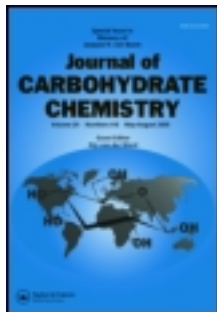


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Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/lcar20>

SN² Displacement of Carbohydrate Triflates by 9-Oximes of Erythromycin A and Of a Tylosin Derivative

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Version of record first published: 22 Aug 2006.

To cite this article: Cyrille Grandjean & Gabor Lukacs (1996): SN² Displacement of Carbohydrate Triflates by 9-Oximes of Erythromycin A and Of a Tylosin Derivative, Journal of Carbohydrate Chemistry, 15:7, 831-855

To link to this article: <http://dx.doi.org/10.1080/07328309608005695>

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S_N2 DISPLACEMENT OF CARBOHYDRATE TRIFLATES BY 9-OXIMES OF ERYTHROMYCIN A AND OF A TYLOSIN DERIVATIVE

Cyrille Grandjean* and Gabor Lukacs

CNRS, Institut de Chimie des Substances Naturelles
Avenue de la Terrasse, 91198 Gif sur Yvette Cedex, France

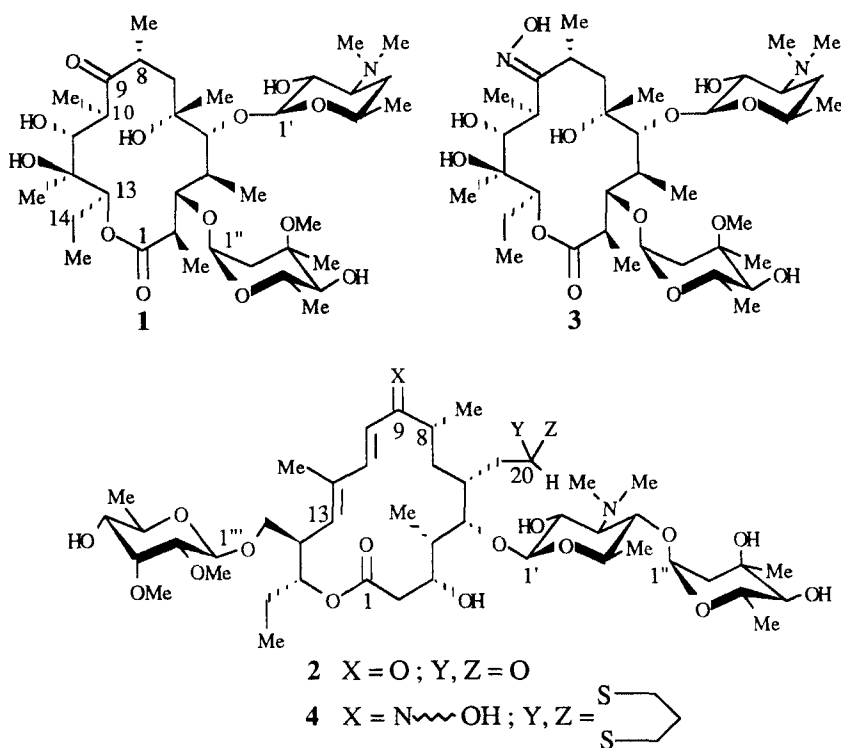
Received February 11, 1996 - Final Form June 17, 1996

ABSTRACT

The preparation of 9-*O*-glycosyloxime derivatives of erythromycin A (**1**) and tylosin (**2**) is reported. Access to this new class of macrolides was achieved from (*E*)-9-oxime of erythromycin A (**3**) and 9-oxime of tylosin 20-(1,3-dithiane) (**4**), by successful displacement of triflates of suitably protected carbohydrates.

INTRODUCTION

Medicinal and veterinary utility of macrolide antibiotics has never been contested since their discovery.¹ However, these antibiotics and their metabolites have been shown to induce hepatic cytochrome P450 and to produce, in some cases, a suicide-type inhibitory complex after *N*-demethylation and oxidation of their *N*-dimethylamino function.² This hepatotoxicity could not simply be avoided by chemical modification of the tertiary amine since this group was recognised as essential for the binding of the antibiotic agent with its ribosomal target in bacteria.³ The ionization state of the tertiary amine (pK_a effect),⁴ the steric hindrance around the dimethylamino group and the hydrophobicity of the molecule⁵ seem to be three factors of significance which affect the interaction of these antibiotics with



Scheme 1

cytochrome P450. Focusing on hydrophobicity, the introduction of additional hydroxyl groups on erythromycin A (**1**) and tylosin (**2**) was thought to contribute to a better tolerance.

