# On the formation of reductic acid from pentoses or hexuronic acids

Tania Ahmad<sup>a</sup>, Rolf Andersson<sup>a</sup>, Kjell Olsson<sup>a,\*</sup> and Eric Westerlund<sup>b</sup>

Departments of <sup>a</sup> Chemistry and <sup>b</sup> Food Science, Swedish University of Agricultural Sciences, S-750 07 Uppsala (Sweden)

(Received October 17th, 1992; accepted March 30th, 1993)

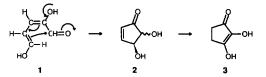
#### ABSTRACT

Careful hydrolysis of  $(\pm)$ -*cis*- or  $(\pm)$ -*trans*-tetrahydro-2,5-dimethoxy-2-furaldehyde dimethyl acetal proceeded via 5,5-dimethoxy-4-oxopentanal to give  $(\pm)$ -*trans*-4-hydroxy-5-methoxy-2-cyclopentenone and  $(\pm)$ -*trans*-4,5-dihydroxy-2-cyclopentenone. The latter product did not isomerize to 2,3-dihydroxy-2-cyclopentenone (reductic acid) on prolonged reaction.

## INTRODUCTION

Although 2,5-dihydroxy-2,4-pentadienal has never been isolated, some of its geometrical isomers have been proposed as key intermediates in the degradation of pentoses or hexuronic acids. Thus, one isomer (1) is believed<sup>1</sup> to cyclize by aldol condensation to the cyclopentenone  $2^{\dagger}$ , which then tautomerizes to the so-called reductic acid (3), see Scheme 1. The stereochemistry of 2 is not known.

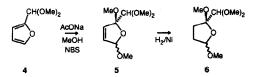
Isomer 1 has also been proposed as a precursor of certain phenolic degradation products<sup>2-4</sup>. In order to confirm this, we needed a compound related to 1 but stable enough to permit isolation. For this purpose, tetrahydro-2,5-dimethoxy-2-furaldehyde dimethyl acetal (6 in Scheme 2) was chosen, because its ethyl analogue had been converted previously into a phenolic compound (3,8-dihydroxy-2-methyl-chromone)<sup>3</sup>. The present paper deals with the preparation and hydrolysis of 6. The hydrolysis invariably produced the *trans* isomer (2t) of 2, but no reductic acid (3).



Scheme 1. Proposed formation mechanism of reductic acid.

<sup>\*</sup> Corresponding author.

<sup>&</sup>lt;sup>†</sup> All chiral compounds were obtained as racemic mixtures, but only one enantiomer is shown.

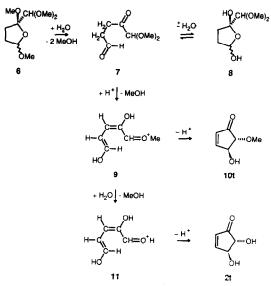


Scheme 2. Preparation of compound 6 (diastereomeric mixture).

## **RESULTS AND DISCUSSION**

Compound 6 has been prepared from 2-furaldehyde dimethyl acetal (4) via the 2,5-dihydro derivative<sup>5</sup> (5), see Scheme 2. After some improvements, we preferred a procedure described for the ethyl analogue<sup>2</sup>. Compound 5 was fractionated by chromatography into the known<sup>6</sup> *cis* and *trans* isomers (5h6c and 5t), which were hydrogenated to the corresponding isomers (6c and 6t) of 6. Since the isomers 6c and 6t behaved in the same way as their mixture (6) on hydrolysis, only the hydrolysis of 6 will be described.

Careful hydrolysis converted 6 into the new 5,5-dimethoxy-4-oxopentanal (7) and its diastereomeric hydrates (8), see Scheme 3. The <sup>1</sup>H and <sup>13</sup>C NMR spectra showed 7 in chloroform solution but 8 in aqueous solution, as evidenced by the presence or absence of the aldehyde proton (H-1) and the two carbonyl carbons. The spin-spin coupling between H-1 and the adjacent methylene protons (H-2) was too weak to cause splitting of the H-1 signal but was revealed by sharpening of the H-2 signal on irradiation at the frequency of H-1.



