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Regioselective hydrostannation of diarylalkynes directed by a labile *ortho* bromine atom: An easy access to stereodefined triarylolefins, hybrids of combretastatin A-4 and *iso*combretastatin A-4

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Dedicated to our colleague Pr. Claude Combet-Farnoux

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1. Introduction

The microtubule system of eukaryotic cells is an important target for the development of anticancer agents. Consequently, perturbation of normal tubulin assembly/disassembly is a popular target for new chemotherapeutic agents [1,2] Examples of clinically used antimitotic agents are paclitaxel, which promoted microtubules polymerization and inhibits microtubules depolymerization, and vincristine which inhibits microtubules assembly [3,4]. However, despite their potent antitumor activities, these drugs have undesirable side effects [5,6] and are subject to multidrug resistance [7,8].

In recent years, there has been more and more interest in the search of antitumor molecules of easy synthesis, with high efficacy and low side effects. Combretastatin A-4 (CA-4), a naturally occurring stilbene, extracted from the South African willow *Combretum*

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ABSTRACT

A series of triarylolefins bearing the combretastatin A-4 and the *iso*combretastatin A-4 cores were synthesized and evaluated. The cooperative *ortho*-effect of a labile bromine atom in the regioselective hydrostannation of unsymmetrical diarylalkynes leading to stereodefined triarylolefins is presented. © 2010 Elsevier Masson SAS. All rights reserved.

caffrum [9,10] is the most studied substance as a highly potent inhibitor of tubulin assembly [11–13]. Moreover, CA-4 displays strong cytotoxicities (nanomolar level) against a large panel of human cancer cells, including multidrug-resistant cell lines [14,15]. Additionally, CA-4P [16–19] the water-soluble prodrug of CA-4, as well as AC-7739 and its amino acid derivative AVE-8062 [20] have been demonstrated to cause a rapid and selective vascular shutdown in established tumors *in vivo*, consistent with an antivascular mechanism of action [21–27]. Currently, CA-4P [28] either as a single agent or in combination therapy is undergoing several advanced clinical trials worldwide for the treatment of age-related macular degeneration or anaplastic thyroid cancer (Fig. 1).

Despite their remarkable anticancer activity, these Z-stilbene compounds are prone to double bond isomerization during storage and administration. The *E*-isomers display dramatically reduced inhibition of cancer cell growth and tubulin assembly [29]. In our efforts to discover novel tubulin assembly inhibitors [30–32], we recently found that *iso*combretastatin A-4 (*iso*CA-4), the third and "forgotten" structural isomer of the natural product, displayed biological activities comparable to that of CA-4 [33,34]. This substance

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Fig. 1. Representative tubulin binding agents and general structure of the synthesized hybrids 1.

having a 1,1-diarylethylene scaffold is easy to synthesize [35,36] at a multi grams scale without the need to control the olefin geometry. By structural modifications on the B ring, we have also identified other promising antiproliferative agents such as *iso*FCA-4 and *iso*-NH₂CA-4 (Fig. 1) [37]. In addition, these apoptosis inductors substances were found to inhibit microtubules formation and to induce cell cycle arrest in G2/M phase. To combine the anticancer effects of CA-4 and *iso*CA-4 within a single substance (e.g.; **1h**), we have synthesized a variety of hybrids **1** that bear, from one hand, the basic skeleton of CA-4 and, on the other hand, the one of *iso*CA-4 (Fig. 1). Moreover, these compounds **1** may be regarded as Tamoxifen [38] or trisubstituted Combretastatin analogues of high antiproliferative activity [39–41]. The potencies of newly synthesized trisubstituted olefins **1** were evaluated for their capacity to inhibit cancer cellular growth and, to act as potential antimitotic agents.

2. Results and discussion

The strategy envisioned to prepare the target triarylolefins **1** involves a three step-sequence by achieving a regioselective hydrostannation of diarylalkynes (Scheme 1). Further direct Stille reaction or sequential iodolysis-Negishi coupling would provide **1**.

At the outset of this work, the synthesis of triarylolefin **1a** (Scheme 2) was first examined from symmetrically diarylalkyne **2** easily available by Sonogashira–Linstrumelle coupling reaction [42,43]. Palladium-catalyzed hydrostannation of **2** was achieved using Bu₃SnH

(1.2 eq) in THF at room temperature in the presence of PdCl₂(PPh₃)₂ (1 mol%) to give vinylstannane **3** in good yield (79%) [44–47]. Subsequent direct coupling of **3** with 3,4,5-trimethoxy-iodobenzene under modified Stille conditions [48] provided a moderate 54% isolated yield of **1a**. In order to increase the overall yield of **1a**, we next evaluated a halodestannylation [49]-Negishi coupling sequence [50,51]. Iododestannylation of **3** with molecular iodine in CH₂Cl₂ at room temperature furnished cleanly and rapidly the desired electrophilic species **4**, which upon reaction with 3,4,5-trimethoxyphenylzinc chloride in the presence of a catalytic amount of PdCl₂(PPh₃)₂ (5 mol%) provided the target olefin **1a** in a quantitative yield. This result clearly indicated that the two steps iododestannylation-Negishi sequence was more efficient than the direct Stille coupling of **3**.

According to this two steps sequence (iodolysis/Negishi coupling) depicted in Scheme 2, the synthesis of other targets triarylolefins **1b**-**h** was envisioned from the hydrostannation of unsymmetrical diarylalkynes. In previous studies, we have reported that in the case of ortho substituted diarylalkynes, the ortho substituent, regardless of its electronic nature, directed the tributyltin addition to afford a single α -vinylstannane. We opted to use this ortho-directing effect (ODE) [52,53] concept to prepare the hybrids **1b**-h according to the retrosynthetic pathway depicted in Scheme 3. To control the regioselectivity of the hydrostannation reaction, we needed first to prepare diarylalkynes 5 bearing an ortho substituent able to exclusively directed the H-Sn bond addition across the triple bond, and therefore to provide a single vinylstannane. To this end we have choosen as ortho-directing group (oDG) a bromine atom, as this substituent would be easy to introduce but also easy to remove at the end of the synthesis by metal-halogen exchange reaction followed by hydrolysis.

Diarylalkynes **5a–d** bearing an ortho bromine atom were prepared according to Scheme 4. **5a** was synthesized from terminal alkyne **6a** by Sonogashira–Linstrumelle (S–L) coupling with 3bromo-4-iodoanisole, readily obtained by regioselective iodination of 3-bromoanisole (94%). For the preparation of diarylalkynes **5b–d**, the synthetic route involves the bromination of 3,4,5-trimethoxy-iodobenzene with molecular Br₂ or MPHT [54] to provide **7** followed by sequential S–L coupling reactions.

The palladium-catalyzed hydrostannation reaction was next applied to ortho substituted diarylalkynes **5a**–**d** (Scheme 5). As expected, a total regiochemical control was achieved by the presence of ortho bromine atom substituent. The reaction provide exclusively α -vinylstannanes **8a**–**d** where the Bu₃Sn group is proximal to the ortho substituted aryl nucleus.

Further iodolysis of **8a–d** was achieved as described for **3** and vinyl iodides **9a–d** were isolated in excellent yields (70–99%) as pure (*E*)-isomers. For the Negishi cross coupling reaction, organozinc species were generated by transmetallation from Grignard reagents at 0 °C using ZnCl₂ except for the synthesis of **10f**.

In this case, the organozinc species was generated after lithiation of 2-fluoro-4-iodoanisole followed by transmetallation with



Scheme 1. Retrosynthetic pathway leading to triarylolefins 1.



Scheme 2. Reagents and conditions: (a) PdCl₂(PPh₃)₂ (2 mol%), Bu₃SnH, (1.2 equiv), THF, 79%. (b) I₂ (1 equiv), CH₂Cl₂, 74%. (c) 3,4,5-(OMe)₃C₆H₂ZnCl (2.2 equiv), PdCl₂(PPh₃)₂ (5 mol%), THF, 99%. (d) 3,4,5-(OMe)₃C₆H₂I (0.9 equiv), Pd(PPh₃)₄ (5 mol%), CuI (10 mol%), CsF (2 equiv), DMF, 60 °C, 54%.

ZnCl₂ as above. As shown in Scheme 5, all organozinc reagents were successfully coupled with the vinyl iodides 9a-d in good yields (50–99%). Finally, removal of the *ortho* bromo directing group was achieved by halogen-metal exchange reaction using *n*-butyl-lithium followed by quenching with aqueous HCl. Subsequent MOM- or TBDMS-ether cleavage furnished the expected triarylolefins 1b-h as pure isomers.

The cytotoxic activity of 12 newly synthesized triarylolefins against human colon carcinoma cell line (HCT-116) was firstly evaluated using *iso*CA-4 [33] and CA-4 [55] as reference compounds. The IC₅₀ values corresponding to the concentration of studied compounds leading to 50% decrease in HCT-116 cell growth are presented in Table 1.

Newly synthesized *ortho*-brominated analogues **10** were not cytotoxic. As it can be observed, most of the triarylolefins **1** retained a light cancer cell growth-inhibitory activity at a micromolar range. In particular, best inhibition results were obtained with hybrids **1e**–**h** bearing the greatest resemblance to CA-4 and *iso*CA-4. However, the cytotoxicities of these triarylolefins are not comparable with those of CA-4 and *iso*CA-4, even though all derivatives of type **1** carry a CA-4 and a *iso*CA-4 (for **1h**) moiety in their structure.

