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# Biological potential of novel methoxy and hydroxy substituted heteroaromatic amides designed as promising antioxidative agents: Synthesis, 3D-QSAR analysis and biological activity

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## Just Accepted

# Biological potential of novel methoxy and hydroxy substituted heteroaromatic amides designed as promising antioxidative agents: Synthesis, 3D-QSAR analysis and biological activity

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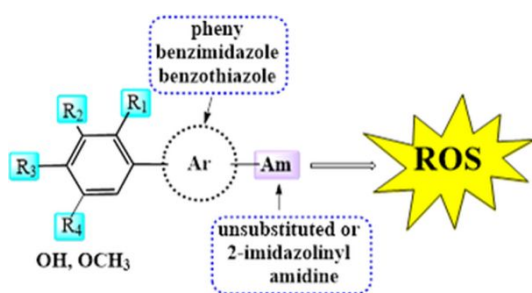
**Abstract**

This paper discusses antioxidative and biological activity of 25 novel amidino substituted benzamides with a variety of heteroaromatic nuclei attached to the benzamide moiety and with a variable number of methoxy and/or hydroxy substituents. Targeted compounds, bearing either amidino or 2-imidazolinyl substituent, were obtained in the Pinner reaction from cyano precursors. 3D-QSAR models were generated to predict antioxidative activity of the 25 novel aromatic and heteroaromatic benzamide derivatives. The compounds were tested for antioxidative activity using *in vitro* spectrophotometric assays. Direct validation of 3D-QSAR approach for predicting activities of novel benzamide derivatives was carried out by comparing experimental and computationally predicted antioxidative activity. Experimentally determined activities for all novel compounds were found to be within standard deviation of error of the models. Following this, structure-activity relationships among the synthesized compounds are discussed. Furthermore, antiproliferative activity *in vitro* against HeLa cells as well as antibacterial and antifungal activity was tested to confirm the other biological activities of the prepared compounds.

**Key words:** amides, benzimidazoles, benzothiazoles, 3D-QSAR, antioxidative activity, antiproliferative activity *in vitro*, antimicrobial activity

## 1. Introduction

Significant biological importance of nitrogen heterocycles, particularly benzimidazoles and benzothiazoles, makes them very useful building blocks in organic and medicinal chemistry.[1,2] They are an essential structural part in numerous natural and synthetic bioactive compounds[3] possessing a wide range of biological characteristics including antibacterial,[4,5] anticancer,[6,7] antifungal,[8] antiviral,[9] antioxidative[10,11] *etc*. In the last decade, particular attention and focus was given to their antioxidative potency. It is widely accepted that reactive oxygen species (ROS) could cause the oxidative damage of important biomolecules including nucleic acids, proteins or lipids. These reactive species could be generated in numerous pathophysiological and biochemical processes in the human body.[12] ROS includes non-free radical species like ozone ($O_3$), singlet oxygen ($^1O_2$), hydrogen peroxide ($H_2O_2$), and free radicals including peroxyl radicals ($ROO^\bullet$), hydroperoxyl radicals ($HOO^\bullet$), hydroxyl radicals ($OH^\bullet$) and superoxide anion radicals ($O^\bullet_2$). Consequently, it could lead to the development of some chronic diseases like cancer, diabetes, inflammation, neurodegenerative syndrome or aging.[13,14] The proper usage of antioxidants could regulate the rate of reactive oxygen species production, at even at low concentration, they prevent oxidation of easily oxidizable substrate.[15] Thus, there is a constant growing interest for the study of antioxidative capacity of numerous natural and synthetic organic molecules with the aim of prevention of the above mentioned diseases.

Recently, our scientific research focused on the antiproliferative activity of versatile benzimidazole and benzothiazole derivatives.[16,17,18] A large number of active derivatives are designed to have cationic amidino substituents at the termini of the molecule, which has allowed additional interactions with the biological targets. Additional biological activity studies of tetracyclic benzimidazole derivatives have been confirmed their strong interaction with DNA as intercalators, while acyclic, either benzimidazole or benzothiazole, derivatives have been proven to be selective DNA minor groove binders.[16,17,19] Recent literature data suggests that tumor cells are accompanied by a measurable emission of ROS which might be regulated by apropriate usage of antioxidant agents.[20] Several publications described antioxidative activity and potential of benzimidazole and benzothiazole derivatives, as well as a broad range of salicylanilide and benzamide derivatives, as antioxidative agents.[21,22,23]

In order to explore the antioxidative potency of aromatic and heteroaromatic benzamide derivatives bearing benzazole nuclei, we developed 3D-QSAR models and used them for computational prediction of antioxidative activity of the presented compounds.

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Application of 3D-QSAR modeling for prediction of activity of novel compounds and identification of molecular properties with the highest influence on the studied activity was already proven as an useful method numerous times.[24,25,26,27,28,29] In addition, antioxidative activity of different classes of compounds has been studied using different approaches of QSAR modeling.[30,31,32,33] The main advantage of Volsurf+[34,35] based approach is usage of molecular descriptors that are independent of molecular alignment and have a clear chemical or physical meaning. In this paper, Volsurf+ approach for generating 3D-QSAR models was experimentally validated on prediction of antioxidative activity of the 25 novel compounds. Combining the demonstrated biological potential of benzamides and amidino substituted benzimidazoles and benzothiazoles, we have designed and synthesized novel derivatives bearing a variable number of the methoxy and hydroxy groups substituted with different amidino substituents. Prepared derivatives were explored for their antioxidative, antimicrobial and antiproliferative activity. The obtained experimental results were compared to the predicted results and a very good alignment was observed.

## 2. Results and Discussion

### 2.1. 3D-QSAR models

Using a library of experimental data for antioxidative activity[36,37,38] of already published compounds, 3D-QSAR models were built. Models with the highest predictive ability, namely model **1** and model **2**, were chosen for the prediction of antioxidative activities of novel compounds. Model **1** was generated using the results obtained from the activity testing against DPPH stable radical while model **2** has been generated using activity from superoxide test. Quality of the models validated by internal cross-validation procedure, using two procedures, leave one out (LOO) and random groups (LMO), as well as by the external prediction (Table 1, Figure 1). Both models show similar predictive ability ($Q^2_{LOO}$, $Q^2_{LMO}$, $SDEP_{EXT}$ in Table 1). Statistical properties of the models are shown in Table 1. Compounds for which activity was expressed only as "≥400 µM" were not included in the training set of the model **2** due to the possible largely decreased quality of model.

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**Table 1.** Statistical properties of 3D-QSAR models.

| Model | nO | LV | $R^2$ | SDEC | $Q^2_{LOO}$ | $SDEP_{LOO}$ | $Q^2_{LMO}$ | $SDEP_{LMO}$ | $SDEP_{EXT}$ | nOE |
|-------|-----|-----|-------|------|-------------|--------------|-------------|--------------|--------------|-----|
| **1** | 59 | 4 | 0.73 | 0.25 | 0.66 | 0.28 | 0.65 | 0.28 | 0.41 | 12 |
| **2** | 32 | 3 | 0.71 | 0.12 | 0.54 | 0.16 | 0.53 | 0.16 | 0.18 | 7 |

[a] Number of objects which was used to build the model (training set). [b] Number of latent variables. [c]SDEC - standard deviation of error of calculation. [d] $Q^2$ is the cross-validated predictive performance, LOO stands for Leave One Out cross-validation procedure, LMO stands for Leave Many Out (*i.e.* Random Groups) cross-validation procedure. [e]SDEP - standard deviation of error of prediction obtained by cross-validation. [f] $SDEP_{EXT}$ stands for standard deviation of error of prediction obtained by external validation. [g] Number of objects used for external prediction validation (test set).
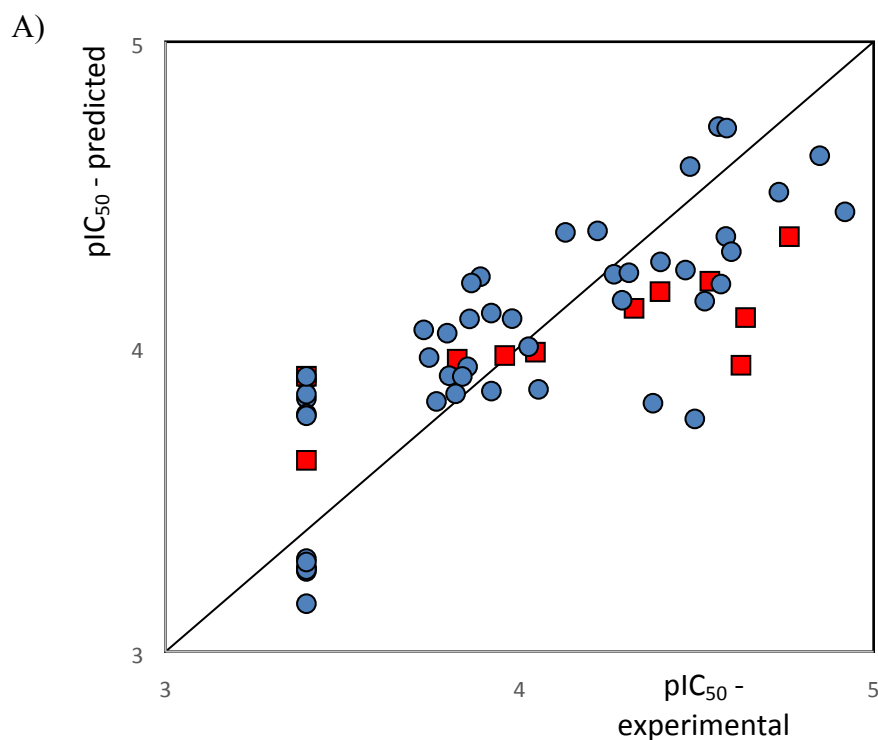
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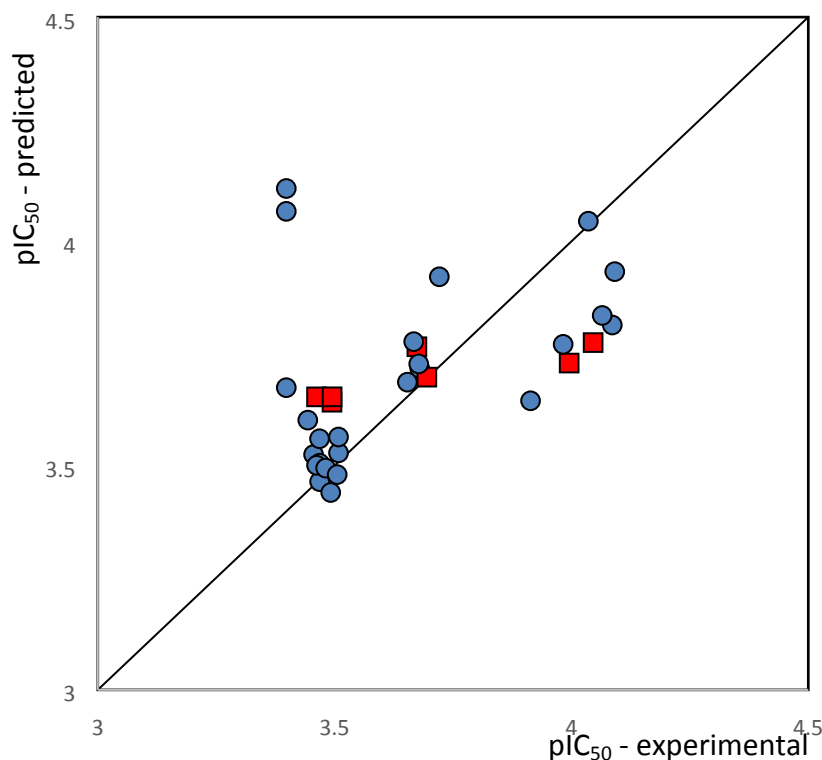


**Figure 1.** Predicted *vs* experimental antioxidative activity expressed as $pIC_{50}$ − negative logarithmic value of concentration that causes 50% of antioxidative activity measured: A) against DPPH stable radical (model **1**) and B) using superoxide test (model **2**). Blue circles present training set compounds; red squares present test set compounds.

