

Relationship of Molecular Structure to in Vivo Scintigraphic Distribution of Carbon-11-Labeled Compounds. 4. Carbon-11-Labeled Mandelonitriles, Mandelic Acids, and Their Esters

Meldrum B. Winstead,* Dennis A. Dougherty,

Bucknell University, Lewisburg, Pennsylvania 17837

Tz-Hong Lin, Archie Khentigan, James F. Lamb, and H. Saul Winchell

Medi-Physics, Inc., Emeryville, California 94608. Received August 26, 1977

¹¹C-Labeled HCN was collected in water containing carrier KCN following bombardment of 99% N₂-1% H₂ with 22 MeV protons. ¹¹C-Labeled mandelonitrile and *p*-methoxy-, *p*-hydroxy-, and 3,4-dihydroxymandelonitrile were synthesized from K¹¹CN and the corresponding benzaldehyde. The initial distribution of ¹¹C activity of these nitriles in dogs was primarily in the region of the heart, liver, and kidneys followed by rapid redistribution to the parotids and stomach with the [¹¹C]hydroxymandelonitriles. ¹¹C-Labeled mandelic acid and *m*-methyl-, *o*-chloro-, and *p*-chloromandelic acid were synthesized from the corresponding [¹¹C]mandelonitrile. Serial scintigraphy of ¹¹C activity of these mandelic acids in dogs showed progressive renal excretion with accumulation of activity in the bladder. ¹¹C-Labeled ethyl and benzyl mandelate were synthesized from [¹¹C]mandelic acid. These esters showed initial accumulation of activity in the lungs with eventual excretion by the kidneys.

Mandelonitrile has been proposed as the "active antitumor" principal arising from the hydrolysis of amygdalin,¹ and mandelonitrile derivatives have been employed as precursors in the synthesis of ¹¹C-labeled neurohumeral transmitters.² Mandelic acid has been used clinically as a urinary antiseptic,³ and esters of mandelic acid have been shown to possess antispasmodic properties.⁴

The diverse pharmacologic properties of this family of related compounds and the ability to achieve their rapid synthesis incorporating positron-emitting, 20.4-min half-life ¹¹C attracted our attention. The results of their synthesis and in vivo evaluation are the subject of this communication. In previous articles we reviewed the rationale for employing serial scintigraphy for studying the distribution of families of related ¹¹C-labeled compounds.⁵⁻⁷

Experimental Section

Materials and Methods. ¹¹C-Labeled mandelonitrile^{8a} and *p*-methoxy-,^{8b} *p*-hydroxy-,⁹ and 3,4-dihydroxymandelonitrile¹⁰ were prepared from K¹¹CN and the corresponding benzaldehyde-bisulfite addition product and were dissolved in ethanol or water prior to intravenous administration to the dog. The ¹¹C-labeled mandelic acids were synthesized by modifying the procedure for preparing [¹⁴C]mandelic acid.¹¹ H¹¹CN was produced on bombardment of 99% N₂-1% H₂ with 30-40 μA of 22 MeV protons for 60-90 min.¹² A solution of K¹¹CN formed by exchange between H¹¹CN and 5 mmol of carrier KCN in water was added to 7.5 mmol of the appropriate benzaldehyde. The reaction mixture was kept below 5 °C while 1.1 g of NaHSO₃ in water was added followed by stirring for 20 min. The mandelonitrile derivative was extracted with benzene and added to 20 mL of concentrated hydrochloric acid. The mixture was boiled to dryness, the residue extracted with hot benzene, and the extract filtered and added to hexane. Upon cooling the [¹¹C]mandelic acid crystallized and was dissolved in 7.5% NaHCO₃ prior to administration to the dog.

¹¹C-Labeled ethyl mandelate was prepared as follows. ¹¹C-Labeled 2,2-dimethyl-5-phenyl-1,3-dioxolan-4-one was prepared from [¹¹C]mandelic acid and acetone.¹³ This product was dissolved in ethanol, hydrogen chloride gas was briefly bubbled into the solution and evaporated, and the residue of [¹¹C]ethyl mandelate was dissolved in Me₂SO prior to its injection to the dog.

¹¹C-Labeled benzyl mandelate was prepared as follows. A solution of 1-*p*-tolyl-3-benzyltriazine in benzene was added to a solution of ¹¹C-labeled mandelic acid in benzene.¹⁴ The reaction mixture was heated to boiling, then cooled, and extracted successively with 10% HCl and 10% Na₂CO₃. Following evaporation of the benzene, the residue of [¹¹C]benzyl mandelate was recrystallized from hexane, dissolved in Me₂SO, and administered to the dog.