These trisubstituted olefinic combretastatin analogues were next evaluated as tubulin polymerization inhibitors. All samples



exclusive α -isomer

Scheme 3. Retrosynthetic pathway leading to triarylolefins 1b-h.

were dissolved in DMSO, incubated at 37 $^{\circ}$ C for 10 min and at 0 $^{\circ}$ C for 5 min before evaluation of the tubulin assembly rate.

Triarylolefins were tested at different concentrations and the IC_{50} was calculated only for compounds inhibiting tubulin assembly by more than 50% at 9.0×10^{-6} M. The tubulin assembly assay was realized according to a slightly modified Guénard's protocol [56]. The tested analogues show a moderate potency (micromolar level) related to the references compounds CA-4 and *iso*CA-4. When comparing the inhibition of tubulin polymerization *versus* the cell growth-inhibitory effect, we do not find a good correlation for most of the active compounds except for the hybrid **1h**. This hybrid most closely resembling CA-4 and *iso*CA-4 in structure displayed a potent antimitotic activity and a weak cytotoxicity (Table 1).

2.1. Conclusion

The aim of this work was to devise a synthetic strategy to prepare stereodefined triarylolefins **1** as CA-4 and *iso*CA-4 hybrids and to evaluate the influence of the double bond substitution on antitumor activity. From a chemical perspective, the "traceless use" of an ortho bromo substituent to regioselectively control the addition of tributyltin hydride on internal alkynes leading to stannylated stilbenes precursors of trisubstituted olefins **1** is notable. The new CA-4 and *iso*CA-4 analogues were evaluated for their cell growth inhibition and antitubulin activity. Some of these compounds showed interesting antitubulin activities (8 μ M) but are less cytotoxic than CA-4 and *iso*CA-4, indicating that the nature of the olefin substitution in combretastatin series plays an important role on activity.

3. Experimental

3.1. *General considerations*

Tetrahydrofuran (THF) and diethylether were distilled from sodium-benzophenone ketyl. Piperidine, and triethylamine were distilled from potassium hydroxide under argon prior to use. The compounds were all identified by usual physical methods, i.e. ¹H NMR, ¹³C NMR, IR, MS and elemental analysis. ¹H and ¹³C NMR spectra were measured in CDCl₃ with a Bruker Avance 300. ¹H chemical shifts are reported in ppm from an internal standard TMS or of residual chloroform (7.27 ppm). The following abreviation are used: m (multiplet), s (singlet), bs (broad singlet), d (doublet). ¹³C chemical shifts are reported in ppm from the central peak of



Scheme 4. Synthesis of ortho bromodiarylalkynes 5. *Reagents and conditions*: (a) (i) HC=CSiMe₃ (1.5 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), piperidine, 20 °C; (ii) K₂CO₃, MeOH, 92%. (b) Arl (0.7 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), Et₃N/THF (1/1), 60 °C, 5a: 80%; 5b: 86%, 5c: 66%, 5d: 80%. (c) Br₂, AcOH, 92%. (d) (i) HC=CSiMe₃ (1.5 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), Et₃N/THF (1/1), 60 °C, 5a: 80%; 5b: 86%, 5c: 66%, 5d: 80%. (c) Br₂, AcOH, 92%. (d) (i) HC=CSiMe₃ (1.5 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), Et₃N/THF (1/1), 60 °C, 5a: 80%; 5b: 86%, 5c: 66%, 5d: 80%. (c) Br₂, AcOH, 92%. (d) (i) HC=CSiMe₃ (1.5 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), Et₃N/THF (1/1), 60 °C, 5a: 80%; 5b: 86%, 5c: 66%, 5d: 80%. (c) Br₂, AcOH, 92%. (d) (i) HC=CSiMe₃ (1.5 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), Et₃N/THF (1/1), 60 °C, 5a: 80%; 5b: 86%, 5c: 66%, 5d: 80%. (c) Br₂, AcOH, 92%. (d) (i) HC=CSiMe₃ (1.5 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), Et₃N/THF (1/1), 60 °C, 5a: 80%; 5b: 86%, 5c: 66%, 5d: 80%. (c) Br₂, AcOH, 92%. (d) (i) HC=CSiMe₃ (1.5 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), Et₃N/THF (1/1), 60 °C, 5a: 80%; 5b: 86%, 5c: 66%, 5d: 80%. (c) Br₂, AcOH, 92%. (d) (i) HC=CSiMe₃ (1.5 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), Et₃N/THF (1/1), 60 °C, 5a: 80%; 5b: 86%, 5c: 66%, 5d: 80%. (c) Br₂, AcOH, 92%. (d) (i) HC=CSiMe₃ (1.5 equiv), Cul (10 mol%), PdCl₂(PPh₃)₂ (5 mol%), Et₃N/THF (1/1), 60 °C, (ii) K₂CO₃, MeOH, 72%.

deuteriochloroform (77.14). IR spectra were measured on a Bruker Vector 22 spectrophotometer (neat, cm⁻¹). Elemental analyses were performed with a Perkin–Elmer 240 analyser. Mass spectra were obtained with a LCT Micromass spectrometer. Analytical TLC was performed on Merck precoated silica gel 60F plates. Merck silica gel 60 (230–400 mesh) was used for column chromatography.

3.1.1. 3-Bromo-4-iodoanisole

A stirred solution of 3-bromoanisole (500 mg, 2.15 mmol), HgO (354 mg, 1.63 mmol), Ac₂O (0.1 mL) in CH₂Cl₂ (10 mL) was refluxed for 30 min. Then, I₂ (709 mg, 2.79 mmol) was added by 6 portions every 30 min. After refluxing for 12 h and filtration over a pad of celite, the filtrate was washed with a saturated Na₂S₂O₃ solution. The aqueous layer was extracted with CH₂Cl₂ (3 × 10 mL) and the



Scheme 5. Synthesis of triarylolefins 1b-h. Reagents and conditions: (a) PdCl₂(PPh₃)₂ (2 mol%), Bu₃SnH, (1.2 equiv), THF. (b) I₂ (1 equiv), CH₂Cl₂, 20 °C. (c) ArZnCl (2.2 equiv), PdCl₂(PPh₃)₂ (5 mol%), THF. (d) (i) n-BuLi (4.5 equiv), THF, -78 °C. (ii) HCl 1 M, 0.5 h, 20 °C.

Table 1

Cytotoxicity of isoCA on HCT-116 human cancer cell line and antitubulin activities.



^a IC_{50} is the concentration of compound needed to reduce cell growth by 50% following 72 h cell treatment with the tested drug (average of three experiments). ^b $ITP = Inhibition of Tubulin Polymerization; <math>IC_{50}$ is the concentration of compound required to inhibit 50% of the rate of microtubule assembly (average of three

experiments).

^c NA, non active.

combined organic layers were dried with MgSO₄ and evaporated to dryness. Purification by flash chromatography (cyclohexane) afforded the titled compound.

Yield: 94%. TLC: Rf 0.68 (cyclohexane/Et₂O: 95/5). IR (neat) $\nu_{max}/$ cm⁻¹: 2933, 2831, 2361, 2183, 2013, 1579, 1556, 1463, 1436, 1384, 1285, 1260, 1225, 1182, 1093, 1034, 1001. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.68 (d, J = 8.8 Hz, 1H), 7.19 (d, J = 2.8 Hz, 1H), 6.59 (dd, J = 2.8 Hz, J = 8.8 Hz, 1H), 3.77 (s, 3H).

3.1.2. 2-Bromo-1-iodo-3,4,5-trimethoxybenzene (7)

 Br_2 (192 mg) was slowly added to a solution of 1-iodo-3,4,5trimethoxybenzene (1.47 g; 5 mmol) in AcOH (10 mL) and the mixture was stirred over night. After neutralization with NaOH (1 N), the reaction product was extracted with EtOAc (3 × 10 mL). The organic solutions were dried with MgSO₄ and evaporated to dryness. Purification by flash chromatography (cyclohexane/EtOAc: 9/1) afforded the titled compound. Yield: 92%. TLC: Rf 0.68 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 2935, 1562, 1472, 1421, 1371, 1297, 1233, 1102, 1003. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.26 (m, 1H), 7.19 (s, 1H), 3.87 (s, 3H), 3.87 (s, 3H), 3.84 (s, 3H).

3.1.3. 1-Ethynyl-3,4,5-trimethoxybenzene (6a)

To a stirred solution of iodo-3,4,5-trimethoxybenzene (10 mmol) in piperidine (15 mL) was added, Cul (190 mg, 1 mmol), PdCl₂(PPh₃)₂ (351 mg, 0.5 mmol) and trimethylsilylacetylene (15 mmol). The stirred mixture was kept for a night at room temperature and was then treated with HCl 10% (20 mL). After extraction with CH₂Cl₂ (3 × 15 mL), the organic layers were dried with MgSO₄ and evaporated to dryness. Next, the crude mixture was diluted in MeOH (15 mL) containing K₂CO₃ (15 mmol). After stirring for 1 h at room temperature, the black mixture was concentrated under vacuum and treated with a diluted HCl solution until pH = 6. The aqueous layer was extracted with CH₂Cl₂

 $(3 \times 30 \text{ mL})$ and the combined organic layers were dried with MgSO₄ and evaporated to dryness. Purification by flash chromatography afforded **6a**.