Scheme 3. Hydrolysis of compound 6.

On prolonged reaction, 7 gradually cyclized to 2t and its monomethyl ether 10t, but no reductic acid (3) was detected. The cyclizations of 7 are apparently aldol condensations via cationic analogues (9 and 11) of 1. Owing to rapid tautomerization, the <sup>1</sup>H NMR spectrum of 3 in D<sub>2</sub>O consists of a single sharp line<sup>7</sup>. The <sup>1</sup>H NMR spectrum of 2t was much more informative and strongly resembled that of its *cis* isomer<sup>8</sup>. However, the two spectra showed different coupling constants (*J*), especially for coupling between the saturated methine protons (H-4 and H-5). Thus,  $J_{4,5}$  2.7 Hz was observed for 2t and  $J_{4,5}$  5.3 Hz for the *cis* isomer. The *cis* configuration of the latter isomer followed from its synthesis via the 4,5-di-O-isopropylidene derivative<sup>8</sup>. Hence, 2t is the *trans* isomer. Since the four coupling constants observed for 10t agreed, within ±0.1 Hz, with those observed for 2t, 10t is also the *trans* isomer. The <sup>13</sup>C NMR spectra of 2t and 10t, together with the <sup>1</sup>H NMR spectra of their acetates, further supported their formulas.

#### **EXPERIMENTAL**

General methods.—Solvent mixtures are defined by volume ratios (y/y). The light petroleum used boiled at 60-70°C. Concentration was conducted under diminished pressure below 40°C. The progress of all reactions was monitored by TLC using Silica Gel HF<sub>254</sub> (Merck) microplates and the following standard spray reagents<sup>9</sup>: A, p-anisidine · HCl; B, resorcinol-aq HCl; and C, aq FeCl<sub>3</sub>. Column chromatography was performed on silica gel (230-400 mesh, Merck). GLC was carried out at 100°C on a Packard 427 instrument fitted with a flame-ionization detector and a DB1 column (30 m  $\times$  0.22 mm; linear flow rate, 0.3 m/s). GLC-EIMS was performed at 70 eV. The low-resolution spectra were recorded with a Finnigan 4021 instrument. The high-resolution spectra were recorded with a magnetic sector instrument<sup>10</sup> at 100  $\mu$ A and 200°C with perfluorotributylamine as a reference. IR spectra were scanned with a Perkin-Elmer 1760X FTIR spectrometer; no solvent was used. NMR spectra were recorded with a Varian VXR-400 instrument at 25°C. The samples were dissolved in CDCl<sub>3</sub> or D<sub>2</sub>O. <sup>1</sup>H NMR spectra were recorded at 400 MHz and referenced to the solvent (CHCl<sub>3</sub>,  $\delta$  7.26) or (in D<sub>2</sub>O) to internal Me<sub>3</sub>SiCH<sub>2</sub>CH<sub>2</sub>CO<sub>2</sub>Na (Me,  $\delta$  0.00). <sup>13</sup>C NMR spectra were recorded at 101 MHz and referenced to the solvent (CDCl<sub>3</sub>,  $\delta$  77.1) or (in D<sub>2</sub>O) to internal 1,4-dioxane ( $\delta$  67.4). For complete assignment of all signals in the di- and tetra-hydrofuran derivatives 5, 6, and 8, H-H COSY, C-H COSY, and long-range C-H COSY experiments were performed, using the standard software from Varian.

*Reductic acid* (3).—This was prepared according to Theander<sup>11</sup>.

2-Furaldehyde dimethyl acetal (4).—This was prepared from the aldehyde according to a general procedure<sup>12</sup>; bp 55–58°C at 2.0 kPa; lit.<sup>13</sup> 57–58°C at 1.9 kPa.

