CHEMISTRY
A EUROPEAN JOURNAL

DOI: 10.1002/chem.200901022

Synthesis of 2-C-Branched Oligo(glyco–amino acid)s (OGAAs) by Ring Opening of 1,2-Cyclopropanecarboxylated Sugar Donors

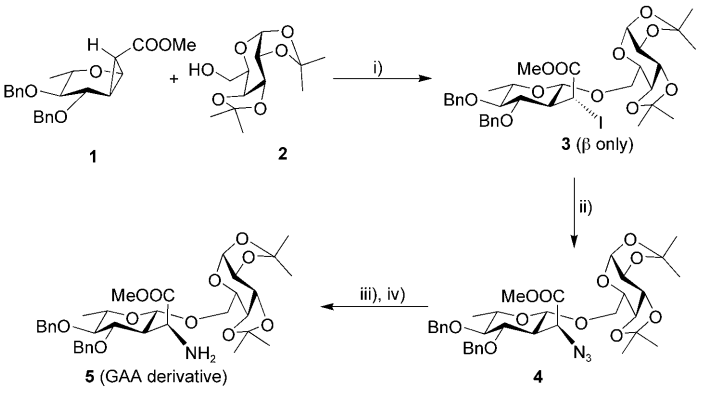
Perali Ramu Sridhar,* Patteti Venu Kumar, Kalapati Seshadri, and Rayala Satyavathi[a]

Dedicated to Professor S. Chandrasekaran

Carbohydrates decorated with amino acids are becoming an important area of glyco-chemistry research.[1] Possessing the architecture of a sugar and an amino acid in a single molecule, these glyco–amino acids (GAAs)[2] are expected to exhibit the characteristics of both carbohydrates and amino acids, which are both biological polymer precursors. C-branched glyco–α-amino acid moieties are found in a variety of nucleoside antibiotics, such as polyoxins,[3] miharamycins,[4] nikkomycin[5] and amipuramycin.[6] Very few methods are available in the literature for the synthesis of monosaccharide-derived C-branched GAA derivatives.[7] Linking an α-amino acid at C-2 or C-4 through a C–C bond has been found to be very difficult. For this reason, the biological importance of these GAAs is not yet fully understood. It has been shown that unnatural 2-C-acetonylsugars serve as metabolic substrates for cell surface engineering by mimicking 2-*N*-acetylsugars.[8] Similarly, a 2-C-*N*-hydroxyacetamide mimic of GlcNAc was synthesised and shown to be an inhibitor of the biosynthesis of lipid A.[9] Herein, we report the first stereoselective synthesis of 2-C-branched oligo(glyco–amino acid)s (OGAAs) by ring opening of 1,2-cyclopropanecarboxylated sugar donors.

The high reactivity and regioselectivity of donor–acceptor cyclopropanes has been well documented in the literature.[10] 1,2-cyclopropanecarboxylated sugars have been used as donor–acceptor cyclopropanes in the synthesis of 2-*C*-branched monosaccharides through electrophilic C1–C7 cyclopropane ring opening or by transition-metal-catalysed glycosylation.[11] Recently, a four-component Pavarov reaction and a transition-metal-mediated radical reaction were developed for the direct synthesis of 2-C-branched carbohydrate derivatives from glucals.[7a,12] These branched glycosides were further derivatised to bicyclic carbohydrate 1,2-lactones.[13] Glucal-derived donor–acceptor cyclopropanes have also been used as 1,3-dipoles under acidic conditions, which result in (3+2) cycloaddition reactions in presence of dipolarophiles.[14] By using the ability of 1,2-cyclopropanecarboxylated sugars to undergo electrophilic ring opening assisted by the adjacent oxygen in presence of an electrophile,[15] we herein present the *N*-iodosuccinimide (NIS)-mediated ring opening of 1,2-cyclopropanecarboxylated glycosyl donors with carbohydrate *O*-nucleophilic glycosyl bond acceptors.

To achieve this novel glycosylation reaction, we began by using 1,5-anhydro-2,6-dideoxy-1,2-C-(*exo*-carbomethoxymethylene)-3,4-di-*O*-benzyl-α-L-rhamnal **1**[7b] as the donor and 1,2-3,4-diisopropylidine-α-D-galactose **2** as the acceptor with NIS as the electrophile at 0 °C in acetonitrile. However, no expected disaccharide was observed under these reaction conditions, even with an excess of acceptor **2** (>3 equiv). Similar reaction conditions with dichloromethane as the



Scheme 1. Synthesis of 2-C-branched GAA disaccharides. i) NIS, TMSOTf, CH_2Cl_2, 0 °C–RT, 74% yield; ii) NaN_3, DMF, 96% yield; iii) Ph_3P, THF; iv) H_2O, reflux, 91% yield.

