Bioorganic & Medicinal Chemistry Letters 22 (2012) 1073-1077

Contents lists available at SciVerse ScienceDirect



Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Synthesis and structure–activity relationship of 5-pyridazin-3-one phenoxypiperidines as potent, selective histamine H₃ receptor inverse agonists

Ming Tao^{*}, Lisa D. Aimone, John A. Gruner, Joanne R. Mathiasen, Zeqi Huang, Jacquelyn Lyons, Rita Raddatz, Robert L. Hudkins

Discovery Research, Cephalon, Inc., 145 Brandywine Parkway, West Chester, PA 19380, USA

ARTICLE INFO

Article history: Received 21 October 2011 Revised 22 November 2011 Accepted 28 November 2011 Available online 4 December 2011

Keywords: Phenoxypiperidine 5-pyridazin-3-one Histamine H₃ receptor CEP-26401 irdabisant Sleep-wake

ABSTRACT

Optimization of the \mathbb{R}^2 and \mathbb{R}^6 positions of (5-{4-[3-(\mathbb{R})-2-methylpyrrolin-1-yl-propoxy]phenyl}-2H-pyridazin-3-one) **2a** with constrained phenoxypiperidines led to the identification of 5-[4-(cyclobutyl-piperidin-4-yloxy)-phenyl]-6-methyl-2H-pyridazin-3-one **8b** as a potent, selective histamine H₃ receptor antagonist with favorable pharmacokinetic properties. Compound **8b** had an excellent safety genotoxocity profile for a CNS-active compound in the Ames and micronucleus tests, also displayed potent H₃R antagonist activity in the brain in the rat dipsogenia model and robust wake activity in the rat EEG/EMG model.

© 2011 Elsevier Ltd. All rights reserved.

Histamine plays an important role in different physiological processes by four G-protein coupled receptors (H_1R-H_4R) and exerts a variety of functions in the central nervous system (CNS).¹ The histamine H_3 receptor (H_3R) is predominantly expressed in the central nervous system (CNS), where it functions both as an autoreceptor to modulate histamine release and as an inhibitory heteroreceptor regulating the release of multiple neurotransmitters including acetylcholine, histamine, norepinephrine, and dopamine.² H_3R antagonists are currently being evaluated as potential therapeutic agents for the treatment of cognitive deficits associated with a variety of CNS disease states including Alzheimer's disease (AD), attention deficit hyperactivity disorder (ADHD) and schizophrenia.³

There have been a number of H₃R antagonists advance to preclinical and clinical development studies (Fig. 1). For example, ABT-239 (**1a**) was nominated for clinical development for cognition promotion, but was halted because of QTc prolongation observed in monkeys.^{1,3g,f} MK-0249 (**1b**) from Merck has completed Phase II trails for ADHD, AD and cognitive impairment associated with schizophrenia.^{1,3g,f} JNJ-31001074 (bavisant, **1c**) is reported in Phase II for ADHD.^{1a} Pfizer has reported PF-03654746 (**1d**) in Phase II trail for ADHD and was discontinued.^{3f} GSK-189254 (**1e**) enhanced cognitive performance pre-clinically and advanced to Phase II for narcolepsy and was in early clinical trails for AD.^{3f} We recently reported a novel class of pyridazin-3-one H₃R antago-

* Corresponding author. E-mail address: mtao@cephalon.com (M. Tao). nists/inverse agonists with excellent drug properties, safety and in vivo profiles and advanced CEP-26401 **1f** as a clinical candidate, which recently completed Phase $I.^5$

As part of our H_3R discovery project studying the structureactivity relationships (SAR) around **1f**, we synthesized and reported the profile of the 5-regiomer **2a** (5-{4-[3-(*R*)-2-methylpyrrolin-1-ylpropoxy]-phenyl}-2*H*-pyridazin-3-one) (Fig. 2). Compound **2a** had



Figure 1. Structure of pre-clinical and clinical H₃R antagonists.