Yield: 82%. TLC: Rf 0.47 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 3242, 2989, 2939, 2023, 1967, 1576, 1501, 1450, 1427, 1410, 1329, 1232, 1180, 1125, 1034, 998, 955, 832, 773. ¹H NMR (300 MHz, CDCl₃) δ ppm 6.72 (s, 2H), 3.85 (s, 6H), 3.83 (s, 3H), 3.02 (s, 1H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.1 (2C), 123.68, 117.05, 114.99, 109.35 (2C), 105.21, 83.73, 76.26, 60.99, 56.17 (2C). MS (APCI) *m/z*: 193.22 (M + H)⁺.

3.1.4. Synthesis of 2-bromo-1-ethynyl-3,4,5-trimethoxybenzene (**6b**)

To a stirred solution of **7** (10 mmol) in THF (15 mL) and Et₃N (15 mL) was added, CuI (190 mg, 1 mmol), $PdCl_2(PPh_3)_2$ (351 mg, 0.5 mmol) and trimethylsilylacetylene (15 mmol). The stirred mixture was kept for a night at room temperature and was then treated with HCl 10% (20 mL). After extraction with CH₂Cl₂ (3 × 15 mL), the organic layers were dried with MgSO₄ and evaporated to dryness. Next, the crude mixture was diluted in MeOH (15 mL) containing K₂CO₃ (20 mmol). After stirring for 1 h at room temperature, the black mixture was concentrated under vacuum and treated with a diluted HCl solution until pH = 6. The aqueous layer was extracted with MgSO₄ and evaporated to dryness. Purification by flash chromatography afforded **6b**.

Yield: 72%. TLC: Rf 0.17 (cyclohexane/EtOAc: 95/5). IR (neat) ν_{max}/cm^{-1} : 3291, 2938, 1555, 1477, 1424, 1383, 1336, 1241, 1195, 1166, 1105, 1041, 1004. ¹H NMR (300 MHz, CDCl₃) δ ppm 6.73 (s, 1H), 3.75 (s, 3H), 3.73 (s, 3H), 3.70 (s, 3H), 3.17 (s, 1H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 155.2, 152.6, 119.3, 112.9, 112.6, 80.9, 61.3, 61.0, 56.2 (two C missing).

3.2. Synthesis of internal alkynes 5

To a mixture of required aryl iodide (0.7 mmol), $PdCl_2(PPh_3)_2$ (0.1 mmol), CuI (0.15 mmol), TEA (10 mL) in THF (10 mL) was added dropwise under an argon atmosphere, a solution of **6a** or **6b** (1.0 mmol). The mixture was stirred at 60 °C for a night. Then Et₂O (20 mL) was added to the crude and the mixture was filtered over a short pad of celite. The organic layer was washed with brine (5 mL) twice, separated, dried over MgSO₄, filtered, and concentrated. Purification by flash chromatography afforded internal alkynes **5a**–**d**.

3.2.1. 5-(2-(2-Bromo-4-methoxyphenyl)ethynyl)-1,2,3trimethoxybenzene (**5a**)

Yield: 80%. Anal. calcd for **5a** ($C_{18}H_{17}BrO_4$): C, 57.31; H, 4.54. Found: C, 57.18; H, 4.39. TLC: Rf 0.14 (cyclohexane/EtOAc: 9/1). IR (neat) ν_{max}/cm^{-1} : 3751, 2935, 1601, 1572, 1506, 1457, 1437, 1409, 1351, 1287, 1268, 1221, 1182, 1123, 1027. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.45 (d, J = 8.6 Hz, 1H), 7.15 (d, J = 2.5 Hz, 1H), 6.83 (dd, J = 2.6 Hz, J = 8.7 Hz, 1H), 6.78 (s, 2H), 3.87 (s, 6H), 3.86 (s, 3H), 3.80 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 159.9, 153.2, 138.9, 133.9, 126.3, 118.3, 117.9, 117.5, 113.6, 108.8, 92.3, 87.1, 61.0, 56.2, 55.7.MS (APCI) m/z: 377.0 ((M + H)⁺, ⁷⁹Br), 379.0 ((M + H)⁺, ⁸¹Br).

3.2.2. 2-Bromo-3,4,5-trimethoxy-1-(2-(4-methoxyphenyl) ethynyl) benzene (**5b**)

Yield: 86%. Anal. calcd for **5b** ($C_{18}H_{17}BrO_4$): C, 57.31; H, 4.54. Found: C, 57.11; H, 4.35. TLC: Rf 0.30 (cyclohexane/Et₂O: 9/1). IR (neat) ν_{max}/cm^{-1} : 2936, 2218, 1736, 1604, 1578, 1551, 1511, 1477, 1462, 1427, 1383, 1358, 1295, 1250, 1174, 1098, 1030. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.51 (d, *J* = 8.3 Hz, 2H), 6.90 (s, 2H), 6.89 (d, *J* = 10.2 Hz, 2H), 3.90 (s, 6H), 3.87 (s, 3H), 3.83 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 159.9, 152.6, 151.3, 143.7, 133.2, 120.9, 115.1, 114.1, 112.6, 111.6, 93.2, 86.9, 61.3, 61.1, 56.3, 55.4. MS (APCI) *m/z*: 377.0 ((M + H)⁺, ⁷⁹Br), 379.0 ((M + H)⁺, ⁸¹Br).

3.2.3. (5-(2-(2-Bromo-3,4,5-trimethoxyphenyl)ethynyl)-2methoxyphen-oxy)(tert-butyl)dimethylsilane (**5c**)

Yield: 66%. TLC: Rf 0.35 (cyclohexane/EtOAc: 9/1). IR (neat) ν_{max}/cm^{-1} : 2931, 2855, 2211, 1566, 1508, 1477, 1443, 1413, 1384, 1360, 1288, 1257, 1233, 1189, 1151, 1132, 1107, 1043, 1023. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.00 (dd, J = 1.9 Hz, J = 8.3 Hz, 1H), 6.88 (d, J = 1.7 Hz, 1H), 6.73 (s, 1H), 6.64 (d, J = 8.4 Hz, 1H), 3.74 (s, 3H), 3.73 (s, 3H), 3.70 (s, 3H), 3.66 (s, 3H), 0.83 (s, 9H), 0.00 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 152.6, 151.9, 151.3, 144.8, 143.6, 125.9, 123.9, 120.8, 115.1, 112.6, 111.7, 111.5, 93.2, 86.5, 61.2, 60.9, 56.2, 55.4, 25.7 (3C), 18.5, -4.6 (2C). MS (APCI) m/z: 509.1 (M + H)⁺. Anal. calcd for **5c** (C₂₄H₃₁BrO₅Si): C, 56.80; H, 6.16. Found: C, 56.65; H, 6.05.

3.2.4. 4-(2-(2-Bromo-3,4,5-trimethoxyphenyl)ethynyl)-1-methoxy-2-(methoxymethoxy)benzene (**5d**)

Yield: 80%. Anal. calcd for **5d** ($C_{20}H_{21}BrO_6$): C, 54.93; H, 4.84. Found: C, 54.83; H, 4.80. TLC: Rf 0.20 (cyclohexane/EtOAc: 9/1). IR (neat) ν_{max}/cm^{-1} : 2942, 1556, 1514, 1478, 1421, 1382, 1364, 1317, 1246, 1198, 1151, 1131, 1104, 1074, 1047. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.28 (d, *J* = 1.9 Hz, 1H), 7.17 (dd, *J* = 2.3 Hz, *J* = 8.8 Hz, 1H), 6.84 (s, 1H), 6.80 (d, *J* = 8.4 Hz, 1H), 5.18 (s, 2H), 3.84 (s, 6H), 3.83 (s, 3H), 3.80 (s, 3H), 3.47 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 152.6, 151.3, 150.5, 146.2, 143.7, 126.6, 120.7, 119.3, 115.3, 112.6, 111.7, 95.5, 93.0, 86.7, 61.3, 60.9, 56.3, 56.2, 55.9. MS (APCI) *m/z*: 437.28 ((M + H)⁺, ⁷⁹Br), 439.0 ((M + H)⁺, ⁸¹Br).

3.3. General procedure for the hydrostannation of diarylalkynes

Tributyltin hydride (12 mmol) was added dropwise at room temperature to a solution of $PdCl_2(PPh_3)_2$ (0.1 mmol) and alkyne (10 mmol) in THF (15 mL). The dark brown reaction mixture was stirred for 90 min. Then, the solution was concentrated *in vacuo*. Purification by flash chromatography on silica gel gave the desired products **3** and **8a**–**d**.

