This article was downloaded by: [USC University of Southern California] On: 23 June 2013, At: 22:16 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Synthetic Communications: An International Journal for Rapid Communication of Synthetic Organic Chemistry

Publication details, including instructions for authors and subscription information: http://www.tandfonline.com/loi/lsyc20

# New Nitrilotriacetic Acid and Ethylenediaminetetraacetic Acid Nitro Derivatives for the Synthesis of Bifunctional and Trifunctional Chelating Agents

Stéphane Meunier <sup>a</sup> , Pierre Cristau <sup>a</sup> & Frédéric Taran <sup>a</sup>

<sup>a</sup> Service de Marquage Moléculaire et de Chimie Bio-Organique, DBJC/DSV CEA Saclay, Gif sur Yvette Cedex, France Published online: 18 Aug 2006.

To cite this article: Stéphane Meunier, Pierre Cristau & Frédéric Taran (2005): New Nitrilotriacetic Acid and Ethylenediaminetetraacetic Acid Nitro Derivatives for the Synthesis of Bifunctional and Trifunctional Chelating Agents, Synthetic Communications: An International Journal for Rapid Communication of Synthetic Organic Chemistry, 35:18, 2415-2425

To link to this article: <u>http://dx.doi.org/10.1080/00397910500189718</u>

# PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <u>http://www.tandfonline.com/page/terms-and-conditions</u>

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

*Synthetic Communications*<sup>®</sup>, 35: 2415–2425, 2005 Copyright © Taylor & Francis, Inc. ISSN 0039-7911 print/1532-2432 online DOI: 10.1080/00397910500189718



# New Nitrilotriacetic Acid and Ethylenediaminetetraacetic Acid Nitro Derivatives for the Synthesis of Bifunctional and Trifunctional Chelating Agents

Stéphane Meunier, Pierre Cristau, and Frédéric Taran

Service de Marquage Moléculaire et de Chimie Bio-Organique, DBJC/DSV CEA Saclay, Gif sur Yvette Cedex, France

**Abstract:** Dehydration of serine derivatives into didehydroamino acids and subsequent conjugate addition of nitromethane led to novel nitro derivatives of nitrilotriacetic acid and ethylenediaminetetraacetic acid. Straightforward transformations of these compounds allowed the synthesis of bifunctional and trifunctional metal-chelating agents via an Henry reaction.

Keywords: Chelates, Henry reaction, lanthanides, ligands, polyaminocarboxylates

#### INTRODUCTION

Polyaminocarboxylate chelates have the ability to form stable metal complexes with many metal ions such as nickel, copper, indium, technetium, and iron, in different oxidation states. Therefore, the development of bifunctional chelating agents that can be attached to macro(bio)molecules offers useful tools such as (1) probes of proteins structure,<sup>[11]</sup> (2) probes for molecular recognition,<sup>[2]</sup> (3) contrast agents for magnetic resonance,<sup>[3]</sup> (4) in vivo tumor imaging reagents,<sup>[4]</sup> and (5) radioimmunotherapy agents.<sup>[5]</sup> The purpose of designing trifunctional chelating agents is to combine these properties to achieve more complex and targeted applications.

Received in the U.K. January 11, 2005

Address correspondence to Frédéric Taran, Service de Marquage Moléculaire et de Chimie Bio-Organique, DBJC/DSV CEA Saclay, 91191 Gif sur Yvette Cedex, France. Fax: +33 (0)3 69 08 79 91; E-mail: frederic.taran@cea.fr

Biotinylated-EDTA-based trifunctional agents have been used for the preparation of radiolabeled antibodies that can be easily removed from blood with the aim to decrease side effects of radiotherapies.<sup>[6]</sup> As a second example, functionalization of phospholipids using a biotinylated-NTAbased trifunctional agent allows the organization of streptavidin and polyhistidine-tagged proteins around tubular structures offering valuable tools to structural biology.<sup>[7]</sup>

Although nitrilotriacetic acid (NTA)-based bifunctional chelating agents are usually synthesized from protected L-Lysine by alkylation with bromoacetic acid,<sup>[8]</sup> the most common method for attaching ethylenediaminetetraacetic acid (EDTA) to macromolecules involves the reaction of EDTA dianhydride with an amine residue on the target molecule. However, in this latter case, the stability constant of the resulting carboxamide-derivatized metal chelate can be lower than that of the parent EDTA. One solution to circumvent this difficulty is to design an EDTA derivative bearing an additional reactive function dedicated to the attachment to the target molecule. Several EDTA-based bifunctional chelating agents have been described, corresponding either to the modification of the ethylenediamino fragment or to the modification at the methylene carbon of one carboxymethyl arm; among them are EDTA derivatives bearing a carboxylic acid,<sup>[9]</sup> a bromoacetate,<sup>[10]</sup> an amine,<sup>[11]</sup> or an isothiocyanobenzyl reactive moiety.<sup>[12]</sup>

Herein is reported the synthesis, starting from serine, of two nitro derivatives of NTA and EDTA, compounds 1 and 2, which are modified on one carboxymethyl arm (Figure 1). These molecules are key intermediates for the straightforward synthesis of bifunctional and trifunctional chelating agents.