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter \odot 2011 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2011.11.118



Figure 2. Pyridazinone H₃R antagonists.

high affinity for both the human (hH₃R K_i = 2.8 nM) and rat H₃Rs (rH₃R K_i = 8.5 nM), but displayed low oral bioavailability (%*F* = 24) in the rat.⁵ To explore the structure–activity relationships (SAR) of 5-pyridazin-3-one core R² and R⁶ positions around **2a** with a focus on improving the pharmacokinetic (PK) properties, we identified 5-{4-[3-(*R*)-2-methyl-pyrrolidin-1-yl)-propoxy]-phenyl}-2-pyridin-2-yl-2H-pyridazin-3-one **2b** (hH₃R K_i = 1.7 nM and rH₃R K_i = 3.7 nM) with favorable oral bioavailability (%*F* = 78 in rat).⁶ However, **2b** showed positive results in the Ames test, and was discontinued for development.

Most non-imidazole H₃R antagonists reported share common structural features: a basic tertiary amine attached to an aromatic ring through a linker-space group; for example, the propyloxy, and its constrained piperidinyloxy moiety.^{7,1a} There have been many reports describing a variety of histamine H₃R receptor antagonist pharmacophores in the past few years.^{4,1a} One design strategy we used in our search for 5-aryl analogs with a clean genotoxicity profile and to improved PK was to rigidify the core to reduce in the number of rotatable bonds.^{7d} Based on this, we synthesized a series of 5-regiomers 8 (5-[4-(1-cyclobutyl-piperidin-4-yloxy)-phenyl]-2H-pyridazin-3-one) where the pyrrolidinylpropyloxy side chain was constrained as the *N*-cyclobutylpiperidinyloxy moiety; a maneuver explored by the Merck and Johnson and Johnson groups on their respective cores.^{7e} In this Letter, we report the synthesis, SAR, selectivity, pharmacokinetics (PK) and the in vivo activities of 5-[4-(1-cyclobutyl-piperidin-4-yloxy)-phenyl]-6-methyl-2Hpyridazin-3-one 8b in the rat dipsogenia model and in the rat EEG/EMG model of wakefulness.

The synthesis of 5-[4-(1-cyclobutyl-piperidin-4-yloxy)-phenyl]-pyridazinone analog 8a is shown in Scheme 1. Alkylation of 4-hydroxy-phenyl ethanol **3** with 4-methanesulfonyloxy-1-Bocpiperidine **4** in the presence of cesium carbonate in DMF afforded the ether 5. Dess-Martin oxidation of 5, followed by condensation with glyoxylic acid, and subsequent cyclization with hydrazine hydrate yielded the pyridazinone **7**.⁸ Deprotection of **7** with HCl in dioxane, followed by reductive amination with cyclobutanone in the presence of NaCNBH₃ gave 8a. The 6-methyl substituted pyridazinones 8b and 8d were synthesized as shown in Scheme 2. Mitsunobu coupling of 4-hydroxyphenyl acetone 9 and 4-hydroxy-1-Boc-piperidine **10** in the presence of 40% of diethylazodicarboxylate in toluene and triphenylphosphine gave ether **11**. Condensation of **11** with glyoxylic acid, followed by cyclization with hvdrazine afforded the 6-methyl pyridazinone 12. Final targets **8b** and **8d** were synthesized following the same steps as described in Scheme 1. The synthesis of 5-[4-(1-cyclobutyl-piperidin-4-yloxy)-phenyl]-5-methyl-4,5-dihydro-2H-pyridazin-3-one 8c is shown in Scheme 3. Alkylation of **11** with ethyl bromoacetate in the presence of KHMDS, and cyclization with hydrazine yielded the Boc-piperidinyl-6-methyl dihydropyridazinone 14. Deprotection and reductive amination as described in Scheme 1 gave 8c. The 2-aryl substituted pyridazinone analogs 8e-h were synthesized as shown in Scheme 4. The 5-(4-methoxy-phenyl)-6methyl-2H-pyridazin-3-one 16 was coupled with aryl bromide in the presence of copper(I) iodide in DMF to give 2-aryl substituted 17.9 Deprotection of 17 with BBr₃, followed by alkylation with 1-Boc-4-methanesulfonyloxy-piperidine **4** afforded **12**. Deprotection and reductive amination as shown in Scheme 1 gave the 2-aryl pyridazinones 8e-h. The synthesis of 24 is shown in Scheme 5. Suzuki coupling 2-chloropyridyl-4-boronic acid **20** with 2-hydroxymethyl-5-iodo-2H-pyridazin3-one, followed by substitution with 4-hydroxy-1-Boc-piperidine afforded the Boc-protected pyridyl pyridazinone 22. Deprotection of 22 with HCl in dioxane, and reductive amination gave compound 24.