Prior to the ¹¹C reaction each mandelonitrile, mandelic acid, and ester was prepared from nonradioactive cyanide under experimental conditions designed to optimize the yield at minimal reaction times. The resulting products were isolated and purified, and their melting points and infrared and NMR spectra were determined as a confirmation of their respective data reported in the literature.

The animals were fasted overnight before being surgically anesthetized with nembutal. The in vivo distribution of the intravenously administered ¹¹C compounds was determined scintigraphically using a Searle Radiographics HP scintillation camera fitted with a pinhole collimator containing a 1-cm diameter hyperbolic aperture. In several cases the dog was sacrificed. Quantitative estimates of organ and tissue distribution of activity were obtained by placing the weighed organs and tissues obtained at necropsy at fixed geometric positions in front of a thallium-activated NaI scintillation detector and analyzing detected scintillation events using routine methods.

Results

A. Synthesis. ¹¹C-Labeled mandelonitrile and *p*-methoxymandelonitrile were prepared from 275 and 417 mCi of K¹¹CN (at *t*₀) in radiochemical yields of 6 and 5% in a total synthesis time of 50 and 60 min, respectively. ¹¹C-Labeled *p*-hydroxymandelonitrile and 3,4-dihydroxymandelonitrile were prepared from 722 and 827 mCi of K¹¹CN in radiochemical yields of 1 and 3% in a total synthesis time of 59 and 65 min, respectively.

The results obtained in the synthesis of ¹¹C-labeled mandelic acids are listed in Table I. In each case the melting point and infrared spectrum of the product agreed with both that obtained in prior nonradioactive preparations and with the respective data reported in the literature.

¹¹C-Labeled 2,2-dimethyl-5-phenyl-1,3-dioxolan-4-one was prepared from 469 mCi of K¹¹CN via [¹¹C]mandelic acid in a radiochemical yield of 17.5% in a total synthesis time of 63 min: mp 42-43 °C (lit.¹³ mp 45 °C). ¹¹C-Labeled ethyl mandelate was prepared in a four-step synthesis from 747 mCi of K¹¹CN via mandelonitrile, mandelic acid, and the dioxolanone in a radiochemical yield of 30% in an overall reaction time of 91 min. ¹¹C-Labeled benzyl mandelate was prepared in an 8.5% radiochemical yield from 925 mCi of K¹¹CN in a total synthesis time of 92 min via [¹¹C]mandelic acid: mp 90.5-92 °C (lit.⁴ mp 90-91 °C). The infrared and NMR spectrum of the dioxolanone and esters agreed with that obtained in prior nonradioactive preparations as well as that reported.

Table I. Preparation of Carbon-11-Labeled Mandelic Acids

| K ¹¹ CN, mCi, t ₀ | Ar (5-mmol rxn scale) | Product | | Radio- chemical yield, % | Chemical yield, % | Total synth time, min | Mp, °C |
|--|---|---------------------|-----------------------|--------------------------------|----------------------|--------------------------|----------------------|
| | | mCi, t ₀ | mCi, t _{inj} | | | | |
| 377 | C ₆ H ₅ | 141 | 16 | 37 | 51 | 65 | 119–120 ^a |
| 519 | <i>m</i> -CH ₃ C ₆ H ₄ | 209 | 20 | 40 | 48 | 70 | 90–91 ^b |
| 1823 | <i>o</i> -ClC ₆ H ₄ | 502 | 56 | 28 | 31 | 65 | 78–80 ^c |
| 353 | <i>p</i> -ClC ₆ H ₄ | 88 | 11 | 25 | 49 | 63 | 118–119 ^d |

^a Lit.¹⁵ mp 119–120 °C. IR spectrum agreed with Sadtler Standard Prism Spectrum No. 13457. ^b Lit.¹⁶ mp 93–94 °C. ^c Lit.¹⁷ mp 84–85 °C. IR spectrum agreed with Sadtler Standard Grating Spectrum No. 13445. ^d Lit.¹⁸ mp 119–120 °C. IR spectrum agreed with Sadtler Standard Prism Spectrum No. 18490.

