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Direct Synthesis of Anthracenes from o-Tolualdehydes and Aryl Iodides through Pd(II)-Catalyzed sp^3 C–H Arylation and Electrophilic Aromatic Cyclization

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ABSTRACT

The first direct synthesis of substituted anthracenes from *o*-tolualdehydes and aryl iodides *via* a Pd(II)-catalyzed C–H arylation using an alcohol-bearing transient directing group and subsequent AgOTf-assisted electrophilic aromatic cyclization is described. New transient directing groups consisting of amino acids and amino alcohols enhanced the reactivity, and the C-H arylation was complete in 12 h at 90 °C. By simply changing the silver salt to silver triflate, the one-pot synthesis of anthracene derivatives was carried out using the present reaction conditions

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1. Introduction

Anthracenes have attracted the attention of many chemists by virtue of their broad utility.¹ They can be utilized as chromophores in various kinds of bio-probes² or light-emitting devices³ and as starting materials or reagents in syntheses of multifunctional metal-organic frameworks⁴ or potential polycyclic aromatic drugs.⁵ Their versatility has made the development of efficient methods for their formation highly desirable. A number of synthetic methods for the preparation of anthracenes have been reported. However, those traditional synthetic strategies, such as Bronsted acid-catalyzed dehydrative cyclizations,6 Au-catalyzed cyclizations of 0alkynyldiarylmethanes, [4+2]cycloadditions with naphthoquinones or benzynes,⁸ and Co-catalyzed [2+2+2] alkyne-cyclotrimerizations,⁹ usually require complex substrates that demand multistep syntheses and show poor functional group tolerance. The recent advances in transition metal-catalyzed C-H functionalization inspired us to develop a direct synthetic method for the preparation of substituted anthracenes from easily chemicals. Transition metal-catalyzed accessible C-H functionalizations have undoubtedly become one of the most direct synthetic strategies¹⁰ and have provided many efficient synthetic routes for total syntheses of natural products.¹¹ A directing group (DG) in a substrate, such as a pyridine, an imine, an amine, a (thio)amide, or a carboxylate, are essential in C-H activation strategies to lower the activation energy via the formation of a five- or six-membered metallacycle.¹² However, requiring a DG hurts the step- and atom-economy of the synthesis of a target compound by adding synthetic steps for DG installation ¹and DG detachment.¹³ The emergence of transient directing groups (TDGs) has become a crucial breakthrough for alleviating the intrinsic problems of DG-promoted C–H activations. Since Jun's group reported Rh-catalyzed sp^2 C–H functionalization with transient amino ligands,¹⁴ many relevant studies have been reported.¹⁵ In the case of sp^3 C–H functionalizations, Yu and co-workers first reported a synthetic protocol for the preparation of 2-benzylbenzaldehydes by Pd-catalyzed alkyl C–H activation using an amino acid as a TDG.¹⁶ Later, the related reports of sp^3 C–H functionalization using a TDG strategy have been published by several groups in 2017, respectively.¹⁷

To construct tricyclic aromatic compounds like anthracenes, Sorensen and co-workers used benzaldehydes and aryl iodides with 40 mol% anthranilic acid as a TDG to obtain fluorenones *via* a Pd-catalyzed *ortho* C–H arylation and migratory insertion (Scheme 1-a).^{15c} An approach to the preparation of substituted acridines by Rh(III)-catalyzed C–N bond formation *via ortho* C– H activation controlled by benzyl amine as a TDG and subsequent electrophilic aromatic cyclization has also been reported (Scheme 1-b).^{15a} Inspired by these elegant strategies, we hypothesized that a similar approach would be applicable for the one-pot synthesis of substituted anthracenes by an sp^3 C–H arylation using a TDG and subsequent electrophilic aromatic

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cyclization (Scheme 1-c). In contrast to the acridine synthesis, a benzene moiety is less reactive than an aniline moiety towards electrophilic aromatic cyclization, so a Lewis acid additive is essential to obtain the final product. In this sense, we studied the efficient synthesis of *o*-benzylbenzaldehydes with a TDG-assisted sp^3 C–H arylation strategy as well as the one-pot synthesis of anthracenes. To the best of our knowledge, there have been no reports on the direct synthesis of anthracenes from simple starting materials.



Scheme 1. Direct Synthesis of Tricyclic Aromatic Compounds *via* TDG-assisted C–H activation strategies

2. Results and discussion

To obtain o-benzylbenzaldehydes by an efficient sp^3 C–H arylation, we assumed that oxazoline-bearing primary alkylamine 1 would be the best TDG to accelerate the oxidative addition of the Pd(II) intermediate to the aryl iodide due to the strong σ donating ability of the oxazoline moiety. Moreover, oxazolinebased TDGs have advantages including easy installation of the chiral fragment and simple synthesis from Cbz-protected natural amino acids (valine 3a, phenylalanine 3b, and leucine 3c) and amino alcohols (DL-phenylglycinol **4**a, 2-amino-2methylpropanol 4b, and tert-butylamine 4c); including CDMTmediated amide synthesis for 5, 5a-5e,¹⁸ oxazoline formation under basic condition for 6, and deprotetion of Cbz moiety by



hydrogenolysis for 1 and 2a-2e (Scheme 2).



Based on this strategy, we commenced our study by treating *o*-tolualdehyde (**7a**) with *p*-iodophenol (**8a**), TDG **1**, and the Pd catalyst. The reaction generated arylated product **9a** in 50% yield in 36 h. We also observed that the ring opening of oxazoline **1** occurred under the reaction conditions, and δ -amino alcohol **2a**

was regenerated since the reaction is conducted under acidic conditions. To clarify that the active TDG is either 1 or 2a, an sp^3 C–H arylation reaction with 2a was conducted, and it was found that 2a allowed a comparable yield (48%) to oxazoline-based TDG 1. As a result, we have modified the TDG strategy from oxazoline derivatives to alcohol-bearing, amino acid-amino alcohol coupled molecules as novel TDGs.

We have successfully synthesized a series of amino acidamino alcohol coupled TDGs (2a-2d) by a simple, 3-step procedure from an easily accessible Cbz-protected amino acid. In addition, at the outset of our studies, o-tolualdehvde (7a) and 4iodophenol (8a) were used as model substrates in the presence of various catalytic palladium systems (Table 1). A low conversion was observed in the absence of a TDG after a short time (entry 1). The addition of a transient group increased the efficiency of the reaction (entries 3-5), and the structure of the employed amino acid-based TDGs impacted the activity of the palladium catalyst. Among the three representative amino acid derivatives (2b, 2c, and 2d), phenylalanine-based TDG 2c showed the best conversion in the sp^3 C–H arylation. To verify the role of the hydroxyl group of 2c, a CH₃-substituted TDG (2e), instead of – CH₂OH, was employed in the Pd-catalyzed reaction. In the absence of a terminal alcohol group (2e), the conversion was lowest among the TDGs (entry 7). Further improvement was observed by changing the palladium catalyst precursor from palladium acetate (Pd(OAc)₂) to palladium trifluoroacetate (Pd(TFA)₂). Although no reactivity was observed with Pd(TFA)₂ alone at 90 °C for 6 h (entry 8), the addition of TDG 2c resulted in a dramatic increase of the efficiently of the reaction and led to 81% of arylated product 9a within 12 h at 90 °C (entry 10). It should be noted that the laboratories of Yu and Hu reported Pd(II)-catalyzed sp³ C-H arylations of o-tolualdehyde and aryl iodide using TDGs; glycine was used by Yu's group and acetohydrazide was used by Hu's group.^{16,19} Nevertheless, extended reaction times (24~36 h) and an elevated reaction temperature (110 °C) undermined the practicality of the method. In this sense, TDG 2c could be one solution to making this system more practical. When the amount of palladium catalyst was further increased, the reaction time could be decreased to 6 h with good yield (entries 10 and 11). Interestingly, in this case, a N_2 or O_2 atmosphere reduced the efficiency of the reaction compared to a regular air atmosphere (entries 10, 12, and 13). Unfortunately, aryl bromide was inert under the reaction conditions (entry 14).