The substituted pyridazinone analogs were tested using in vitro binding assays by displacement of $[^{3}H]N-\alpha$ -methylhistamine ($[^{3}H]NAMH$) in membranes isolated from CHO cells transfected with cloned human H₃ or rat H₃ receptors.¹⁰ Rat and human H₃R binding data for the analogs is shown in Table 1 in comparison with **2a**.⁵ The *N*-cyclobutyl was established to be optimum and was fixed for comparison while exploring SAR.¹¹ As previously reported, the 5-pyridazin-3-one regiomer of **2a** had high affinity (hH₃R K_i = 2.8 nM, rH₃R K_i = 8.5 nM), but displayed low oral



Scheme 1. Reagents and conditions: (a) Cs₂CO₃, DMF, 100 °C, 59%; (b) Dess–Martin [O], CH₂Cl₂, 0 °C to rt, 35%; (c) (i) HCOCOOH·H₂O, 135 °C; (ii) NH₂NH₂, EtOH, 90 °C, 34% in 2 steps; (d) (i) 4 N HCl in dioxane, 50 °C; (ii) cyclobutanone, NaCNBH₃, DMF/MeOH/acetic acid, 60 °C, 1 h, 56%.



Scheme 2. Reagents and conditions: (a) DEAD, PPh₃, THF, 0 °C→rt, 63%; (b) (i) HCOCOOH·H₂O, 135 °C; (ii) R²NHNH₂, EtOH, 90 °C, 23% in 2 steps; (c) 4 N HCl in dioxane, dioxane, 50 °C; (d) cyclobutanone, NaCNBH₃, DMF/MeOH/acetic acid, 60 °C, 2 h, 80%.



Scheme 3. Reagents and conditions: (a) (i) KHMDS, $BrCH_2COOEt$, THF, -78 °C; (ii) NH_2NH_2 , EtOH, 90 °C, 56% in 2 steps; (b) TFA, CH_2CI_2 , 0 °C to rt; (c) cyclobutanone, $NaCNBH_3$, DMF/MeOH/acetic acid, 60 °C, 2 h, 30%.

bioavailability in the rat (%F = 24).⁵ Replacement of (R)-2-methylpyrrolidinylpropoxy of 5-pyridazin-3-one with the constrained cyclobutylpiperidinyloxy (**8a**) showed high affinity in both human and rat H₃R (hH₃R $K_i = 2$ nM, rH₃R $K_i = 2$ nM), but did not show improvement on rat oral bioavailability (%F = 21) compared to **2a**. Further profiling showed **8a** had moderate selectivity for the hERG channel in a patch clamp assay (IC₅₀ = 2 μ M). Compound **8b**, with a methyl substitution at the 6-position, designed to increase the torsional angle between the two rings, retained high affinity in both human and rat H₃R (hH₃R $K_i = 5$ nM, rH₃R $K_i = 6$ nM), and displayed acceptable in vitro metabolic stability in liver microsomes from rat, dog and human ($t_{1/2} > 40$ min), and showed weak CYP inhibition