3.3.1. (E)-(1,2-bis(4-Methoxyphenyl)vinyl)tributylstannane (3)

Yield: 79%. TLC: Rf 0.82 (cyclohexane/Et₂O: 95/5). IR (neat) ν_{max}/cm^{-1} : 2954, 2920, 2361, 2162, 1965, 1604, 1503, 1462, 1376, 1279, 1242, 1173, 1106, 1071, 1038. ¹H NMR (300 MHz, CDCl₃) δ 6.87 (d, J = 8.7 Hz, 2H), 6.82 (d, J = 8.8 Hz, 2H), 6.73 (d, J = 8.7 Hz, 2H), 6.82 (d, J = 8.8 Hz, 2H), 6.73 (d, J = 8.7 Hz, 2H), 6.55 (d, J = 8.7 Hz, 2H), 6.48 (s, 1H), 3.68 (s, 3H), 3.60 (s, 3H), 1.45–1.35 (m, 6H), 1.29–1.15 (m, 6H), 0.92–0.76 (m, 5H). ¹³C NMR (75 MHz, CDCl₃) δ 158.1, 157.2, 146.8, 138.1, 137.8, 130.6, 130.4 (2C), 127.5 (2C), 114.1 (2C), 113.3 (2C), 55.2, 55.1, 30.6 (3C), 29.0 (3C), 27.5 (3C), 13.7 (3C).

3.3.2. (E)-(1-(2-Bromo-4-methoxyphenyl)-2-(3,4,5-

trimethoxyphenyl) *vinyl*)*tributylstannane* (**8a**)

Yield: 78%. TLC: Rf 0.29 (cyclohexane/EtOAc: 9/1). ¹H NMR (300 MHz, CDCl₃) δ ppm 7.17 (d, J = 2.2 Hz, 1H), 6.87 (d, J = 8.4 Hz, 1H), 6.82 (dd, J = 2.2 Hz, J = 8.4 Hz, 1H), 6.57 (s, 1H), 6.26 (s, 2H), 3.78 (s, 6H), 3.59 (s, 6H), 1.49 (m, 6H), 1.29 (m, 6H), 0.95 (m, 6H), 0. 87 (m, 9H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 158.0, 152.8 (2C), 148.9, 139.1, 138.7, 137.2, 133.2, 128.4, 121.7, 117.7, 114.5, 106.0, 60.9, 55.8 (3C), 28.9 (3C), 27.5 (3C), 13.8 (3C), 10.9 (3C).

3.3.3. (E)-(1-(2-Bromo-3,4,5-trimethoxyphenyl)-2-(4-

methoxyphenyl) vinyl)tributylstannane (**8b**)

Yield: 92%. TLC: Rf 0. 2 (cyclohexane/Et₂O: 95/5). IR (neat) ν_{max}/cm^{-1} : 2928, 2847, 2168, 2029, 1973, 1604, 1554, 1508, 1476, 1420, 1378, 1319, 1286, 1248, 1175, 1161, 1106. ¹H NMR (300 MHz, CDCl₃)

δ ppm 6.95 (d, J = 8.7 Hz, 2H), 6.68 (d, J = 8.7 Hz, 2H), 6.57 (s, 1H), 6.31 (s, 1H), 3.91 (s, 6H), 3.74 (s, 3H), 3.73 (s, 3H), 1.52–1.43 (m, 6H), 1.35–1.23 (m, 6H), 0.98–0.93 (m, 6H), 0.87 (t, J = 7.3 Hz, 9H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 158.4, 153.1, 151.2, 146.5, 142.0, 140.4, 137.9, 130.2, 129.9 (2C), 113.4 (2C), 107.2, 105.9, 61.3, 60.9, 55.9, 55.1, 28.9 (3C), 27.3 (3C), 13.7 (3C), 10.8 (3C). MS (APCI) *m/z*: 669.0 (M + H)⁺.

3.3.4. (E)-(5-(2-(2-Bromo-3,4,5-trimethoxyphenyl)-2-

(tributylstannyl) vinyl)-2-methoxyphen-oxy)(tert-butyl)dimethyl silane (**8c**)

Yield: 75%. TLC: Rf 0.58 (cyclohexane/EtOAc: 9/1). IR (neat) ν_{max}/cm^{-1} : 3836, 2929, 2854, 1556, 1506, 1463, 1423, 1380, 1281, 1231, 1139, 1109, 1013. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.00 (dd, J = 1.9 Hz, J = 8.3 Hz, 1H), 6.88 (d, J = 1.7 Hz, 1H), 6.73 (s, 1H), 6.64 (d, J = 8.4 Hz, 1H), 3.74 (s, 3H), 3.73 (s, 3H), 3.70 (s, 3H), 3.66 (s, 3H), 0.83 (s, 9H), 0.00 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 152.5, 151.5, 149.7, 146.2, 144.6, 142.8, 142.3, 138.9, 127.9, 123.3, 120.1, 111.2, 107.3, 61.3, 60.6, 56.4, 55.4, 29.1 (3C), 27.5 (3C), 25.9 (3C), 18.6, 13.5 (3C), 8.07 (3C), -2.7 (2C).

3.3.5. (*E*)-(1-(2-Bromo-3,4,5-trimethoxyphenyl)-2-(4-methoxy-3-(methoxymethoxy)phenyl)vinyl)tributyl stannane (**8d**)

Yield: 75%. TLC: Rf 0.28 (cyclohexane/EtOAc: 9/1). IR (neat) $\nu_{max}/$ cm⁻¹: 2916, 1737, 1373, 1234, 1044. ¹H NMR (300 MHz, CDCl₃) δ ppm 6.76 (s, 1H), 6.64 (s, 2H), 6.47 (s, 1H), 6.25 (s, 1H), 4.85 (s, 2H), 3.82 (s, 3H), 3.80 (s, 3H), 3.72 (s, 3H), 3.65 (s, 3H), 3.27 (s, 3H), 1.45–1.36 (m, 6H), 1.27–1.15 (m, 6H), 0.91–0.85 (m, 6H), 0.79 (t, J = 7.2 Hz, 9H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.1, 151.2, 148.7, 147.1, 145.8, 141.8, 140.4, 137.7, 130.5, 123.4, 116.1, 111.0, 107.0, 105.8, 95.1, 60.9, 60.7, 55.9, 55.6, 28.8 (3C), 27.2 (3C), 13.5 (3C), 10.8 (3C).

3.4. General procedure for the iodination of **3** and **8a**-**d**

To a CH_2Cl_2 (15 mL) solution containing vinylstannane (0.39 mmol) was added in one portion I_2 (0.39 mmol) at 0 °C. The mixture was then stirred at rt until the disappearance of the starting material (judged by TLC). Then a saturated Na₂S₂O₃ solution (10 mL) was added to the mixture which was extracted with CH₂Cl₂ (3 × 10 mL). The combined organic layers were dried with MgSO₄ and evaporated to dryness. Purification by flash chromatography afforded terminal alkynes **4** and **9a–d**.

3.4.1. (E)-1-Iodo-1,2-bis(4-methoxyphenyl)ethene (4)

Yield: 74%. Anal. calcd for 4 ($C_{16}H_{15}IO_2$): C, 52.48; H, 4.13. Found: C, 52.24; H, 4.07. TLC: Rf 0.68 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 2932, 2938, 1596, 1568, 1504, 1462, 1440, 1421, 1286, 1245, 1174, 1161, 1109, 1024. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.22 (s, 1H), 7.18 (d, J = 8.7 Hz, 2H), 6.80 (d, J = 8.9 Hz, 2H), 6.74 (d, J = 8.7 Hz, 2H), 6.57 (d, J = 8.9 Hz, 2H), 3.73 (s, 3H), 3.65 (m, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 159.3, 158.8, 140.3, 135.6, 130.2 (2C), 130.1, 129.9 (2C), 114.0 (2C), 113.5 (2C), 96.1, 55.3, 55.2. MS (CI) *m/z*: 365.9 (M + H⁺).

3.4.2. (E)-2-Bromo-1-(1-iodo-2-(3,4,5-trimethoxyphenyl)vinyl)-4methoxy benzene (**9**)

Yield: 70%. Anal. calcd for **9a** ($C_{18}H_{18}BrIO_4$): C, 42.80; H, 3.59. Found: C, 42.74; H, 3.57. TLC: Rf 0.13 (cyclohexane/EtOAc: 9/1). ¹H NMR (300 MHz, CDCl₃) δ ppm 7.32 (s, 1H), 7.19 (d, *J* = 8.5 Hz, 1H), 7.15 (d, *J* = 2.6 Hz, 1H), 6.86 (dd, *J* = 8.5 Hz, *J* = 2.6 Hz, 1H), 6.14 (s, 2H), 3.78 (s, 3H), 3.77 (s, 3H), 3.57 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 160.0, 152.9, 142.8, 137.8, 135.9, 132.2, 130.7, 123.2118.4, 114.4, 105.4 (2C), 94.5, 60.8, 55.8, 55.7 (2C). MS (APCI) *m/z*: 500.0 (M + H⁺).

3.4.3. (E)-1-(2-(2-Bromo-3,4,5-trimethoxyphenyl)-2-iodovinyl)-4methoxy benzene (**9b**)

Yield: 85%. Anal. calcd for **9b** ($C_{18}H_{18}BrIO_4$): C, 42.80; H, 3.59. Found: C, 42.59; H, 3.37. TLC: Rf 0.21 (cyclohexane/EtOAc: 95/5). ¹H NMR (300 MHz, CDCl₃) δ ppm 7.31 (s, 1H,), 6.81 (d, *J* = 8.7 Hz, 2H), 6.65 (d, *J* = 8.7 Hz, 2H), 6.62 (s, 1H), 3.93 (s, 3H), 3.91 (s, 3H), 3.79 (s, 3H), 3.74 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 159.5, 151.9, 145.6, 138.8, 136.0, 133.6, 130.3 (2C), 128.6, 113.5 (2C), 107.2, 106.5, 90.7, 60.93, 60.6, 56.1, 55.4. MS (ESI) *m/z*: 528.8 (M + Na⁺).