Table 1. Optimization of the reaction conditions for the palladium-catalyzed sp^{3} C-H arylation^a

\wedge		OH Pd Cat. / Ligar	nd	_СНОС
		AgTFA (1.5 eq AcOH/H ₂ O=9	uiv) 1/1	I, I
7a	a 8:	90 °C, Time		9a
Ent	Cat.	TDG (mol%)	Time	Yield ^b
ry	(mol%)		(h)	(%)
1	$Pd(OAc)_2$	_	6	0
	(10)			
2	$Pd(OAc)_2$	1	36	50
	(10)			
3	$Pd(OAc)_2$	2a, Val-PhAA	36	48
	(10)	(40)		
4	$Pd(OAc)_2$	2b , Val-Me ₂ AA	36	50
	(10)	(40)		
5	$Pd(OAc)_2$	2c, Phe-Me ₂ AA	36	59
	(10)	(40)		

6	$Pd(OAc)_2$	2d , Leu-Me ₂ AA	36AC	CE43TEI
	(10)	(40)		
7	$Pd(OAc)_2$	2e, Val-AM- <i>t</i> Bu	36	25
	(10)	(40)		
8	Pd(TFA) ₂	-	6	0
	(10)			
9	Pd(TFA) ₂	2c, Phe-Me ₂ AA	6	60
	(10)	(40)		
10	Pd(TFA) ₂	2c, Phe-Me ₂ AA	12	81(63 ^f)
	(10)	(40)		
11	Pd(TFA) ₂	2c, Phe-Me ₂ AA	6	77
	(15)	(40)		
12 ^c	Pd(TFA) ₂	2c, Phe-Me ₂ AA	12	35
	(10)	(40)		
13 ^d	Pd(TFA) ₂	2c, Phe-Me ₂ AA	12	29
	(10)	(40)		
14 ^e	Pd(TFA) ₂	2c, Phe-Me ₂ AA	36	<10
	(10)	(40)		

^aAll reactions were performed with 0.2 mmol (1 equiv.) of **8a** and 0.24 mmol of **7a**. ^b The yield was determined by ¹H NMR analysis of the crude product using CH_2Cl_2 as the internal standard. ^cConditions: O₂ atmosphere ^dConditions: N₂ atmosphere ^e4-Bromotoluene was used instead of 4-iodophenol. ^fIsolated yield.

Under the optimized reaction conditions using 10 mol% Pd(TFA)₂ and 40 mol% Phe-Me₂AA (**2c**), a range of iodoarenes (**8a-8p**) were explored with *o*-tolualdehyde (**3a**) in the sp^3 C–H bond arylation (Table 2). Iodobenzenes **8b**, **8c**, and **8d** with simple non-coordinating substituents (–H or –Me) showed the best performance (87~91% yield). Coordinating substituents were detrimental to the efficiency (54~68% yield), but electronic variations of the substituents on the iodobenzenes did not have a substantial influence on the reaction efficiency (56% of **9f** vs 54% of **9p**). We observed that changes to the steric environment had significant effects on the sp^3 C-H bond arylation (43% of **9e**). In contrast to Ar–I, Ar–F and Ar–Cl were well tolerated in the Pd-catalyzed reaction and allow the preparation of fluoro- and chloroiodobenzene derivatives (**9l**, **9m**, and **9n**).

Table 2. Pd-catalyzed sp^3 C–H arylation with various iodobenzenes^a

		⁴ ³ ⁴ ^R ⁴ ^R ⁴ ^R ^A
7a	8a–8	p 90 °C, 12 h 9a–9p
Entry	R	Yield (%) ^b
1	4-OH	9a , 63
2	Н	9b , 87 ^b
3	3-Me	9c , 91 ^b
4	4-Me	9d , 89 ^b
5	2-OMe	9e , 43 ^b
6	4-OMe	9f , 56 ^b
7	4-Ac	9 g, 55
8	3-CO ₂ H	9h , 68
9	4-CO ₂ H	9i , 66
10	3-CO ₂ Me	9j , 62
11	4-CO ₂ Me	9k , 59
12	3-F	91 , 63 ^b
13	4-F	9m , 54 ^b
14	4-Cl	9n , 62 ^b
15	3-NO ₂	90 , 54

^bsee footnotes a-b of Table 1.

4-NO

16

Next, the scope of benzaldehyde derivatives was examined in the reaction with 1-iodo-3-nitrobenzene (**80**, Scheme 3). It was clearly that this palladium-catalyzed sp^3 C-H bond arylation is highly sensitive to the steric environment. 2,3-Dimethylsubstituted benzaldehyde showed a lower yield of the desired product than 2,4-dimethyl- or 2,5-dimethyl-substituted benzaldehydes (**9q**, **9r**, and **9s**). In addition, when an aldehyde with an additional fluoro substituent was subjected to the reaction conditions, the arylated product was isolated in moderate yield, which suggests electron-rich benzaldehydes are more favorable

9p, 54





Scheme 3. Pd-catalyzed sp^3 C-H arylation with various benzaldehydes.

A putative reaction mechanism of the TDG-assisted sp^3 C–H arylation is proposed as shown in Scheme 4.15c,20 The catalytic cycle begins with the formation of the transient directing group from an amino alcohol and o-tolualdehyde. After complexation of PdX₂ with the in situ-generated imine to form Pd(II)-complex A, the sp^3 C–H proximal to the metal center undergoes palladation with the loss of CF_3COOH .²¹ Oxidative addition of Pd(II)-complex **B** to iodobenzene produces Pd(IV)-intermediate \mathbf{D}^{22} and subsequent reductive elimination allows the formation of anylated imine E along with the regeneration of PdX₂ via a ligand exchange with AgTFA and trifluoroacetic acid (or acetic acid). Hydration of imine E produces the desired obenzylbenzaldehyde and the TDG, which goes on to participate in the next catalytic cycle. A hydroxyl group on the TDG presumably coordinates to the metal center, which may enhance the rate of oxidative addition of Pd(II)-intermediate B and make the reaction more practical than the reaction with -OH-absent TDGs. Moreover, the dramatic effect of the counterion (X = TFA)vs OAc) on Pd(II) was presumably caused by the basicity of acetate and trifluoroacetate. The basic acetate ion was prone to deprotonate the -OH of the amino alcohol to generate intermediate C, inhibiting the catalytic cycle, but the less basic trifluoroacetate favors the formation of **B**, leading to reduced



reaction times and superior yields.