2,5-Dihydro-2,5-dimethoxy-2-furaldehyde dimethyl acetal (5).—2-Furaldehyde dimethyl acetal (4; 7.1 g, 50 mmol) and NaOAc (10 g) were dissolved in MeOH (15

mL) and Et<sub>2</sub>O (15 mL). NBS (*N*-bromosuccinimide, 8.9 g, 50 mmol) was added in small portions to the stirred solution (which was cooled in an ice bath) kept below 5°C. After 30 min, the mixture was concentrated, aq KOH (50 mL, 7 g, 0.12 mol) added, and the mixture extracted with  $CH_2Cl_2$  (4 × 50 mL). The extract was dried (K<sub>2</sub>CO<sub>3</sub>) overnight and filtered, the solvent evaporated, and the residue distilled to give a colourless liquid (5.49 g, 58% yield); bp 100–105°C at 2.0 kPa. This liquid was fractionated on a silica gel column (80 × 5 cm) with 5:1 light petroleum –EtOAc. Two chromatographically pure compounds (5c and 5t) were obtained, giving a violet colour with spray *B*, and having the same mass spectrum, m/z (*rel* int.): 173 (M – MeO, 2) 129 (10), 101 (10), 75 (100). Their <sup>1</sup>H NMR spectra were in accordance with those reported<sup>6</sup>.

The cis isomer 5c (yield 14%) had  $R_f$  0.24; GLC retention time, 9.0 min.

The trans isomer 5t (yield 13%) had  $R_f$  0.16; GLC retention time, 9.3 min.

Tetrahydro-2,5-dimethoxy-2-furaldehyde dimethyl acetal (6).—A mixture (27.35 g, 0.14 mol) of 5c and 5t in dry MeOH (400 mL) was shaken with Raney nickel under  $H_2$  gas overnight under ambient conditions. The solvent was evaporated and a colourless liquid isolated by distillation (25 g, 91% yield); bp 100–110°C at 2.0 kPa. This liquid was fractionated on a silica gel column with 5:1 light petroleum-EtOAc. Two chromatographically pure compounds (6c and 6t) were obtained, giving a greyish-red colour with spray B and having the same mass spectrum, m/z (rel int.): 206 (M, 0.5), 175 (3), 144 (5), 131 (42), 115 (12), 99 (18), 75 (100), 71 (88). Their configurations were established by hydrogenation of 5c and 5t separately on a smaller scale.

The *cis* isomer **6c** had  $R_f$  0.35; GLC retention time, 9.3 min; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.88–2.18 (m, 4 H, H-3,4), 3.36 (s, 3 H, OMe-2), 3.43 (s, 3 H, OMe-5), 3.49 and 3.52 (2 s, each 3 H, OMe- $\alpha$ ), 4.22 (s, 1 H, H- $\alpha$ ), 5.05 (dd, 1 H, J 1.9 and 4.9 Hz, H-5); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  27.0 (C-3), 32.4 (C-4), 49.6 (OMe-2), 55.4 (OMe-5), 56.4 and 58.1 (OMe- $\alpha$ ), 106.3 (C- $\alpha$ ), 106.5 (C-5), 111.2 (C-2).

The *trans* isomer **6t** had  $R_f$  0.19; GLC retention time, 8.9 min; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.82–2.28 (m, 4 H, H-3,4), 3.30 (s, 3 H, OMe-2), 3.39 (s, 3 H, OMe-5), 3.52 and 3.53 (2 s, each 3 H, OMe- $\alpha$ ), 4.27 (s, 1 H, H- $\alpha$ ), 5.17 (dd, 1 H, J 1.0 and 5.3 Hz, H-5). <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  26.0 (C-3), 31.6 (C-4), 49.7 (OMe-2), 53.3 (OMe-5), 54.8 and 55.2 (OMe- $\alpha$ ), 106.25 (C- $\alpha$ ), 106.3 (C-5), 110.0 (C-2).