(IC₅₀>30 µM, 5 CYP isoforms). Compound 8b had an improved PK profile in rat (%F = 68, Table 2). The dihydropyridazinone analog **8c** showed \sim 2–5-fold weaker affinity compared to **2a** (hH₃R $K_i = 15 \text{ nM}$, rH₃R $K_i = 13 \text{ nM}$). Compound **8c** had acceptable in vitro metabolic stability in liver microsomes from rat, dog and human ($t_{1/2}$ >40 min) and weak CYP inhibition (IC₅₀ >30 μ M, 5 CYP isoforms). However, plasma levels of 8c were low following oral administration (%F = 6, C_{max} = 73 ng/mL, AUC = 370 ng*h/mL), and 8c was extensively aromatized to the metabolite 8b (~40% in plasma, C_{max} = 34 ng/mL, AUC = 152 ng*h/mL) in a rat PK experiment. The N-2-methyl analog 8d, with high affinity, acceptable in vitro metabolic stability in liver microsomes from rat, dog and human $(t_{1/2} > 40 \text{ min})$ and weak CYP inhibition (IC₅₀ > 30 μ M, 5 CYP isoforms), was profiled in a rat PK experiment. Following oral administration of **8d** (%*F* = 19, *C*_{max} = 107 ng/mL, AUC = 337 ng*h/mL) in the rat, high levels of the demethylated active metabolite 8b (~95% in plasma, $C_{\text{max}} = 68 \text{ ng/mL}$, AUC = 320 ng*h/mL) was formed in plasma, therefore the compound was not further advanced. The N-2-pyridyl analog 8e was synthesized with the goal to find metabolically stable R^2 analogs while maintaining a low log*P* value (<3).⁶ Compound **8e** had high affinity in both human and rat H₃R and displayed acceptable in vitro metabolic stability in liver microsomes from rat, dog and human ($t_{1/2}$ >40 min), and weak CYP inhibition (IC₅₀ >30 μ M for 5 CYP isoforms). Further profiling showed 8e displayed an acceptable PK profile (Table 2), weak hERG activity in a patch clamp assay (IC₅₀>30 µM), and showed high selectivity against hH₁, hH₂, and hH₄ receptor subtypes (<30% inhibition at 10 μ M). However, it had moderate affinity for the muscarinic M₂ subtype (IC₅₀ = 0.34μ M) when screened against



Scheme 4. Reagents and conditions: (a) K₂CO₃, ArBr, Cul, DMF, 150 °C, 36–87%; (b) BBr₃, CH₂Cl₂, 0 °C to rt, ~80%; (c) 4-methanesulfonyloxy-piperidine-1-carboxylic acid *tert*butyl ester, Cs₂CO₃, DMF, 100 °C, 40–70%; (d) (i) 4 N HCl in dioxane, dioxane, 50 °C, (ii) cyclobutanone, NaCNBH₃, DMF/MeOH/acetic acid, 60 °C, 1 h, ~70%.



Scheme 5. Reagents and conditions: (a) Pd(PPh₃)₄, K₂CO₃, DME/water, 85 °C, 36%; (b) 4-hydroxy-piperidine-1-carboxylic acid *tert*-butyl ester, K₂CO₃, DMSO, 110 °C, 54%; (c) 4 N HCl in dioxane, dioxane, 50 °C; (d) cyclobutanone, NaCNBH₃, DMF/MeOH/acetic acid, 60 °C, 1 h, 56%.

Table 1

5-Pyridazin-3-one in vitro binding data

-							
	Compd	Bond	Х	R ²	\mathbb{R}^{6}	$hH_3(K_i, nM)$	$rH_3(K_i, nM)$
	8a	Double	СН	Н	Н	2	2
	8b	Double	CH	Н	Me	5	6
	8c	Single	CH	Н	Me	15	13
	8d	Double	CH	Me	Me	4	8
	8e	Double	CH	2-Py	Me	3	9
	8f	Double	CH	6-Me-2-py	Me	4	10
	8g	Double	CH	3-Me-2-py	Me	6	17
	8h	Double	CH	3-Thiophenyl	Me	3	8
	24	Double	Ν	Н	Н	14	20

 K_i values are an average of two or more determinations. The assay-to-assay variation was typically within 2.5-fold.

71 GPCRs, ion channels and enzymes (MDS Pharma Services, Lead Profiler). The methyl substituted N^2 -2-pyridine analogs **8f**, and **8g** also showed high affinity for both human and rat H₃R, good in vitro metabolic stability, weak CYP inhibition (IC₅₀ >30 µM, 5 CYP isoforms), and acceptable hERG selectivity (IC₅₀ >30 µM) in a patch clamp assay. However, it showed no improvement in rat PK compared to **2a** (Table 2). Further R² modification, the *N*-2-(3)-thiophene analog **8h** also showed high affinity for both human and rat H₃R but had significantly lower solubility. Changing the central phenyl ring to pyridyl **24** was designed to lower the *c*log*P*

Table 2

Pharmacokinetic properties in rat

and potentially improve hERG activity. However, this maneuver showed much weaker H_3R affinity compared to **8a**.