Tetrahedron ACCEPTED M/4. Experimental section

Scheme 4. Proposed Reaction Mechanism

Based on the results of sp^3 C–H arylation, a cascade reaction of the Pd-catalyzed o-benzylbenzaldehyde formation and Lewis acid-mediated electrophilic aromatic cyclization (Friedel-Crafts cyclization) was studied. Based on our screening of various Lewis acids, the silver triflate (AgOTf) oxidant was found to produce substituted anthracenes in a one-pot reaction. Although the efficiency of silver triflate for sp^3 C–H arylations is much lower than that of silver trifluoroacetate (i.e., the best silver oxidant for the present system), silver triflate was the sole reagent performing the cyclization step following the sp^3 C–H arylation. Once again, by simply changing the silver salt from AgTFA to AgOTf, the one-pot synthesis of anthracene derivatives was successively achieved in the present $Pd(TFA)_2$ -2c system (Scheme 5). In addition, the addition of 4-nitroaniline enhances the reactivity and reproducibility of the reaction. Under the newly developed one-pot reaction conditions, the substrate scope was then examined with a special focus on functional group tolerance. This cyclization to generate anthracene derivatives was significantly affected by both electronic and steric environments. Iodobenzenes or tolualdehydes bearing electron-withdrawing substituents showed very poor conversion in the synthesis of anthracenes. Thus, a variety of methyl-substituted and fluorosubstituted anthracenes were synthesized from methyl-substituted iodobenzene and tolualdehyde derivatives. In general, <50% of sp^3 C-H arylated product was cyclized to the corresponding anthracene derivative.



Scheme 5. One-pot synthesis of anthracenes by Pd-catalyzed sp^3 C-H arylations

3. Conclusion

In summary, we have reported an efficient palladiumcatalyzed sp^3 C–H bond arylation using amino acid-amino alcohol-type transient directing groups with a silver oxidant. The reaction was efficiently carried out with a wide range of substrates in short reaction times. By simply changing the silver salt to AgOTf, the sp^3 -arylated compound is then cyclized. Therefore, various anthracene derivatives can be successfully synthesized by a one-pot reaction using the present reaction conditions.

4.1. General Methods.

Concentration of solution was carried out by using a rotary evaporator with a water aspirator, and generally followed by removal of residual solvents on a vacuum line held at 0.1-1 torr. Unless otherwise stated, all commercial reagents and solvents were used without additional purification. All chemicals were purchased from Sigma-Aldrich, TCI, Alfa Aesar chemical company. Analytical thin layer chromatography (TLC) was performed on precoated silica gel 60 F254 plates. Visualization on TLC was achieved by the use of UV light (254 nm). Flash column chromatography was undertaken on silica gel (400-630 mesh). Proton nuclear magnetic resonance spectra (¹H NMR) were recorded on FT AM 400 or 500 (400 or 500 MHz). Chemical shifts were quoted in parts per million (ppm) referenced to the appropriate solvent peak. The following abbreviations were used to describe peak patterns when appropriate: br = broad, s = singlet, d = doublet, t = triplet, q =quartet, and m = multiplet. Coupling constants, J, were reported in Hertz unit (Hz). Carbon 13 nuclear magnetic resonance spectroscopy (¹³C NMR) was recorded on FT AM 400 or 500 (100 or 125 MHz) and was fully decoupled by broad band decoupling. Chemical shifts were reported in ppm referenced to the center line of a triplet at 77.0 ppm of chloroform-d. or 39.5 ppm of DMSO-d₆.

4.2. Preparation of Oxazoline-based TDG 1

N-Cbz-L-valine **3a** (2.0 mmol, 503 mg) was reacted with 2chloro-4,6-dimethoxy-1,3,5-triazine (CDMT, 2.2 mmol, 386 mg) and 4-methylmorpholine (NMM, 6.0 mmol, 0.66 mL) in THF 5 mL at room temperature for 1 h. To mixture solution was added *DL*-phenylglycinol **4a** (2.0 mmol, 274 mg) in THF 5 mL and the mixture was stirred at room temperature for 12 h. The mixture was extracted with ethyl acetate (25 mL) and organic layer was dried with MgSO₄, filtered and evaporated. Purification by silica gel column chromatography afforded the corresponding amide **5a**.

5a (1.0 mmol, 370 mg) was reacted with *p*-toluenesulfonylchloride (1.1mmol, 210 mg), trimethylamine (3.0mmol, 0.42 mL) and 4-dimethylaminopyridine (0.10 mmol, 12 mg) in DCM 20 mL at room temperature for 24 h. The mixture was extracted with DCM (25 mL) and organic layer was dried with MgSO₄, filtered and evaporated. Purification by silica gel column chromatography afforded the corresponding Cbz-protected oxazoline **6**.

6 (0.17 mmol, 60 mg) was stirred under hydrogen atmosphere (1 atm) in the presence of 10% Pd/C (6.0 mg) in MeOH (5 mL) at room temperature for 1 h. After filtering the mixture through celite followed by evaporation of the volatile materials in vacuo. Purification by silica gel column chromatography afforded the corresponding oxazoline-based TDG **1**.

4.3. Typical Procedure for the Preparation of Amino Alcoholbased TDGs 2a-2e

N-Cbz-protected amino acid **3a**, **3b** or **3c** (2 mmol) was reacted with 2-chloro-4,6-dimethoxy-1,3,5-triazine (CDMT, 2.2 mmol, 386 mg) and 4-methylmorpholine (NMM, 6 mmol, 0.66 mL) in THF 5 mL at room temperature for 1 h. To mixture solution was added amino alcohol **4a**, **4b** or **4c** (2 mmol) in THF 5 mL and the mixture was stirred at room temperature for 12 h. The mixture was extracted with ethyl acetate (25 mL) and

organic layer was dried with MgSO4, filtered and evaporated.	$MS(+) \subset m/z^{-}$	calcd.	For $C_{17}H_{27}N_2O_4$	$[M+H]^+$:
Purification by silica gel column chromatography afforded the	323.1965,	found	$[M+H]^+$:	323.1965.
corresponding amide 5a–5e.				

The amide **5a–5e** was stirred under hydrogen atmosphere (1 atm) in the presence of 10% Pd/C (6 mg) in MeOH (10 mL) at room temperature for 2 h. After filtering the mixture through celite followed by evaporation of the volatile materials in vacuo. Purification by silica gel column chromatography afforeded the corresponding TDGs **2a–2e**.

4.4. Typical Procedure for the Pd-Catalyzed sp³ C–H Arylation 9a–9u

o-Tolualdehyde (0.24 mmol), iodobenzene (0.2 mmol), silver trifluoroacetate (0.3 mmol, 66 mg), palladium trifluoroacetate (0.02 mmol, 7 mg), Phe-Me₂ ligand (0.08 mmol, 19 mg), acetic acid (1.8 mL) and H₂O (0.2 mL) were added to a vial. The mixture was heated to 90 °C for 12 h. The reaction mixture was allowed to cool to room temperature and filtered through a silica gel pad, concentrated in vacuum. The desired product was isolated by a silica gel column chromatography or preparative TLC.