### 2.1.2. 3D-QSAR analysis

Principal component analysis (PCA) was performed on the overall dataset of 59 compounds. The first two principal components explained 95% of variance (the first three PCs explained 98% of variance) of the descriptors (X-matrix). Molecular descriptors including hydrophilic (W1-W3) and hydrophobic regions (D1) related to the polarizability and dispersion forces, molecular volume (V), and possibility of accepting (WN1) and donating H-bonds (WO1) were identified, from the PCA loadings plot (Figure S63 in the Supplement), as the ones with the highest variation among the dataset compounds

These results indicated to the mentioned descriptors as the ones with the highest variation between the dataset compounds which might be related to the variation in antioxidative activities of the dataset compounds. The PLS analysis could estimate the quantitative impact of each descriptor on studied activities.

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The influence of each molecular descriptor to the compound's antioxidative activity could be evaluated from the analyses of the PLS coefficients of the obtained 3D-QSAR. Since both models were generated using raw data, the real impact of a molecular descriptor on the antioxidative activity is given as the product of the descriptor's value and it's PLS coefficient. Such QSAR analysis performed for both models (Figure 2), enabled the finding of molecular properties with the highest positive or negative impact on antioxidative activity.
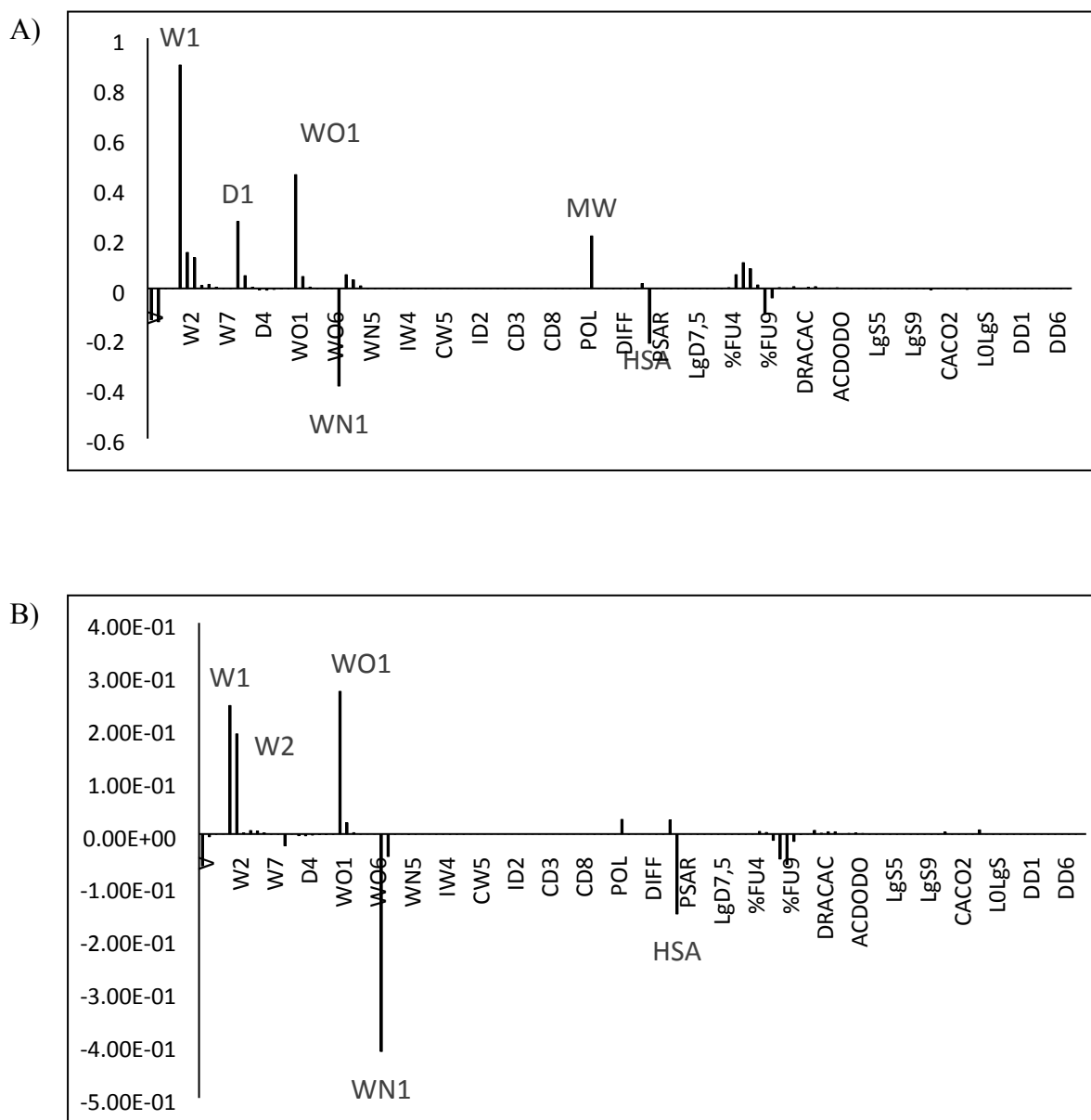


**Figure 2**. Products of the descriptor's average value calculated for the dataset used to derive model and the associated PLS coefficient of 3D-QSAR model for: A) model **1** and B) model **2**.

Descriptors with the highest positive influence on antioxidative activity for both models have proven to be hydrophilic volume related to polarizability (W1) and possibility of a compound to donate H-bond (WO1). Consequently, their increase should lead to increase of activity measured by both tests. Possibility of a compound to accept H-bond (WN1) and sum of hydrophobic surface areas (HSA) are negatively correlated for both models and their decrease should cause the increase of antioxidatve activity in both tests.

## 2.2. Chemistry

All chosen amide derivatives were synthesized following two synthetic procedures shown in Scheme 1 and 2.[39] Cyano substituted benzamides **4-6** and 2-benzimidazolyl/2-benzothiazolyl substituted benzamides **20-25** were obtained from the methoxy substituted acyl-halogenides **1-3** and *p*-cyanoaniline **2** or cyano substituted 2-aminobenzimidazole **18** or 2-aminobenzothiazole **19** prepared by previously published synthetic procedures.[40]
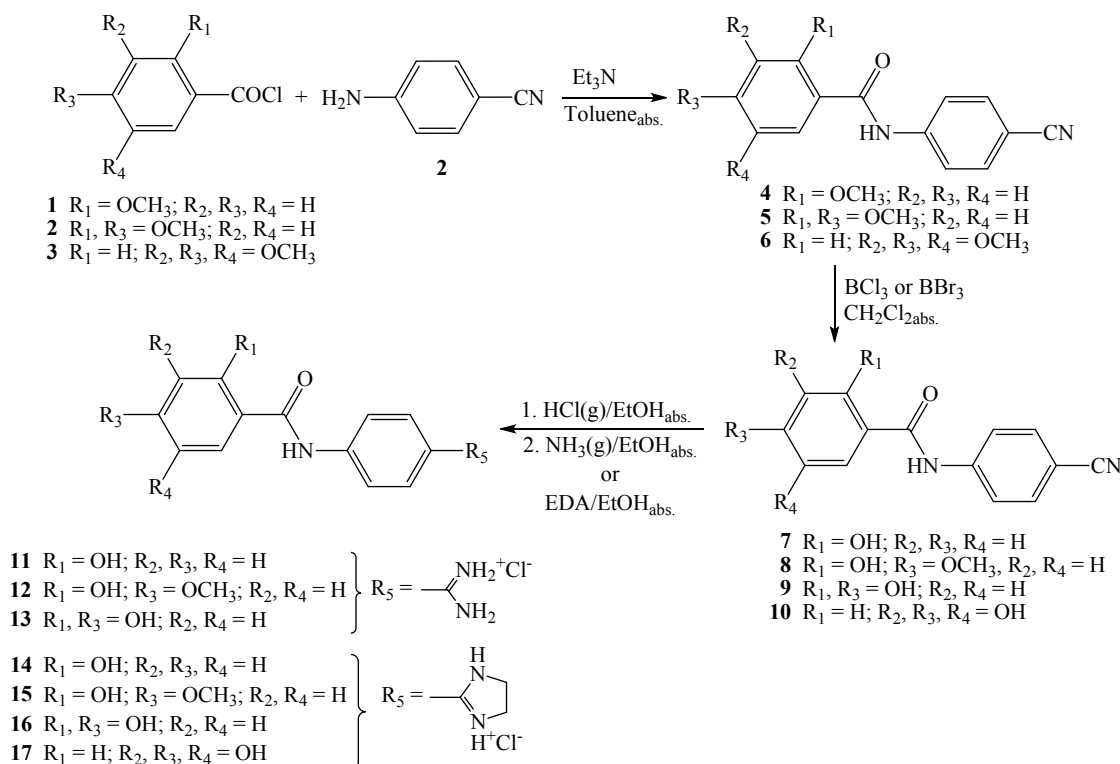


**Scheme 1.**

In order to obtain corresponding hydroxy substituted benzamides **7-10** and **26-29** bearing cyano substituent, the removal of methoxy protecting groups was accomplished by using either boron

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trichloride or boron tribromide in absolute dichloromethane at -75 °C. Reaction yields of all compounds were moderate.

By using boron trichloride, only one methoxy group has been removed from the compounds **5**, **21** and **24** which allowed the synthesis of compounds **8**, **27** and **29** with one methoxy and one hydroxy group. 2D NOESY NMR spectra confirmed that the methoxy group at the position 2 ($R_1$) was successfully deprotected. Target compounds with variable number of hydroxy **11-17** and **30-37** and/or methoxy groups **38-47** bearing amidino or 2-imidazolinyl substituent were prepared in acidic twostep Pinner reaction from corresponding cyano precursors due to reaction with $NH_{3(g)}$ or ethylenediamine (EDA).



**1** $R_1 = OCH_3$; $R_2, R_3, R_4 = H$
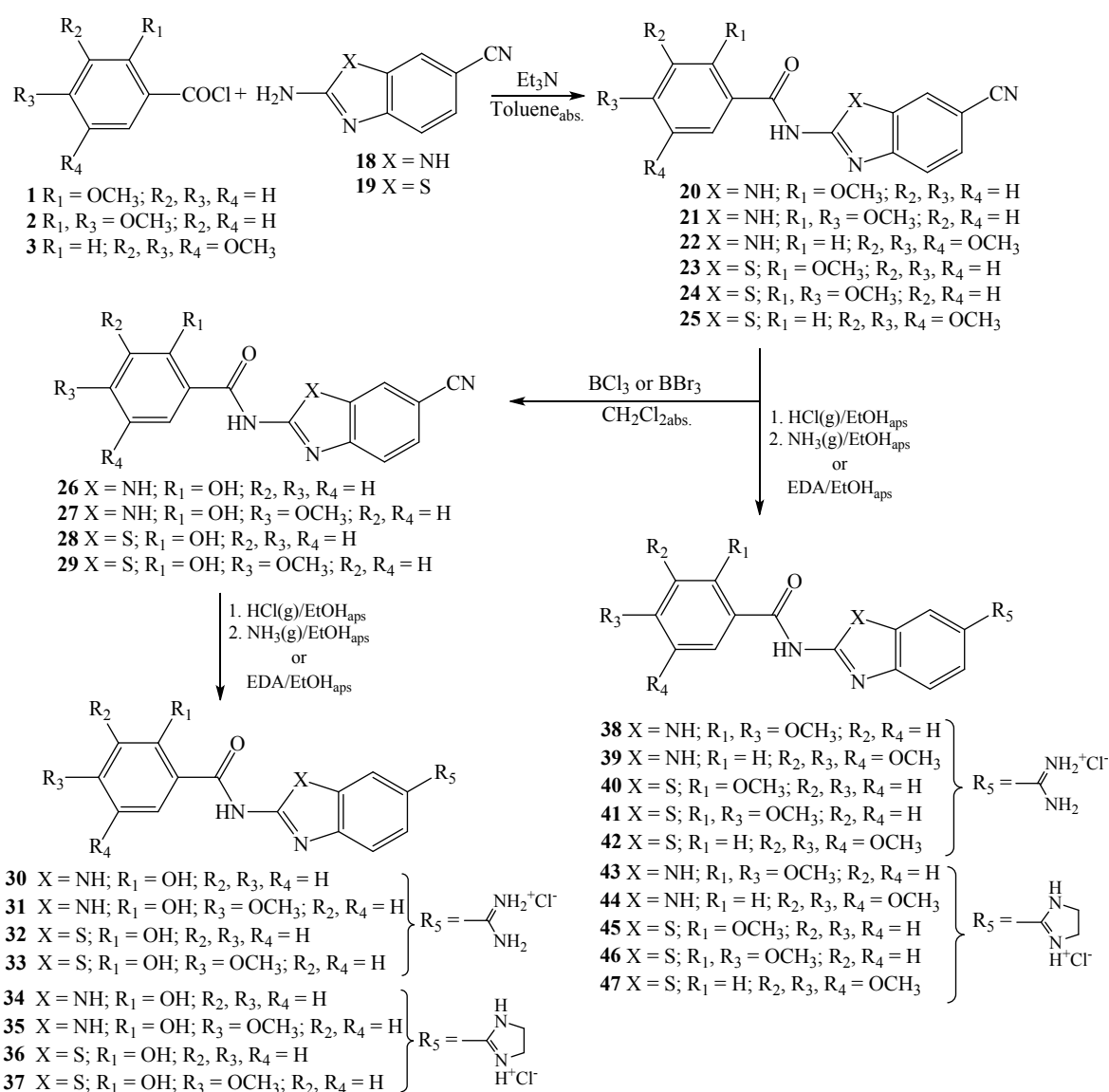**2** $R_1, R_3 = OCH_3$; $R_2, R_4 = H$
**3** $R_1 = H$; $R_2, R_3, R_4 = OCH_3$