Compound **8b** was further tested for H₃R functional activity using a [35S]GTPγS hH₃R binding assay and displayed full inverse agonist activity with an $EC_{50} = 1.1 \text{ nM.}^{10}$ Compound **8b** showed high selectivity against hH₁, hH₂, and hH₄ receptor subtypes (<20% inhibition at 10 μ M) and in a panel of 101 GPCRs, ion channels and enzymes inhibited the α -adrenergic receptor subtype greater than 50% at 10 μ M. It showed acceptable hERG selectivity with an IC_{50} value of 7 μ M and displayed acceptable drug-like properties with high water solubility (pH 2 and pH 7.4 >0.14 mg/ mL), low lipophilicity ($c\log P = 1.5$), and high permeability in the Caco-2 assay ($P_{app} = 14.9 \times 10^{-6}$ cm/s). Compound **8b** had low binding to plasma proteins of rat (54%), dog (43%), and human (58%) in vitro. In genotoxicity profiling, **8b** did not induce mutation in the Ames assay (Salmonella typhimurium strains TA98, TA100, TA102, TA1535 and TA1537) both in the presence and absence of rat liver S9 metabolic activation and did not induce micronuclei in vitro in a micronucleus test in mouse lymphocytes with and without metabolic activation.

The rat dipsogenia model was used as an in vivo surrogate measure of H₃R functional inhibition in the brain following peripheral administration, and the activity in this model may be predictive of efficacy in cognitive models. Histamine and the H₃R-selective agonist, R- α -methylhistamine (RAMH), induce water drinking in the rat when administered either peripherally or centrally, an effect that is blocked by H₃R antagonists.¹² Compound **8b** inhibited RAMH-induced water drinking, demonstrating potent inhibition of central H₃R following ip administration of compound with an ED₅₀ value of 0.02 mg/kg, consistent with the high brain exposure

	2a ^a	8b ^a	8e ^b	8f ^b	8g ^b
iv ^c					
$t_{\frac{1}{2}}(h)$	1.0 ± 0.1	1.5 ± 0.5	1.3 ± 0.0	1.1 ± 0.1	1.1 ± 0.1
Vd (L/kg)	0.8 ± 0.1	3.4 ± 1.5	1.4 ± 0.1	1.7 ± 0.6	1.1 ± 0.2
CL (mL/min/kg)	9.6 ± 1.0	25 ± 4	13 ± 1.3	16 ± 4	11 ± 2.4
ро ^с					
AUC (ng h/mL)	4209 ± 258	4869 ± 524	1502 ± 103	1273 ± 300	1860 ± 347
$C_{\rm max} (ng/mL)$	446 ± 65	673 ± 44	287 ± 31	332 ± 129	505 ± 137
$T_{\rm max}$ (h)	2 ± 0	1 ± 0	3.2 ± 0.7	2.0 ± 1.0	2.3 ± 0.9
F (%)	24 ± 2	68 ± 7	22 ± 2	20 ± 5	23 ± 4
B/P^{d}	1.8 ± 0.2	2.3 ± 0.1	1.6 ± 0.2	1.6 ± 0.1	1.0 ± 0.0

^a Administration at 1 mg/kg iv and 10 mg/kg po.

^b Administration at 1 mg/kg iv and 5 mg/kg po.

^c iv formulation (3% DMSO, 30% solutol, 67% phosphate buffered saline) oral formulation (50% Tween 80, 40% propylene carbonate and 10% propylene glycol).

^d B/P = brain to plasma ratio measured 6 h post 10 mg/kg ip dose.



Figure 3. Compound **8b**-induced wake promotion. Cumulative wake time (min) for 4 h post dosing following administration of vehicle (Veh), Compound **8b** in rats chronically implanted with electrodes for recording EEG and EMG activity. Mean + SEM; n = 6-7/group. *p < 0.05, Dunnett's test versus vehicle.