5,5-Dimethoxy-4-oxopentanal (7).—A mixture (5 g, 24.27 mmol) of **6c** and **6t** was heated with water (50 mL) in a water bath at 96°C for 3 h, cooled, and extracted with CHCl<sub>3</sub>. The extract was evaporated. The syrupy residue was fractionated on a column of silica gel with 10:1 CH<sub>2</sub>Cl<sub>2</sub>–Me<sub>2</sub>CO to give 7 in 38% yield;  $R_f$  0.16 (10:1 CH<sub>2</sub>Cl<sub>2</sub>–Me<sub>2</sub>CO). It gave a yellow colour with spray A and a red colour with spray B; GLC retention time, 5.3 min;  $\nu_{max}$  1730 (C=O); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  2.78 (t, 2 H, H-2), 2.88 (t, 2 H, H-3), 3.42 (s, 6 H, Me), 4.54 (s, 1 H, H-5), 9.8 (s, 1 H, H-1); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  29.9 (C-2), 36.9 (C-3), 54.8 (Me), 103.7 (C-5), 200.2 (C-4), 203.9 (C-1); MS, m/z (rel int.): 159 (M – H<sup>+</sup>, 0.1), 129 (M – MeO<sup>+</sup>, 2), 97 (2), 85 (3), 75 (100); Calcd for C<sub>2</sub>H<sub>11</sub>O<sub>4</sub> (M – H<sup>+</sup>): 159.066. Found: 159.061.

Tetrahydro-2,5-dihydroxy-2-furaldehyde dimethyl acetal (8).—Compound 7 was dissolved in water.

*Major isomer.* <sup>1</sup>H NMR (D<sub>2</sub>O):  $\delta$  1.79–2.30 (m, 4 H, H-3,4), 3.558 and 3.561 (2 s, each 3 H, Me), 4.34 (s, 1 H, H- $\alpha$ ), 5.60 (m, 1 H, H-5); <sup>13</sup>C NMR (D<sub>2</sub>O):  $\delta$  30.9 (C-3), 31.8 (C-4), 58.3 and 58.5 (Me), 99.6 (C-5), 106.9 (C-2), 107.9 (C- $\alpha$ ).

*Minor isomer.* <sup>1</sup>H NMR (D<sub>2</sub>O):  $\delta$  1.94–2.25 (m, 4 H, H-3,4), 3.54 and 3.55 (2 s, each 3 H, Me), 4.30 (s, 1 H, H- $\alpha$ ), 5.52 (m, 1 H, H-5); <sup>13</sup>C NMR (D<sub>2</sub>O):  $\delta$  31.8 (C-3), 32.8 (C-4), 58.3 and 58.5 (Me), 100.4 (C-5), 107.0 (C-2), 107.5 (C- $\alpha$ ).

Preparation of compounds 2t and 10t.—Compound 7 (450 mg, 2.8 mmol) was heated at 80°C in a water bath with Me<sub>2</sub>CO (25 mL) and 2% H<sub>2</sub>SO<sub>4</sub> (500  $\mu$ L) for 6 h. The Me<sub>2</sub>CO was evaporated. The residue was neutralized with aq Na<sub>2</sub>CO<sub>3</sub> and extracted with CHCl<sub>3</sub>. The extract was evaporated. The resulting syrup was fractionated on a column of silica gel (20 × 5 cm) and three main fractions were collected by elution with 3:2 CH<sub>2</sub>Cl<sub>2</sub>–Me<sub>2</sub>CO. The first fraction gave 2t in 12% yield;  $R_f$  0.46 (3:2 CH<sub>2</sub>Cl<sub>2</sub>–Me<sub>2</sub>CO), brown colour with spray A and red with B; GLC retention time, 3.6 min;  $\nu_{max}$  1730 (C=O), 1100 (C–OH), and 3400 cm<sup>-1</sup> (OH); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  3.2 and 3.5 (2 bs, each 1 H, OH), 4.21 (d, 1 H, J 2.7 Hz, H-5), 4.82 (m, 1 H, J 1.4, 1.9, and 2.7 Hz, H-4), 6.28 (dd, 1 H, J 1.4 and 6.1 Hz, H-2), 7.47 (dd, 1 H, J 1.9 and 6.1 H, H-3); <sup>13</sup>C NMR (D<sub>2</sub>O):  $\delta$  78.7 (C-4), 83.1 (C-5), 134.6 (C-3), 164.9 (C-2), 209.7 (C-1); MS, *m/z* (*rel* int.): 114 (2, M), 113 (3, M – H ), 96 (80), 68 (100); Calcd for C<sub>5</sub>H<sub>5</sub>O<sub>3</sub> (M – H ): 113.024. Found: 113.019.