4.5. Typical Procedure for the Preparation of Anthracenes 10a-10f

o-Tolualdehyde (0.24 mmol), iodobenzene (0.2 mmol), silver trifluoromethanesulfonate (0.3 mmol, 77 mg), Phe-Me₂ ligand (0.08 mmol, 19 mg), 4-nitroaniline (0.4 mmol, 55 mg), palladium trifluoroacetate (0.02 mmol, 7 mg), acetic acid (1.8 mL) and H₂O (0.2 mL) were added to a vial. The mixture was heated to 120 °C for 24 h. The reaction mixture was allowed to cool to room temperature and filtered through a silica gel pad, concentrated in vacuum. The desired product was isolated by a silica gel column chromatography or preparative TLC.

4.6. The Characterization data of products

4.6.1. Benzyl ((S)-1-(((R, or S)-2-hydroxy-1phenylethyl)amino)-3-methyl-1-oxobutan-2yl)carbamate $(5a)^{23}$

¹H NMR (400 MHz, DMSO-d₆): δ 8.36-8.34 (d, 1H, J = 8.1 Hz, 5a(S,S)), 8.23-8.21(d, 1H, J = 8.1 Hz, **5a**(S,R)), 7.37-7.22 (m, 11H, **5a**(S,S) + **5a**(S,R)), 5.05 (s, 1H, **5a**(S,S)), 5.02(s, 1H, **5a**(S,R)), 4.86-4.83 (m, 2H, **5a**(S,S)), 5.02(s, 1H, **5a**(S,R)), 4.86-4.83 (m, 2H, **5a**(S,S)), 3.57-3.54 (m, 2H, **5a**(S,S) + **5a**(S,R)), 3.57-3.54 (m, 2H, **5a**(S,S) + **5a**(S,R)), 2.00-1.92 (m, 1H, **5a**(S,S) + **5a**(S,R)), 0.89-0.84 (m, 6H, **5a**(S,R)), 0.80-0.76 (m, 6H, **5a**(S,S)); ¹³C NMR (100 MHz, DMSO-d₆): δ 171.3, 156.6, 141.7, 141.5, 137.6, 128.8, 128.5, 128.4, 128.2, 128.1, 128.1, 127.5, 127.4, 127.2, 65.8, 65.1, 60.8, 60.6, 55.44, 55.38, 31.1, 30.8, 19.75, 19.69, 18.7, 18.5.

4.6.2. (S)-Benzyl (1-((1-hydroxy-2-methylpropan-2yl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (5b)

¹H NMR (400 MHz, DMSO-d₆): δ 7.36-7.30 (m, 6H), 7.17-7.15 (d, 1H, J = 8.8 Hz), 5.03 (s, 2H), 4.85-4.82 (t, 1H, J = 5.5 Hz), 3.84-3.80 (t, 1H, J = 7.9 Hz), 3.40-3.34 (m, 3H) 1.92-1.87 (m, 1H), 1.17 (s, 6H), 0.86-0.81 (m, 6H); ¹³C NMR (100 MHz, DMSO-d₆): δ 171.0, 156.1, 137.2, 128.3, 127.8, 127.6, 67.6, 65.3, 60.4, 54.4, 40.1, 40.0, 39.7, 39.5, 39.3, 39.1, 38.9, 30.6, 23.5, 23.3, 19.2, 18.1; ESI-

4.6.3. (S)-Benzyl (1-((1-hydroxy-2-methylpropan-2-
yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate
(5c)

¹H NMR (500 MHz, CDCl₃): δ 7.37-7.19 (m, 11H), 5.66 (br s, 1H), 5.45 (br s, 1H), 5.08 (s, 2H), 4.29-4.28 (br d, 1H), 3.50-3.43 (m, 2H), 3.13-3.09 (dd, 1H, J = 13.5 Hz, J = 6.1 Hz), 2.97-2.93 (dd, 1H, J = 13.5 Hz, J = 8.3 Hz), 1.11 (s, 3H), 1.09 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): 171.2, 156.0, 136.3, 136.0, 129.3, 128.7, 128.5, 128.2, 128.0, 127.1, 69.3, 67.0, 56.8, 56.0, 38.9, 24.3, 23.9; ESI-MS(+) m/z calcd. For C₂₁H₂₇N₂O₄ [M+H]⁺: 371.1965, found [M+H]⁺: 371.1965.

4.6.4. (S)-Benzyl (1-((1-hydroxy-2-methylpropan-2yl)amino)-4-methyl-1-oxopentan-2-yl)carbamate (5d)

¹H NMR (400 MHz, CDCl₃): δ 7.36-7.34 (m, 5H), 6.10 (br s, 1H), 5.18-5.10 (m, 3H), 4.08-4.02 (m, 1H), 3.62-3.51 (m, 2H,), 1.69-1.58 (m, 2H, 1.51-1.47 (m, 1H), 1.27-1.24 (d, 6H, *J* = 10.9 Hz), 0.92-0.94 (m, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 172.7, 156.4, 136.0, 128.6, 128.3, 128.1, 69.8, 67.2, 56.2, 54.1, 41.1, 24.7, 24.6, 24.3, 22.9, 22.0; ESI-MS(+) m/z calcd. For C₁₈H₂₉N₂O₄ [M+H]⁺: 337.2122, found [M+H]⁺: 337.2122.

4.6.5. (S)-Benzyl $(1-(tert-butylamino)-1-oxo-3-phenylpropan-2-yl)carbamate (5e)^{24}$

¹H NMR (400 MHz, CDCl₃): δ 7.29-7.14 (m, 10H), 5.42 (br s, 1H), 5.08-5.02 (m, 3H), 4.15 (br s, 1H), 3.08-3.04 (dd, 1H, J = 12.8 Hz, J = 5.3 Hz), 2.81-2.79 (m, 1H), 1.09 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 169.5, 155.8, 136.8, 136.2, 129.4, 128.7, 128.5, 128.1, 128.0, 127.0, 66.9, 56.8, 51.3, 39.3, 28.4.

4.6.6. Benzyl ((1S)-2-methyl-1-((R or S)-4-phenyl-4,5-dihydrooxazol-2-yl)propyl)carbamate (**6**)

¹H NMR (400 MHz, CDCl₃): δ 7.37-7.21 (m, 10H, **6**(S,S) + **6**(S,R)), 6.03-5.87 (d, 2H, *J* = 9.0 Hz, **6**(S,S) + **6**(S,R)), 5.18-5.08 (m, 3H, **6**(S,S) + **6**(S,R)), 4.65-4.59 (m, 1H, **6**(S,S) + **6**(S,R)), 4.56-4.48 (m, 1H, **6**(S,S) + **6**(S,R)), 4.12-4.02 (m, 1H, **6**(S,S) + **6**(S,R)), 2.24-2.16 (m, 1H, **6**(S,S) + **6**(S,R) + **6**(S,R)), 1.04-1.02 (d, 3H, *J* = 6.6 Hz, **6**(S,S) + **6**(S,R)), 0.96-0.95 (d, 3H, *J* = 6.8 Hz, **6**(S,S) + **6**(S,R)); ¹³C NMR (100 MHz, CDCl₃): δ 167.78, 167.61, 158.1, 141.7, 141.6, 136.3, 128.6, 128.6, 128.3, 127.94, 127.91, 127.53, 127.50, 126.50, 126.46, 69.11, 69.07, 66.7, 54.5, 54.4, 31.4, 29.6, 18.84, 18.78, 17.5; ESI-MS(+) m/z calcd. For C₂₁H₂₅N₂O₃ [M+H]⁺: 353.1860, found [M+H]⁺: 353.1860.