#### **RESULTS AND DISCUSSION**

As outlined in Scheme 1, the NTA derivative **3** was built by double *N*-alkylation of serine *tert*-butyl ester using *tert*-butyl bromoacetate at the reflux of acetonitrile. Under similar conditions, but in the presence of 1.2 equivalent of *tert*butyl bromoacetate, the product of mono *N*-alkylation **4** was obtained in a 67% yield. This compound was further *N*-alkylated using *N*, *N*-bis[(*tert*-butoxycalbonyl)methyl]-2-bromoethylamine<sup>[13]</sup> to afford the EDTA derivative **5**. It should be underlined that *tert*-butyl esters were chosen as protective groups



Figure 1. Structure of key compounds 1 and 2.



Scheme 1. (a)  $BrCH_2CO_2t$ -Bu (4 equiv), DIPEA (6 equiv),  $CH_3CN$ , reflux, 20 h (70%); (b)  $BrCH_2CO_2t$ -Bu (1.2 equiv), DIPEA (4 equiv),  $CH_3CN$ , reflux, 4 h (67%); (c)  $Br(CH_2)_2N(CH_2CO_2t$ -Bu)\_2 (3 equiv), DIPEA (5 equiv),  $CH_3CN$ , reflux, 20 h (87%).

for the carboxylate moieties because of the ease of deprotection in the presence of TFA, and to avoid the lactonization that was observed when serine was submitted to *N*-alkylation with methyl bromoacetate.

The nitro derivatives 1 and 2 were obtained by a two-step reaction from the primary alcohols 3 and 5, as shown in Scheme 2. The use of Mitsunobu reagents, tris(n-butyl)phosphine and diethyl azodicarboxylate, allowed the smooth conversion of the serine derivatives 3 and 5 into the didehydroamino acids 6 and 7, presumably through  $\alpha$ -deprotonation.<sup>[14]</sup> NMR analysis of the crude mixtures obtained after evaporation of the solvent under reduced pressure demonstrated that these reactions proceed very cleanly and in excellent yield; however, the didehydroaminoacids 6 and 7 could not be purified because of their instability on silica gel. Michael additions of nitromethane to 6 and 7 in the presence of fluoride ions afforded the nitro derivatives 1 and 2 in 50% and 64% yields. The addition of nitromethane on several dehvdroalanines (phthalimido, benzyloxycarbonylamino, protected acetamido) has been already extensively studied.<sup>[15]</sup> In the case of 6 and 7, despite a reduced reactivity resulting from the absence of electron-

 $3 \xrightarrow{a} \begin{bmatrix} t-BuO_2C \\ t-BuO_2C \end{bmatrix} \xrightarrow{N} \begin{bmatrix} CO_2t-Bu \\ G \end{bmatrix} \xrightarrow{b} 1$   $5 \xrightarrow{a} \begin{bmatrix} t-BuO_2C \\ t-BuO_2C \\ t-BuO_2C \\ CO_2t-Bu \end{bmatrix} \xrightarrow{b} 2$ 

Scheme 2. (a)  $(n-Bu)_3P$  (1.5 equiv), DEAD (1.5 equiv), Et<sub>2</sub>O, from  $-10^{\circ}$ C to rt, 3 h; (b)  $n-Bu_4NF \cdot 3H_2O$  (6 equiv), CH<sub>3</sub>NO<sub>2</sub>, 50°C, 20 h (1 50%, 2 64%).

withdrawing groups on the amino site of the didehydroaminoacid moiety, smooth additions occurred in the presence of an excess of nitromethane.

As shown in Scheme 3, the nitro derivatives 1 and 2 are readily reduced by ammonium formate in the presence of palladium on activated carbon to afford the bifunctional chelating agents 8 and 9 in good yields. At this stage the carboxylic acids are quantitatively deprotected by the treatment of 8 or 9 in pure trifluoroacetic acid at room temperature for 6 h. Although the protocols are not emphasized in this report, these bifunctional polyaminocarboxylate chelates can be conjugated to proteins using, for instance, a commercially available bis-N-hydroxysuccinimide linker.