**18** X = NH
**19** X = S

**20** X = NH; $R_1 = OCH_3$; $R_2, R_3, R_4 = H$
**21** X = NH; $R_1, R_3 = OCH_3$; $R_2, R_4 = H$
**22** X = NH; $R_1 = H$; $R_2, R_3, R_4 = OCH_3$
**23** X = S; $R_1 = OCH_3$; $R_2, R_3, R_4 = H$
**24** X = S; $R_1, R_3 = OCH_3$; $R_2, R_4 = H$
**25** X = S; $R_1 = H$; $R_2, R_3, R_4 = OCH_3$

**26** X = NH; $R_1 = OH$; $R_2, R_3, R_4 = H$
**27** X = NH; $R_1 = OH$; $R_3 = OCH_3$; $R_2, R_4 = H$
**28** X = S; $R_1 = OH$; $R_2, R_3, R_4 = H$
**29** X = S; $R_1 = OH$; $R_3 = OCH_3$; $R_2, R_4 = H$

**38** X = NH; $R_1, R_3 = OCH_3$; $R_2, R_4 = H$
**39** X = NH; $R_1 = H$; $R_2, R_3, R_4 = OCH_3$
**40** X = S; $R_1 = OCH_3$; $R_2, R_3, R_4 = H$
**41** X = S; $R_1, R_3 = OCH_3$; $R_2, R_4 = H$
**42** X = S; $R_1 = H$; $R_2, R_3, R_4 = OCH_3$

**43** X = NH; $R_1, R_3 = OCH_3$; $R_2, R_4 = H$
**44** X = NH; $R_1 = H$; $R_2, R_3, R_4 = OCH_3$
**45** X = S; $R_1 = OCH_3$; $R_2, R_3, R_4 = H$
**46** X = S; $R_1, R_3 = OCH_3$; $R_2, R_4 = H$
**47** X = S; $R_1 = H$; $R_2, R_3, R_4 = OCH_3$

**30** X = NH; $R_1 = OH$; $R_2, R_3, R_4 = H$
**31** X = NH; $R_1 = OH$; $R_3 = OCH_3$; $R_2, R_4 = H$
**32** X = S; $R_1 = OH$; $R_2, R_3, R_4 = H$
**33** X = S; $R_1 = OH$; $R_3 = OCH_3$; $R_2, R_4 = H$

**34** X = NH; $R_1 = OH$; $R_2, R_3, R_4 = H$
**35** X = NH; $R_1 = OH$; $R_3 = OCH_3$; $R_2, R_4 = H$
**36** X = S; $R_1 = OH$; $R_2, R_3, R_4 = H$
**37** X = S; $R_1 = OH$; $R_3 = OCH_3$; $R_2, R_4 = H$

**Scheme 2.**

The progress of the reaction was monitored by IR spectroscopy. In order to ensure their better water solubility, all amidino substituted derivatives were prepared as hydrochloride salts. Due

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to some synthetic issues in the Pinner reaction, monomethoxy substituted analogues of amidino compounds **38-47** have not been successfully synthesized.

## 2.3. Antioxidative activity

Results obtained from DPPH test revealed that all tested compounds displayed improved or similar activity in comparison to used standard, ascorbic acid (Table 2).

**Table 2.** Antioxidative activity of tested amidino substituted compounds.

| Compound | DPPH ($\mu$M) | Superoxide test ($\mu$M) |
|---|---|---|
| **11** | 59±4 | 137±4 |
| **12** | 66±5 | 158±8 |
| **13** | 39±3 | 123±4 |
| **14** | 89±6 | 170±8 |
| **15** | 70±6 | 156±7 |
| **16** | 51±6 | 138±5 |
| **17** | 42±5 | 133±7 |
| **30** | 42±3 | 135±4 |
| **31** | 38±3 | 130±5 |
| **32** | 49±4 | 139±5 |
| **33** | 46±7 | 141±6 |
| **34** | 56±5 | 148±6 |
| **35** | 65±6 | 163±4 |
| **36** | 80±3 | 166±3 |
| **37** | 58±3 | 154±5 |
| **38** | 59±6 | 142±8 |
| **39** | 52±3 | 145±4 |
| **40** | 66±7 | 149±7 |
| **41** | 73±4 | 171±7 |
| **42** | 57±2 | 151±6 |
| **43** | 71±5 | 163±7 |
| **44** | 85±4 | 180±9 |
| **45** | 82±4 | 169±5 |
| **46** | 94±2 | 195±9 |
| **47** | 73±4 | 172±8 |
| **Ascorbic acid** | 79±3 | 95±2 |

Interestingly, a regularly behavior was noticed in the pairs of tested bioisosteric compounds. The radical scavenging test for the compounds with amidino moiety always resulted in ehanced

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activity when compared to compound having imidazole moiety in the same position (pairs are as goes: **11** and **14**, **30** and **34**, **32** and **36**, **40** and **45**, **12** and **15**, **31** and **35**, **33** and **37**, **38** and **43**, **41** and **46**, **13** and **16**, **39** and **44**, **42** and **47**).

According to the presented results, the compound with the highest antioxidative potential has proven to be **13**, with the activity almost 50% better in comparison to the control compound (ascorbic acid). The pair of compounds including **13** and **16**, is the only one having two hydroxy groups in positions 2 and 4, and thus has a highest potential for radical transfer to stable DPPH radical and still maintain his stability by resonance stabilization. Compared to compound **13**, its bioisosteric compounds **11**, having hydroxy group on first phenyl ring in position 2, and compound **12**, with hydroxy and methoxy groups placed in positions 2 and 4 respectively, showed significantly lower activity, although still better than control compound. From this result we can conclude that hydroxy groups are important for activity in order to promote radical scavenging activity. This is in accordance with QSAR analysis showing that the values of descriptors WO1 and W1 are higher for compounds **11-13** comparing to ascorbic acid, while the opposite is true for the descriptors WN1 and HSA (Figure 3).

In second performed antioxidative test, which included measuring capability of compounds for superoxide anion radical scavenging, all compounds showed lower activity compared to ascorbic acid. Nevertheless, compound **13** is the most active in this test as well.



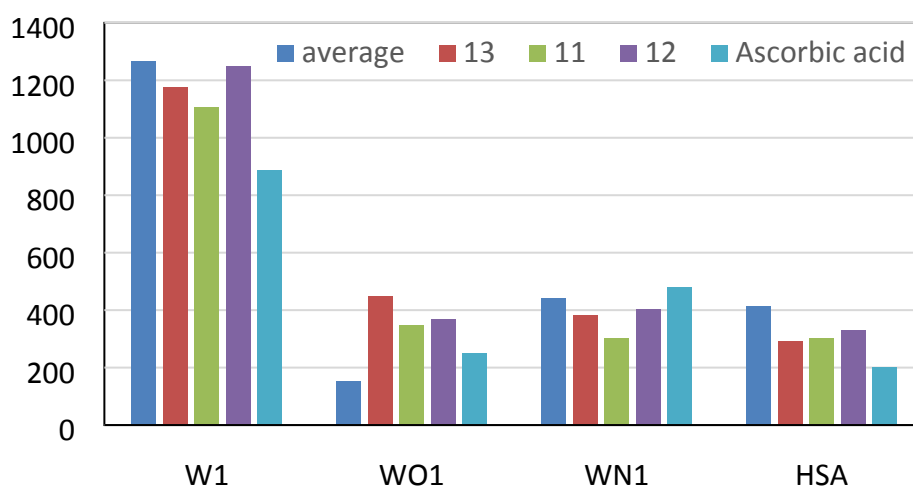**Figure 3.** Comparison of values of the descriptors with the highest influence on activity for the compounds **11-13**, average value and ascorbic acid.

## 2.4. Comparison of predicted and experimental antioxidative activities

Internal validation using cross validation procedure, as well as validation by external prediction, are the most often used methods for validation of QSAR models.
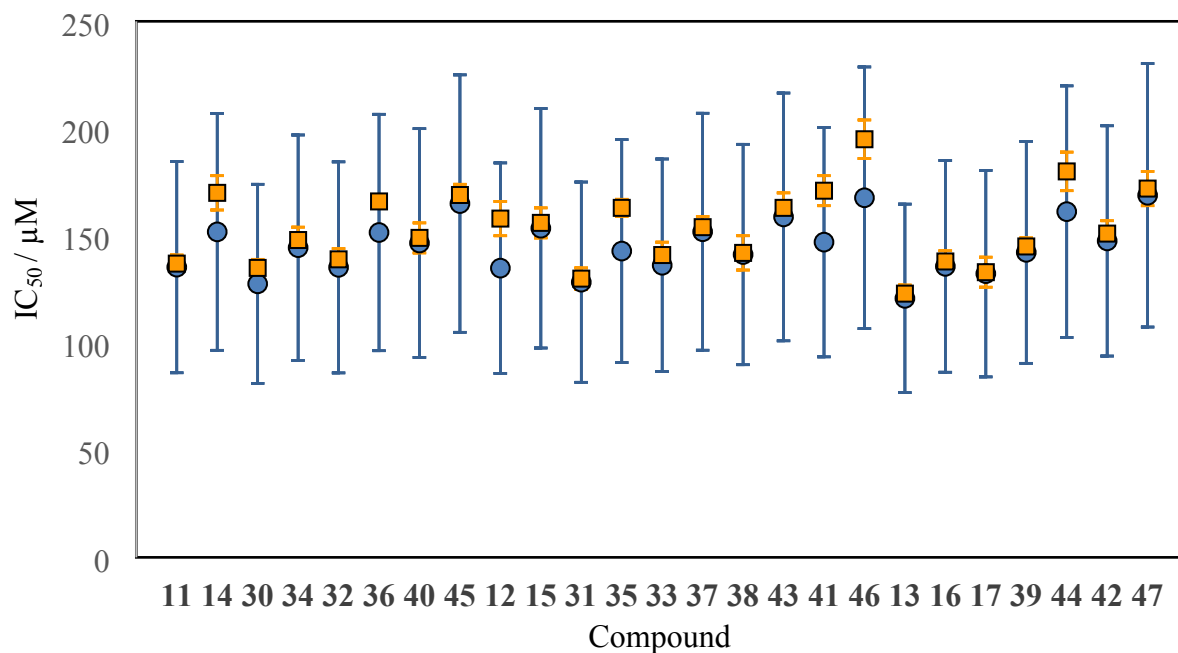
A)
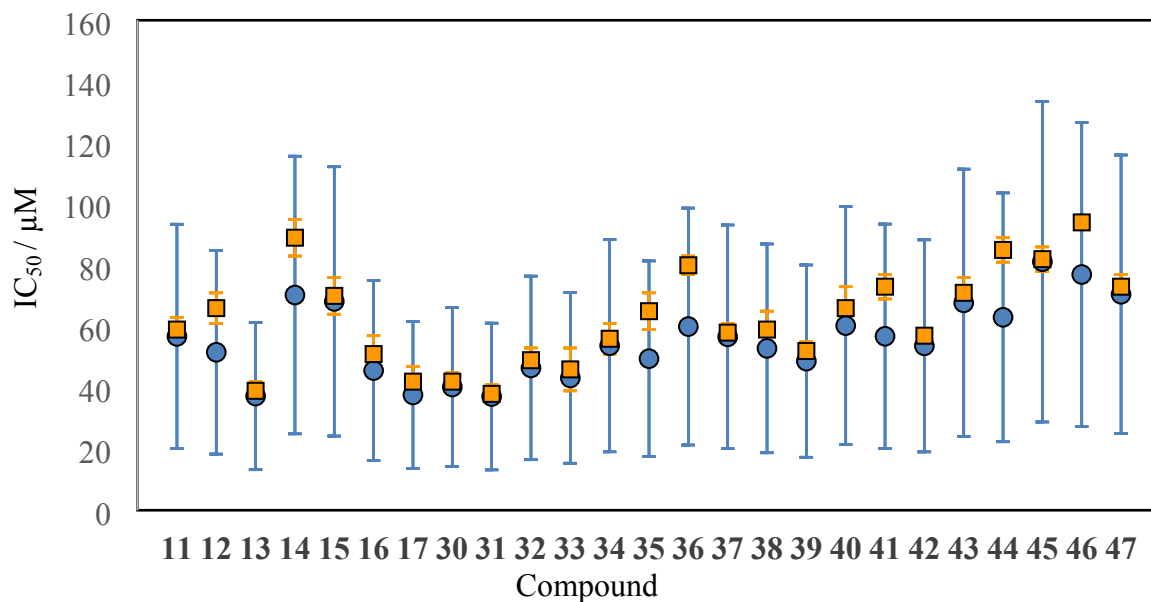