and potent rat H₃R affinity. Histamine-producing neurons are an important part of the monoaminergic arousal system and H₃R antagonists have been documented to increase wakefulness in a number of species. However, potency of H₃R antagonist in the rat dipsogenia model has been reported to correlate with lower levels of receptor occupancy compared with wake promotion, which required much higher doses, and 80-100% receptor occupy for robust wake activity.^{13,14} Compound **8b** was evaluated in the rat EEG/EMG sleep-wake model. Wake-promoting activity in the rat was measured as previously described using male Sprague Dawley rats surgically implanted for chronic recording of EEG (electroencephalographic) and EMG (electromyographic) signals.¹⁵ Cumulative wake time for 4 h post dosing was evaluated during the normal quiet period of the rat. While the wake activity at 1 and 3 mg/kg was significantly greater than vehicle, the corresponding slow-wave sleep onset values $(34.9 \pm 5.1, 35.3 \pm 7.0, \text{ and } 45.0 \pm 7.5 \text{ for vehicle, 1, and 3 mg/kg}$ groups, respectively) were not different. Compound 8b significantly increased waking at 10 and 30 mg/kg ip (167 ± 11 min, and 226 ± 2.9 min) by 4 h AUC values (Fig. 3). Maximal cumulative wake surplus at 30 mg/kg was 163 min at 6 h post dosing, and dropped by only 20 min over the next 11 h (data not shown). At 30 mg/kg ip, 8b demonstrated robust wake promotion, with the treated animals being awake 94% of the time up to 4 h post dose and the increase in wake time over vehicle = 145 min which is 178% of the vehicle wake time (Fig. 3). No hypersomnolence was observed in any group (data not shown).

In summary, optimization of the 5-pyridazin-3-one R^2 and R^6 positions of **8** with constrained phenoxypiperidine amine led to the identification of 5-[4-(cyclobutyl-piperidin-4-yloxy)-phenyl]-6-methyl-2*H*-pyridazin-3-one **8b** as a potent, selective histamine H₃ receptor antagonist. Compound **8b** showed favorable pharma-cokinetic properties with improved oral bioavailability (%*F* = 61 in rat), and displayed an excellent safety genotoxicity profile for a CNS-active compound in the Ames and mouse lymphocyte micronucleus in vitro tests. Compound **8b** displayed potent H₃R antagonist activity in the brain in the rat dipsogenia model and robust wake activity in the rat EEG/EMG model.

Acknowledgments

The authors acknowledge the support and contributions from Edward R. Bacon, Mark A. Ator, Michael J. Marino, Mehran Yazdanizn, Amy Decamillo, Bob Bendesky, Nathalie Bourrit, and Debra Galinis.