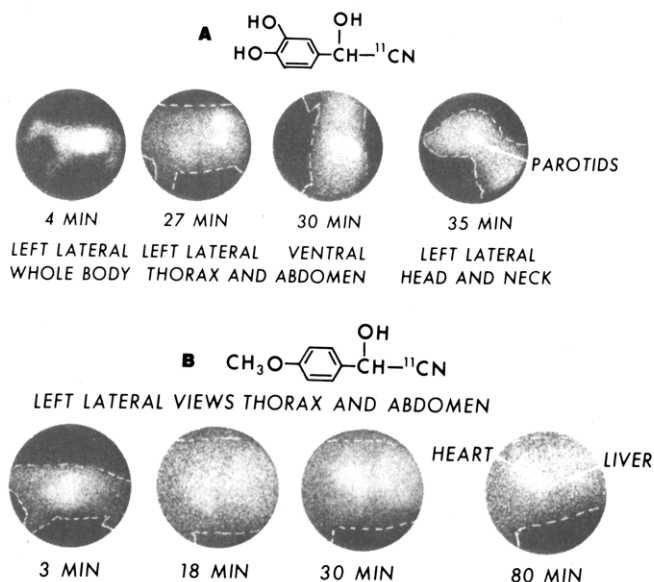


Figure 1. Distribution of ¹¹C activity of 3,4-dihydroxymandelonitrile (A) and *p*-methoxymandelonitrile (B) in dog. With [¹¹C]-3,4-dihydroxymandelonitrile early images show activity predominantly in the blood pool followed by progressive concentration of activity in the parotids and stomach. Activity is seen with this compound in the region of the heart at 27 min in the left view of the thorax. With [¹¹C]-*p*-methoxymandelonitrile scintiphotos at 3–5 min show activity predominately in the blood pool followed by progressive concentration of activity in the region of the heart greater than that in the liver in the left lateral views of the thorax at 18, 30, and 80 min.

B. In Vivo Distribution. Serial scintiphotos of ¹¹C activity in the dog following intravenous administration of [¹¹C]-3,4-dihydroxymandelonitrile and [¹¹C]-*p*-methoxymandelonitrile are shown in Figure 1. The in vivo distribution of [¹¹C]-*p*-hydroxymandelonitrile is comparable to that of [¹¹C]-3,4-dihydroxymandelonitrile in which scintiphotos at 1–4 min show activity predominately in the blood pool followed by progressive concentration of activity in the parotids and stomach. Greater activity is seen in the region of the heart than in the liver in the left lateral view of the thorax at 27 min. This pattern is perhaps more apparent in the left lateral views of the thorax at 18, 30, and 80 min in the study with [¹¹C]-*p*-methoxymandelonitrile. Since reduction of [¹¹C]-3,4-dihydroxymandelonitrile yields [¹¹C]norepinephrine, which has been shown to localize in the myocardium,¹⁹ it is intriguing to consider whether reduction of [¹¹C]mandelonitriles can occur in the body resulting in endogenous production and myocardial localization of ¹¹C-labeled neurohumeral amine-like agents.

Serial scintiphotos of ¹¹C activity in the dog following intravenous administration of [¹¹C]-*m*-methylmandelic acid are shown in Figure 2. Initial distribution of activity in

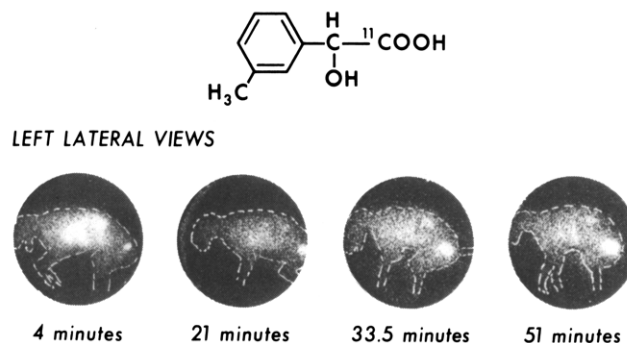


Figure 2. Urinary clearance of ¹¹C activity of *m*-methylmandelic acid as a function of time. Initial distribution of activity in the blood pool is followed by progressive renal excretion with accumulation of activity in the bladder. Similar distributions were observed with *o*- and *p*-chloromandelic acid and mandelic acid itself.