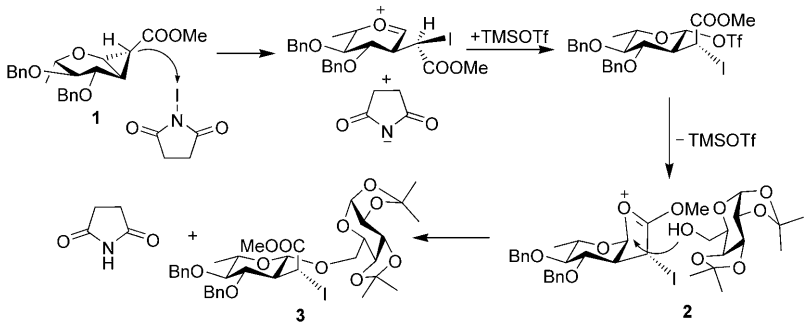
[a] Dr. P. R. Sridhar, P. V. Kumar, K. Seshadri, R. Satyavathi
School of Chemistry
University of Hyderabad
Hyderabad, 500 046 (India)
Fax: (+91) 040-23012460
E-mail: prssc@uohyd.ernet.in

Supporting information for this article is available on the WWW under http://dx.doi.org/10.1002/chem.200901022.

solvent did not improve the glycosylation reaction. We then looked for promoters and after several attempts found that trimethylsilyl trifluoromethanesulfonate (TMSOTf; 15 mol%) is the best promoter for this glycosylation reaction.[16] Treatment of **1** and **2** with NIS/TMSOTf in dichloromethane (0–28°C, 8 h) afforded 2′-C-branched disaccharide **3** in 74% yield as a single diastereomer[17] in which two new stereocenters were introduced at C1′ and C7′ in a single reaction. It is worth noting that only 1.1 equivalents of acceptor, with respect to the donor, were used for this glycosylation reaction. Substitution of α-iodocarboxylate **3** with NaN_3/DMF (28°C, 24 h; DMF = *N*,*N*-dimethylformamide) afforded azidocarboxylate **4** in 96% yield. Reduction of the azide under Staudinger reaction conditions (Ph_3P/THF/H_2O) produced disaccharide GAA derivative **5** in 91% isolated yield (Scheme 1).

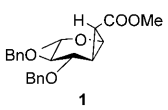
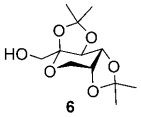
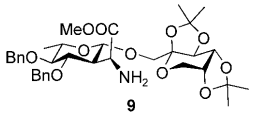
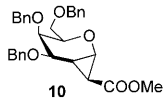
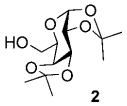
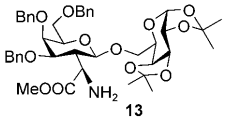
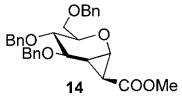
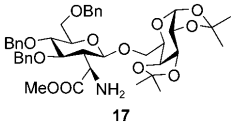
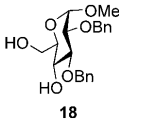
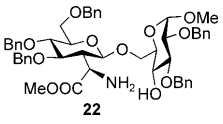
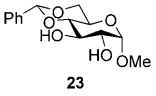
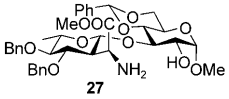
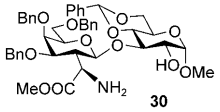
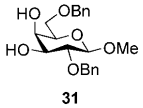
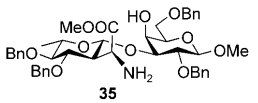
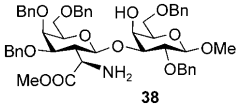
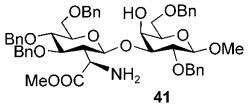
The proposed mechanism of NIS-mediated ring opening involves a stereospecific "edge attack" of iodine on 1,2-cyclopropanecarboxylate **1** to generate an oxocarbenium ion that is immediately trapped with triflate. The triflate is released by neighbouring-group participation of the C7-carboxylate to generate a second oxocarbenium ion intermediate, which is sufficiently long lived that it can be intercepted with a nucleophile. Nucleophilic attack by a glycosyl acceptor oxygen at the anomeric carbon gives disaccharide product **3** (Scheme 2).