To illustrate the preparation of trifunctional chelating agents starting from the nitro derivatives 1 or 2, the synthesis of such a molecule was investigated (Scheme 4). In the course of our research project a trifunctional molecule was needed that would contain (1) an EDTA moiety to chelate iron, (2) a histamine-like fragment to be recognized by an antihistamine monoclonal antibody, and (3) a Bolton-Hunter fragment allowing the introduction of radioisotopes (compound 13, Scheme 4). In a first step the EDTA derivative 2 was condensed on benzyl 6-oxohexanoate via an Henry reaction. Among the different sets of conditions or catalysts reported to promote the nitroaldol reaction,<sup>[16]</sup> the use of neutral alumina,<sup>[17]</sup> and the *n*-Bu<sub>4</sub>NF/triethylamine/ tert-butyldimethylchlorosilane reagent system were found unacceptably slow or inefficient in that case.<sup>[18]</sup> However, the adduct 10 was obtained with an excellent yield using one equivalent of *n*-Bu<sub>4</sub>NF in THF after 6 h at room temperature. The use of a catalytic amount of n-Bu<sub>4</sub>NF led to a slow reaction and a moderate yield (for 0.2 equiv, yield was 63% after 2 days at room temperature). The  $\beta$ -nitroalcohol 10 was then converted to the nitro compound 11 by a two-step, one-pot reaction, corresponding to the dehydration of 10 by the use of tris(n-butyl)phosphine and diethyl azodicarboxylate. This led to the intermediate nitroalkene and its subsequent reduction in the presence of sodium borohydride.

Hydrogenolysis of the benzyl ester on compound **11** in the presence of palladium on activated carbon under an atmospheric pressure of hydrogen



*Scheme 3.* (a) Pd/C (50 wt.%), HCO<sub>2</sub>NH<sub>4</sub> (40 equiv), MeOH, rt, 1 h (175%, 280%).



Scheme 4. (a)  $HCO(CH_2)_4CO_2Bn$  (1.5 equiv), n-Bu<sub>4</sub>NF · 3H<sub>2</sub>O (1 equiv), THF, rt, 6 h (95%); (b) (n-Bu<sub>3</sub>P (2 equiv), DEAD (2 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 0°C, 1 h, then dilution with EtOH, NaBH<sub>4</sub> (6 equiv), 0°C, 1 h (49%); (c) Pd/C (10 wt.%), H<sub>2</sub> (1 atm.), MeOH, rt, 1 h (96%); (d) NHS (1.1 equiv), DCC (1.1 equiv), CH<sub>2</sub>Cl<sub>2</sub>, rt, 4 h, then histamine (1.3 equiv), DMF, rt, 1.5 h (91%); (e) Pd/C (10 wt.%), HCO<sub>2</sub>NH<sub>4</sub> (40 equiv), MeOH, rt, 6 h (77%); (f) Bolton–Hunter reagent (1.2 equiv), TEA (3 equiv), CH<sub>2</sub>Cl<sub>2</sub>, rt, 20 h (57%); (g) TFA, rt, 6 h (95%).

was carried out quantitatively without reducing the nitro group. Activation of the obtained carboxylic acid as an *N*-hydroxysuccinimidyl ester and subsequent coupling with histamine led to **12** in high yield (Scheme 4). Then, the nitro group was reduced using ammonium formate and palladium on activated carbon, and the corresponding amine was coupled with the Bolton–Hunter reagent. Finally, quantitative removal of the *tert*-butyl ester protective groups by treatment with trifluoroacetic acid at room temperature led to the trifunctional chelating agent **13**.

In conclusion, the synthesis of new nitro derivatives of NTA and EDTA have been reported. These compounds are versatile building blocks for the preparation of bi- or trifunctional chelating agents, taking advantage of the nitro moiety reactivity. This was illustrated by the preparation of a bifunctional metal-chelating agent via the reduction of the nitro group and the design of a trifunctional metal-chelating agents via the coupling with an aldehyde using a Henry reaction.

#### **EXPERIMENTAL**

Analytical thin-layer chromatography (TLC) was performed using 0.25-mm silica gel-coated MERCK 60  $F_{254}$  plates. Visualization of the chromatogram was by UV absorbance and ethanolic phosphomolybdic acid. Flash chromatography was performed using compressed air with the indicated solvent system and silica gel 60 (MERCK, 230–400 mesh). <sup>1</sup>H NMR (300 MHz) and <sup>13</sup>C NMR (75 MHz) spectra were recorded on a BRUKER AC 300 spectrometer. Chemical shifts are reported in parts per million (ppm) on the  $\delta$  scale from an internal standard. Mass spectra (MS) by chemical ionisation in ammonia

 $(CI/NH_3)$  were recorded on a quadripolar FINNIGAN-MAT 4600 spectrometer and by electrospray on an ESI/TOF MARINER spectrometer.