B)

**Figure 4.** Comparison of experimental (orange squares) and computationally predicted (blue circles) antioxidative activities of the synthetized compounds expressed as $IC_{50}$. Error bars coloured blue on the blue circles present standard deviation of error of prediction (SDEP) of the 3D-QSAR model used to predict activities: A) model **1** and B) model **2**, while orange error bars on orange squares present standard deviation of experimental measurements.

However, the experimental measurements of the activities of novel compounds whose activities were previously predicted by the QSAR model should be the most direct and reliable method for validation of prediction ability of built QSAR model. The antioxidative activities of 25 novel compounds (**11-17** and **30-47)**, were predicted by using model **1** and model **2**. Therefore, experimental measurements of their antioxidative activity enabled direct validation of the obtained 3D-QSAR models (Table 3 and Figure 4). Antioxidative activities of all 25 newly synthesised compounds are correctly predicted by both models *i.e.* experimentally measured activities for all novel compounds are within standard deviation of error of prediction (SDEP) of the models estimated by cross-validation procedure (Figure 3). Correlation coefficients ($R^2$) between experimental and predicted values of antioxidative activities were 0.82 and 0.78 for the models **1** and **2**, respectively. Furthermore, comparison of experimental and predicted activities shows that for majority of the novel compounds (18 out of 25) predicted antioxidative activity is within experimental error (Figure 4). For 7 compounds for which discrepancy of predicted and experimental activity is larger than standard deviation of experimental measurements, predicted values are smaller than experimental, showing that models slightly underestimated antioxidative activity of these compounds.

## 2.4. Antiproliferative, antibacterial and antifungal activity

Additionally, to explore and confirm the biological potential of newly prepared compounds, antiproliferative activity *in vitro*, antibacterial and antifungal activities were evaluated. As the antiproliferative potency of suchlike compounds was previously confirmed in our published papers, the synthesized compounds were tested against HeLa cancer cells. Obtained results are depicted in the Table 3 while cisplatin was used as a standard anticancer drug. Obtained results indicated that the majority of tested compounds showed moderate to low activity at micromolar inhibition concentrations.

In general, it could be concluded that benzothiazole derivatives showed stronger antiproliferative activity in comparison to their benzimidazole analogues with the exception of

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2,4-dimethoxy substituted benzimidazole **43** bearing 2-imidazolinyl group. Thus, the strongest antiproliferative activity against HeLa cells displayed 2-hydroxy **36** ($IC_{50}$ = 23.0 µM) and 2,4-dimethoxy **45** ($IC_{50}$ = 13.6 µM) substituted benzothiazoles bearing 2-imidazolinyl group and 2-methoxy **40** ($IC_{50}$ = 20.1 µM) and 3,4,5-trimethoxy **42** ($IC_{50}$ = 32.8 µM) substituted benzothiazoles bearing amidino substituent. All tested amidino substituted benzamides **11-17** showed very low antiproliferative activity.

**Table 3.** Antiproliferative activity *in vitro* against HeLa cells.

| Comp. | $IC_{50}$ (µM) |
|---|---|
| **11** | >200 |
| **12** | >200 |
| **13** | 172.9±15.44 |
| **14** | >200 |
| **15** | >200 |
| **16** | 195.1±6.99 |
| **17** | 78.0±2.22 |
| **30** | 167.2±10.93 |
| **31** | 133.6±12.25 |
| **32** | 192.0±7.99 |
| **33** | 41.4±5.15 |
| **34** | 194.6±7.65 |
| **35** | >200 |
| **36** | 23.0±0.40 |
| **37** | >200 |
| **38** | 129.2±5.61 |
| **39** | 175.6±1.28 |
| **40** | 20.1±5.71 |
| **41** | >200 |
| **42** | 32.8±0.21 |
| **43** | 40.9±7.57 |
| **44** | 117.4±10.90 |
| **45** | 13.6±1.24 |
| **46** | >200 |
| **47** | 55.4±0.54 |
| **Cisplatin** | 2.5±0.24 |

Furthermore, the *in vitro* antibacterial activity of the prepared compounds was evaluated against eight different bacterial strains fully sensitive to antibiotics. Gram-positive bacterial strains

included key representatives of human pathogens including *S. aureus*, *M. luteus*, *B. subtilis* and *C. sporogenes*, while the panel of Gram-negative bacteria included *E. coli*, *P. hauseri*, *P. aeruginosa* and *K. pneumonia*. Obtained results were displayed in Table 4 as the minimal inhibitory concentrations (MICs) and are compared to antibiotic amikacin used as a standard. In Table 4 are shown only results for compounds which showed some activity.

**Table 4.** Antibacterial activity of tested compounds.

| Comp. | MIC (mM) | | | | | | | |
|---|---|---|---|---|---|---|---|---|
| | *P. hauseri* | *P. aeruginosa* | *E. coli* | *K. pneumoniae* | *M. luteus* | *S. aureus* | *B. subtilis* | *C. sporogenes* |
| **11** | 4.23 | 2.14 | 2.14 | 2.14 | 4.23 | 2.14 | 0.13 | 0.27 |
| **13** | 2.03 | 2.03 | 2.03 | 4.06 | 4.06 | 2.03 | 1.02 | 2.03 |
| **17** | 0.89 | 0.89 | 1.77 | 0.89 | 1.77 | 1.77 | 0.89 | 0.89 |
| **31** | 3.46 | 3.46 | 3.46 | 3.46 | 6.91 | 6.91 | 3.46 | 3.46 |
| **32** | 7.17 | 7.17 | 7.17 | 7.17 | 7.17 | 7.17 | 7.17 | 7.17 |
| **34** | 6.99 | 3.50 | 6.99 | 6.99 | 6.99 | 6.99 | 6.99 | 3.50 |
| **39** | 0.37 | 0.37 | 0.77 | 0.77 | 0.77 | 0.77 | 0.37 | 1.54 |
| **43** | 3.11 | 6.22 | 6.22 | 6.22 | 6.22 | 3.11 | 1.56 | 0.78 |
| **47** | 1.39 | 1.39 | 0.70 | 1.39 | 1.39 | 1.39 | 0.70 | 1.39 |
| **Amikacin** | 0.012 | 0.085 | 0.009 | 0.003 | 0.014 | 0.019 | 0.072 | 0.026 |

The obtained results revealed that the compound with the most significant antibacterial activity proved to be 3,4,5-trimethoxy substituted benzimidazole derivative **39** bearing amidino group (MICs value 0.37-1.54 µg/mL) without conspicuous selectivity between Gram-negative and Gram-positive bacteria. The highest activity was observed against *P. hauseri*, *P. aeruginosa* and *B. subtilus* bacteria with MIC value 0.37 µg/mL. 3,4,5-trihydroxy substituted benzamide derivative **17** bearing 2-imidazolinyl group showed also promising antibacterial activity with MICs values 0.89-1.77 µg/mL. Interestingly, compounds **36**, **40**, **42** and **45** which showed the most pronounced antiproliferative activity were not active on tested bacterial strains at all. In addition, the antifungal activity of prepared compounds was also tested against tree strains including *C. albicans*, *A. brasiliensis* and *S. cerevisiae*. The obtained results for compounds which displayed some activity are presented in Table 5 including the nystatin which was used as a standard antifungal drug.

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**Table 5.** Antifungal activity of tested compounds.

| Comp. | MIC (mM) | | |
|---|---|---|---|
| | *C. albicans* | *A. brasiliensis* | *S. cerevisiae* |
| **11** | 0.54 | 1.07 | 0.54 |
| **12** | 0.25 | 1.94 | 0.25 |
| **13** | 0.26 | 8.12 | 0.51 |
| **15** | 0.90 | 1.80 | 0.45 |
| **16** | 0.12 | 0.94 | 0.12 |
| **17** | 0.19 | 0.94 | 0.19 |
| **31** | 0.22 | 1.73 | 0.11 |
| **32** | 0.90 | 1.79 | 1.79 |
| **33** | 0.82 | 3.30 | 0.41 |
| **34** | 0.11 | 1.75 | 0.11 |
| **38** | 0.21 | 0.83 | 0.11 |
| **39** | 0.19 | 1.54 | 0.39 |
| **40** | 0.11 | 3.45 | 0.43 |
| **43** | 0.20 | 0.78 | 0.20 |
| **44** | 0.18 | 1.44 | 0.18 |
| **45** | 0.20 | 3.21 | 0.10 |
| **47** | 0.69 | 1.34 | 0.35 |
| **Nystatin** | 2.70 | 1.35 | 1.35 |

In general, obtained results revealed that some tested compounds showed better and improved antifungal activity in comparison to antibacterial activity. The most pronounced and significantly improved antifungal activity of tested compounds was observed against *C. albicans* as well as against *S. cerevisiae* strains compared to the standard drug nystatin. Only 2-hydroxy substituted benzothiazole derivative bearing amidino group **32** showed weaker antifungal activity against *S. cerevisiae* in comparison to nystatin. The most active compounds with high antifungal potential have proven to be 2-hydroxy substituted benzimidazole **34** and 2,4-dihydroxy substituted benzamide **16** both bearing 2-imidazolinyl group with MICs values 0.11 and 0.12 μg/mL. Also, among all compounds showing antifungal activity, benzimidazole derivatives displayed more pronounced antifungal activity in comparison to their benzothiazole analogues. In conclusion, it is obvious that this type of amidino substituted benzimidazole and benzothiazole derivatives have a high potential for further optimization and design of very efficient antifungal agents.

## 3. Conclusion

Within this paper, we present the design and synthesis of 25 novel amidino substituted benzamides that differ in the heteroaromatic nuclei attached to the benzamide moiety as well as in the number of methoxy and hydroxy groups. All synthesized benzamide derivatives, were also, besides with methoxy and hydroxy groups, substituted with either amidino or 2-imidazolinyl group and are prepared within the acidic Pinner reaction from corresponding cyano substituted precursors. In order to ensure better solubility, all amidino derivatives were prepared as their hydrochloride salts.

For prediction of the antioxidative activities of 25 novel compounds, 3D-QSAR models were generated and validated. Comparison of computationally predicted and experimentally obtained activities indicated that 3D-QSAR method can be successfully applied for design of novel methoxy and hydroxy substituted heteroaromatic amides as well as to successfully predict their antioxidative potential.