The generality of this method has been proved by successfully applying it to a number of 1,2-cyclopropanated glycosyl donors and differentially protected sugar acceptors. Thus, the reaction of cyclopropanecarboxylates **1**, **10** and **14** with acceptors **6** and **2** gave the ring-



Scheme 2. Proposed mechanism for the NIS-mediated ring opening of 1,2-cyclopropanecarboxylated sugar derivatives.

Table 1. Ring opening of 1,2-cyclopropanecarboxylated carbohydrate donors with sugar acceptors; synthesis of 2-C-branched GAA disaccharides.

Entry	Donor cyclopropane	Acceptor	Iodide ([%])	Azide ([%])	GAA derivative	Yield[a] [%]
1	 **1**	 **6**	**7** (75)	**8** (95)	 **9**	92
2	 **10**	 **2**	**11** (72)	**12** (96)	 **13**	90
3	 **14**	**2**	**15** (72)	**16** (92)	 **17**	89
4	**14**	 **18**	**19** (70)	**20** (95) **21**[b] (98)	 **22**	85
5	**1**	 **23**	**24** (67)	**26** (92) **25**[c] (96)	 **27**	94
6	**10**	**23**	**28** (63)	**29** (90)	 **30**	92
7	**1**	 **31**	**32** (70)	**33** (93) **34**[d] (97)	 **35**	97
8	**10**	**31**	**36** (69)	**37** (92)	 **38**	90
9	**14**	**31**	**39** (65)	**40** (88)	 **41**	88

[a] Yield of GAA derivative. Only β-glycosides were formed, and no trace of α product was observed. [b] The free hydroxyl group of azide **20** was acetylated. [c] The free hydroxyl group of iodide **24** was acetylated. [d] The free hydroxyl group of azide **33** was acetylated.

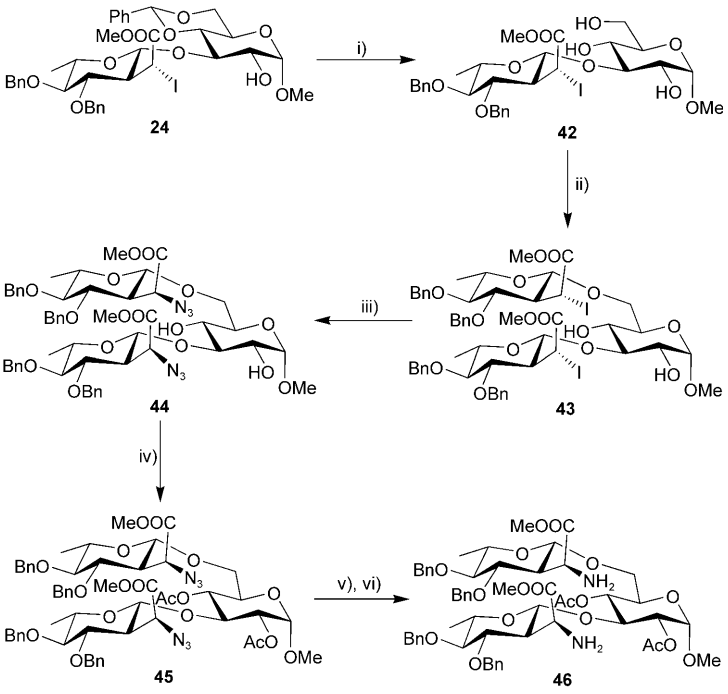
opened 2-C-branched GAA disaccharide derivatives **9**, **13** and **17**, respectively, in good yields with very high diastereoselectivity at the newly formed C1′ and C7′ stereocenters (Table 1, entries 1, 2 and 3). The stereochemistry at C1′ was confirmed by observing a large coupling constant ($J \approx 8.8$ Hz) for the C1′ proton, which indicates a 1,2-trans configuration for all the ring-opened disaccharide derivatives. The stereochemistry at C2′ was defined on the basis of the stereochemistry present in the 1,2-cyclopropanecarboxylated sugar precursor. The stereochemistry at C7′ was assigned based on the proposed mechanism and on one of the GAA derivative crystal structures we reported previously.[7b]