4.6.7. (S)-2-Methyl-1-((R or S)-4-phenyl-4,5dihydrooxazol-2-yl)propan-1-amine $(1)^{25}$.

¹H NMR (400 MHz, CDCl₃): δ 7.37-7.24 (m, 5H, **1**(S,S)+**1**(S,R)), 5.22-5.18 (m, 1H, **1**(S,S)+**1**(S,R)), 5.22-5.18 (t, 1H, J = 9.9 Hz, **1**(S,S)+**1**(S,R)), 4.69-4.65, (dd, 1H, J = 8.5 Hz, J = 10.2 Hz, **1**(S,S)), 4.68-4.63 (dd, 1H, J = 8.5 Hz, J = 10.3 Hz, **1**(S,R)), 4.15-4.11 (t, 1H, J = 8.4 Hz, **1**(S,R)), 4.14-4.10 (t,

1H, J = 8.5 Hz, 1(S,S), 3.48-3.45 (dd, 1H, J = 5.4 M 4.6.12, (S)-2-Amino-N-(tert-butyl)-3-Hz, J = 1.2 Hz, 1(S,S)+1(S,R)), 2.12-2.10. (m, 1H, phenylpropanamide (2e)²⁷

Hz, J = 1.2 Hz, $\mathbf{1}(S,S)+\mathbf{1}(S,R)$), 2.12-2.10. (m, 1H, $\mathbf{1}(S,S)+\mathbf{1}(S,R)$), 1.051-1.034(d, 3H, J = 6.8 Hz, $\mathbf{1}(S,S)$), 1.044-1.027 (d, 3H, J = 6.8 Hz, $\mathbf{1}(S,R)$), 1.017-1.000 (d, 3H, J = 6.8 Hz, $\mathbf{1}(S,S)$), 1.012-0.995 (d, 3H, J = 6.8 Hz, $\mathbf{1}(S,R)$); ¹³C NMR (100 MHz, CDCl₃): δ 171.0, 170.8, 142.01, 141.97, 128.56, 128.54, 127.40, 127.38, 126.44, 126.40, 74.8, 74.7, 69.13, 69.06, 55.4, 55.3, 32.1, 32.0, 19.3, 17.5, 17.3.

4.6.8. (S)-2-Amino-N-((R or S)-2-hydroxy-1phenylethyl)-3-methylbutanamide $(2a)^{25, 26}$

¹H NMR (400 MHz, CDCl₃): δ 8.16-8.14 (d, 1H, J = 7.4 Hz, **2a**(S,S)), 8.09-8.08 (d, 1H, J = 7.2 Hz, **2a**(S,R)) 7.37-7.28 (m, 5H, **2a**(S,S)+**2a**(S,R)), 5.10-5.04 (m, 1H, **2a**(S,S)+**2a**(S,R)), 3.87-3.83 (m, 2H, **2a**(S,S)+**2a**(S,R)), 3.41-3.40 (d, 1H, J = 4.6 Hz, **2a**(S,S)) 3.32-3.31 (d, 1H, J = 3.9 Hz, **2a**(S,R)), 2.86 (br s, 3H, **2a**(S,S)+**2a**(S,R)), 2.33-2.21 (m, 1H, **2a**(S,S)+**2a**(S,R)), 0.98-0.96 (dd, 6H, J = 6.9 Hz, J = 1.4 Hz, **2a**(S,S)), 0.91-0.90 (d, 3H, J = 6.9 Hz, J = 1.4 Hz, **2a**(S,S)), 0.91-0.90 (d, 3H, J = 6.9 Hz, **2a**(S,R)), 0.79-0.78(d, 3H, J = 6.9 Hz, 2a(S,R)); ¹³C NMR (100 MHz, CDCl₃): δ 175.1, 174.1, 139.0, 138.8, 128.8, 127.80, 127.77, 126.74, 126.67, 67.0, 66.8, 60.1, 60.0, 56.2, 56.0, 31.0, 30.8, 19.7, 19.4, 16.6, 16.1.

4.6.9. (S)-2-Amino-N-(1-hydroxy-2-methylpropan-2yl)-3-methylbutanamide (**2b**)

¹H NMR (400 MHz, CDCl₃): δ 7.59 (s, 1H), 3.62-3.51 (m, 2H), 3.29-3.28 (d, 1H, *J* = 4.1 Hz), 2.33-2.25 (m, 1H), 1.29-1.28 (d, 6H, *J* = 3.2 Hz), 1.00-0.98 (d, 3H, *J* = 7.0 Hz), 0.85-0.83 (d, 3H, *J* = 6.9 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 174.5, 59.8, 55.8, 30.6, 24.8, 24.6, 19.5, 16.1; ESI-MS(+) m/z calcd. For C₉H₂₁N₂O₂ [M+H]⁺: 189.1598, found [M+H]⁺: 189.1598.

4.6.10. (S)-2-Amino-N-(1-hydroxy-2-methylpropan-2-yl)-3-phenylpropanamide (2c)

¹H NMR (500 MHz, CDCl₃): δ 7.38 (br s, 1H), 7.33-7.22 (m, 5H), 3.68-3.65 (dd, 1H, *J* = 8.4 Hz, *J* = 4.9 Hz), 3.63-3.61 (d, 1H, *J* =12 Hz), 3.52-3.50 (d, 1H, *J* = 12 Hz), 3.20-3.16 (dd, 1H, *J* =13.7 Hz, *J* = 4.9 Hz), 2.85-2.81 (dd, 1H, *J* = 13.7 Hz, *J* = 8.4 Hz), 1.25 (s, 3H), 1.21 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 174.2, 137.1, 129.4, 128.7, 127.0, 70.4, 56.1, 55.7, 40.4, 24.8, 24.4; ESI-MS(+) m/z calcd. For C₁₃H₂₁N₂O₂ [M+H]⁺: 237.1598, found [M+H]⁺: 237.1598.

4.6.11. (S)-2-Amino-N-(1-hydroxy-2-methylpropan-2-yl)-4-methylpentanamide (2d)

¹H NMR (400 MHz, CDCl₃): δ 7.60 (s, 1H), 3.62-3.51 (m, 2H), 3.41-3.37 (m, 1H), 3.11 (br s, 2H), 1.70-1.66 (m, 2H,), 1.36-1.26 (m, 7H), 0.96-0.91 (dd, 6H, J = 12.5 Hz, J = 6.3 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 176.0, 70.6, 55.6, 53.3, 43.6, 24.84, 24.78, 24.6, 23.3, 21.3; ESI-MS(+) m/z calcd. For C₁₀H₂₃N₂O₂ [M+H]⁺: 203.1754, found [M+H]⁺: 203.1754.