The experimental evaluation of antioxidative activity revealed that hydroxy groups are essential and promote radical scavenging activity. The most pronounced antioxidative activity was observed for the compound **13**. DPPH test showed that all novel compounds showed improved or similar antioxidative activity compared to standard, ascorbic acid. Furthermore, the majority of tested compounds displayed moderate to low antiproliferative activity at micromolar inhibitory concentrations. In general, it could be concluded that benzothiazole derivatives showed stronger antiproliferative activity in comparison to their benzimidazole analogues. The strongest antiproliferative activity against HeLa cancer cells showed the compound **45**. Additionally, to confirm the great biological potential of suchlike derivatives, the antibacterial and antifungal activities were evaluated. The compound with the most significant *in vitro* antibacterial activity proved to be 3,4,5-trimethoxy substituted benzimidazole derivative **39**. Antifungal activity of the tested compounds was significantly better than their antibacterial activity, while the majority of tested compounds showed improved activity in comparison to standard.

To conclude, all presented results of biological activities evaluation, confirmed the high biological potential of amidino substituted benzimidazole and benzothiazole derivatives, with several compounds which could be chosen as lead compounds for a further structure modification and optimization of suchlike scaffold in order to developed more efficient biologically active molecules with enhanced antioxidant potential.

## 4. Experimental part

### 4.1. 3D-QSAR modelling

#### 4.1.1. Dataset

3D-QSAR models were generated using the data on antioxidative activities of methoxy and hydroxy substituted heteroaromatic amides found in the literature.[41,42] For each compound, antioxidative activities were measured using two tests, DPPH stable radical and superoxide tests, and expressed as $IC_{50}$ values *i.e.* concentration that causes 50% antioxidative activity. For purpose of generating 3D-QSAR models, $pIC_{50}$ (negative logarithm of $IC_{50}$ value expressed in mol dm$^{-3}$) were calculated. Dataset for model **1**, generated using activities against DPPH stable radical, consisted of 59 compounds from the literature, while dataset for model **2**, generated using activities obtained with superoxide test, consisted of 32 compounds from the literature.

#### 4.1.2. Molecular descriptors

Molecular descriptors for each compound were generated with program Volsurf+.[33,34] Starting from the SMILES codes, 3D structures were generated using VolSurf+ 3D structure generator. Considering high rigidity of the dataset compounds, VolSurf+ 3D structure generator was considered to be powerful enough to produce the most relevant conformers and no additional conformational search was applied. Using following probes: H2O (the water molecule), O (sp$^2$ carbonyl oxygen atom), N1 (neutral NH group (*e.g.* amide)), and DRY (the hydrophobic probe), Molecular Interaction Fields (MIFs) were calculated. From the MIFs, 128 descriptors with clear chemical and physical meaning were calculated for each compound. The detailed description of all 128 VolSurf+ descriptors is given in the VolSurf+ manual.[35,36]

#### 4.1.3. Partial Least Square and Principal Component analysis

Partial Least Square (PLS) analysis was applied with goal of finding the relationship between the 3D structure-based molecular descriptors and antioxidative activities. The number of significant latent variables (nLV) and quality of the models were determined using the leave-one-out (LOO) cross-validation procedures and leave many out (LMO) cross-validation procedure with eight random groups. Also, for each model, standard deviation of error of calculation (SDEC) and standard deviation of error of prediction (SDEP) were calculated. Validation of predictive ability of each model was also performed using external validation. For that purpose, dataset was divided into the training set, that was used to build the model, and

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the test set, that was used to validate predictive ability of the model. Standard deviation of error of prediction (SDEP) for external prediction was calculated for each model.

In order to identify molecular descriptors with the highest positive or negative impact on antioxidative activities of studied compounds, analyses of PLS coefficients of the 3D-QSAR models were performed. Models were built using raw data (without autoscaling pretreatment of descriptors) and the quantitative influence of each descriptor on the anti-proliferative activity is given as product of the descriptor's value and it's PLS coefficient.

Principal Component Analysis (PCA) was performed on the molecular descriptors of the complete dataset (all 59 compounds). PCA loadings were used for identification of the descriptors with the highest contribution to the overall variance in the X-space.

## 4.2. Synthesis

### 4.2.1. General Methods

All chemicals and solvents were purchased from commercial available suppliers including Aldrich, Fluka and Acros. Melting points were recorded on an Original Keller Mikroheitztisch apparatus (Reichert, Wien), SMP11 Bibby and Büchi 535 apparatus and are not corrected. ^{1}H and ^{13}C NMR spectra were recorded on a Varian Gemini 300 or Varian Gemini 600 spectrophotometers at 300, 600, 150 and 75 MHz, respectively. All NMR spectra were measured in DMSO-$d_6$ solutions using TMS as an internal standard. Chemical shifts are reported in ppm (δ) relative to TMS. All compounds were routinely checked by thin layer chromatography (TLC) using precoated Merck silica gel 60F-254 plates and the spots were detected under UV light (254 nm). Column chromatography (CC) was performed using silica gel (0.063–0.2 mm) Fluka; glass column was slurry-packed under gravity.

### 4.2.2. General method for the preparation of carboxamides 20-25

To a solution of mono-, di- or tri-methoxy substituted benzoyl-chloride in dry toluene corresponding amine was added followed by the addition of Et₃N. The reaction mixture was refluxed for 24 hours. After cooling the resulting product was filtered off and washed with diluted HCl and water to obtain pure solid.

***2-Methoxy-N-[5(6)-cyano-1H-benzimidazol-2-yl]benzamide 20***

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Following the general method, from 0.32 g (1.9 mmol) 2-methoxybenzoylchloride **1**, 0.30 g (1.9 mmol) 2-amino-5(6)-cyano-*1H*-benzimidazole **18** and 0.27 g (2.7 mmol) Et$_3$N in dry toluene (15 ml), 0.47 g (86%) of white powder was obtained; mp 234–237 °C; ^{1}H NMR (300 MHz, DMSO-*d$_6$*) ($\delta$/ppm): 11.55 (s, 1H, NH$_{amide}$), 7.92 (s, 1H, H$_{arom}$), 7.79 (dd, $J_1$ = 7.6 Hz, $J_2$ = 1.6 Hz, 1H, H$_{arom}$), 7.63 (t, $J$ = 7.2 Hz, 2H, H$_{arom}$), 7.55 (t, $J$ = 8.2 Hz, 1H, H$_{arom}$), 7.25 (d, $J$ = 8.4 Hz, 1H, H$_{arom}$), 7.13 (t, $J$ = 7.5 Hz, 1H, H$_{arom}$), 3.96 (s, 3H, CH$_3$); ^{13}C NMR (150 MHz, DMSO-*d$_6$*) ($\delta$/ppm): 165.1, 157.1, 148.6, 133.6, 130.2, 125.1, 121.9, 120.7 (2C), 120.2, 112.3 (2C), 56.2.

### 2-Methoxy-N-(6-cyanobenzothiazol-2-yl)benzamide 23

Following the general method, from 0.29 g (1.7 mmol) 2-methoxybenzoylchloride **1**, 0.30 g (1.7 mmol) 2-amino-6-cyanobenzothiazole **19** and 0.24 g (2.4 mmol) Et$_3$N in dry toluene (15 ml), 0.40 g (77%) of white powder was obtained; mp 238–241 °C; ^{1}H NMR (300 MHz, DMSO-*d$_6$*) ($\delta$/ppm): 12.40 (s, 1H, NH$_{amide}$), 8.62 (d, $J$ = 1.1 Hz, 1H, H$_{arom}$), 7.92 (d, $J$ = 8.4 Hz, 1H, H$_{arom}$), 7.86 (dd, $J_1$ = 8.4 Hz, $J_2$ = 1.6 Hz, 1H, H$_{arom}$), 7.75 (dd, $J_1$ = 7.7, $J_2$ = 1.7 Hz, 1H, H$_{arom}$), 7.62 (t, $J$ = 7.0 Hz, 1H, H$_{arom}$), 7.25 (d, $J$ = 8.3 Hz, 1H, H$_{arom}$), 7.13 (t, $J$ = 7.5 Hz, 1H, H$_{arom}$), 3.95 (s, 3H); ^{13}C NMR (75 MHz, DMSO-*d$_6$*) ($\delta$/ppm): 165.9, 162.1, 157.8, 152.2, 134.4, 132.8, 130.8, 130.1, 127.6, 121.7, 121.2, 119.6, 112.8, 105.9, 56.6.

### 2,4-Dimethoxy-N-(6-cyanobenzothiazol-2-yl)benzamide 24

Following the general method, from 1.94 g (9.7 mmol) 2,4-dimethoxybenzoylchloride **2**, 1.7 g (9.7 mmol) 2-amino-6-cyanobenzothiazole **19** and 1.37 g (13.6 mmol) Et$_3$N in dry toluene (30 ml), 2.58 g (78%) of white powder was obtained; mp > 280 °C; ^{1}H NMR (300 MHz, DMSO-*d$_6$*) ($\delta$/ppm): 11.89 (s, 1H, NH$_{amide}$), 8.60 (d, $J$ = 1.0 Hz, 1H, H$_{arom}$), 7.93 − 7.82 (m, 3H, H$_{arom}$), 6.78 − 6.70 (m, 2H, H$_{arom}$), 4.00 (s, 3H, CH$_3$), 3.88 (s, 3H, CH$_3$); ^{13}C NMR (150 MHz, DMSO-*d$_6$*) ($\delta$/ppm): 164.5, 164.1, 161.6, 159.4, 151.7, 132.7, 132.4, 129.6, 127.0, 121.1, 119.2, 112.5, 106.5, 105.3, 98.7, 56.5, 55.8.

### 3,4,5-Trimethoxy-N-(6-cyanobenzothiazol-2-yl)benzamide 25

Following the general method, from 2.95 g (13.0 mmol) 3,4,5-trimethoxybenzoylchloride **3**, 2.24 g (13.0 mmol) 2-amino-6-cyanobenzothiazole **19** and 1.88 g (13.0 mmol) Et$_3$N in dry toluene (30 ml), 3.30 g (69%) of white powder was obtained; mp 251–253 °C; ^{1}H NMR (600 MHz, DMSO-*d$_6$*) ($\delta$/ppm): 13.16 (brs, 1H, NH$_{amide}$), 8.62 (s, 1H, H$_{arom}$), 7.92 (d, $J$ = 8.3 Hz, 1H, H$_{arom}$), 7.87 (dd, $J_1$ = 8.4 Hz, $J_2$ = 1.5 Hz, 1H, H$_{arom}$), 7.54 (s, 2H, H$_{arom}$), 3.90 (s, 6H, OCH$_3$), 3.77 (s, 3H, OCH$_3$); ^{13}C NMR (75 MHz, DMSO-*d$_6$*) ($\delta$/ppm): 165.7, 163.3, 153.3 (2C), 142.0, 130.2, 127.6, 126.5, 121.6, 119.7, 106.5 (2C), 105.9, 60.6, 56.6 (2C).

### 4.2.3. General method for the preparation of hydroxy substituted compounds 7-10 and 26-29

Corresponding methoxy substituted compound was dissolved in dry dichloromethane, cooled to -78 °C and $BCl_3$ or $BBr_3$ was added under argon atmosphere. The reaction mixture was stirred on that temperature for several hours and kept refrigerated overnight. The reaction was quenched by the addition of methanol, solvent was evaporated to dryness and the residue was purified by column chromatography ($CH_2Cl_2$/$CH_3OH$).

### *2-Hydroxy-N-(4-cyanophenyl)benzamide 7*

Following the general method, from 0.37 g (1.5 mmol) 2-methoxy-*N*-(4-cyanophenyl)benzamide **4** and 5.0 ml $BCl_3$ (2.2 mmol) in absolute dichloromethane (20 ml) under the argon to obtain 0.31 g (85%) of white powder; mp 177–180 °C; ^{1}H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 11.38 (brs, 1H, OH), 10.65 (s, 1H, $NH_{amide}$), 7.94 (d, $J$ = 8.8 Hz, 2H, $H_{arom}$), 7.88 (dd, $J_1$ = 7.9 Hz, $J_2$ = 1.6 Hz, 1H, $H_{arom}$), 7.83 (d, $J$ = 8.8 Hz, 2H, $H_{arom}$), 7.45 (m, 1H, $H_{arom}$), 7.02 – 6.95 (m, 2H, $H_{arom}$); ^{13}C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 166.5, 157.5, 142.7, 133.7, 133.2 (2C), 129.5, 120.4 (2C), 119.2, 119.0, 118.5, 117.1, 105.6.