Our next investigations focused on regioselective glycosylation reactions based on the relative reactivity between two hydroxyls on a single sugar acceptor. Towards this goal, cyclopropanecarboxylate **14** was treated with methyl-2,3-di-*O*-benzyl-α-D-glucopyranoside **18** in the presence of NIS/TMSOTf in dichloromethane at 0 °C. The reaction produced a single product, **19**, that was converted to azide **20**. The regioselectivity at the 6-*O* position was assigned by acetylating the free hydroxyl group in **20** with Ac_2O/pyridine and observing a downfield shift in the signal of the C4 proton in acetylated disaccharide **21**. Similarly, reactivity-based glycosylation of 1,2-cyclopropanecarboxylates **1** and **10** with methyl-4,6-*O*-benzylidine-α-D-glucopyranoside **23** produced the 2-C-branched disaccharide derivatives **24** and **28**, respectively, in good yield. Interestingly, C3−OH was involved in the glycosylation step of these reactions.[18]

The aforementioned acceptor-reactivity-based glycosylation of 1,2-cyclopropanecarboxylated sugar donors could also be extended to the other sugar derivatives. Thus, treatment of cyclopropanecarboxylated donors **1**, **10** and **14** with methyl-2,6-di-*O*-benzyl-β-D-galactopyranoside **31** gave disaccharide derivatives **32**, **36** and **39**, respectively, in good yields. All the disaccharide α-iodocarboxylates (**24**, **28**, **32**, **36** and **39**) were converted to the corresponding azides (**26**, **29**, **33**, **37** and **40**, respectively) by using NaN_3/DMF to give excellent yields (≈90%). All these azides were further converted to the corresponding 2-C-branched GAA derivatives **27**, **30**, **35**, **38** and **41**, respectively, under Staudinger reaction conditions (Table 1, entries 5, 6, 7, 8, and 9).

Keeping the above-mentioned acceptor-reactivity-based regio- and stereoselective glycosylation of 1,2-cyclopropanecarboxylated sugar donors in mind, we further planned to synthesise an OGAA derivative. Towards this goal, the benzylidine protecting group in α-iodocarboxylate **24** was deprotected by using *p*-TsOH·H_2O/MeOH to give disaccharide triol **42**. A second acceptor-reactivity-based glycosylation was performed by treating **1** with triol **42** in presence of NIS/TMSOTf to give trisaccharide **43** in good yield as the only isolated product. Treatment of **43** with NaN_3/DMF gave diazide **44**. The free hydroxyls were acetylated to give compound **45**, which gave OGAA derivative **46** under Staudinger reaction conditions (Scheme 3).

In summary, a new glycosylation method that uses carbohydrate-derived donor–acceptor cyclopropanes as glycosyl acceptors has been developed. To the best of our knowledge, this method is the first report of the use of 1,2-cyclopropanecarboxylated sugars in traditional oligosaccharide synthesis. The novel glycosidation method was successfully applied to the synthesis of a number of 2-C-branched GAA disaccharides and to the preparation of an OGAA derivative. Mimicking natural glycosides with carbon-branched GAAs and determining the biological importance of these hybrid biomolecules are in progress.



Scheme 3. Synthesis of 2-C-branched OGAA derivatives. i) *p*-TsOH·H_2O, MeOH, 92% yield; ii) **1**, NIS, TMSOTf, CH_2Cl_2, 0 °C–RT, 62% yield; iii) NaN_3, DMF, 85% yield; iv) Ac_2O, pyridine, 93% yield; v) Ph_3P, THF; vi) H_2O, reflux, 80% yield.

Experimental Section

General procedure for the glycosylation of 1,2-cyclopropanecarboxylated sugar donors: *N*-iodosuccinimide (0.55 mmol) and trimethylsilyl trifluoromethane sulfonate (0.01 mmol) were added to a stirred suspension of 1,2-cyclopropanecarboxylated sugar derivative (0.50 mmol), glycosyl acceptor (0.55 mmol), and a 4-Å molecular sieve in dichloromethane (5 mL) at 0 °C under a nitrogen atmosphere. The temperature was slowly raised to 25 °C and the mixture was stirred for 6 h (until reaction completion, as determined by using TLC). The reaction mixture was diluted with dichloromethane, filtered and washed with aqueous sodium thiosulfate (5%), then the organic layer was dried over anhydrous sodium sulphate and concentrated under vacuum. Column chromatography of the crude product with ethyl acetate/hexane afforded the pure glycosidation product.

Acknowledgements

This work was supported by the Department of Science and Technology (DST) FAST track project grant.

Keywords: carbohydrates · donor–acceptor systems · glyco–amino acids · glycosylation · oligosaccharides

[1] For recent reviews, see: a) F. Schweizer, *Angew. Chem.* **2002**, *114*, 240; *Angew. Chem. Int. Ed.* **2002**, *41*, 230; b) S. A. W. Gruner, E. Locardi, E. Lohof, H. Kessler, *Chem. Rev.* **2002**, *102*, 491; c) A. Dondoni, A. Marra, *Chem. Rev.* **2000**, *100*, 4395. For a compendium of sugar–amino acids, see: M. D. P. Risseeuw, M. Overhand, G. W. J. Fleet, M. I. Simone, *Tetrahedron: Asymmetry* **2007**, *18*, 2001.

