂O, Et₃N, CH₂Cl₂; (iii) *L*-Cys-OMe, Et₃N, MeOH; (iv) 0.9 equiv 1 M LiOH, 0 °C to room temperature, 1 h; (v) EtOH, H₂O, room temperature, 18 h.

to see whether our inhibitors have a broad spectrum of activity. *B. anthracis* is the causative agent of anthrax and expression of Bla2 can result in resistance to β -lactam antibiotics. Therefore, inhibitors of this enzyme have potential therapeutic value. The results are shown in Tables 1 and 2. Captopril was also found to be a weak inhibitor of Bla2 with an IC_{50} value of 25.8 μ M, or a K_i value of 17.9 μ M, while EDTA is almost inactive against this enzyme (IC_{50} >1 mM). Although compound 4 is a good IMP-1 inhibitor, it was observed to have essentially no activity against Bla2 with an IC_{50} of >200 μ M. However, analogous compound 10 with a 3-NH₂ substituent showed improved inhibitory activity against Bla2 as compared to captopril, with an IC_{50} value of 4.9 μ M (Table 2) or a K_i value of 5.1 μ M. All other compounds, except for compound 12 being a weak inhibitor (IC_{50} = 74.1 μ M), are also inactive against Bla2. Nevertheless, the activity of compound 10 shows that it is also possible to develop 4,5-dihydrothiazole-4-carboxylic acids to find a compound that have a broad activity against other MBLs.

General methods for synthesizing compounds 3–17 are illustrated in Scheme 1.²⁵ For 4,5-dihydrothiazole compounds 3–5 and 8–13, an aromatic nitrile, 1.2 equiv amount of *L*-cysteine (or *D*-cysteine) hydrochloride and NaHCO₃ was refluxed in methanol and phosphate buffer (pH 6) for 72 h,²⁶ to give these compounds in 40–70% yield. However, this method does not work for making analogous compounds 14–17. Rather, the methyl esters of these compounds were synthesized using a similar procedure,²⁷ which were carefully undergone a mild hydrolysis using 0.9 equiv of LiOH to afford compounds 14–17 in 70–80% yield. Excess amount of LiOH caused decomposition of the products. Thiazolidine compounds 6 and 7 were prepared in ~80% yield by reacting benzaldehyde or phenylacetaldehyde with *L*-cysteine at room temperature in a mixture of ethanol and water.²⁸

In summary, this work is of interest for several reasons. First, using rational compound screening followed by medicinal chemistry, 2-phenyl-4,5-dihydrothiazole-4-carboxylic acid (4) was identified to be a novel inhibitor of *P. aeruginosa* MBL IMP-1 with an IC_{50} value of 5.5 μ M. Second, docking studies were explored to rationalize the structure activity relationships of this class of compounds and provide implications for future inhibitor design. Third, 2-(3-aminophenyl)-4,5-dihydrothiazole-4-carboxylic acid (10) was identified to be an inhibitor of *B. anthracis* MBL Bla2 (IC_{50} = 4.9 μ M), showing the promise of further developing this class of compounds in the context of overcoming β -lactam resistance.

Acknowledgments

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.bmcl.2012.08.012>.