3.4.4. (E)-(5-(2-(2-Bromo-3,4,5-trimethoxyphenyl)-2-iodovinyl)-2methoxyphen-oxy)(tert-butyl)dimethyl silane (**9**c)

Yield: 99%. TLC: Rf 0.50 (cyclohexane/EtOAc: 9/1). ¹H NMR (300 MHz, CDCl₃) δ ppm 7.26 (s, 1H), 6.67 (d, *J* = 8.4 Hz, 1H), 6.62 (s, 1H), 6.56 (dd, *J* = 8.4 Hz, *J* = 2.2 Hz, 1H), 6.36 (d, *J* = 2.2 Hz, 1H), 3.91 (s, 3H), 3.80 (s, 3H), 3.78 (s, 3H), 3.74 (s, 3H), 0.99 (s, 9H), 0.15 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.3, 151.8, 151.14, 144.6, 143.1, 142.1, 138.7, 129.7, 123.1, 119.9, 111.3, 109.0, 107.9, 92.2, 61.2, 61.1, 56.1, 55.3, 25.6 (3C), 16.4, -4.8 (2C). MS (ESI) *m/z*: 659.0 (M + Na⁺). Anal. calcd for **9c** (C₂₄H₃₂BrIO₅Si): C, 45.37; H, 5.08. Found: C, 45.17; H, 4.98.

3.4.5. (E)-2-Bromo-1-(1-iodo-2-(4-methoxy-3-(methoxymethoxy) phenyl) vinyl)-3,4,5-trimethoxybenzene (**9d**)

Yield: 88%. TLC: Rf 0.49 (cyclohexane/EtOAc: 8/2). IR (neat) $\nu_{max}/$ cm⁻¹: 2984, 1737, 1373, 134, 1044. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.27 (s, 1H), 6.72–6.61 (m, 3H), 6.63 (s, 1H), 4.92 (s, 2H), 3.92 (s, 6H), 3.82 (s, 3H), 3.80 (s, 3H), 3.34 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.4, 151.8, 149.4, 146.1, 143.1, 141.9, 138.5, 129.7, 123.4, 115.2, 110.9, 108.8, 107.8, 95.2, 92.6, 61.1, 61.0, 56.2, 55.8 (2C). Anal. calcd for **9d** (C₂₀H₂₂BrIO₆): C, 42.50; H, 3.92. Found: C, 42.38; H, 3.88.

3.5. General procedure for the PdCl₂(PPh₃)-catalyzed cross coupling reaction of vinyl iodides with arylzinc reagents

To a solution of vinyl iodides **4** or **9a**–**d**, $PdCl_2(PPh_3)_2$ (5 mol%) in THF was added at room temperature, ArZnCl (2 equiv.) prepared by transmetallation from the corresponding Grignard reagent (2.2 equiv.) and anhydrous ZnCl₂ (3 equiv.). The reaction was stirred at room temperature and monitored by TLC until complete consumption of starting materials. The reaction was hydrolyzed at 0 °C with aqueous HCl (1 N), extracted with Et₂O, the organic extract was dried over MgSO₄ and the solvent was removed *in vacuo*. Purification by flash chromatography afforded olefins **1a**, **10a**–**g**.

3.5.1. (E)-5-(1,2-Bis(4-methoxyphenyl)vinyl)-1,2,3-

trimethoxybenzene (**1a**)

Yield: 99%. Anal. calcd for **1a** ($C_{25}H_{26}O_5$): C, 73.87; H, 6.45. Found: C, 73.77; H, 6.38. TLC: Rf 0.31 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 2930, 1603, 1576, 1507, 1463, 1412, 1332, 1285, 1238, 1173, 1119, 1028. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.13 (d, J = 8.7 Hz, 2H), 6.99 (d, J = 8.7 Hz, 2H), 6.87 (d, J = 8.8 Hz, 2H), 6.81 (s, 1H), 6.69 (d, J = 8.8 Hz, 2H), 6.53 (s, 2H), 3.86 (s, 3H), 3.85 (s, 3H), 3.79 (s, 6H), 3.76 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 158.9, 158.3, 152.8 (2C), 140.4, 139.9, 137.6, 132.5, 131.7 (2C), 130.7 (2C), 130.2, 126.9, 113.9 (2C), 113.4 (2C), 104.9 (2C), 60.9, 56.1 (3C), 55.2. MS (CI) *m/z*: 407.2 (M + H⁺).

3.5.2. (Z)-5-(1-(2-Bromo-4-methoxyphenyl)-2-(3,4,5-

trimethoxyphenyl) vinyl)-1,2,3-trimethoxybenzene (10a)

Yield: 95%. Anal. calcd for **10a** ($C_{27}H_{29}BrO_7$): C, 59.46; H, 5.36. Found: C, 59.29; H, 5.24. TLC: Rf 0.13 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 2936, 2836, 1578, 1506, 1452, 1418, 1334, 1283, 1229, 1186, 1122, 1032, 1004. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.23 (d, *J* = 2.5 Hz, 1H), 7.13 (d, *J* = 8.5 Hz, 1H), 6.91 (dd, *J* = 8.5 Hz, $J = 2.5 \text{ Hz}, 1\text{H}, 6.71 \text{ (s, 1H)}, 6.54 \text{ (s, 2H)}, 6.26 \text{ (s, 2H)}, 3.92 \text{ (s, 3H)}, 3.81 \text{ (s, 3H)}, 3.80 \text{ (s, 6H)}, 3.79 \text{ (s, 6H)}, 3.60 \text{ (s, 3H)}. {}^{13}\text{C} \text{ NMR} (75 \text{ MHz}, \text{CDCl}_3) \delta \text{ ppm 159.6}, 153.4 \text{ (2C)}, 153.0 \text{ (2C)}, 152.8, 139.9, 137.6, 137.3, 133.1, 132.5, 132.3, 128.7, 124.9, 118.7, 114.2, 106.2 \text{ (2H)}, 104.0 \text{ (2C)}, 60.9, 60.8, 56.3, 56.1 \text{ (2C)}, 55.6 \text{ (2C)}. \text{ MS} (\text{APCI) } m/z: 547.0 \text{ (M} + \text{H}^+).$

3.5.3. (Z)-1-(1-(2-Bromo-3,4,5-trimethoxyphenyl)-2-(4-methoxyphenyl) vinyl)-4-methoxybenzene (**10b**)

Yield: 99%. Anal. calcd for **10b** ($C_{25}H_{25}BrO_5$): C, 61.86; H, 5.19. Found: C, 61.65; H, 5.04. TLC: Rf 0.41 (cyclohexane/EtOAc: 8/2). ¹H NMR (300 MHz, CDCl₃) δ ppm 7.20 (d, J = 8.4 Hz, 2H), 6.90 (s, 1H), 6.82 (d, J = 8.7 Hz, 2H), 6.78 (d, J = 8.7 Hz, 2H), 6.61 (d, J = 8.4 Hz, 2H), 6.48 (s, 1H), 3.96 (s, 3H), 3.92 (s, 3H), 3.81 (s, 3H), 3.75 (s, 3H), 3.72 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 159.0, 158.5, 153.4, 151.8, 142.5, 138.2, 136.9, 134.0, 130.1 (2C), 129.8, 127.5 (2C), 126.8, 113.8 (2C), 113.6 (2C), 110.8, 110.2, 61.4, 61.2, 56.2, 55.3, 55.2. MS (ESI) *m/z*: 485.2 (M + H⁺).

3.5.4. (Z)-(5-(2-(2-Bromo-3,4,5-trimethoxyphenyl)-2-(4methoxyphenyl) vinyl)-2-methoxyphen-oxy)(tert-butyl)dimethyl silane (**10c**)

Yield: 77%. Anal. calcd for **10c** ($C_{31}H_{39}BrO_6Si$): C, 60.48; H, 6.39. Found: C, 60.32; H, 6.30. TLC: Rf 0.14 (cyclohexane/EtOAc: 9/1). IR (neat) ν_{max}/cm^{-1} : 2203, 2149, 2073, 2020, 1985, 1605, 1570, 1497, 1440, 1388, 1275, 1243, 1178, 1136, 1109, 1040, 1012. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.22 (d, *J* = 7.9 Hz, 2H), 6.88 (s, 1H), 6.84 (d, *J* = 7.9 Hz, 2H), 6.67 (s, 2H), 6.56 (s, 1H), 6.49 (s, 1H), 3.95 (s, 3H), 3.91 (s, 3H), 3.80 (s, 3H), 3.75 (s, 3H), 3.72 (s, 3H), 0.92 (s, 9H), 0.03 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 158.9, 153.3, 151.8, 150.3, 144.4, 142.4, 138.1, 136.8, 134.1, 130.0, 127.5 (2C), 126.9, 123.6, 120.8, 113.8 (2C), 111.4, 110.7, 110.0, 61.2, 61.1, 56.1, 55.3, 55.3, 25.6 (3C), 18.3, -6.1 (2C). MS (APCI) *m/z*: 617.1 (M + H⁺).