### *2-Hydroxy-4-methoxy-N-(4-cyanophenyl)benzamide 8*

Following the general method, from 0.50 g (1.8 mmol) 2,4-dimethoxy-*N*-(4-cyanophenyl)benzamide **5** and 10.0 ml $BCl_3$ (5.3 mmol) in absolute dichloromethane (30 ml) under the argon to obtain 0.21 g (44%) of beige powder; mp 222–225 °C; ^{1}H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 11.99 (brs, 1H, OH), 10.50 (s, 1H, $NH_{amide}$), 7.95 (d, $J$ = 8.0 Hz, 1H, $H_{arom}$), 7.92 (d, $J$ = 9.1 Hz, 1H, $H_{arom}$), 7.82 (d, $J$ = 8.8 Hz, 1H, $H_{arom}$), 6.58 (dd, $J_1$ = 8.9 Hz, $J_2$ = 2.4 Hz, 1H, $H_{arom}$), 6.52 (d, $J$ = 2.4 Hz, 1H, $H_{arom}$), 3.80 (s, 3H, $OCH_3$); ^{13}C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 167.0, 164.0, 161.0, 142.7, 133.1 (2C), 130.7, 120.7 (2C), 119.0, 109.5, 106.5, 105.5, 101.3, 55.5.

### *2,4-Dihydroxy-N-(4-cyanophenyl)benzamide 9*

Following the general method, from 0.50 g (1.8 mmol) 2,4-dimethoxy-*N*-(4-cyanophenyl)benzamide **5** and 10.0 ml $BBr_3$ (2.7 mmol) in absolute dichloromethane (30 ml) under the argon to obtain 0.06 g (13%) of beige powder; mp 250–253 °C; ^{1}H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 11.87 (brs, 1H, OH), 10.44 (s, 1H, $NH_{amide}$), 10.26 (brs, 1H, OH), 7.91 (d, $J$ = 8.8 Hz, 2H, $H_{arom}$), 7.87 (d, $J$ = 8.7 Hz, 1H, $H_{arom}$), 7.81 (d, $J$ = 8.7 Hz, 2H, $H_{arom}$), 6.41 (dd, $J_1$ = 8.6 Hz, $J_2$ = 2.3 Hz, 1H, $H_{arom}$), 6.38 (d, $J$ = 2.3 Hz, 1H, $H_{arom}$); ^{13}C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 167.5, 163.3, 161.4, 143.3, 133.6 (2C), 131.5, 121.0 (2C), 119.5, 108.7, 108.2, 105.8, 103.3.

### 3,4,5-Trihydroxy-N-(4-cyanophenyl)benzamide 10

Following the general method, from 0.50 g (1.6 mmol) 3,4,5-trimethoxy-$N$-(4-cyanophenyl)benzamide **6** and 15.0 ml BBr$_3$ (15.0 mmol) in absolute dichloromethane (30 ml) under the argon to obtain 0.31 g (71%) of white powder; mp 273–275 °C; $^1$H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 10.30 (s, 1H, NH$_{amide}$), 9.22 (s, 2H, OH), 8.91 (s, 1H, OH), 7.97 (d, $J$ = 8.8 Hz, 2H, H$_{arom}$), 7.78 (d, $J$ = 8.7 Hz, 2H, H$_{arom}$), 6.97 (s, 2H, H$_{arom}$); $^{13}$C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 166.6, 146.0, 144.4, 137.8, 133.5 (2C), 124.7, 120.4 (2C), 119.7, 107.9 (2C), 105.2.

### 2-Hydroxy-N-[5(6)-cyano-1H-benzimidazol-2-yl]benzamide 26

Following the general method, from 0.50 g (1.7 mmol) 2-methoxy-$N$-[5(6)-cyano-1$H$-benzimidazol-2-yl]benzamide **20** and 10.0 ml BBr$_3$ (10.0 mmol) in absolute dichloromethane (25 ml) under the argon to obtain 0.35 g (74%) of white powder; mp > 280 °C; $^1$H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 12.91 (brs, 2H, NH$_{benzimidazole}$, OH), 8.02 (dd, $J_1$ = 7.7 Hz, $J_2$ = 1.3 Hz, 1H, H$_{arom}$), 7.90 (s, 1H, H$_{arom}$), 7.69 – 7.57 (m, 2H, H$_{arom}$), 7.44 (t, $J$ = 7.0 Hz, 1H, H$_{arom}$), 6.94 (m, 2H, H$_{arom}$); $^{13}$C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 200.7, 173.8, 165.9, 164.3, 150.2, 124.2, 116.1, 108.9.

### 2-Hydroxy-4-methoxy-N-[5(6)-cyano-1H-benzimidazol-2-yl]benzamide 27

Following the general method, from 0.20 g (0.6 mmol) 2,4-dimethoxy-$N$-[5(6)-cyano-1$H$-benzimidazol-2-yl]benzamide **21** and 3.6 ml BBr$_3$ (3.6 mmol) in absolute dichloromethane (15 ml) under the argon to obtain 0.14 g (59%) of white powder; mp > 280 °C; $^1$H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 12.05 (brs, 2H, NH$_{benzimidazole}$, NH$_{amide}$), 8.58 (s, 1H, H$_{arom}$), 8.01 (d, $J$ = 8.9 Hz, 1H, H$_{arom}$), 7.86 (s, 2H, H$_{arom}$), 6.63 (dd, $J_1$ = 8.9 Hz, $J_2$ = 2.4 Hz, 1H, H$_{arom}$), 6.58 (d, $J$ = 2.3 Hz, 1H, H$_{arom}$), 6.47 (s, 1H, H$_{arom}$), 3.82 (s, 3H, CH$_3$); $^{13}$C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 167.6, 164.2, 131.7 (2C), 129.6, 126.5, 119.6 (2C), 106.5, 104.1, 101.0 (2C), 55.4.

### 2-Hydroxy-N-(6-cyanobenzothiazol-2-yl)benzamide 28

Following the general method, from 0.50 g (1.6 mmol) 2-methoxy-$N$-[5(6)-cyano-1$H$-benzothiazol-2-yl]benzamide **23** and 10.0 ml BBr$_3$ (10.0 mmol) in absolute dichloromethane (25 ml) under the argon to obtain 0.37 g (78%) of white powder; mp > 280 °C; $^1$H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 12.30 (brs, 1H, NH$_{amide}$), 11.77 (brs, 1H, OH), 8.61 (s, 1H, H$_{arom}$), 7.99 (dd, $J_1$ = 7.9 Hz, $J_2$ = 1.6 Hz, 1H, H$_{arom}$), 7.93 – 7.84 (m, 2H, H$_{arom}$), 7.53 (t, $J$ = 7.7 Hz, 1H, H$_{arom}$), 7.13 – 6.99 (m, 2H, H$_{arom}$); $^{13}$C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 144.0, 134.9, 130.6, 129.8, 127.2, 119.9, 119.1, 117.6, 117.2, 105.5.

### 2-Hydroxy-4-methoxy-N-(6-cyanobenzothiazol-2-yl)benzamide 29

Following the general method, from 0.50 g (1.5 mmol) 2,4-dimethoxy-*N*-(6-cyanobenzothiazol-2-yl)benzamide **24** and 0.65 g (5.6 mmol) $BCl_3$ in dichloromethane (30 ml) under the argon to obtain 0.12 g (25%) of yellow powder; mp > 280 °C; ^{1}H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 12.21 (brs, 2H, OH, NH$_{amide}$), 8.60 (s, 1H, H$_{arom}$), 8.03 (d, *J* = 8.8 Hz, 1H, H$_{arom}$), 7.88 (s, 2H), 6.65 (dd, *J$_1$* = 8.9 Hz, *J$_2$* = 2.2 Hz, 1H, H$_{arom}$), 6.59 (d, *J* = 2.1 Hz, 1H, H$_{arom}$), 3.84 (s, 3H); ^{13}C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 165.2, 132.7, 130.3, 127.6, 119.6, 107.7, 105.9, 101.8, 56.0.

### 4.2.4. General method for the preparation of amidines

Dry HCl gas was bubbled for 4 h into a cooled suspension (0 °C) of nitrile compound in anhydrous ethanol or methoxyethanol. The suspension was stirred at room temperature until the –CN band was undetectable (IR). After anhydrous diethylether was added, the corresponding imidate ester was filtered off and dried *in vacuo*. The product was then suspended in anhydrous ethanol and the corresponding amine was added. For the synthesis of unsubstituted amidines, the suspension was treated with anhydrous ammonia gas and stirred for 24 h at room temperature. For the synthesis of imidazolinyl-substituted amidines, excess of ethylenediamine (3.5 molar equivalents) was added to a suspension of imidate ester and the reaction mixture was stirred at reflux for 24 h. The crude product was then filtered off and washed with diethylether to give powder products which were suspended in anhydrous ethanol and saturated with HCl gas. The reaction mixture was stirred at room temperature for 24 h. The products were filtered off.

### *2-Hydroxy-N-(4-amidinophenyl)benzamide 11*

The compound was prepared following the general method. A suspension of 0.19 g (0.8 mmol) 2-hydroxy-*N*-(4-cyanophenyl)benzamide **7** in anhydrous ethanol (15 ml) was saturated with dry HCl gas and stirred for 5 days. The crude imidate was filtered off, suspended in absolute ethanol (8 ml) and dry NH$_3$ was bubbled into the suspension. The mixture was worked up as it is described to give 0.98 g (87%) of light yellow powder; mp 270–273 °C; ^{1}H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 15.69 (brs, 1H, NH$_{amide}$), 8.92 (brs, 4H, NH$_{amidine}$), 7.84 (s, 4H, H$_{arom}$), 7.2 (dd, *J$_1$* = 7.8 Hz, *J$_2$* = 1.8 Hz, 1H, H$_{arom}$), 7.07 – 7.01 (m, 1H, H$_{arom}$), 6.49 (d, *J* = 8.2 Hz, 1H, H$_{arom}$), 6.27 (t, *J* = 7.3 Hz, 1H, H$_{arom}$), 4.35 (brs, 1H, OH); ^{13}C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 169.9, 167.3, 164.8, 145.8, 132.6, 129.6, 129.2 (2C), 121.0, 120.9, 120.3, 118.9 (2C), 118.0, 111.2.

### *2-Hydroxy-4-methoxy-N-(4-amidinophenyl)benzamide 12*

The compound was prepared following the general method. A suspension of 0.12 g (0.4 mmol) 2-hydroxy-4-methoxy-*N*-(4-cyanophenyl)benzamide **8** in anhydrous ethanol (10 ml) was saturated with dry HCl gas and stirred for 4 days. The crude imidate was filtered off, suspended in absolute ethanol (10 ml) and dry $NH_3$ was bubbled into the suspension. The mixture was worked up as it is described to give 0.07 g (51%) of light rose powder; mp 256–259 °C; ^{1}H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 15.47 (brs, 1H, $NH_{amide}$), 9.13 (brs, 4H, $NH_{amidine}$), 7.84 (d, *J* = 8.8 Hz, 2H, $H_{arom}$), 7.81 (d, *J* = 8.8 Hz, 2H, $H_{arom}$), 7.61 (d, *J* = 8.7 Hz, 1H, $H_{arom}$), 5.94 (d, *J* = 2.1 Hz, 1H, $H_{arom}$), 5.87 (dd, $J_1$ = 8.7 Hz, $J_2$ = 2.3 Hz, 1H, $H_{arom}$), 4.36 (brs, 1H, OH), 3.67 (s, 3H, $OCH_3$); ^{13}C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 172.8, 167.3, 164.9, 163.6, 146.6, 130.5, 129.2 (2C), 119.4, 118.6 (2C), 111.9, 103.5, 99.3, 54.4.

### *2,4-Dihydroxy-N-(4-amidinophenyl)benzamide 13*

The compound was prepared following the general method. A suspension of 0.30 g (1.2 mmol) 2,4-dihydroxy-*N*-(4-cyanophenyl)benzamide **9** in anhydrous ethanol (15 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off, suspended in absolute ethanol (15 ml) and dry $NH_3$ was bubbled into the suspension. The mixture was worked up as it is described to give 0.06 g (17%) of light brown powder; mp > 280 °C; ^{1}H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 15.67 (brs, 1H, $NH_{amide}$), 8.94 (brs, 6H, $NH_{amidine}$, OH), 7.82 – 7.78 (m, 4H, $H_{arom}$), 7.53 (d, *J* = 8.6 Hz, 1H, $H_{arom}$), 5.82 (s, 1H, $H_{arom}$), 5.78 (d, *J* = 8.5 Hz, 1H, $H_{arom}$); ^{13}C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 172.8, 167.7, 165.2, 162.4, 146.9, 131.2, 129.7 (2C), 120.0, 119.0 (2C), 111.5, 105.8, 101.6.