References and notes

- Leeb, M. *Nature* **2004**, *431*, 892.
- Fisher, J. F.; Meroueh, S. O.; Mobashery, S. *Chem. Rev.* **2005**, *105*, 395.
- Crowder, M. W.; Spencer, J.; Vila, A. J. *Acc. Chem. Res.* **2006**, *39*, 721.
- Wang, Z.; Fast, W.; Valentine, A. M.; Benkovic, S. J. *Curr. Opin. Chem. Biol.* **1999**, *3*, 614.
- Walsh, T. R.; Toleman, M. A.; Poirel, L.; Nordmann, P. *Clin. Microbiol. Rev.* **2005**, *18*, 306.
- Bebrone, C. *Biochem. Pharmacol.* **2007**, *74*, 1686.
- Walsh, T. R. *Clin. Microbiol. Infect.* **2005**, *11*, 2.
- Toney, J. H.; Moloughney, J. G. *Curr. Opin. Investig. Drugs* **2004**, *5*, 823.
- Garcia-Saez, I.; Hopkins, J.; Papamical, C.; Franceschini, N.; Amicosante, G.; Rossolini, G. M.; Galleni, M.; Frere, J. M.; Dideberg, O. *J. Biol. Chem.* **2003**, *278*, 23868.
- Concha, N. O.; Janson, C. A.; Rowling, P.; Pearson, S.; Cheever, C. A.; Clarke, B. P.; Lewis, C.; Galleni, M.; Frere, J. M.; Payne, D. J.; Bateson, J. H.; Abdel-Meguid, S. S. *Biochemistry* **2000**, *39*, 4288.
- Walter, M. W.; Felici, A.; Galleni, M.; Soto, R. P.; Adlington, R. M.; Baldwin, J. E.; Frère, J.-M.; Gololobov, M.; Schofield, C. J. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 2455.
- Toney, J. H.; Fitzgerald, P. M.; Grover-Sharma, N.; Olson, S. H.; May, W. J.; Sundelof, J. G.; Vanderwall, D. E.; Cleary, K. A.; Grant, S. K.; Wu, J. K.; Kozarich, J. W.; Pompliano, D. L.; Hammond, G. G. *Chem. Biol.* **1998**, *5*, 185.
- Toney, J. H.; Hammond, G. G.; Fitzgerald, P. M.; Sharma, N.; Balkovec, J. M.; Rouen, G. P.; Olson, S. H.; Hammond, M. L.; Greenlee, M. L.; Gao, Y. D. *J. Biol. Chem.* **2001**, *276*, 31913.
- Watanabe, M.; Iyobe, S.; Inoue, M.; Mitsunashi, S. *Antimicrob. Agents Chemother.* **1991**, *35*, 147.
- Deng, L.; Sundriyal, S.; Rubio, V.; Shi, Z.; Song, Y. *J. Med. Chem.* **2009**, *52*, 6539.
- Jacobsen, F. E.; Lewis, J. A.; Cohen, S. M. *J. Am. Chem. Soc.* **2006**, *128*, 3156.
- Jacobsen, F. E.; Lewis, J. A.; Cohen, S. M. *ChemMedChem* **2007**, *2*, 152.
- Brown, N. G.; Horton, L. B.; Huang, W.; Vongpunsawad, S.; Palzkill, T. *Antimicrob. Agents Chemother.* **2011**, *55*, 5696.
- The enzyme assay was performed using 2 nM IMP-1 (or 10 nM Bla2) and 25 μ M nitrocefin in 50 mM HEPES buffer (pH 7.0) containing 20 μ g/mL BSA and 0.01% Triton in a final volume of 300 μ L. For inhibition assay, compounds were pre-incubated with the enzyme for 20 min at room temperature, before initiation of the reaction by adding nitrocefin. The increasing absorbance at 482 nm was monitored using a Beckman DU640 UV spectrometer. The initial velocities for different concentrations of an inhibitor were imported into Prism (version 5.0, GraphPad, La Jolla, CA). The IC_{50} as well as K_i values were calculated by using a standard dose response curve fitting in the software.
- Deng, L.; Endo, K.; Kato, M.; Cheng, G.; Yajima, S.; Song, Y. *ACS Med. Chem. Lett.* **2011**, *2*, 165.
- Deng, L.; Diao, J.; Chen, P.; Pujari, V.; Yao, Y.; Cheng, G.; Crick, D. C.; Prasad, B. V.; Song, Y. *J. Med. Chem.* **2011**, *54*, 4721.
- Glide, version 5.5, Schrödinger, LLC., New York, NY, 2010.
- Schrödinger Suite, version 2010, Schrödinger, LLC, New York, NY, 2010.
- Materon, I. C.; Queenan, A. M.; Koehler, T. M.; Bush, K.; Palzkill, T. *Antimicrob. Agents Chemother.* **2003**, *47*, 2040.
- Experimental Section can be found in Supplementary data on-line.
- Lu, Y.; Li, C.; Wang, Z.; Chen, J.; Mohler, M. L.; Li, W.; Dalton, J. T.; Miller, D. D. *J. Med. Chem.* **2011**, *54*, 4678.
- Merino, P.; Tejero, T.; Unzurrunzaga, F. J.; Franco, S.; Chiacchio, U.; Saita, M. G.; Iannazzo, D.; Piperno, A.; Romeo, G. *Tetrahedron: Asymmetry* **2005**, *16*, 3865.
- Lu, Y.; Li, C.; Wang, Z.; Ross, C. R.; Chen, J.; Dalton, J. T.; Li, W.; Miller, D. D. *J. Med. Chem.* **2009**, *52*, 1701.