### *tert*-Butyl-2-di(*tert*-butyloxycarbonylmethyl)amino-3hydroxypropanoate (3)

To a suspension of serine *tert*-butyl ester hydrochloride (600 mg, 3.04 mmol) in acetonitrile (25 mL), di-*iso*-propylethylamine (3.17 mL, 18.23 mmol) and *tert*-butyl bromoacetate (1.79 mL, 12.15 mmol) were added. The mixture was heated under reflux for 20 h. After cooling at room temperature and evaporation of the solvent under reduced pressure, the residue was purified by column chromatography (silica gel, hexane–EtOAc 8:2) to afford **3** (826 mg, 70%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.42 (s, 27H), 3.38–3.55 (m, 6H), 3.67–3.78 (m, 1H), 4.28 (d, J = 10.3 Hz, 1H). MS (CI, NH<sub>3</sub>): m/z 390 [M + 1].

# *tert*-Butyl-2-(*tert*-butyloxycarbonylmethylamino)-3hydroxypropanoate (4)

To a suspension of serine *tert*-butyl ester hydrochloride (3.58 g, 18.14 mmol) in acetonitrile (150 mL), di-*iso*-propylethylamine (11.42 mL, 72.55 mmol) and *tert*-butyl bromoacetate (3.21 mL, 21.76 mmol) were added. The mixture was heated under reflux for 4 h. After cooling at room temperature and evaporation of the solvent under reduced pressure, the residue was purified by column chromatography (silica gel, hexane–EtOAc 5:5) to afford **4** (3.33 g, 67%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.46 (s, 9H), 1.50 (s, 9H), 3.25 (dd ABX-system, J = 4.3 and 6.1 Hz, 1H), 3.27 and 3.39 (2d AB-system, J = 17.1 Hz, 2H), 3.60 (dd ABX-system, J = 6.1 and 11.0 Hz, 1H), 3.72 (dd ABX-system, J = 4.3 and 11.0 Hz, 1H). <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 27.8, 49.5, 62.3, 62.6, 81.3, 81.6, 171.2, 171.3. MS (CI, NH<sub>3</sub>): m/z 276 [M + 1].

# *tert*-Butyl-2-*tert*-butyloxycarbonylmethyl[2-di(*tert*butyloxycarbonylmethyl)aminoethyl]amino-3-hydroxypropanoate (5)

To a solution of **4** (300 mg, 1.09 mmol) in acetonitrile (10 mL) di-*iso*-propylethylamine (0.87 mL, 5.45 mmol) and *N*,*N*-bis[(*tert*-butoxycalbonyl)methyl]-2-bromoethylamine (1.15 g, 3.27 mmol)<sup>[13]</sup> were added. The mixture was heated under reflux for 20 h. After cooling at room temperature and evaporation of the solvent under reduced pressure, the residue was purified by column chromatography (silica gel, hexane–EtOAc 7:3) to afford **5** (520 g, 87%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.43 (s, 36H), 2.74–2.98 (m, 4H),

#### Synthesis of Bifunctional and Trifunctional Chelating Agents

3.30–3.55 (m, 8H), 3.65–3.80 (m, 1H). <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 27.7, 27.8, 27.9, 52.0, 53.2, 53.7, 55.9, 59.7, 67.2, 80.7, 81.1, 81.2, 170.2, 170.4, 172.1. MS (CI, NH<sub>3</sub>): m/z 547 [M + 1].

# Typical Procedure for the Synthesis of *tert*-Butyl-2-di(*tert*butyloxycarbonylmethyl)amino-4-nitrobutanoate (1) and *tert*-Butyl-2-*tert*-butyloxycarbonylmethyl[2-di(*tert*butyloxycarbonylmethyl)aminoethyl]amino-4-nitrobutanoate (2)