[2] A glyco–amino acid (GAA) is a saccharide attached to a single amino acid by any kind of covalent bond, see: J. P. McDevitt, P. T. Lansbury, Jr., *J. Am. Chem. Soc.* **1996**, *118*, 3818.

[3] a) A. Rosenthal, K. Shudo, *J. Org. Chem.* **1972**, *37*, 4391; b) T. Goto, Y. Toya, T. Ohgi, T. Kondo, *Tetrahedron Lett.* **1982**, *23*, 1271; c) S. Haranda, T. Kishi, *J. Antibiot.* **1977**, *30*, 11; d) P. Garner, J. M. Park, *J. Org. Chem.* **1990**, *55*, 3772.

[4] H. Seto, M. Koyama, H. Ogino, T. Tsuruoka, *Tetrahedron Lett.* **1983**, *24*, 1805.

[5] C. S. Stauffer, P. Bhaket, A. W. Fothergill, M. G. Rinaldi, A. Datta, *J. Org. Chem.* **2007**, *72*, 9991.

[6] G. Casiraghi, L. Colombo, G. Rassu, P. Spanu, *J. Org. Chem.* **1991**, *56*, 6523.

[7] a) O. Jiménez, G. de La Rosa, R. Lavilla, *Angew. Chem.* **2005**, *117*, 6679; *Angew. Chem. Int. Ed.* **2005**, *44*, 6521; b) P. R. Sridhar, K. C. Ashalu, S. Chandrasekaran, *Org. Lett.* **2004**, *6*, 1777.

[8] H. C. Hang, C. R. Bertozzi, *J. Am. Chem. Soc.* **2001**, *123*, 1242.

[9] a) J. E. Jackman, C. A. Fierke, L. N. Tumey, M. Pirrung, T. Uchiyama, S. H. Tahir, O. Hindsgaul, C. R. H. Raetz, *J. Biol. Chem.* **2000**, *275*, 11002; b) X. Li, T. Uchiyama, C. R. H. Raetz, O. Hindsgaul, *Org. Lett.* **2003**, *5*, 539.

[10] For a recent review on sugar-derived substrates, see: a) M. Yu, B. L. Pagenkopf, *Tetrahedron* **2005**, *61*, 321; for leading reviews, see: b) H. U. Reissig, R. Zimmer, *Chem. Rev.* **2003**, *103*, 1151; c) H. U. Reissig, *Top. Curr. Chem.* **1988**, *144*, 73; d) E. Wenkert, *Acc. Chem. Res.* **1980**, *13*, 27.

[11] a) J. Beyer, R. Madsen, *J. Am. Chem. Soc.* **1998**, *120*, 12137; b) J. Beyer, P. R. Skaanderup, R. Madsen, *J. Am. Chem. Soc.* **2000**, *122*, 9575.

[12] a) T. Linker, K. Hartmann, T. Sommermann, D. Scheutzow, E. Ruckdeschel, *Angew. Chem.* **1996**, *108*, 1819; *Angew. Chem. Int. Ed. Engl.* **1996**, *35*, 1730; b) T. Linker, T. Sommermann, F. Kahlenberg, *J. Am. Chem. Soc.* **1997**, *119*, 9377.

[13] J. Yin, T. Linker, *Chem. Eur. J.* **2009**, *15*, 49.

[14] a) M. Yu, B. L. Pagenkopf, *J. Am. Chem. Soc.* **2003**, *125*, 8122; b) M. Yu, B. L. Pagenkopf, *Org. Lett.* **2003**, *5*, 5099.

[15] a) C. V. Ramana, R. Murali, M. Nagarajan, *J. Org. Chem.* **1997**, *62*, 7694, and references therein; b) C. Kim, T. Brady, S. H. Kim, E. A. Theodarakis, *Synth. Commun.* **2004**, *34*, 1951.

[16] TMSOTf has been used as an activator in various reactions of donor–acceptor cyclopropanes.

[17] Based on the ^{1}H and ^{13}C NMR spectra.

[18] The site of reactivity was found by acetylating the free hydroxyl group in disaccharide **24** and observing a downfield shift in the signal of the C2 proton in compound **25**. Please see the Supporting Information.

Received: April 17, 2009
Published online: July 8, 2009