¹H NMR (400 MHz, CDCl₃): δ 7.35-7.24 (m, 5H), 7.00 (br s, 1H), 3.57-3.53 (dd, 1H, J = 8.7 Hz, J = 4.7 Hz), 3.25-3.20 (dd, 1H, J = 13.7 Hz, J = 4.7Hz), 2.81-2.75 (dd, 1H, J = 13.7 Hz, J = 8.7 Hz), 2.02 (br s, 2H), 1.34 (s, 9H); ¹³C NMR (125 MHz, CDCl₃): δ 172.9, 137.9, 129.4, 128.6, 126.7, 56.8, 50.5, 40.8, 28.6.

4.6.13. 2-(4-Hydroxybenzyl)benzaldehyde $(9a)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.24 (s, 1H), 7.87-7.85 (dd, 1H, J = 7.6 Hz, 1.4 Hz), 7.55-7.51 (dd, 1H, J = 7.5 Hz, 1.5 Hz), 7.43-7.39 (td, 1H, J = 7.5 Hz, 0.9 Hz), 7.28-7.26 (d, 1H, J = 7.4 Hz), 7.00-6.98 (d, 2H, J = 8.7 Hz), 6.75-6.73 (d, 2H, J = 8.6 Hz), 4.36 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 192.9, 154.1, 143.6, 134.1, 133.7, 132.3, 132.0, 131.5, 129.8, 126.9, 115.4, 37.2.

4.6.14. 2-Benzylbenzaldehyde $(9b)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.26 (s, 1H), 7.85-7.87 (dd, 1H, J = 7.6 Hz, 1.4 Hz), 7.55-7.51 (td, 1H, J = 7.5 Hz, 1.5 Hz), 7.44-7.42 (td, 1H, J = 7.6 Hz, 1.0 Hz), 7.30-7.26 (m, 3H), 7.21-7.14 (m, 3H), 4.46 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 192.4, 143.0, 140.2, 133.9, 132.0, 131.6, 128.8, 128.6, 127.0, 126.3, 38.0.

4.6.15. 2-(3-Methylbenzyl)benzaldehyde $(9c)^{19}$

¹H NMR (400 MHz, CDCl₃): δ 10.27 (s, 1H), 7.89-7.86 (dd, 1H, J = 7.6 Hz, 1.5 Hz), 7.55-7.51 (td, 1H, J = 7.5 Hz, 1.5 Hz), 7.44-7.40 (td, 1H, J =7.4 Hz, 0.9 Hz), 7.28-7.26 (d, 1H, J = 7.7 Hz), 7.19-7.15 (t, 1H, J = 7.6 Hz), 7.03-7.00 (d, 1 H, J =7.6 Hz), 6.97-6.93 (m, 2H), 4.42 (s, 2H), 2.30 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 192.4, 143.1, 140.2, 138.2, 133.9, 131.7, 131.6, 129.5, 128.4, 127.0, 126.9, 125.8, 37.9, 21.4.

4.6.16. $2 - (4 - Methylbenzyl)benzaldehyde (9d)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.26 (s, 1H), 7.87-7.85 (dd, 1H, J = 7.6 Hz, 1.4 Hz), 7.54-7.50 (td, 1H, J = 7.5 Hz, 1.4 Hz), 7.42-7.38 (td, 1H, J = 7.5 Hz, 0.7 Hz), 7.27-7.25 (d, 1H, J = 7.4 Hz), 7.09-7.07 (d, 2H, J = 7.9 Hz), 7.03-7.02 (d, 2H, J = 8.0 Hz), 4.40 (s, 2H), 2.30 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 192.4, 143.4, 137.2, 135.8, 133.9, 131.8, 131.6, 129.3, 128.7, 126.9, 37.6, 21.0.

4.6.17. 2-(2-Methoxybenzyl)benzaldehyde $(9e)^{28}$

¹H NMR (400 MHz, CDCl₃): δ 10.36 (s, 1H), 7.90-7.87 (dd, 1H, J = 7.7 Hz, 1.4 Hz), 7.49-7.47 (td, 1H, J = 7.5 Hz, 1.6 Hz), 7.39-7.35 (td, 1H, J =7.5 Hz, 0.6 Hz), 7.24-7.19 (m, 2H), 6.95-6.93 (dd, 1H, J = 7.3 Hz), 6.88-6.84 (m, 2H), 4.40 (s, 2H), 3.81 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 192.4, 157.0, 143.3, 134.0, 133.8, 131.3, 130.2, 130.0, 128.7, 127.7, 126.6, 120.6, 110.3, 55.2, 31.8.

4.6.18. 2-(4-Methoxybenzyl)benzaldehyde $(9f)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.26 (s, 1H), 7.87-7.85 (dd, 1H, J = 7.6 Hz, 1.4 Hz), 7.54-7.50

(td, 1H, J = 7.5 Hz, 1.5 Hz), 7.43-7.39 (td, TIH, $J = M \ 16412$ (td, 7.7142.85, 142.78, 142.0, 134.0, 133.0, 7.6 Hz, 0.9 Hz), 7.27-7.25 (d, 1H, J = 7.6 Hz), 7.07-7.05 (d, 2H, J = 8.8 Hz), 6.83-6.81 (d, 2H, J =8.8 Hz), 4.39 (s, 2H), 3.77 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 192.4, 158.0, 133.9, 132.3, 131.9, 131.5, 129.7, 126.9, 113.9, 55.2, 37.1.

4.6.19. 2-(4-Acetylbenzyl)benzaldehyde (9g)

¹H NMR (400 MHz, CDCl₃): δ 10.17 (s, 1H), 7.88-7.84 (m, 3H), 7.57-7.53 (td, 1H, J = 7.5 Hz, 1.6 Hz), 7.48-7.34 (td, 1H, J = 7.5 Hz, 1.2 Hz), 7.27-7.22 (m, 3H), 4.51 (s, 2H), 2.56 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 197.7, 192.5, 145.9, 141.6, 135.3, 134.0, 133.9, 133.5, 131.8, 128.9, 128.6, 127.3, 38.2, 26.5; ESI-MS(+) m/z calcd. For $C_{16}H_{14}NaO_2$ [M+Na]⁺: 261.0886, found [M+Na]⁺: 261.0891.

4.6.20. $3 - (2 - Formylbenzyl)benzoic acid (9h)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.20, (s, 1H), 7.95-7.85 (m, 3H), 7.58-7.54(td, 1H, J = 7.5 Hz, 1.5 Hz), 7.48-7.44(td, 1H, J = 7.5 Hz, 1.2 Hz), 7.42-7.36(m, 2H), 7.29-7.7.26(d, 1H, J = 7.6 Hz), 4.52(s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 192.6, 171.0, 142.0, 140.8, 134.3, 134.0, 133.8, 133.4, 131.7, 130.4, 129.4, 128.7, 128.2, 127.3, 37.9.