Table II. Comparative Carbon-11 Activity in Tissues Relative to Temporal Muscle Following Intravenous Administration of [¹¹C]-*o*-Chloromandelic Acid

| Organ | (cpm/g of tissue)/ (cpm/g of temporal muscle) | |
|------------------|--|-------------|
| | I (34 min) | II (40 min) |
| Brain | 0.23 | 0.16 |
| Lungs | 1.08 | 1.60 |
| Heart (washed) | 1.15 | 1.06 |
| Temporal muscle | 1.00 | 1.00 |
| Liver | 1.38 | 1.79 |
| Bile | 1.23 | 0.85 |
| Spleen | 1.08 | 1.46 |
| Stomach | 1.35 | 1.17 |
| Pancreas | 4.54 | 1.18 |
| Kidneys | 4.46 | 6.26 |
| Blood | 13.77 | |
| Mesenteric nodes | | 2.28 |
| Mesenteric fat | | 0.47 |
| Urine | 36.77 | 295 |
| Adrenal | | 3.73 |
| CSF | 2.00 | |

the blood pool is followed by progressive renal excretion with accumulation of activity in the bladder. Comparable results were seen with mandelic acid and with *p*-chloro- and *o*-chloromandelic acid.

Organ distribution of ¹¹C activity in two dogs sacrificed at 34 and 40 min following administration of [¹¹C]-*o*-chloromandelic acid is shown in Table II. Activity in lungs, heart, liver, bile, spleen, and stomach is more or less comparable. Activity in brain is substantially lower and that in kidneys, blood, and urine substantially higher than activity in other tissues.

The water-insoluble [¹¹C]ethyl and [¹¹C]benzyl mandelates were administered in Me₂SO. Scintiphotos taken within 3 min showed substantial activity retained

in the lungs, presumably related to intravascular precipitation of the agent and mechanical lodging of the precipitate in the pulmonary capillary bed. Subsequent serial scintiphotos showed clearance of activity from the lungs in 5–10 min and slow renal excretion of activity which appeared to be delayed 15–20 min in time relative to the corresponding [¹¹C]mandelic acid.

¹¹C-Labeled 2,2-dimethyl-5-phenyl-1,3-dioxolan-4-one was administered in Me₂SO and was initially retained in the lungs. However, clearance from the lungs was rapid and concentration of ¹¹C activity in the kidneys and bladder was prompt and comparable to that seen with [¹¹C]mandelic acid. Moreover, absorption of expired air from the dog in concentrated Ba(OH)₂ showed the presence of ¹¹CO₂.

Discussion

It has been postulated that certain nitriles such as amygdalin liberate cyanide in vivo and that such regional release of cyanide might be of use in cancer chemotherapy.¹ The scintigraphic evaluation of the mandelonitriles supports this notion in that the eventual distribution of [¹¹C]-hydroxymandelonitrile derivatives is similar to that noted with [¹¹C]cyanide,²⁰ the dissociation product of the [¹¹C]mandelonitriles. It is not clear whether certain nitriles such as *p*-methoxymandelonitrile or mandelonitrile itself can be reduced in vivo to form the corresponding amine. Our results are consistent with but do not establish this possibility. We are not aware of any literature reports describing such a metabolic pathway in animals. However, reduction of nitriles in plants has been suggested.^{21,22}

The findings of similar scintigraphic distribution following administration of *m*-methyl-, *p*-chloro-, and *o*-chloromandelic acid and mandelic acid itself suggest that the metabolism of these compounds does not involve aromatic ring hydroxylation, as is the case for many aromatic pharmaceuticals. The –CH(OH)COOH moiety of these mandelic acids appears to determine their renal excretion kinetics, probably through the kidneys' organic acid excretory pathways.

The key role of the free carboxyl group of the mandelic acids is underscored by the delay in renal excretion of ¹¹C activity of the ethyl and benzyl esters. The prompt excretion of activity with the dioxolanone derivative further suggests that hydrolysis of these esters proceeds far more slowly than hydrolysis of the dioxolanone.

It has been reported that phenylglyoxylic acid is a metabolite of mandelic acid.²³ Consequently, the detection of activity in the expired breath of the dog suggests that [¹¹C]mandelic acid produced by in vivo hydrolysis of the [¹¹C]dioxolanone may be undergoing metabolic oxidation to [¹¹C]phenylglyoxylic acid with subsequent decarboxylation to ¹¹CO₂.

The tissue distribution study performed after administration of *o*-chloromandelic acid showed a disproportionately high blood level of activity suggesting the presence of binding sites for this mandelic acid. The relatively high concentration of activity in the pancreas

of one dog and the adrenal of another is unexplained.