To a solution of 5 (150 mg, 0.27 mmol) in diethyl ether (6 mL) under argon and at  $-10^{\circ}$ C, tris(*n*butyl)phosphine (100  $\mu$ L, 0.41 mmol) and diethyl azodicarboxylate (65 µL, 0.41 mmol) were added. The mixture was stirred for 3 h, while the temperature was allowed to reach 20°C. After evaporation of the solvent under reduced pressure, the crude material was diluted with nitromethane (8 mL), and tributylammonium fluoride trihydrate (520 mg, 1.65 mmol) was added. The mixture was stirred at room temperature for 1 h, and at 50°C overnight. After cooling at room temperature and evaporation of the solvent under reduced pressure, the residue was purified by column chromatography (silica gel, hexane-EtOAc 8:2) to afford 2 (103 mg, 64%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.41 (s, 27H), 1.43 (s, 9H), 2.08–2.40 (m, 2H), 2.67–2.93 (m, 4H), 3.21 (d AB-system, J = 17.1 Hz, 1H), 3.33 (d AB-system, J = 17.1 Hz, 1H), 3.37 (d AB-system, J = 17.7 Hz, 2H), 3.45 (d AB-system, J = 17.7 Hz, 2H), 3.62 (dd ABX-system, J = 4.3 and 11.6 Hz, 1H), 4.60–4.88 (m, 2H). <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) δ: 27.7, 27.8, 28.0, 50.6, 51.2, 53.8, 55.6, 60.5, 72.1, 80.7, 80.8, 81.5, 170.3, 170.6. HRMS: m/z calcd.: 590.3652; found: 590.3657 [M + 1]. Analytical data for compound 1: <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.42 (s, 18H), 1.45 (s, 9H), 2.10-2.45 (m, 2H), 3.34 (d AB-system, J = 17.1 Hz, 2H), 3.37 (d ABsystem, J = 17.1 Hz, 2H), 3.47 (dd ABX-system, J = 4.3 and 11.6 Hz, 1H). 4.68–4.90 (m, 2H). <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) δ: 27.3, 27.8, 27.9, 54.0, 61.9, 71.9, 80.9, 81.8, 169.9, 170.4. HRMS: m/z calcd.: 433.2550; found: 433.2547 [M+1].

# Typical Procedure for the Synthesis of *tert*-Butyl-4-amino-2-di(*tert*butyloxycarbonylmethyl)aminobutanoate (8) and *tert*-Butyl-4-amino-2-*tert*-butyloxycarbonylmethyl[2-di(*tert*butyloxycarbonylmethyl)aminoethyl]aminobutanoate (9)

To a solution of 2 (100 mg, 0.17 mmol) in methanol (3 mL), palladium on activated carbon (20 mg) and ammonium formate (428 mg, 40 eq) were added in four portions every hour. The catalyst was filtered and washed with methanol. The filtrate was evaporated under reduced pressure. The residue was diluted with methylene chloride and washed with a saturated

solution of sodium hydrogenocarbonate. The organic phase was dried over magnesium sulfate and evaporated under reduced pressure to afford **9** (76 mg, 80%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.30–1.50 (m, 36H), 1.80–2.10 (m, 2H), 2.58–2.90 (m, 4H), 3.18–3.50 (m, 9H), 3.67–3.74 (m, 1H). <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 25.6, 27.7, 27.8, 27.9, 49.7, 50.1, 55.0, 60.8, 81.4, 81.7, 82.0, 170.4, 172.0. MS (CI, NH<sub>3</sub>): *m/z* 560 [M + 1]. Analytical data for compound 8: <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.41 (bs, 27H), 1.90–2.30 (m, 2H), 3.05–3.60 (m, 7H). <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 25.0, 27.8, 27.9, 38.6, 53.7, 64.1, 81.9, 82.0, 169.6, 170.6. MS (CI, NH<sub>3</sub>): *m/z* 403 [M + 1].

#### Procedure for the Synthesis of 10

To a solution of **2** (800 mg, 1.36 mmol) in THF (20 mL), benzyl 6-oxohexanoate (400 mg, 2.04 mmol) and tributylammonium fluoride trihydrate (428 mg, 1.36 mmol) were added. The mixture was stirred for 6 h at room temperature. After evaporation of the solvent under reduced pressure, the crude material was diluted with methylenechloride and washed twice with water and once with brine. The organic phase was dried over magnesium sulfate and evaporated under reduced pressure. The residue was purified by column chromatography (silica gel, hexane–EtOAc 8:2) to afford **10** (1.5 g, 95%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 0.90–1.80 (m, 40H), 1.92–2.50 (m, 6H), 2.55–2.95 (m, 4H), 3.00–3.78 (m, 7H), 3.80–4.25 (m, 1H), 4.85–5.35 (m, 3H), 7.20–7.45 (m, 5H). <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 24.4, 25.0, 27.8, 27.9, 28.0, 29.3, 29.9, 31.4, 32.5, 33.9, 51.3, 52.0, 54.0, 55.0, 55.7, 60.8, 61.2, 65.9, 70.5, 71.4, 72.2, 80.9, 81.0, 81.6, 87.2, 88.2, 127.9, 128.3, 135.8, 170.5, 170.7, 173.1. MS (CI, NH<sub>3</sub>): *m/z* 810 [M + 1]. MS (TOF): *m/z* 832 [M + 23].