3.5.5. (*Z*)-2-Bromo-3,4,5-trimethoxy-1-(2-(4-methoxy-3-(methoxymethoxy) phenyl)-1-p-tolylvinyl)benzene (**10d**)

Yield: 72%. TLC: Rf 0.50 (cyclohexane/EtOAc: 7/3). IR (neat) ν_{max}/cm^{-1} : 2190, 1512, 1263, 1106, 1008. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.22 (d, J = 8.3 Hz, 1H), 7.12 (d, J = 8.1 Hz, 1H), 6.99 (s, 1H), 6.80 (d, J = 1.6 Hz, 1H), 6.76 (t, J = 5.2 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 6.58 (s, 1H), 4.95 (s, 2H), 3.95 (s, 3H), 3.92 (s, 3H), 3.83 (s, 3H), 3.74 (s, 3H), 3.38 (s, 3H), 2.04 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.5, 151.8, 148.8, 145.9, 142.4, 138.7, 138.3, 137.1, 136.6, 130.0 (2C), 129.1, 127.5 (2C), 126.2, 124.1, 116.1, 111.1, 110.6, 110.1, 95.2, 61.1, 61.0, 56.2, 55.9, 55.8. MS (APCI) m/z: 551.0 (M + Na⁺, ⁷⁹Br), 553.1 (M + Na⁺, ⁸¹Br). Anal. calcd for **10d** (C₂₇H₂₉BrO₆): C, 61.25; H, 5.52. Found: C, 61.31; H, 5.52.

3.5.6. (*Z*)-2-(1-(2-Bromo-3,4,5-trimethoxyphenyl)-2-(4-methoxy-3-(methoxymethoxy)phenyl)vinyl) naphthalene (**10e**)

Yield: 72%. Anal. calcd for **10e** $(C_{30}H_{29}BrO_6)$: C, 63.72; H, 5.17. Found: C, 63.45; H, 5.00. TLC: Rf 0.13 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 2989, 1737, 1373, 1236, 1045. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.81 (d, J = 9.2 Hz, 1H), 7.81–7.50 (m, 2H), 7.63 (d, J = 1.8 Hz, 1H), 7.61 (dd, J = 1.9 Hz, J = 3.7 Hz, 1H), 7.44 (d, J = 9.4 Hz, 1H), 7.44 (dd, J = 1.9 Hz, J = 3.1 Hz, 1H), 6.86 (d, J = 1.9 Hz, 1H), 6.82 (dd, J = 1.9 Hz, J = 8.5 Hz, 1H), 6.66 (s, 1H)6.76 (d, J = 8.4 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.6, 151.9, 149.0, 146.0, 142.6, 138.8, 138.6, 136.4, 133.5, 132.7, 129.9, 128.9, 128.3, 127.9, 127.5, 126.1, 125.9, 125.5, 124.4, 124.3, 116.3, 111.2, 110.8, 110.2, 95.2, 61.2, 61.1, 56.2, 55.9, 55.8. MS (APCI) m/z: 587.2 (M + Na⁺, ⁷⁹Br), 589.1 (M + Na⁺, ⁸¹Br).

3.5.7. (*Z*)-2-Bromo-1-(1-(3-fluoro-4-methoxyphenyl)-2-(4methoxy-3-(methoxymethoxy)phenyl)vinyl)-3,4,5trimethoxybenzene (**10**f)

Yield: 50%. Anal. calcd for **10f** (C₂₇H₂₈BrFO₇): C, 57.56; H, 5.01. Found: C, 57.50; H, 4.98. TLC: Rf 0.33 (cyclohexane/EtOAc: 7/3). IR (neat) ν_{max}/cm^{-1} : 2937, 1514, 1481, 1385, 1262, 1156, 1132, 1106, 1079, 1009. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.09–7.03 (m, 1H), 7.02–6.98 (m, 1H), 6.92 (s, 1H), 6.87 (d, J = 8.6 Hz, 1H), 6.76 (d, J = 10.4 Hz, 1H), 6.74 (s, 1H), 6.56 (s, 1H), 4.94 (s, 2H), 3.95 (s, 3H), 3.92 (s, 3H), 3.88 (s, 3H), 3.83 (s, 3H), 3.74 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.6, 152.3 (1C, d, J = 244.9 Hz), 151.9, 148.9, 146.9, 146.8, 146.0, 142.6, 137.4, 136.0, 134.8, 129.7, 127.6, 124.2, 122.2, 116.1, 113.9 (1C, d, J = 19.3 Hz), 113.1, 111.2, 110.6, 110.0, 95.2, 61.2, 61.1, 56.3, 56.2, 55.9, 55.8. MS (APCI) m/z: 441.0 (M + H⁺).

3.5.8. (*Z*)-1-(1,2-Bis(4-methoxy-3-(methoxymethoxy)phenyl) vinyl)-2-bromo-3,4,5-trimethoxybenzene (**10g**)

Yield: 54%. Anal. calcd for **10g** ($C_{29}H_{33}BrO_9$): C, 57.53; H, 5.49. Found: C, 57.08; H, 5.10. TLC: Rf 0.20 (cyclohexane/EtOAc: 7/3). IR (neat) ν_{max}/cm^{-1} : 2984, 1737, 1373, 1235, 1045. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.22 (d, *J* = 1.5 Hz, 1H), 6.91 (s, 1H), 6.81 (d, *J* = 2.0 Hz, 2H), 6.80–6.70 (m, 3H), 6.56 (s, 1H), 5.19 (d, *J* = 0.9 Hz, 2H), 4.92 (s, 2H), 3.93 (s, 3H), 3.89 (s, 3H), 3.84 (s, 3H), 3.80 (s, 3H), 3.72 (s, 3H), 3.48 (s, 3H), 3.35 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.5, 151.8, 149.5, 148.8, 146.3, 145.9, 142.5, 138.2, 136.5, 134.2, 129.9, 126.9, 124.1, 121.0, 116.1, 115.0, 111.5, 111.2, 110.7, 110.2, 95.9, 95.2, 61.1, 60.9, 56.2, 56.2, 55.9, 55.8, 55.8. MS (ESI) *m/z*: 627.2 (M + Na⁺, ⁷⁹Br), 629.2 (M + Na⁺, ⁸¹Br).

3.6. Typical procedure for the synthesis of triarylolefins **1b-h**

To a cooled solution $(-78 \ ^{\circ}\text{C})$ of **10a**–**g** (0.25 mmol) in THF (5 mL) was added a 2.5 M hexane solution of *n*-BuLi (0.5 mL, 1.25 mmol) over 2 min. After 30 min at $-78 \ ^{\circ}\text{C}$ the solution was allowed to warm to rt then HCl 1 N (5 mL) was added and stirring was continued for 1 h. The aqueous phase was separated and extracted with EtOAc, the organic extract was dried over MgSO₄ and the solvent was removed *in vacuo*. Purification by flash chromatography afforded triarylolefins **1b**–**h**.

3.6.1. (E)-1,2,3-Trimethoxy-5-(1-(4-methoxyphenyl)-2-(3,4,5-trimethoxy phenyl)vinyl)benzene (**1b**)

Yield: 91%. Anal. calcd for **1b** ($C_{27}H_{30}O_7$): C, 69.51; H, 6.48. Found: C, 69.41; H, 6.35. TLC: Rf 0.46 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 2954, 2922, 1573, 1505, 1482, 1463, 1417, 1376, 1327, 1280, 1223, 1184, 1125, 1030, 1010. ¹H NMR (400 MHz, CDCl₃) δ ppm: 7.16 (d, J = 8.7 Hz, 2H), 6.89 (d, J = 8.7 Hz, 2H), 6.71 (s, 1H), 6.55 (s, 2H), 6.29 (s, 2H), 3.93 (s, 3H), 3.86 (s, 3H), 3.82 (s, 3H), 3.80 (s, 6H), 3.59 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 159.1, 153.4, 152.9 (2C), 152.6 (2C), 141.8, 139.3, 137.6, 132.9, 132.4, 131.7 (2C), 127.5, 114.1 (2C), 106.7 (2C), 104.9 (2C), 60.9, 56.3, 56.1 (2C), 55.7 (2C), 55.3. MS (ESI) *m/z*: 467.3 (M + H⁺).

3.6.2. (Z)-1-Methoxy-4-(2-(4-methoxyphenyl)-1-(3,4,5-

trimethoxyphenyl) vinyl)benzene (**1c**)

Yield: 65%. Anal. calcd for **1c** ($C_{25}H_{26}O_5$): C, 73.87; H, 6.45. Found: C, 73.73; H, 6.36. TLC: Rf 0.40 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 2934, 1605, 1577, 1508, 1461, 1410, 1280, 1237, 1175, 1124, 1032. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.28 (d, J = 8.8 Hz, 2H), 6.97 (d, J = 8.8 Hz, 2H), 6.86 (d, J = 8.8 Hz, 2H), 6.81 (s, 1H), 6.68 (d, J = 8.8 Hz, 2H), 6.42 (s, 2H), 3.91 (s, 3H), 3.82 (s, 3H), 3.76 (s, 3H), 3.69 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 159.1, 158.3, 153.5 (2C), 139.9, 137.3, 136.2, 135.9, 130.8 (2C), 130.6, 128.5 (2C), 125.9, 113.6 (2C), 113.4 (2C), 107.4 (2C), 61.1, 56.1 (2C), 55.3, 55.2. MS (APCI) m/z: 407.3 (M + H⁺).