References and notes

- For reviews see (a) Berlin, M.; Boyce, C. W.; de Lera Ruiz, M. J. Med. Chem. 2011, 54, 26; (b) Brown, R.; Stevens, D. R.; Haas, H. L. Prog. Neurobiol. 2001, 63, 637; (c) Leurs, R.; Bakker, R. A.; Timmerman, H.; de Esch, J. Nat. Rev. Drug Disc. 2005, 4, 107; (d) Wijtmans, M.; Leurs, R.; de Esch, I. Expert Opin. Investig. Drugs 2007, 16, 967; (e) Esbenshade, T. A.; Fox, G. B.; Cowart, M. D. Mol. Interv. 2006, 6, 77; (f) Esbenshade, T. A.; Browman, K. E.; Bitner, R. S., et al Br. J. Pharmacol. 2008, 154, 1166; (g) Hudkins, R. L.; Raddatz, R. Annu. Rep. Med. Chem. 2007, 42, 49; (h) Raddatz, R.; Tao, M.; Hudkins, R. L. Curr. Top. Med. Chem. 2010, 10, 153; (i) Brioni, J. D.; Esbenshade, T. A.; Garrison, T. R.; Bitner, S. R.; Cowart, M. D. J. Pharmacol. Exp. Ther. 2011, 336, 38.
- (a) Oda, T.; Morikawa, N.; Saito, Y.; Masuho, Y.; Matsumoto, S. J. Biol. Chem.
 2000, 275, 36781; (b) Liu, C.; Ma, X.; Jiang, X.; Wilson, S. J.; Hofstra, C. L.; Blevitt, J.; Pyati, J.; Li, X.; Chai, W.; Carruthers, N.; Lovenberg, T. W. Mol. Pharmacol.
 2001, 59, 420; (c) Cowart, M. D.; Altenbach, R. J.; Liu, H.; Hsieh, G. C.; Drizin, I.; Milicic, I.; Miller, T. R.; Witte, D. G.; Wishart, N.; Fix-Stenzel, S. R.; McPherson, M. J.; Adair, R. M.; Wetter, J. M.; Bettencourt, B. M.; Marsh, K. C.; Sullivan, J. P.; Honore, P.; Esbenshade, T. A.; Brioni, J. D. J. Med. Chem. 2008, 51, 6547; (d) Cowart, M. D.; Faghih, R.; Curtis, M. P.; Gfesser, G. A.; Bennani, Y. L.; Black, L. A.; Pan, L.; Marsh, K. C.; Sullivan, J. P.; Esbenshade, T. A.; Fox, G. B.; Hancock, A. J. Med. Chem. 2005, 48, 38; (e) Nagase, T.; Mizutani, T.; Ishikawa, S.; Sekino, E.; Sasaki, T.; Fujimura, T.; Ito, S.; Mitobe, Y.; Miyamoto, Y.; Yoshimoto, R.; Tanaka, T.; Ishihara, A.; Takenaga, N.; Tokita, S.; Fukami, T.; Sato, N. J. Med. Chem. 2008, 51, 4780.
- (a) Arrang, J. M.; Garbarg, M.; Schwartz, J. C. Nature **1983**, 302, 832; (b) Lovenberg, T. W.; Roland, B. L.; Wilson, S. J.; Jiang, X.; Pyati, J.; Huvar, A.; Jackson, M. R.; Erlander, M. G. *Mol. Pharmacol.* **1999**, 55, 1101; (c) Bongers, G.; Bakker, R. A.; Leurs, R. *Biochem. Pharmacol.* **2007**, 73, 1195; (d) Wulff, B. S.; Hastrup, S.; Rinvall, K. *Eur. J. Pharmacol.* **2002**, 453, 33; (g) Lin, J.; Sergeeva, O. A.; Haas, H. L. *J. Pharmacol. Exp. Ther.* **2011**, 336, 17; (f) http://clinicaltrails.gov (assessed July 2011).
- 4. Raddatz, R.; Tao, M.; Hudkins, R. L. Curr. Top. Med. Chem. 2010, 10, 153.
- Hudkins, R. L.; Raddatz, R.; Tao, M.; Mathiasen, J. R.; Aimone, L. D.; Becknell, N. C.; Prouty, C. P.; Knutsen, L.; Yazdanian, M.; Moachon, G.; Ator, M. A.; Mallamo, J. P.; Marino, M. J.; Bacon, E. R.; Williams, M. J. Med. Chem. 2011, 54, 4781.
- Tao, M.; Aimone, L. D.; Huang, Z.; Mathiason, J.; Raddatz, R.; Lyons, J.; Hudkins, R. L. J. Med. Chem., in press. Nov. 22. [Epub ahead of print].
- (a) Apodaca, R.; Dvorak, C. A.; Xiao, W.; Barbie, A. J.; Boggs, J. D.; Wilson, S. J.; Lovenberg, T. W.; Carruthers, N. I. J. Med. Chem. 2003, 46, 3938; (b) Stark, H. Expert Opin. Ther. Pat. 2003, 13, 851; (c) Celanire, S.; Wijtmans, M.; Talaga, P.; Leurs, R.; de Esch, I. J. P. Drug Discovery Today 2005, 10, 1613; (d) Dvorak, C. A.; Apodaca, R.; Barbier, A. J.; Berridge, C. W.; Wilson, S. J.; Boggs, J. D.; Xiao, W.; Lovenberg, T. W.; Carruthers, N. I. J. Med. Chem. 2005, 48, 2229; (e) Nagase, T.; Mizutani, T.; Sekino, E.; Ishikawa, S.; Ito, S.; Mitobe, Y.; Miyamoyo, Y.; Yoshimoto, R.; Tanaka, T.; Ishihara, A.; Takenaga, N.; Tokita, S.; Sato, N. J. Med. Chem. 2008, 51, 6889.