#### Procedure for the Synthesis of 11

To a solution of **10** (270 mg, 0.33 mmol) in methylene chloride (2 mL) were added, under argon and at  $-15^{\circ}$ C, tris(*n*butyl)phosphine (165 µL, 0.66 mmol) and diethyl azodicarboxylate (105 µL, 0.66 mmol). The mixture was stirred for 1 h at 0°C, then cooled at  $-5^{\circ}$ C, diluted with ethanol (7 mL), and sobium borohydride (75 mg, 1.99 mmol) was added. The mixture was stirred for 1 h at 0°C, and quenched by addition of water. The solution was extracted twice with ethyl acetate, the organic phases were dried over magnesium sulfate and the solvent was evaporated under reduced pressure. The residue was purified by column chromatography (silica gel, hexane–EtOAc 8:2) to afford **11** (126 mg, 49%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.23–2.25 (m, 44H), 2.22–2.43 (m, 4H), 2.67–2.98 (m, 4H), 3.12–3.60 (m, 7H), 4.82 (quint., J = 7.2 Hz, 0.5H), 5.00–5.34 (m, 2.5H), 7.28–7.43 (sl, 5H). <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 24.3, 25.0, 27.8, 27.9,

28.3, 33.1, 33.3, 33.4, 33.8, 34.1, 50.1, 52.0, 52.2, 52.5, 53.5, 53.9, 55.7, 55.8, 60.6, 61.7, 65.8, 80.6, 81.5, 84.4, 85.3, 127.9, 128.3, 135.8, 170.3, 170.7, 170.8, 170.9, 173.0. MS (TOF): *m*/*z* 794 [M + 1].

#### Procedure for the Synthesis of 12

To a solution of 11 (112 mg, 0.14 mmol) in methanol (3 mL) was added palladium on activated carbon (10 mg). The mixture was stirred for 1 h under hydrogen (1 atm). The catalyst was filtered and washed with methanol. The filtrate was evaporated under reduced pressure to afford the corresponding benzyl-deprotected carboxylic acid (96 mg, 96%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) & 1.20-2.15 (m, 44H), 2.22-2.42 (m, 4H), 2.62-3.05 (m, 4H), 3.18-3.80 (m, 7H), 4.80-4.92 (m, 0.5H), 4.98-5.12 (m, 0.5H). MS (TOF): m/z 704 [M+1]. To the solution of this compound in methylene chloride (2 mL), N-hydroxysuccinimide (18 mg, 0.15 mmol) and dicyclocarbodiimide (31 mg, 0.15 mmol) were added. The mixture was stirred for 4h and filtered. The filtrate was evaporated under reduced pressure. The residue obtained was diluted with DMF (2 mL), and histamine (20 mg, 0.17 mmol) was added. The mixture was stirred for 1.5 h, and then the solvent was evaporated under reduced pressure. The residue was purified by column chromatography (silica gel, methylene chloridemethanol 9:1) to afford 12 (99 mg, 91%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.12-1.30 (m, 4H), 1.40 (sl, 36H), 1.45-2.02 (m, 5H), 2.10 (t, J = 7.3 Hz, 2H), 2.20–2.35 (m, 1H), 2.62–2.92 (m, 6H), 3.10–3.50 (m, 9H), 4.77 (quint., J = 6.7 Hz, 0.5H), 4.93–5.08 (m, 0.5H), 6.68 (t, J = 6.5 Hz, 1H), 6.76 (s, 1H), 7.54 (s, 1H). MS (TOF): m/z 797 [M + 1].