Compound **2t** (10 mg) was acetylated with dry pyridine (1 mL) and Ac<sub>2</sub>O (1 mL) overnight at room temperature. The solution was poured into ice-water and the resulting mixture extracted with CHCl<sub>3</sub>. The extract was washed with 0.1 M HCl (2 × 25 mL), 0.1 M NaHCO<sub>3</sub> (2 × 5 mL), and water (2 × 2 mL), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and evaporated. The diacetate was obtained in 90% yield; GLC retention time, 7.8 min;  $R_f$  0.28 (3:1 light petroleum –EtOAc), red colour with spray B; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  2.10 and 2.17 (2 s, each 3 H, Ac), 5.18 (d, 1 H, J 3.0 Hz, H-5), 5.90 (m, 1 H, J 1.4, 2.2, and 3.0 Hz, H-4), 6.41 (dd, 1 H, J 1.4 and 6.1 Hz, H-2), 7.45 (dd, 1 H, J 2.2 and 6.1 Hz, H-3); MS, m/z (*rel* int.): 198 (M, 0.1), 156 (2), 138 (M – AcOH, 2), 96 (25), 68 (22), 43 (100); Calcd for C<sub>9</sub>H<sub>10</sub>O<sub>5</sub> (M): 198.053. Found: 198.048.

The second fraction above gave **10t** in 10% yield;  $R_f 0.76$  (3 : 2 CH<sub>2</sub>Cl<sub>2</sub>-Me<sub>2</sub>CO), no colour with spray *A*, red colour with spray *B*; GLC retention time, 4.3 min; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  3.4 (bs, 1 H, OH), 3.65 (s, 3 H, Me), 3.85 (d, 1 H, *J* 2.7 Hz, H-5), 4.82 (m, 1 H, *J* 1.3, 2.0, and 2.7 Hz, H-4), 6.20 (dd, 1 H, *J* 1.3 and 6.2 Hz, H-2), 7.40 (dd, 1 H, *J* 2.0 and 6.2 Hz, H-3); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  57.5 (Me), 75.6 (C-4), 88.4 (C-5), 133.2 (C-3), 159.0 (C-2), 199.0 (C-1); MS, m/z (rel int.): 128 (M, 54), 127 (13), 111 (7), 100 (39), 85 (57), 71 (75), 57 (100), 43 (71); Calcd for C<sub>6</sub>H<sub>8</sub>O<sub>3</sub> (M): 128.047. Found: 128.046.

Compound 10t was acetylated as described for 2t. The yield of the acetate was 90%;  $R_f$  0.35 (3:1 light petroleum-EtOAc), red colour with spray B; GLC retention time, 6.5 min; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  2.10 (s, 3 H, Ac), 3.65 (s, 3 H, OMe),

3.96 (d, 1 H, J 2.7 Hz, H-5), 5.76 (m, 1 H, J 1.4, 2.1, and 2.7 Hz, H-4), 6.30 (dd, 1 H, J 1.4 and 6.2 Hz, H-2), 7.40 (dd, 1 H, J 2.1 and 6.2 Hz, H-3); MS, m/z (rel int.): 128 (M – CH<sub>2</sub>CO, 19), 111 (M – AcO', 18), 110 (M – AcOH, 13), 96 (34), 68 (28), 55 (15), 43 (100); Calcd for C<sub>8</sub>H<sub>10</sub>O<sub>4</sub> (M): 170.058. Found: 170.058.