3.6.3. (Z)-2-Methoxy-5-(2-(4-methoxyphenyl)-2-(3,4,5-

trimethoxyphenyl) vinyl)phenol (1d)

Yield: 99%. Anal. calcd for **1d** (C₂₅H₂₆O₆): C, 71.07; H, 5.20. Found: C, 70.95; H, 6.14. TLC: Rf 0.36 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 3335, 1561, 1411, 803. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.12 (d, J = 8.8 Hz, 2H), 6.7 (d, J = 8.8 Hz, 2H), 6.61 (s, 1H), 6.51 (d, J = 2 Hz, 1H), 6.47 (d, J = 8.5 Hz, 1H), 6.38 (dd, J = 8.5 Hz, J = 2 Hz, 1H), 6.27 (s, 2H), 3.76 (s, 3H), 3.68 (s, 3H), 3.67 (s, 3H), 3.55 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 159.1, 153.2 (2C), 145.4, 144.9, 140.4, 137.4, 135.9, 135.8, 131.1, 128.5 (2C), 125.9, 121.5, 115.5, 113.6 (2C), 110.2, 107.5 (2C), 61.0, 56.1 (2C), 55.7, 55.3. MS (APCI) m/z: 423.2 (M + H⁺).

3.6.4. (Z)-2-Methoxy-5-(2-p-tolyl-2-(3,4,5-trimethoxyphenyl) vinyl)phenol (**1e**)

Yield: 92%. Anal. calcd for **1e** ($C_{25}H_{26}O_5$): C, 73.87; H, 6.45. Found: C, 73.68; H, 6.34. TLC: Rf 0.33 (cyclohexane/EtOAc: 6/4). IR (neat) ν_{max}/cm^{-1} : 3650, 2227, 2173, 2082, 2004, 1581, 1510, 1279, 1126. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.17 (d, J = 9.0 Hz, 2H), 7.05 (d, J = 8.0 Hz, 2H), 6.74 (s, 1H), 6.59 (d, J = 2.0 Hz, 1H), 6.56 (d, J = 8.4 Hz, 1H), 6.47 (dd, J = 2.0 Hz, J = 8.8 Hz, 1H), 6.35 (s, 2H), 3.84 (s, 3H), 3.77 (s, 3H), 3.65 (m, 6H), 2.29 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.6 (2C), 145.6, 145.1, 140.9, 140.5, 137.4 (2C), 136.1, 131.2, 129.1 (2C), 127.4 (2C), 126.9, 121.8, 115.8, 110.3, 107.6 (2C), 61.2, 56.3 (2C), 56.0, 21.3. MS (APCI) m/z: 407.0 (M + H⁺).

3.6.5. (*Z*)-2-Methoxy-5-(2-(naphthalen-2-yl)-2-(3,4,5trimethoxyphenyl) vinyl)phenol (**1f**)

Yield: 95%. Anal. calcd for **1f** ($C_{28}H_{26}O_5$): C, 76.00; H, 5.92. Found: C, 75.84; H, 5.97. TLC: Rf 0.35 (cyclohexane/EtOAc: 8/2). IR (neat) ν_{max}/cm^{-1} : 2983, 1737, 1373, 1235, 1054. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.81 (d, J = 9.2 Hz, 1H), 7.81–7.50 (m, 2H), 7.63 (d, J = 1.8 Hz, 1H), 7.61 (dd, J = 1.9 Hz, J = 3.7 Hz, 1H), 7.44 (d, J = 9.4 Hz, 1H), 7.44 (dd, J = 1.9 Hz, J = 3.1 Hz, 1H), 6.86 (d, J = 1.9 Hz, 1H), 6.82 (dd, J = 1.9 Hz, J = 8.5 Hz, 1H), 6.66 (s, 1H), 6.76 (d, J = 8.4 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.7 (2C), 145.8, 145.2 (2C), 141.0, 140.8, 137.7, 135.9, 133.5, 132.9, 131.1, 128.4, 127.8, 127.7, 126.6, 126.3, 126.0, 125.6, 121.9, 115.8, 110.3, 107.7 (2C), 61.3, 56.3 (2C), 56.0.

3.6.6. (*E*)-5-(2-(3-Fluoro-4-methoxyphenyl)-2-(3,4,5trimethoxyphenyl) vinyl)-2-methoxyphenol (**1g**)

Yield: 53%. Anal. calcd for **1g** ($C_{25}H_{25}FO_6$): C, 68.17; H, 5.72. Found: C, 68.01; H, 5.66. TLC: Rf 0.20 (cyclohexane/EtOAc: 7/3). IR (neat) ν_{max}/cm^{-1} : 3650, 2983, 2190, 1739, 1373, 1239, 1047. ¹H NMR (300 MHz, CDCl₃) δ ppm 7.10–7.03 (m, 2H), 6.9–6.8 (m, 2H), 6.75 (s, 1H), 6.63 (d, J = 8.3 Hz, 1H), 6.53 (dd, J = 1.9 Hz, J = 8.4 Hz, 1H), 6.40 (s, 2H), 3.91 (s, 3H), 3.90 (s, 3H), 3.83 (s, 3H), 3.71 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.5 (2C), 152.1 (1C, d, J = 244.49 Hz), 145.6, 144.9, 139.4, 137.5, 136.6, 136.5, 135.4, 130.6, 126.9, 123.1, 121.7, 115.5, 114.6 (1C, d, J = 19.19 Hz), 112.9, 110.1, 107.3 (2C), 61.1, 56.3, 56.1, 55.9. MS (ESI) m/z: 587.1 (M + Na⁺).

3.6.7. (*Z*)-5-(2-(3-Hydroxy-4-methoxyphenyl)-2-(3,4,5trimethoxyphenyl) vinyl)-2-methoxyphenol (**1h**)

Yield: 97%. Anal. calcd for **1h** ($C_{25}H_{26}O_7$): C, 68.48; H, 5.98. Found: C, 68.34; H, 5.88. TLC: Rf 0.29 (cyclohexane/EtOAc: 6/4). IR (neat) ν_{max}/cm^{-1} : 2935, 1736, 1580, 1508, 1127, 1045. ¹H NMR (300 MHz, CDCl₃) δ ppm 6.88 (d, J = 1.6 Hz, 1H), 6.75–6.73 (m, 2H), 6.69 (s, 1H), 6.57 (d, J = 1.6 Hz, 1H), 6.55 (d, J = 8.3 Hz, 1H), 6.44 (dd, J = 1.6 Hz, J = 8.4 Hz, 1H), 6.34 (s, 2H), 5.53 (s, 1H), 5.38 (s, 1H), 3.83 (s, 3H), 3.82 (s, 3H), 3.75 (s, 3H), 3.63 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ ppm 153.6 (2C), 16.3, 145.6, 145.4, 145.1, 140.5, 137.5, 136.9, 135.9, 131.1, 126.5, 121.7, 119.4, 115.7, 113.7, 110.4, 110.3, 107.5 (2C), 61.2, 56.3, 56.1, 56.0.

3.7. Biolology

3.7.1. Cell culture and proliferation assay

Cancer cell lines were obtained from the American type Culture Collection (Rockville, MD) and were cultured according to the supplier's instructions. HCT-116 colorectal carcinoma cells were grown in RPMI 1640 containing 10% FCS and 1% glutamine. Cell lines were maintained at 37 °C in a humidified atmosphere containing 5% CO₂. Cell viability was assessed using Promega CellTiter-Blue TM reagent according to the manufacturer's instructions. Cells were seeded in 96-well plates (5×10^3 cells/well) containing 50 µL growth medium. After 24 h of culture, the cells were supplemented with 50 µL of the tested compound dissolved in DMSO (less than 0.1% in each preparation). After 72 h of incubation, 20 µL of resazurin was added for 2 h before recording fluorescence (λ ex = 560 nm, λ em = 590 nm) using a Victor microtiter plate fluorimeter (Perkin–Elmer,USA). The IC₅₀ corresponds to the concentration of the tested compound that caused a decrease of 50% in fluorescence of drug treated cells compared with untreated cells. Experiments were performed in triplicate.

3.7.2. Tubulin binding assay

Sheep brain tubulin was purified according to the method of Shelanski [57] by two cycles of assembly–disassembly and then dissolved in the assembly buffer containing 0.1 M MES, 0.5 mM MgCl₂, 1 mM EGTA, and 1 mM GTP, pH 6.6 (the concentration of tubulin was about 2–3 mg/mL). Tubulin assembly was monitored and recorded continuously by turbidimetry at 350 nm in a UV spectrophotometer equipped with a thermostatted cell at 37 °C. The GI₅₀ value of each compound was determined as the concentration which decreased the maximum assembly rate of tubulin by 50% compared to the rate in the absence of compound. The GI₅₀ values for all compounds were compared to the GI₅₀ of Ca-4 and *iso*CA-4 and measured the same day under the same conditions.