#### Procedure for the Synthesis of 13

To a solution of **12** (83 mg, 0.10 mmol) in methanol (3 mL) were added palladium on activated carbon (20 mg) and ammonium formate (264 mg, 40 eq) in four portions every 90 min. The catalyst was filtered and washed with methanol. The filtrate was evaporated under reduced pressure. The residue was diluted with methylene chloride and washed with a saturated solution of sodium hydrogenocarbonate. The organic phase was dried over magnesium sulfate and evaporated under reduced pressure to afford the expected amine (62 mg, 77%) as the product of the nitro function reduction. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.00–1.93 (m, 46H), 2.11 (t, *J* = 7.3 Hz), 2.58–2.90 (m, 6H), 3.00–3.80 (m, 10H), 6.74 (s, 1H), 7.45 (s, 1H). MS (TOF): *m/z* 767 [M + 1]. To the solution of the previous compound in methylene chloride (2 mL), triethylamine (35 µL, 0.24 mmol) and the Bolton–Hunter reagent (26 mg, 0.10 mmol) were added. The mixture was stirred for 20 h, and then diluted with methylene chloride and washed with a saturated solution of sodium hydrogenocarbonate. The organic phase was dried over magnesium sulfate and evaporated under reduced pressure. The residue was purified by column chromatography (silica gel, methylene chloride–methanol 9:1) to afford the expected coupling adduct (42 mg, 57%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.25–1.90 (m, 46H), 2.03 (t, J = 7.3 Hz, 2H), 2.42–2.60 (m, 2H), 2.62–3.00 (m, 8H), 3.12–3.92 (m, 10H), 6.68–6.82 (m, 3H), 7.02 (d, J = 6.7, 1H), 7.04 (d, J = 8.5, 1H), 7.53 (s, 1H). MS (TOF): m/z 937 [M + 23]. The solution of the previous compound in trifluoroacetic acid (3 mL) was stirred for 6 h. Diethyl ether was added, and the precipitate was filtered, washed with diethyl ether, and dried under vacuum to afford 13-(tris-trifluoroacetate salt) (40 mg, 95%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.20–2.00 (m, 10H), 2.02–2.15 (m, 2H), 2.38–2.52 (m, 2H), 2.70–3.20 (m, 8H), 2.35–2.70 (m, 4H), 3.80–4.48 (m, 6H), 6.62 (d, J = 7.9 Hz, 2H), 6.97 (d, J = 7.9 Hz, 1H), 6.99 (d, J = 7.9 Hz, 1H), 7.28 (s, 1H). MS (TOF): m/z 691 [M + 1].

#### REFERENCES

- (a) Rana, T. M.; Meares, C. F. Specific cleavage of a protein by an attached iron chelate. J. Am. Chem. Soc. 1990, 112 (6), 2457–2458; (b) Meares, C. F.; Wensel, T. G. Metal chelates as probes of biological systems. Acc. Chem. Res. 1984, 17 (6), 202–209.
- (a) Dorn, I. T.; Neumaier, K. R.; Tampé, R. Molecular recognition of histidinetagged molecules by metal-chelating lipids monitored by fluorescence energy transfer and correlation spectroscopy. J. Am. Chem. Soc. 1998, 120 (12), 2753–2763; (b) Min, C.; Verdine, G. L. Immobilized metal affinity chromatography of DNA. Nucleic Acids Res. 1996, 24 (19), 3806–3810.
- Storrs, R. W.; Tropper, F. D.; Li, H. Y.; Song, C. K.; Kuniyoshi, J. K.; Sipkins, D. A.; Li, K. C. P.; Bednarski, M. D. Paramagnetic polymerized liposomes: Synthesis, characterization, and applications for magnetic resonance imaging. J. Am. Chem. Soc. 1995, 117 (28), 7301–7306.
- (a) Scheinberg, D. A.; Strand, M.; Gansow, O. A. Tumor imaging with radioactive metal chelates conjugated to monoclonal antibodies. *Science* 1982, 215 (4539), 1511–1513;
  (b) De Riemer, L. H.; Meares, G. F.; Goodwin, D. A.; Diamanti, C. I. BLEDTA: Tumor localization by a bleomycin analog containing a metal-chelating group. *J. Med. Chem.* 1979, 22 (9), 1019–1023.
- (a) Hnatowich, D. J.; Layne, W. W.; Childs, T. W.; Doherty, P. W. Radioactive labeling of antibody: A simple and efficient method. *Science* 1983, 220 (4597), 613–615; (b) Fritzberg, A. R.; Abrams, P. G.; Beaumier, P. L.; Kasina, S.; Morgan, A. C.; Rao, T. N.; Reno, J. M.; Sanderson, J. A.; Srinivasan, A.; Wilbur, D. S.; Vanderheyden, J. L. Specific and stable labeling of antibodies with technetium-99 m with a diamide dithiolate chelating agent. *Proc. Natl. Acad. Sci. USA* 1988, 85 (11), 4025–4029; (c) Westerberg, D. A.; Carney, P. L.; Rogers, P. E.; Kline, S. J.; Johnson, D. K. J. Synthesis of novel bifunctional chelators and their use in preparing monoclonal antibody conjugates for tumor targeting. *J. Med. Chem.* 1989, *32* (1), 236–243.