- (a) Wermuth, C. G.; Schlewer, G.; Bourguignon, J. J.; Maghioros, G.; Bouchet, M. J.; Moire, C.; Kan, J. P.; Worms, P.; Biziere, K. *J. Med. Chem.* **1989**, *32*, 528; (b) Coates, W. J.; McKillop, A. *Synthesis* **1993**, 334.
- 9. Sugahara, M.; Ukita, T. Chem. Pharm. Bull. 1997, 45, 719.
- Bacon, E. R.; Bailey, T. R.; Becknell, N. C.; Chatterjee, S.; Dunn, D.; Hostetler, G. A.; Hudkins, R. L.; Josef, K. A.; Knutsen, L.; Tao, M.; Zulli, A. L. US2010273779, 2010.
- Hudkins, R. L.; Zulli, A. L.; Dandu, R.; Tao, M.; Josef, K. A.; Aimone, L. D.; Haltwanger, C.; Huang, Z.; Lyons, J. A.; Raddatz, R.; Mathiasen, J. R.; Grunner, J. A. *Bioorg. Med. Chem. Lett.*, submitted for publication.
- Clapham, J.; Kilpatrick, G. J. Eur. J. Pharmacol. **1993**, 232, 99; (b) Medhurst, A. D.; Atkins, A. R.; Beresford, I. J.; Brackenborough, K.; Briggs, M. A.; Calver, A. R.; Cilia, J.; Cluderay, J. E.; Crook, B.; Davis, J. B.; Davis, R. K.; Davis, R. P.; Dawson, L. A.; Foley, A. G.; Gartlon, J.; Gonzalez, M. I.; Heslop, T.; Hirst, W. D.; Jennings, C.; Jones, D. N.; Lacroix, L. P.; Martyn, A.; Ociepka, S.; Ray, A.; Regan, C. M.; Roberts, J. C.; Schogger, J.; Southam, E.; Stean, T. O.; Trail, B. K.; Upton, N.; Wadsworth, G.; Wald, J. A.; White, T.; Witherington, J.; Woolley, M. L.; Worby, A.; Wilson, D. M. J. Pharmacol. Exp. Ther. **2007**, 321, 1032; (c) Le, S.; Gruner, J. A.; Mathiasen, J. R.; Marino, M. J.; Schaffhauser, H. J. Pharmacol. Exp. Ther. **2008**, 325, 902.
- (a) Fox, G. B.; Esbenshade, T. A.; Pan, J. B.; Radek, R. J.; Krueger, K. M.; Yao, B. B.; Browman, K. E.; Buckley, M. J.; Ballard, M. E.; Komater, V. A.; Miner, H.; Zhang, M.; Faghih, R.; Rueter, L. E.; Bitner, R. S.; Drescher, K. U.; Wetter, J.; Marsh, K.; Lemaire, M.; Porsolt, R. D.; Bennani, Y. L.; Sullivan, J. P.; Cowart, M. D.; Decker, M. W.; Hancock, A. A. J. Pharmacol. Exp. Ther. 2005, 313, 176; (b) Medhurst, A. D.; Atkins, A. R.; Beresford, I. J.; Brackenborough, K.; Briggs, M. A.; Calver, A. R.; Cilia, J.; Cluderay, J. E.; Crook, B.; Davis, J. B.; Davis, R. K.; Davis, R. P.; Dawson, L. A.; Foley, A. G.; Garlton, J.; Gonzalez, M. I.; Heslop, T.; Hirst, W. D.; Jennings, C.; Jones, D. N.; Lacroix, L. P.; Martyn, A.; Ociepka, S.; Ray, A.; Regan, C. M.; Roberts, J. C.; Schogger, J.; Southam, E.; Stean, T. O.; Trail, B. K.; Upton, N.; Wadsworth, G.; Wald, J. A.; White, T.; Witherington, J.; Woolley, M. L.; Worby, A.; Wilson, D. M. J. Pharmacol. Exp. Ther. 2007, 321, 1032; (c) Raddatz, R.; Hudkins, R. L.; Mathiasen, J. R.; Gruner, J. A.; Flood, D. G.; Aimone, L. D.; Le, S.; Schaffhauser, H.; Gasior, M.; Bozyczko-Coyne, D.; Marino, M. J.; Ator, M. A.; Bacon, E. R.; Mallamo, J. P.; Williams, M. J. Pharmacol. Exp. Ther. 2011, 340, in press. doi:10.1124/jpet.111.186585.
- Le, S.; Grunner, J. A.; Mathiasen, J. R.; Marino, M. J.; Schaffhauser, H. J. Pharmacol. Exp. Ther. 2008, 325, 902.
- 15. Edgar, D. M.; Seidel, W. F. J. Pharmacol. Exp. Ther. 1997, 283, 757.