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References

- [1] A. Jordan, J.A. Hadfield, N.J. Lawrence, A.T. Mc Gown, Med. Res. Rev. 18 (1998) 259-296.
- [2] T. Beckers, S. Mahboobi, Drugs Future 28 (2003) 767–785.
- [3] F. Gueritte, J. Fahy, in: G.M. Cragg, D.G.I. Kingston, D.J. Newman (Eds.), Anticancer Agents from Natural Products, CRC Press, L. Boca Raton, 2005, pp. 123–135.
- [4] D.G.I. Kingston, in: G.M. Cragg, D.G.I. Kingston, D.J. Newman (Eds.), Anticancer Agents from Natural Products, CRC Press, L, Boca Raton, 2005, pp. 89–122.
- [5] R.J. Freilich, C. Balmaceda, A.D. Seidman, M. Rubin, L.M. DeAngelis, Neurology 47 (1996) 115–118.
- [6] P.H. Hilkens, J. Verweij, C.J. Vecht, G. Stoter, M.J. van den Bent, Ann. Oncol. 8 (1997) 187–190.
- [7] A.T. Fojo, M. Menefee, Semin. Oncol. 32 (2005) S3–S8.
- [8] C. Dumontet, B.I. Sikic, J. Clin. Oncol. 17 (1999) 1061-1070.
- [9] G.R. Pettit, S.B. Singh, E. Hamel, C.M. Lin, D.S. Alberts, D. Garcia-Kendall, Experientia 45 (1989) 209–211.
- [10] G.R. Pettit, S.B. Singh, M.R. Boyd, E. Hamel, R.K. Pettit, J.M. Schmidt, F. Hogan, J. Med. Chem. 38 (1995) 1666–1672.
- [11] R. Singh, H. Kaur, Synthesis (2009) 2471–2497.
- [12] G.C. Tron, T. Pirali, G. Sorba, F. Pagliai, S. Busacca, A.A. Genazzani, J. Med. Chem. 49 (2006) 3033–3044.
- [13] A. Cirla, J. Mann, Nat. Prod. Rep. 20 (2003) 558–564.
- [14] G.R. Pettit, M.R. Rhodes, D.L. Herald, E. Hamel, J.M. Schmidt, R.K. Pettit, J. Med. Chem. 48 (2005) 4087–4099.
- [15] A.T. McGown, B.W. Fox, Cancer Chemother. Pharmacol. 26 (1990) 79-81.
- [16] G.R. Pettit, C. Temple, V.L. Narayanan, R. Varma, M.J. Simpson, M.R. Boyd, G. A. Rener, N. Bansal, Anticancer Drug Des. 10 (1995) 299–309.
- [17] D.W. Siemann, D.J. Chaplin, P.A. Walicke, Expert. Opin. Investig. Drugs 18 (2009) 189–197.
- [18] N.E. Mealy, B. Lupone, M. Balcell, Drugs Future 31 (2006) 547-548.
- [19] D.M. Patterson, G.J.S. Rustin, Drugs Future 32 (2007) 1025–1032.
- [20] K. Oshumi, R. Nakagawa, Y. Fukuda, T. Hatanaka, T. Tsuji, J. Med. Chem. 41 (1998) 3022–3032.
- [21] G.M. Tozer, C. Kanthou, C.S. Parkins, S.A. Hill, Int. J. Exp. Pathol. 83 (2002) 21-38.
- [22] G.D. Dark, S.A. Hill, V.E. Prise, G.M. Tozer, G.R. Pettit, D.J. Chaplin, Cancer Res. 57 (1997) 1829–1834.
- [23] G.M. Tozer, C. Kanthou, B.C. Baguley, Nat. Rev. Cancer 5 (2005) 423-435.

- [24] J. Griggs, J.C. Metcalfe, R. Hesketh, Lancet. Oncol. 2 (2001) 82-87.
- [25] G.M. Tozer, V.E. Prise, J. Wilson, R.J. Locke, B. Vojnovic, M.R.L. Stratford, M.
- F. Dennis, D.J. Chaplin, Cancer Res. 59 (1999) 1626–1634. [26] D.J. Chaplin, M.R. Horsman, D.W. Siemann, Curr. Opin. Invest. Drugs 7 (2006)
- 522-528. [27] A.M. Gaya, G.J.S. Rustin, Clin. Oncol. 17 (2005) 277-290.
- [28] J.W. Lippert, Bioorg. Med. Chem. 15 (2007) 605-615.
- [29] G.R. Pettit, B. Toki, D.L. Herald, M.R. Boyd, E. Hamel, R.K. Pettit, G.C. Chapuis, I. Med. Chem. 42 (1999) 1459–1465.
- [30] O. Provot, A. Giraud, J.-F. Peyrat, M. Alami, J.-D. Brion, Tetrahedron Lett. 46 (2005) 8547-8550.
- [31]] C. Mousset, A. Giraud, J. O.ProvotA.HamzeBignon, J.M. Liu, S. Thoret, J. Dubois,
- J.-D. Brion, M. Alami, Bioorg. Med. Chem. Lett. 18 (2008) 3266-3271. [32]] C. Mousset, O. Provot, A. Hamze, J. Bignon, J.-D. Brion, M. Alami, Tetrahedron 64 (2008) 4287-4294.
- [33] S. Messaoudi, B. Tréguier, A. Hamze, O. Provot, J.-F. Peyrat, J.R. Rodrigo De Losada, J.-M. Liu, J. Bignon, J. Wdzieczak-Bakala, S. Thoret, J. Dubois, J.-D. Brion, M. Alami, J. Med. Chem. 52 (2009) 4538–4542.
- [34] M. Alami, J.-D. Brion, O. Provot, J.-F. Peyrat, S. Messaoudi, A. Hamze, A. Giraud, J. Bignon, J. Bakala, J.-M. Liu, WO 122620 A1, 2008.
- [35] A. Hamze, D. Veau, O. Provot, J.-D. Brion, M. Alami, J. Org. Chem. 74 (2009) 1337 - 1340
- [36] B. Tréguier, A. Hamze, O. Provot, J.-D. Brion, M. Alami, Tetrahedron Lett. 50 (2009) 6549 - 6552.
- [37] A. Hamze, A. Giraud, S. Messaoudi, O. Provot, J.-F. Peyrat, J. Bignon, J.-M. Liu, J. Wdzieczak-Bakala, S. Thoret, J. Dubois, J.-D. Brion, M. Alami, ChemMedChem 4 (2009) 1912–1924.
- [38] M. Clarke, R. Collins, C. Davies, J. Godwin, R. Gray, R. Peto, The Lancet 351 (1998) 1451-1467 (and references therein).

- [39] C. Borrel, S. Thoret, X. Cachet, D. Guénard, F. Tillequin, M. Koch, S. Michel, Bioorg. Med. Chem. 13 (2005) 3853-3864.
- [40] K. Ohsumi, R. Nakagawa, Y. Fukuda, T. Hatanaka, Y. Morinaga, Y. Nihei, K. Ohishi, Y. Suga, Y. Akiyama, T. Tsuji, J. Med. Chem. 41 (1998) 3022-3032.
- [41] D. Alloatti, G. Giannini, W. Cabri, I. Lustrati, M. Marzi, A. Ciacci, G. Gallo, M. O. Tinti, M. Marcellini, M.B. Gugliemi, P. Carminati, C. Pisano, J. Med. Chem. 51 (2008) 2708-2771.
- [42] K. Sonogashira, Y. Tohda, N. Hagigara, Tetrahedron Lett. 16 (1975) 4467-4470.
- [43] M. Alami, F. Ferri, G. Linstrumelle, Tetrahedron Lett. 34 (1993) 6403-6406.
- F. Liron, P. Le Garrec, M. Alami, Synlett (1999) 246-248. [44]
- [45] M. Alami, F. Liron, M. Gervais, J.-F. Peyrat, J.-D. Brion, Angew. Chem., Int. Ed. 41 (2002) 1578-1580.
- [46] A. Hamze, O. Provot, J.-D. Brion, M. Alami, J. Org. Chem. 72 (2007) 3868–3874.
- M. Alami, F. Ferri, Y. Gaslain, Tetrahedron Lett. 37 (1996) 57-60. Ì47Ì
- [48] S.P.H. Mee, V. Lee, J.E. Baldwin, Chem. Eur. J. 11 (2005) 3294-3308.
- [49] S.-M. Chen, R.E. Schaub, C. Grudzinskas, J. Org. Chem. 72 (2007) 3868-3874. [50] F. Liron, M. Gervais, J.-F. Peyrat, M. Alami, J.-D. Brion, Tetrahedron Lett. 44 (2003) 2789–2792.
- M. Bujard, F. Ferri, M. Alami, Tetrahedron Lett. 39 (1998) 4243-4246. [51]
- [52] A. Hamze, O. Provot, M. Alami, J.-D. Brion, Org. Lett. 7 (2005) 5625–5628.
 [53] A. Hamze, O. Provot, M. Alami, J.-D. Brion, Synthesis (2007) 2025–2036.
- [54] A. Bekaert, O. Provot, O. Rasolojahona, M. Alami, J.-D. Brion, Tetrahedron Lett. 46 (2005) 4187-4191.
- [55] A. Giraud, O. Provot, A. Hamze, J.-D. Brion, M. Alami, Tetrahedron Lett. 49 (2008) 1107 - 1110
- [56] F. Zavala, D. Guénard, J.-P. Robin, E. Brown, J. Med. Chem. 23 (1980) 546-549. [57] M.L. Shelanski, F. Gaskin, C.R. Cantor, Proc. Natl. Acad. Sci. U.S.A. 70 (1973) 765 - 768