### *2-Hydroxy-N-[4-(imidazolin-2-yl)phenyl]benzamide 14*

The compound was prepared following the general method. A suspension of 0.19 g (0.8 mmol) 2-hydroxy-*N*-(4-cyanophenyl)benzamide **7** in anhydrous ethanol (15 ml) was saturated with dry HCl gas and stirred for 5 days. The crude imidate was filtered off and suspended in absolute ethanol. EDA (0.09 ml, 1.4 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.09 g (37%) of beige powder; mp 290–293 °C; ^{1}H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 11.52 (brs, 1H, $NH_{amide}$), 10.79 (s, 1H, OH), 10.64 (brs, 2H, $NH_{amidine}$), 8.07 (d, *J* = 8.8 Hz, 2H, $H_{arom}$), 8.01 (d, *J* = 8.7 Hz, 2H, $H_{arom}$), 7.93 (d, *J* = 7.7 Hz, 1H, $H_{arom}$) 7.45 (t, *J* = 7.6 Hz, 1H, $H_{arom}$), 7.07 (d, *J* = 8.2 Hz, 1H, $H_{arom}$), 6.98 (t, *J* = 7.5 Hz, 1H, $H_{arom}$), 3.99 (s, 4H, $CH_2$); ^{13}C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 166.4, 164.1, 157.5, 144.0, 133.7, 129.8 (2C), 129.7, 120.0 (2C), 119.2, 118.5, 117.1, 116.7, 44.2 (2C).

### *2-Hydroxy-4-methoxy-N-[4-(imidazolin-2-yl)phenyl]benzamide 15*

The compound was prepared following the general method. A suspension of 0.20 g (0.8 mmol) 2-hydroxy-4-methoxy-*N*-(4-cyanophenyl)benzamide **8** in anhydrous ethanol (35 ml) was saturated with dry HCl gas and stirred for 4 days. The crude imidate was filtered off and suspended in absolute ethanol. EDA (0.07 ml, 1.1 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.06 g (24%) of white powder; mp 292–295 °C; $^1$H NMR (600 MHz, DMSO-$d_6$) (δ/ppm): 12.13 (s, 1H, NH$_{amide}$), 10.72 (s, 3H, NH$_{amidine}$, OH), 8.09 (d, $J$ = 8.7 Hz, 2H, H$_{arom}$), 8.06 (d, $J$ = 8.8 Hz, 1H, H$_{arom}$), 8.01 (d, $J$ = 8.7 Hz, 2H, H$_{arom}$), 6.61 (d, $J$ = 1.9 Hz, 1H, H$_{arom}$), 6.58 (dd, $J_1$ = 8.9 Hz, $J_2$ = 1.9 Hz, 1H, H$_{arom}$), 3.99 (s, 4H, CH$_2$), 3.81 (s, 3H, OCH$_3$); $^{13}$C NMR (75 MHz, DMSO-$d_6$) (δ/ppm): 167.4, 164.5, 164.4, 161.3, 144.5, 131.5, 130.2 (2C), 120.7 (2C), 117.1, 110.1, 106.9, 101.8, 55.9, 44.7 (2C).

### 2,4-Dihydroxy-N-[4-(imidazolin-2-yl)phenyl]benzamide 16

The compound was prepared following the general method. A suspension of 0.14 g (0.6 mmol) 2,4-dihydroxy-*N*-(4-cyanophenyl)benzamide **9** in anhydrous ethanol (10 ml) was saturated with dry HCl gas and stirred for 10 days. The crude imidate was filtered off and suspended in absolute ethanol. EDA (0.16 ml, 2.4 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.10 g (53%) of beige powder; mp > 280 °C; $^1$H NMR (300 MHz, DMSO-$d_6$) (δ/ppm): 11.96 (s, 1H, OH), 10.63 – 10.62 (m, 3H, NH$_{amidine}$, OH), 10.38 (s, 1H, NH$_{amide}$), 8.06 (d, $J$ = 8.7 Hz, 2H, H$_{arom}$), 7.99 (d, $J$ = 8.8 Hz, 2H, H$_{arom}$), 7.94 (d, $J$ = 8.6 Hz, 1H, H$_{arom}$), 6.45 – 6.41 (m, 2H, H$_{arom}$), 3.99 (s, 4H, CH$_2$); $^{13}$C NMR (75 MHz, DMSO-$d_6$) (δ/ppm): 167.4, 164.6, 163.4, 161.2, 144.6, 131.7, 130.1 (2C), 120.6 (2C), 116.9, 108.7, 108.3, 103.3, 44.7 (2C).

### 3,4,5-Trihydroxy-N-[4-(imidazolin-2-yl)phenyl]benzamide 17

The compound was prepared following the general method. A suspension of 0.14 g (0.5 mmol) 3,4,5-trihydroxy-*N*-(4-cyanophenyl)benzamide **10** in anhydrous ethanol (15 ml) was saturated with dry HCl gas and stirred for 10 days. The crude imidate was filtered off and suspended in absolute ethanol. EDA (0.05 ml, 0.8 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.03 g (24%) of yellow powder; mp > 280 °C; $^1$H NMR (300 MHz, DMSO-$d_6$) (δ/ppm): 10.50 (s, 2H, NH$_{amidine}$), 10.38 (s, 1H, NH$_{amide}$), 9.24 (s, 2H, OH), 9.01 (s, 1H, OH), 8.02 (d, $J$ = 9.2 Hz, 2H, H$_{arom}$), 7.98 (d, $J$ = 9.2 Hz, 2H, H$_{arom}$), 6.99 (s, 2H, H$_{arom}$), 3.98 (s, 4H, CH2); $^{13}$C NMR (75 MHz, DMSO-$d_6$) (δ/ppm): 166.6, 164.8, 146.01, 145.8, 138.0, 130.0 (2C), 124.6, 120.0 (2C), 116.3, 107.9 (2C), 44.7 (2C).

### 2-Hydroxy-N-[5(6)-amidino-1H-benzimidazol-2-yl]benzamide 30

The compound was prepared following the general method. A suspension of 0.14 g (0.5 mmol) 2-hydroxy-*N*-[5(6)-cyano-*1H*-benzimidazol-2-yl]benzamide **26** in anhydrous ethanol (15 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off, suspended in absolute ethanol (10 ml) and dry $NH_3$ was bubbled into the suspension. The mixture was worked up as it is described to give 0.04 g (58%) of light yellow powder; mp > 280 °C; $^1$H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 17.49 (brs, 1H, OH), 12.44 (brs, 1H, NH$_{amide}$), 9.08 (s, 2H, NH$_{amidine}$), 8.53 (s, 2H, NH$_{amidine}$), 7.87 − 7.80 (m, 2H, H$_{arom}$), 7.50 (s, 1H, H$_{arom}$), 7.45 (d, *J* = 8.0 Hz, 1H, H$_{arom}$), 7.15 (t, *J* = 7.5 Hz, 1H, H$_{arom}$), 6.63 (d, *J* = 8.2 Hz, 1H, H$_{arom}$), 6.53 (t, *J* = 7.2 Hz, 1H, H$_{arom}$); $^{13}$C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 170.6, 166.9 (2C), 166.4, 132.8, 129.5 (2C), 120.6, 119.4, 119.1 (2C), 114.4.

### 2-Hydroxy-4-methoxy -N-[5(6)-amidino-1H-benzimidazol-2-yl]benzamide 31

The compound was prepared following the general method. A suspension of 0.08 g (0.3 mmol) **27** in anhydrous ethanol (10 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off, suspended in absolute ethanol (10 ml) and dry $NH_3$ was bubbled into the suspension. The mixture was worked up as it is described to give 0.05 g (57%) of light brown powder; mp 255–260°C; $^1$H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 17.12 (brs, 1H, OH), 12.23 (brs, 1H, NH$_{benzimidazole}$), 8.76 (brs, 4H, NH$_{amidine}$), 7.83 (s, 1H, H$_{arom}$), 7.70 (s, 1H, H$_{arom}$), 7.48 (s, 2H, H$_{arom}$), 6.09 (s, 2H, H$_{arom}$), 3.70 (s, 3H, CH$_3$); $^{13}$C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 166.1, 165.5, 163.9, 163.1, 154.8, 132.9, 131.4, 130.5, 121.4, 121.1, 119.3, 118.4, 101.2, 55.2.

### 2-Hydroxy-N-(6-amidinobenzothiazol-2-yl)benzamide 32

The compound was prepared following the general method. A suspension of 0.20 g (0.7 mmol) **28** in anhydrous ethanol (15 ml) was saturated with dry HCl gas and stirred for 10 days. The crude imidate was filtered off, suspended in absolute ethanol (10 ml) and dry $NH_3$ was bubbled into the suspension. The mixture was worked up as it is described to give 0.03 mg (12%) of beige powder; mp > 280 °C; $^1$H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 15.75 (brs, 1H, OH), 9.14 (brs, 2H, NH$_{amidine}$), 8.63 (brs, 2H, NH$_{amidine}$), 8.26 (d, *J* = 1.3 Hz, 1H, H$_{arom}$), 7.88 (dd, $J_1$ = 7.7 Hz, $J_2$ = 1.7 Hz, 1H, H$_{arom}$), 7.73 − 7.62 (m, 2H, H$_{arom}$), 7.25 (t, *J* = 8.5 Hz, 1H, H$_{arom}$), 6.83 − 6.69 (m, 2H, H$_{arom}$); $^{13}$C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 171.3, 169.4, 165.8, 162.2, 155.4, 133.6, 132.9, 129.8, 125.4, 122.0, 120.3, 119.9, 119.0, 117.5, 117.4.

### 2-Hydroxy-4-methoxy-N-(6-amidinobenzothiazol-2-yl)benzamide 33

The compound was prepared following the general method. A suspension of 0.08 g (0.2 mmol) 2-hydroxy-4-methoxy-*N*-(6-cyanobenzothiazol-2-yl)benzamide **29** in anhydrous ethanol (10 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off,

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suspended in absolute ethanol (10 ml) and dry $NH_3$ was bubbled into the suspension. The mixture was worked up as it is described to give 0.04 mg (40%) of white powder; mp > 280 °C; ^{1}H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 13.02 (brs, 2H, OH, NH$_{amide}$), 9.29 (brs, 2H, NH$_{amidine}$), 8.99 (brs, 2H, NH$_{amidine}$), 8.47 – 8.43 (m, 1H, H$_{arom}$), 7.96 – 7.88 (m, 1H, H$_{arom}$), 7.85 – 7.75 (m, 2H, H$_{arom}$), 6.52 (s, 2H, H$_{arom}$), 3.80 (s, 3H, CH$_3$); ^{13}C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 165.5, 164.4, 160.6, 131.9, 129.6, 126.8, 126.0, 122.6, 122.3, 119.3, 109.5, 106.7, 104.9, 102.8, 101.4, 55.4.

### 2-Hydroxy-N-[5(6)-(imidazolinil-2-yl)-1H-benzimidazol-2-yl]benzamide 34

The compound was prepared following the general method. A suspension of 0.14 g (0.5 mmol) 2-hydroxy-*N*-[5(6)-cyano-*1H*-benzimidazol-2-yl]benzamide **26** in anhydrous methoxyethanol (15 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off and suspended in absolute ethanol (10 ml). EDA (0.05 ml, 0.8 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.05 g (68%) of white powder; mp > 280 °C; ^{1}H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 17.36 (brs, 1H, OH), 12.41 (brs, 1H, NH$_{benzimidazole}$), 9.63 (brs, 2H, NH$_{amidine}$), 7.92 (s, 1H, H$_{arom}$), 7.86 (dd, $J_1$ = 7.7, $J_2$ = 1.5 Hz, 1H, H$_{arom}$), 7.57 (d, $J$ = 8.3 Hz, 1H, H$_{arom}$), 7.51 (d, $J$ = 8.0 Hz, 1H, H$_{arom}$), 7.17 (t, $J$ = 7.6 Hz, 1H, H$_{arom}$), 6.65 (d, $J$ = 8.2 Hz, 1H, H$_{arom}$), 6.56 (t, $J$ = 7.4 Hz, 1H, H$_{arom}$), 3.94 (s, 4H, CH$_2$); ^{13}C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 165.7, 134.7, 130.7, 123.6, 119.4, 117.7 (2C), 116.4, 113.6, 44.8 (2C).