4.6.21. 4-(2-Formylbenzyl)benzoic acid $(9i)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.18, (s, 1H), 8.02-8.00 (d, 2H, J = 8.4 Hz), 7.87-7.85 (dd, 1H, J =7.6 Hz, 1.5 Hz), 7.58-7.54 (td, 1H, J = 7.5 Hz, 1.6 Hz), 7.49-7.45(td, 1H, J = 7.5 Hz, 1.2 Hz), 7.28-7.7.24(m, 3H), 4.53 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 192.6, 171.0, 146.7, 141.6, 134.0, 133.9, 133.5, 131.8, 130.5, 128.9, 127.4, 127.2, 38.3.

4.6.22. Methyl 3-(2-formylbenzyl)benzoate $(9j)^{16}$

¹H NMR (400 MHz, $CDCl_3$): δ 10.20 (s, 1H), 7.88-7.84 (m, 3H), 7.56-7.52 (td, 1H, J = 7.5 Hz, 1.6 Hz), 7.60-7.44 (td, 1H, J = 7.5 Hz, 1.2 Hz), 7.35-7.33 (m, 2H), 7.27-7.25 (d, 1H, J = 7.6 Hz), 4.50 (s, 2H), 3.88 (s, 3H); 13 C NMR (100 MHz, CDCl₃): δ 192.5, 167.0, 142.1, 140.6, 134.0, 133.8, 133.4, 133.1, 131.7, 130.4, 129.8, 128.6, 127.6, 127.2, 52.1, 37.9.

4.6.23. Methyl 4-(2-formylbenzyl)benzoate $(9k)^{16}$

¹H NMR (400 MHz, $CDCl_3$): δ 10.18 (s, 1H), 7.95-7.92 (d, 2H, J = 8.5 Hz), 7.86-7.84 (dd, 1H, J = 7.6 Hz, 1.5 Hz), 7.56-7.52 (td, 1H, J = 7.5 Hz, 1.6 Hz), 7.47-7.43 (td, 1H, J = 7.5 Hz, 1.2 Hz), 7.26-7.24 (d, 1H, J = 7.5 Hz), 7.22-7.20 (d, 2H, J =8.6 Hz), 4.50 (s, 2H), 3.88 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 192.5, 166.9, 145.7, 141.7, 133.93, 133.86, 133.3, 131.7, 129.8, 128.8, 128.2, 127.3, 52.0, 38.2.

4.6.24. 2-(3-Fluorobenzyl)benzaldehyde $(91)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.20 (s, 1H), 7.87-7.85 (dd, 1H, J = 7.6 Hz, 1.4 Hz), 7.57-7.53(td, 1H, J = 7.5 Hz, 1.5), 7.47-7.43 (td, 1H, J = 7.5 Hz, 1.1 Hz), 7.27-7.21 (m, 2H), 6.95-6.81 (m, 3H), 4.45 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 192.5, 131.7, 129.9, 129.8, 127.3, 124.43, 124.41, 115.8, 115.5, 113.3, 113.1, 37.83, 37.82.

4.6.25. 2-(4-Fluorobenzyl)benzaldehyde $(9m)^{19}$

¹H NMR (400 MHz, CDCl₃): δ 10.21 (s, 1H), 7.87-7.84 (dd, 1H, J = 7.6 Hz, 1.4 Hz), 7.56-7.52(td, 1H, J = 7.5 Hz, 1.5 Hz), 7.45-7.42 (td, 1H, J =7.5 Hz, 1.0 Hz), 7.26-7.24 (d, 1H, J = 7.7 Hz), 7.12-7.09 (m, 2H), 6.98-6.93 (m, 2H), 4.42 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 192.5, 162.6, 160.2, 133.9, 132.8, 131.5, 130.2, 130.1, 127.1, 115.4, 115.2, 37.3.

4.6.26. 2-(4-Chlorobenzyl)benzaldehyde $(9n)^{19}$

¹H NMR (400 MHz, CDCl₃): δ 10.19 (s, 1H), 7.86-7.84 (dd, 1H, J = 7.6 Hz, 1.5 Hz), 7.56-7.52(td, 1H, J = 7.5 Hz, 1.5 Hz), 7.46-7.42 (td, 1H, J =(di, 11, 0 11, 12, 12, 12, 11, 12, 11, 12), (di, 11, 0 7.5 Hz, 1.1 Hz), 7.26-7.22 (m, 3H), 7.09-7.07 (d, 2H, J = 8.6 Hz), 4.42 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 192.5, 142.3, 138.7, 133.9, 133.8, 133.1, 132.0, 131.6, 130.1, 128.6, 127.2, 37.5

4.6.27. 2-(3-Nitrobenzyl)benzaldehyde $(9o)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.13 (s, 1H), 8.05-8.03 (d, 1H, J = 8.1 Hz), 7.99 (s, 1H), 7.87-7.85 (dd, 1H, J = 7.5 Hz, 1.4 Hz), 7.61-7.57 (td, 1H, J = 7.5 Hz, 1.5 Hz), 7.53-7.49 (m, 2H), 7.45-7.41 (t, H, J = 7.9 Hz), 7.31-7.29 (d, 1H, J = 7.5Hz), 4.55 (s, 2H); 13 C NMR (100 MHz, CDCl₃): δ 192.8, 148.3, 142.4, 140.7, 135.1, 134.7, 134.1, 133.8, 131.9, 129.2, 127.7, 123.5, 121.4, 38.0.

4.6.28. 2 - (4 - Nitrobenzyl) benzaldehyde (**9p** $)^{19}$

¹H NMR (400 MHz, CDCl₃): δ 10.11 (s, 1H), 8.13-8.11 (m, 2H), 7.87-7.85 (dd, 1H, J = 7.5 Hz, 1.5 Hz), 7.61-7.57 (td, 1H, J = 7.5 Hz, 1.5 Hz), 7.53-7.49 (td, 1H, J = 7.5 Hz, 1.3 Hz), 7.32-7.28 (m, 3H), 4.55 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 192.7, 148.1, 146.4, 140.5, 134.7, 134.0, 133.8, 131.9, 129.5, 127.7, 123.6, 38.3.

4.6.29. 5-Methyl-2-(3-nitrobenzyl)benzaldehyde $(9q)^{1\hat{6}}$

¹H NMR (400 MHz, CDCl₃): δ 10.08 (s, 1H), 8.04-8.02 (m, 1H), 7.97 (s, 1H), 7.65 (s, 1H), 7.52-7.50 (d, 1H, J = 7.6 Hz), 7.44-7.38 (m, 2H), 7.20-7.18 (d, 1H, J = 7.7 Hz), 2.44 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 192.9, 148.3, 142.7, 137.7, 137.5, 135.2, 135.0, 134.8, 133.6, 131.9, 129.2, 123.4, 121.3, 37.6, 20.8.