Acknowledgment. The authors thank Eric Knutson, Michael Rogal, and Richard Stechel of Bucknell University and Michael Garrett, Yu Cheun Lee, Chris Coyle, and Dennis Lum of Medi-Physics for their contribution to this study. This work was supported in part by Atomic Energy Commission Contract AT(04-3)-849 to Medi-Physics, Inc., and by the National Science Foundation Undergraduate Research Participation Program, the Camille and Henry Dreyfus Foundation, Inc., and the National Institutes of Health Grant GM 20073 to Bucknell University. This paper was presented at the 9th Middle Atlantic Regional Meeting of the American Chemical Society, Wilkes-Barre, Pa., April 23–26, 1974.

References and Notes

- (1) B. J. Culliton, *Science*, **182**, 1000 (1973).
- (2) J. S. Fowler, R. R. MacGregor, A. N. Ansari, H. L. Atkins, and A. P. Wolf, *J. Med. Chem.*, **17**, 246 (1974).
- (3) H. F. Helmholz, *J. Am. Med. Assoc.*, **109**, 1039 (1937).
- (4) A. B. H. Funcke, M. J. E. Ernsting, R. F. Rekker, and W. T. Navta, *Arzneim.-Forsch.*, **3**, 503 (1953).
- (5) M. B. Winstead, J. F. Lamb, and H. S. Winchell, *J. Nucl. Med.*, **14**, 747 (1973).
- (6) M. B. Winstead, P. J. Widner, J. L. Means, M. A. Engstrom, G. E. Graham, A. Khentigan, T. H. Lin, J. F. Lamb, and H. S. Winchell, *J. Nucl. Med.*, **16**, 1049 (1975).
- (7) M. B. Winstead, S. J. Parr, M. J. Rogal, P. S. Brockman, R. Lubcher, A. Khentigan, T. H. Lin, J. F. Lamb, and H. S. Winchell, *J. Med. Chem.*, **19**, 279 (1976).
- (8) B. Prager, P. Jacobson, P. Schmidt, and D. Stern, Ed., "Beilsteins Organische Chemie", Vol. 10, Springer-Verlag, Berlin, 1927, (a) p 206, (b) p 411.
- (9) K. Ladenberg, K. Folkers, and R. T. Major, *J. Am. Chem. Soc.*, **58**, 1294 (1936).
- (10) K. Shaw, A. McMillan, and M. D. Armstrong, *J. Org. Chem.*, **23**, 27 (1958).
- (11) A. Murray, and D. L. Williams, "Organic Syntheses with Isotopes. Part I. Compounds of Isotopic Carbon", Interscience, New York, N.Y., 1958, p 146.
- (12) J. F. Lamb, R. W. James, and H. S. Winchell, *Int. J. Appl. Radiat. Isot.*, **22**, 475 (1971).
- (13) L. F. Audrieth and M. Sveda, "Organic Syntheses", Collect. Vol. III, Wiley, New York, N.Y., 1955, p 536.
- (14) E. H. White and H. Scherrer, *Tetrahedron Lett.*, **21**, 758 (1961).
- (15) P. G. Stecher, Ed., "The Merck Index", 8th ed, Merck and Co., Rahway, N.J., 1968, p 641.
- (16) D. Papa, E. Schwenk, and H. F. Ginsberg, *J. Org. Chem.*, **14**, 723 (1949).
- (17) F. Richter, Ed., "Beilsteins Organische Chemie", Vol. 10, 2nd Suppl., Springer-Verlag, Berlin, 1949, p 124.
- (18) C. H. Fisher, *J. Am. Chem. Soc.*, **55**, 5003 (1933).
- (19) A. N. Ansari in "Cardiovascular Nuclear Medicine", H. W. Strauss, B. Pitt, and A. E. James, Ed., C. V. Mosby Co., St. Louis, Mo., 1974, Chapter 14, pp 234–240.
- (20) W. G. Myers, J. F. Lamb, R. W. James, and H. S. Winchell, *Nucl. Med.*, **12**, 154 (1973).
- (21) B. Tschiersch, *Phytochemistry*, **3**, 365 (1964).
- (22) C. Ressler, P. A. Redstone, and R. H. Erenberg, *Science*, **134**, 188 (1961).
- (23) W. J. P. Neish, *Arch. Int. Pharmacodyn.*, **107**, 315 (1956).