The third fraction above mainly contained unreacted 7, which was recovered in 7% yield.

Hydrolysis and isomerization experiments. -(a) Compound 6 was heated in 0.1 M DCl (in D<sub>2</sub>O) at 90°C in a water bath. The <sup>1</sup>H NMR spectrum was recorded after 1, 2, and 4 h, and showed increasing amounts of 7 and 8 at the expense of 6. The last spectrum also showed traces of 2t. No reductic acid (3) was detected on comparison with an authentic sample.

(b) Compound 7 was treated as for 6 in (a). Compounds 2t and 10t formed gradually at the expense of 7. Again, no 3 was detected.

(c) Compound 2t (100 mg) was heated at 50, 80, or 90°C in a water bath with 1 mL of 0.1 M  $H_2SO_4$ , glacial AcOH, 3.5 M HCl, or 0.5 M NaHCO<sub>3</sub>. Each mixture was analyzed by TLC (3:2 CH<sub>2</sub>Cl<sub>2</sub>-Me<sub>2</sub>CO) after 0.5, 1, 2, 3, and 4 h, using sprays A, B, and C. No 3 was detected, but the amount of 2t decreased with time.

(d) Reductic acid (3) was treated as in (c). Its amount decreased with time, but no 2t was formed.

## ACKNOWLEDGMENTS

We thank Professor Olof Theander for his kind interest in the present work. This was supported by a grant from the Swedish Council for Planning and Coordination of Research and the Swedish Council for Forestry and Agricultural Research.

## REFERENCES

- 1 H.S. Isbell, J. Res. Natl. Bur. Stand., 33 (1944) 45-61.
- 2 K. Aso, H. Sugisawa, and T. Iso, J. Agric. Chem. Soc. Jpn., 29 (1955) 985-987; Chem. Abstr., 51 (1957) 8658b.
- 3 K. Aso and H. Sugisawa, J. Agric. Chem. Soc. Jpn., 30 (1956) 387-392; Chem. Abstr., 52 (1958) 9963f.
- 4 P. Anderegg and H. Neukom, *Mitt. Geb. Lebensmittelunters. Hyg.*, 63 (1972) 81-87; P. Anderegg, Dissertation, ETH, Zürich, 1973, pp 1-64; E. Olsson, Lic. Thesis, Swedish University of Agricultural Sciences, Uppsala, 1989, pp 1-50; T. Ahmad, R. Andersson, K. Olsson, and O. Theander, *Carbohydr. Res.*, 247 (1993) 211-215.
- 5 V.P. Peresada, R.U. Ostrovskaya, A.M. Likhosherstov, A.P. Rodionov, S.G. Rozenberg, and A.P. Skoldinov, *Khim.-Farm. Zh.*, 16 (1982) 537-542; *Chem. Abstr.*, 97 (1982) 109802s.
- 6 O. Achmatowicz, Jr., P. Bukowski, G. Grynkiewicz, B. Szechner, A. Zamojski, and Z. Zwierzchowska, Rocz. Chem., 46 (1972) 879-888.
- 7 M.S. Feather and J.F. Harris, Adv. Carbohydr. Chem. Biochem., 28 (1973) 161-224.
- 8 F.G. Cocu and T. Posternak, Helv. Chim. Acta, 55 (1972) 2838-2844.
- 9 T. Popoff and O. Theander, Carbohydr. Res., 22 (1972) 135-149.
- 10 G. Stenhagen, Int. Mass Spectrom. Conf., 12th, Amsterdam, 1991, Abstr., FrA-D32.
- 11 O. Theander, Acta Chem. Scand., 12 (1958) 1897-1905.
- 12 L. Claisen, Ber. Dtsch. Chem. Ges., 40 (1907) 3903-3914.
- 13 N. Clauson-Kaas, F. Limborg, and P. Dietrich, Acta Chem. Scand., 6 (1952) 545-550.