4.6.30. 4-Methyl-2-(3-nitrobenzyl)benzaldehyde $(9r)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.13 (s, 1H), 8.05-8.03 (m, 1H), 7.99 (s, 1H), 7.75-7.73 (d, 1H, J = 7.7 Hz), 7.53-7.51 (m, 1H), 7.45-7.41 (t, 1H, J =7.9 Hz), 7.31-7.26 (d, 1H, J = 7.8 Hz), 7.10 (s, 1H), 4.51(s, 2H), 2.42(s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 192.4, 148.3, 145.2, 142.5, 140.6, 135.11, 135.07, 132.7, 131.5, 129.2, 128.3., 123.4, 121.3, 38.0, 21.8.

4.6.31. 3-Methyl-2-(3-nitrobenzyl)benzaldehyde M (10d) (7.30-7.32 (dd, 1H, J = 8.7 Hz, J = 1.6 Hz, $(9s)^{16}$ (0b), 2.83 (s, 3H, 10d), 2.56 (s, 3H, 10b); 13 C NMR

¹H NMR (400 MHz, CDCl₃): δ 10.13 (s, 1H), 8.04-8.01 (m, 1H), 7.85 (s, 1H), 7.76-7.74 (dd, 1H, J = 7.4 Hz, J = 1.4 Hz), 7.51-7.35 (m, 4H), 4.62 (s, 2H), 2.31(s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 193.2, 148.3, 134.8, 139.0, 138.2, 136.3, 134.5, 134.3, 132.7, 129.2, 127.6, 122.8, 121.2, 33.3, 19.6.

4.6.32. 5-Fluoro-2-(3-nitrobenzyl)benzaldehyde $(9t)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.10 (d, 1H, J = 1.0 Hz), 8.07-8.04 (m, 1H), 7.97 (s, 1H), 7.57-7.54 (td, 1H, J = 8.5 Hz, J = 1.7 Hz), 7.50-7.43 (m, 2H), 7.30-7.29 (dd, 1H, J = 6.6 Hz, J = 1.7 Hz), 4.5 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 190.9, 190.9, 163.1, 160.6, 148.4, 142.1, 136.52, 136.49, 135.3, 135.2, 134.9, 133.7, 133.6, 129.4, 123.4, 121.6, 121.3, 121.1, 120.0, 119.7, 37.2.

4.6.33. 4-Fluoro-2-(3-nitrobenzyl)benzaldehyde $(9u)^{16}$

¹H NMR (400 MHz, CDCl₃): δ 10.08 (s, 1H), 8.08-8.70 (d, 1H, J = 8.0 Hz), 8.00 (s, 1H), 7.90-7.87 (dd, 1H, J = 8.6 Hz, J = 5.9 Hz), 7.53-7.45 (m, 2H), 7.20-7.15 (td, 1H, J = 8.1 Hz, J = 2.5 Hz), 6.97-6.94 (dd, 1H, J = 9.4 Hz, J = 2.4 Hz), 4.55 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 191.1, 167.0, 164.4, 148.4, 144.34, 144.25, 131.4, 137.5, 137.4, 135.1, 130.40, 130.37, 129.5, 123.6, 121.7, 119.0, 118.8, 114.8, 114.6, 37.8.

4.6.34. Anthracene (10a)²⁹

¹H NMR (500 MHz, CDCl₃): δ 8.44 (s, 2H), 8.00-8.03 (m, 4H), 7.49-7.46 (m, 4H); ¹³C NMR (125 MHz, CDCl₃): δ 131.7, 128.1, 126.2, 125.3.

4.6.35. 2-Methylanthracene (10b)³⁰

¹H NMR (500 MHz, CDCl₃): δ 8.38 (s, 1H), 8.32 (s, 1H), 7.98-8.00 (m, 2H), 7.91-7.93(d, 1H, *J* = 8.7 Hz), 7.76 (s, 1H), 7.43-7.45 (m, 2H), 7.30-7.32 (dd, 2H, *J* = 8.7 Hz, *J* = 1.6 Hz), 2.56 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 134.9, 132.0, 131.8, 131.2, 130.3, 128.24, 128.16, 128.0, 127.9, 126.3, 125.9, 125.2, 125.1, 124.9, 22.0.

4.6.36. 1,3-Dimethylanthracene $(10c)^{30}$

¹H NMR (500 MHz, CDCl₃): δ 8.49 (s, 1H), 8.32 (s, 1H), 8.04-8.02 (m, 1H), 7.99-7.97 (m, 1H), 7.63 (s, 1H), 7.47-7.44 (m, 2H), 7.17 (s, 1H), 2.80 (s, 3H), 2.52 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 134.6, 134.0, 132.3, 131.6, 131.1, 130.0, 128.6, 128.5, 127.8, 125.7, 125.2, 124.84, 124.81, 122.6, 21.9, 19.6.

4.6.37. 1-Methylanthracene (**10d**)²⁹, 2-Methylanthracene (**10b**)²⁹

¹H NMR (500 MHz, CDCl₃): δ 8.55 (s, 1H, 10d), 8.43 (s, 1H, 10d), 8.38 (s, 1H, 10b), 8.32 (s, 1H, 10b), 7.98-8.07 (m, 2H, 10d + 10b), 7.91-7.93(d, 1H, J = 8.7 Hz, 10b), 7.88-7.89 (d, 1H, J = 8.5 Hz, 10d), 7.76 (s, 1H, 10b), 7.42-7.49 (m, 2H, 10d + 10b), 7.36-7.39(dd, 1H, J = 8.5 Hz, J = 6.7 Hz, **10d**) (7.30-7.32 (dd, 1H, J = 8.7 Hz, J = 1.6 Hz, 10b), 2.83 (s, 3H, 10d), 2.56 (s, 3H, 10b); ¹³C NMR (125 MHz, CDCl₃, for 10b): δ 134.9, 132.0, 131.8, 131.2, 130.3, 128.24, 128.16, 128.0, 127.9, 126.3, 125.9, 125.2, 125.1, 124.9, 22.0.

4.6.38. 2-Fluoroanthracene $(10e)^{29}$, 1-Fluoroanthracene $(10f)^{31}$

¹H NMR (400 MHz, CDCl₃): δ 8.69 (s, 1H, **10e**), 8.43 (s, 1H, **10f**), 8.35 (s, 1H, **10f**), 8.07-8.06 (m, 1H, **10e**), 8.02-7.97 (m, 3H, **10f**), 7.80-7.79 (d, 1H, J = 8.6 Hz, **10e** + **10f**), 7.59-7.57 (dd, 1H, J = 10.2Hz, J = 2.3 Hz, **10f**), 7.52-7.45 (m, 2H, **10e** + **10f**), 7.40-7.35 (m, 1H, **10e**), 7.30-7.26 (m, 1H, **10f**), 7.14-7.10 (1H, **10e**); ¹³C NMR (100 MHz, CDCl₃): δ 161.3, 158.9, 132.2, 131.8, 131.7, 131.1, 131.0, 130.9, 129.0, 128.3, 127.7, 126.64, 126.63, 126.0, 125.4, 125.32, 125.25, 117.4, 117.1, 109.8, 109.6.

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6. Appendix A. Supplementary data

¹H and ¹³C spectra for all obtained compounds. This material is available free of charge via the Internet at https://

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