#### Synthesis of Bifunctional and Trifunctional Chelating Agents

- Wilbur, D. S.; Chyan, M. K.; Hamlin, D. K.; Kegley, B. B.; Nilsson, R.; Sandberg, B. E. B.; Brechbiel, M. Trifunctional conjugation reagents. Reagents that contain a biotin and a radiometal chelation moiety for application to extracorporeal affinity adsorption of radiolabeled antibodies. *Bioconjugate Chem.* 2002, *13* (5), 1079–1092.
- Drakopoulou, E.; Tsivgoulis, G. M.; Mukhopadhyay, A.; Brisson, A. Design and synthesis of multifunctional phospholipids. *Tetrahedron Lett.* 2000, *41* (21), 4131–4134.
- Roy, B. C.; Mallik, S. Synthesis of new polymerizable metal-chelating lipids. J. Org. Chem. 1999, 64 (8), 2969–2974.
- (a) Rana, M. T.; Ban, M.; Hearst, J. E. Synthesis of a metal-ligating amino acid suitable for solid phase assembly of peptides. *Tetrahedron Lett.* **1992**, *33* (32), 4521–4524; (b) Kahana, N.; Arad-Yellin, R.; Warshawsky, A. A conceptual approach to the synthesis of bifunctional EDTA analogs: EDTA-extended polyamides. *J. Org. Chem.* **1994**, *59* (17), 4832–4837.
- Hayward, M. M.; Adrian, J. C.; Schepartz, A. Convenient syntheses of bifunctional metal chelates. J. Org. Chem. 1995, 60 (12), 3924–3927.
- (a) Studer, M.; Meares, C. F. A convenient and flexible approach for introducing linkers on bifunctional chelating agents. *Bioconjugate Chem.* 1992, 3 (5), 420–423; (b) Warshawsky, A.; Altman, J.; Kahana, N.; Arad-Yellin, R.; Deshe, A.; Hasson, H.; Shoef, N.; Gottlieb, H. Ring cleavage of *N*-acyl- and *N*-(arylsulfonyl)histamines with di-*tert*-butyl dicarbonate. A one-pot synthesis of 4-acylamino- and 4-arylsulfonylamino-1,2-diaminobutanes. *Synthesis* 1989 (11), 825–829.
- (a) Keana, J. F. W.; Mann, J. S. Chelating ligands functionalized for facile attachment to biomolecules. A convenient route to 4-isothiocyanatobenzyl derivatives of diethylenetriaminepentaacetic acid and ethylenediaminetetraacetic acid. J. Org. Chem. 1990, 55 (9), 2868–2871; (b) Kline, S. J.; Betebenner, D. A.; Johnson, D. K. Carboxymethyl-substituted bifunctional chelators: Preparation of arylisothiocyanate derivatives of 3-(carboxymethyl)-3-azapentanedioic acid, 3,12-bis(carboxymethyl)-6,9-dioxa-3,12-diazatetradecanedioic acid, and 1,4,7,10-tetraazacyclododecane-N,N',N"'.tetraacetic acid for use as protein labels. Bioconjugate Chem. 1991, 2 (1), 26–31.
- Amedio, J. C., Jr.; Van Wagenen, G., Jr.; Zavlin, G.; Gyorkos, A.; Peterson, S. A. Preparation of N,N-bis[2-[N',N'-bis[(tert-butoxycarbonyl)methyl]-amino]ethyl-Laspartic acid: An intermediate in the synthesis of MRI contrast agents. *Synth. Commun.* 2000, 30 (20), 3755–3763.
- Schimdt, U.; Lieberknecht, A.; Wild, J. Didehydroamino acids (DDAA) and didehydropeptides (DDP). *Synthesis* 1988 (3), 159–172.
- Crossley, M. J.; Fung, Y. M.; Potter, J. J.; Stamford, A. W. Convenient route to γ-Nitro-α-amino acids: Conjugate addition of nitroalkanes to dehydroalanine derivatives. J. Chem. Soc., Perkin Trans. 1 1998 (6), 1113–1122.
- Luzzio, F. A. The Henry reaction: Recent examples. *Tetrahedron* 2001, 57 (6), 915–945.
- Rosini, G.; Ballini, R.; Sorrenti, P. Synthesis of 2-nitroalkanols on alumina surfaces without solvent: A simple, mild and convenient method. *Synthesis* 1983 (11), 1014–1016.
- Fernández, R.; Gasch, C.; Gómez-Sánchez, A.; Vilchez, J. E. Highly efficient nitroaldol reaction promoted by trialkylsilyl chlorides. *Tetrahedron Lett.* 1991, 32 (27), 3225–3228.