### 2-Hydroxy-4-methoxy -N-[5(6)-(imidazolinil-2-yl)-1H-benzimidazol-2-yl]benzamide 35

The compound was prepared following the general method. A suspension of  0.08 mg (0.02 mmol)   2-hydroxy-4-methoxy-*N*-[5(6)-cyano-1H-benzimidazol-2-yl]benzamide  **27**  in anhydrous metoxyethanol (10 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off and suspended in absolute ethanol (10 ml). EDA (0.02 ml, 0.4 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.02 g (38%) of white powder; mp > 280 °C; ^{1}H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 10.02 (brs, 2H, NH$_{amidine}$), 7.92 (s, 1H, H$_{arom}$), 7.74 (d, $J$ = 8.4 Hz, 1H, H$_{arom}$), 7.59 – 7.49 (m, 2H, H$_{arom}$), 6.13 – 6.09 (m, 2H, H$_{arom}$), 3.95 (s, 4H, CH$_2$), 3.71 (s, 3H, CH$_3$); ^{13}C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 169.7, 166.5, 163.9, 155.8, 154.8, 133.7, 130.7, 125.6, 121.9, 121.1, 119.0, 114.6, 113.4, 112.1, 102.4, 55.2, 45.0 (2C).

### 2-Hydroxy-N-[6-(imidazolinil-2-yl)benzothiazol-2-yl]benzamide 36

The compound was prepared following the general method. A suspension of 0.20 g (0.7 mmol) **28** in anhydrous ethanol (15 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off and suspended in absolute ethanol (10 ml). EDA (0.08 ml, 1.2

mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.08 g (56%) of beige powder; mp > 280 °C; $^1$H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 15.69 (brs, 1H, OH), 9.49 (brs, 2H, NH$_{amidine}$), 8.37 (s, 1H, H$_{arom}$), 7.90 (d, $J$ = 6.9 Hz, 1H, H$_{arom}$), 7.82 (d, $J$ = 8.0 Hz, 1H, H$_{arom}$), 7.70 (d, $J$ = 8.5 Hz, 1H, H$_{arom}$), 7.28 (t, $J$ = 6.7 Hz, 1H, H$_{arom}$), 6.80 – 6.76 (m, 2H, H$_{arom}$), 3.99 (s, 4H, CH$_2$); $^{13}$C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 171.2, 165.6, 161.8, 155.7, 133.7, 133.1, 129.9, 125.7, 122.3, 120.1, 119.4, 118.0, 117.7, 117.5, 114.2, 44.7 (2C).

### 2-Hydroxy-4-methoxy-N-[6-(imidazolinil-2-yl)benzothiazol-2-yl]benzamide 37

The compound was prepared following the general method. A suspension of 0.08 g (0.2 mmol) 2-hydroxy-4-methoxy-*N*-(6-cyanobenzothiazol-2-yl)benzamide **29** in anhydrous ethanol (10 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off and suspended in absolute ethanol (10 ml). EDA (0.03 ml, 0.5 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.02 g (23%) of white powder; mp > 280 °C; $^1$H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 16.21 (brs, 1H, OH), 9.69 (brs, 2H, NH$_{amidine}$), 8.33 (s, 1H, H$_{arom}$), 7.81 – 7.77 (m, 2H, H$_{arom}$), 7.66 (d, $J$ = 8.4 Hz, 1H, H$_{arom}$), 6.33 (d, $J$ = 8.7 Hz, 1H, H$_{arom}$), 6.31 (s, 1H, H$_{arom}$), 3.98 (s, 4H, CH$_2$), 3.75 (s, 3H, CH$_3$).

### 2-Methoxy-N-(6-amidinobenzothiazol-2-yl)benzamide 40

The compound was prepared following the general method. A suspension of 0.15 g (0.5 mmol) 2-methoxy-*N*-(6-cyanobenzothiazol-2-yl)benzamide **23** in anhydrous ethanol (20 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off, suspended in absolute ethanol (10 ml) and dry NH$_3$ was bubbled into the suspension. The mixture was worked up as it is described to give 0.14 g (81%) of white powder; mp 258–261°C; $^1$H NMR (300 MHz, DMSO-$d_6$) ($\delta$/ppm): 8.76 (brs, 4H, NH$_{amidine}$), 8.22 (s, 1H, H$_{arom}$), 7.70 (d, $J$ = 7.7 Hz, 1H, H$_{arom}$), 7.58 – 7.53 (m, 2H, H$_{arom}$), 7.32 (t, $J$ = 7.1 Hz, 1H, H$_{arom}$), 7.03 (d, $J$ = 8.2 Hz, 1H, H$_{arom}$), 6.95 (t, $J$ = 7.3 Hz, 1H, H$_{arom}$), 3.79 (s, 3H, OCH$_3$); $^{13}$C NMR (75 MHz, DMSO-$d_6$) ($\delta$/ppm): 165.7, 157.4, 133.5, 130.0, 125.1, 121.7, 120.3, 118.3, 112.5, 56.0.

### 2,4-Dimethoxy-N-(6-amidinobenzothiazol-2-yl)benzamide 41

The compound was prepared following the general method. A suspension of 0.23 g (6.6 mmol) 2,4-dimethoxy-*N*-(6-cyanobenzothiazol-2-yl)benzamide **24** in anhydrous methoxyethanol (20 ml) was saturated with dry HCl gas and stirred for 27 days. The crude imidate was filtered off, suspended in absolute ethanol (15 ml) and dry NH$_3$ was bubbled into the suspension. The

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mixture was worked up as it is described to give 0.09 mg (33%) of white powder; mp 261–264 °C; $^1$H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 11.86 (s, 1H, NH$_{amide}$), 8.58 (s, 1H, H$_{arom}$), 7.88 (d, $J$ = 8.3 Hz, 1H, H$_{arom}$), 7.85 – 7.83 (m, 2H, H$_{arom}$), 6.75 (s, 1H, H$_{arom}$), 6.72 (d, $J$ = 8.5 Hz, 1H, H$_{arom}$), 3.99 (s, 3H, OCH$_3$), 3.86 (s, 3H, OCH$_3$); $^{13}$C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 164.7, 163.9, 161.6, 159.6, 151.8, 132.8, 132.6, 129.5, 126.8, 121.1, 119.0, 112.5, 106.8, 105.5, 99.0, 56.6, 55.8.

### 3,4,5-Trimethoxy-*N*-(6-amidinobenzothiazol-2-yl)benzamide 42

The compound was prepared following the general method. A suspension of 0.18 g (0.5 mmol) 3,4,5-trimethoxy-*N*-(6-cyanobenzothiazol-2-yl)benzamide **25** in anhydrous methoxyethanol (20 ml) was saturated with dry HCl gas and stirred for 70 days. The crude imidate was filtered off, suspended in absolute ethanol (8 ml) and dry NH$_3$ was bubbled into the suspension. The mixture was worked up as it is described to give 0.05 mg (26%) of white powder; mp > 280 °C; $^1$H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 8.55 (s, 1H, H$_{arom.}$), 7.89 (s, 2H, H$_{arom.}$), 7.53 (s, 2H, H$_{arom.}$), 7.39 (s, 4H, NH$_{amidine}$), 3.87 (s, 6H, OCH$_3$), 3.74 (s, 3H, OCH$_3$).

### 2-Methoxy-*N*-[6-(imidazolinil-2-yl)benzothiazol-2-yl]benzamide 45

The compound was prepared following the general method. A suspension of 0.15 g (0.5 mmol) 2-hydroxy-4-methoxy-*N*-(6-cyanobenzothiazol-2-yl)benzamide **29** in ethanol (20 ml) was saturated with dry HCl gas and stirred for 9 days. The crude imidate was filtered off and suspended in absolute ethanol (10 ml). EDA (0.11 ml, 3.4 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.15 g (81%) of beige powder; mp > 280 °C; $^1$H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 10.84 (brs, 2H, NH$_{amidine}$), 8.76 (s, 1H, H$_{arom}$), 8.09 (d, $J$ = 8.5 Hz, 1H, H$_{arom}$), 7.96 (d, $J$ = 8.5 Hz, 1H, H$_{arom}$), 7.77 (d, $J$ = 6.3 Hz, 1H, H$_{arom}$), 7.60 (t, $J$ = 7.2 Hz, 1H, H$_{arom}$), 7.25 (d, $J$ = 8.4 Hz, 1H, H$_{arom}$), 7.12 (t, $J$ = 7.5 Hz, 1H, H$_{arom}$), 4.00 (s, 4H, CH$_2$), 3.96 (s, 3H, OCH$_3$); $^{13}$C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 165.8, 165.4, 162.1, 158.0, 153.4, 134.5, 132.7, 130.9, 127.0, 123.7, 121.7, 121.4, 121.3, 117.6, 113.0, 56.8, 44.9 (2C).

### 2,4-Dimethoxy-*N*-[6-(imidazolinil-2-yl)benzothiazol-2-yl]benzamide 46

The compound was prepared following the general method. A suspension of 0.23 g (6.6 mmol) 2-hydroxy-4-methoxy-*N*-(6-cyanobenzothiazol-2-yl)benzamide **29** in anhydrous methoxyethanol (20 ml) was saturated with dry HCl gas and stirred for 27 days. The crude imidate was filtered off and suspended in absolute ethanol (10 ml). EDA (0.06 ml, 0.9 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.09 g (34%) of light yellow powder; mp 261–264 °C; $^1$H NMR (600 MHz,

DMSO-$d_6$) ($\delta$/ppm): 10.64 (brs, 2H, NH$_{amidine}$), 8.58 (d, $J$ = 1.3 Hz, 1H, H$_{arom}$), 7.88 (d, $J$ = 8.4 Hz, 1H, H$_{arom}$), 7.85 − 7.82 (m, 2H, H$_{arom}$), 6.75 (d, $J$ = 2.1 Hz, 1H, H$_{arom}$), 6.71 (dd, $J_1$ = 8.7, $J_2$ = 2.2 Hz, 1H, H$_{arom}$), 3.99 (s, 3H, OCH$_3$), 3.86 (s, 3H, OCH$_3$); $^{13}$C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 164.7, 164.0, 161.6, 159.5, 151.8, 132.8, 129.6, 126.9, 121.1, 119.0, 112.5, 106.7, 105.5, 98.9, 56.6, 55.8.

### *3,4,5-Trimethoxy-N-[6-(imidazolin-2-yl)benzothiazol-2-yl]benzamide 47*

The compound was prepared following the general method. A suspension of 0.18 g (0.5 mmol) 3,4,5-trimethoxy-*N*-(6-cyanobenzothiazol-2-yl)benzamide **25** in anhydrous methoxyethanol (20 ml) was saturated with dry HCl gas and stirred for 70 days. The crude imidate was filtered off and suspended in absolute ethanol (8 ml). EDA (0.05 ml, 1.7 mmol) was added and the mixture was stirred at reflux for 24 h. The mixture was worked up as it is described to give 0.04 g (45%) of light yellow powder; mp > 280 °C; $^1$H NMR (600 MHz, DMSO-$d_6$) ($\delta$/ppm): 13.32 (brs, 1H, NH$_{amide}$), 10.79 (s, 2H, NH$_{amidine}$), 8.79 (s, 1H, H$_{arom}$), 8.14 (d, $J$ = 8.2 Hz, 1H, H$_{arom}$), 8.01 (d, $J$ = 8.6 Hz, 1H, H$_{arom}$), 7.57 (s, 2H, H$_{arom}$), 4.04 (s, 3H, OCH$_3$), 3.91 (s, 4H, CH$_2$), 3.11 (s, 6H, OCH$_3$); $^{13}$C NMR (150 MHz, DMSO-$d_6$) ($\delta$/ppm): 165.7, 153.3, 142.0, 127.7, 124.8, 124.5, 106.5 (2C), 60.6, 56.6 (2C), 43.2 (2C).

## Acknowledgments

## Supporting Information

general methods for synthesis, figures of NMR spectrum, pharmacology methods, antimicrobial activity tests, antioxidative tests, structures and activities of dataset compounds used for QSAR models, computationally predicted activities of novel compounds, PCA plots

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