These hydroxyl groups could be brought by an additional carbohydrate, linked to the aglycone moiety of these macrolides *via* an oxime at the C-9 position. Choice of an oximic linkage was justified considering the potential antibacterial activity of previously reported 9-*O*-ether oxime derivatives of tylosin⁶ (**2**) and erythromycin A⁷ (**1**). In the latter case, these derivatives were prevented from extensive acidic decomposition at biological pH.⁸ Only a few methods of preparation of *O*-glycosyloxime derivatives are reported in the literature. Pozo and Gotor⁹ have disclosed an enzymatic approach, but their method is limited to a small range of substrates and its application, if possible, would have required full protection of the hydroxy groups of the macrolides to avoid competitive glycosylation. Glycosyloxime derivatives can also be obtained by condensation of *O*-glycosylhydroxylamine derivatives¹⁰ with aldehydes or ketones¹¹ (Figure 1, route A). In view of both the versatility of this

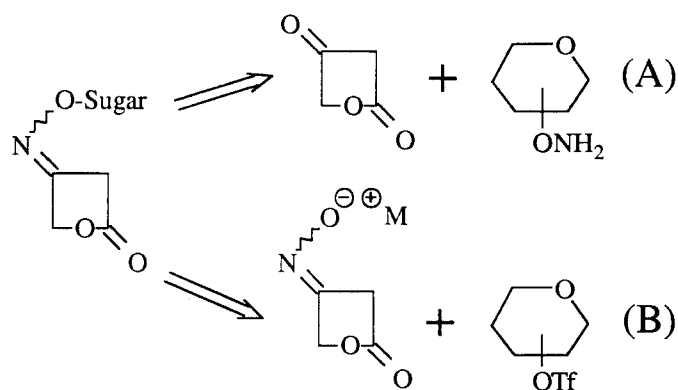


Figure 1. Strategies towards *O*-glycosyloxime derivatives of macrolides.

methodology and the high nucleophilicity of *O*-glycosylhydroxylamines, as demonstrated by Tronchet *et al.*,^{11a} we decided to first explore this route although the ketone function of tylosin (**2**) and especially erythromycin A (**1**) were only poorly reactive. Whatever the experimental conditions applied,¹³ the desired *O*-glycosyloxime derivatives were not detected in the crude reaction mixtures. Thus we alternatively designed a new strategy based on a coupling reaction between carbohydrate bearing triflates and oximate ions of compounds **3** and **4** (Figure 1, route B).

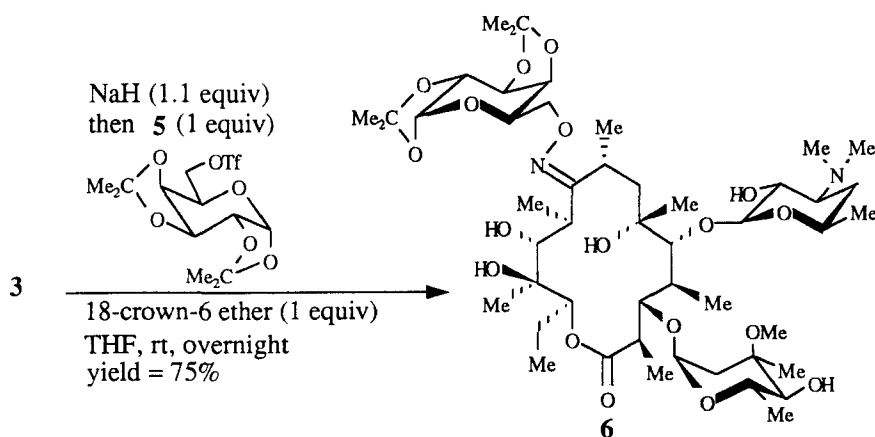
RESULTS AND DISCUSSION

The successful reaction between the (*E*)-9-oxime of erythromycin A⁷ (**3**) and the readily accessible 1,2;3,4-di-*O*-isopropylidene-6-*O*-[(trifluoromethyl)sulfonyl]- α -D-galactopyranoside¹⁴ (**5**) confirmed the feasibility of this approach (Scheme 2).

Thus, the synthesis of various triflated carbohydrates was undertaken (Scheme 3). The choice of the protecting groups was suggested by:

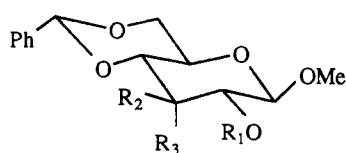
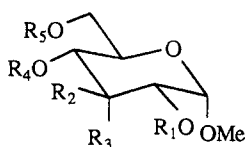
- (a) the putative stability of the triflates,
- (b) the compatibility of their removal with the functionalities of acid and base sensitive macrolides,
- (c) the requirement of a single deprotection step.

Primary triflate **8** was obtained from methyl 2,3,4-tri-*O*-acetyl- α -D-glucopyranoside¹⁵ using Binkley's procedure¹⁶ in 84% yield. Methyl 2,3,6-tri-*O*-benzoyl-4-*O*-[(trifluoromethyl)sulfonyl]- α -D-galactopyranoside (**10**) was obtained from the



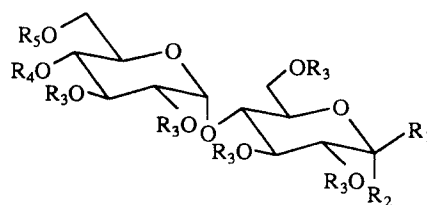
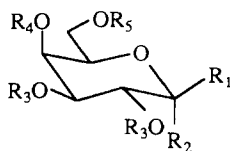
Scheme 2

corresponding alcohol **9** as mentioned in the literature.¹⁷ For the preparation of triflate **14**, we switched from ester to 4-methoxybenzyl ether protective groups to avoid possible intramolecular displacement of the triflate by the 3-*O* ester group.¹⁸ Etherification¹⁹ of diol **11**²⁰ with 4-methoxybenzyl chloride (MPMCl) in DMSO afforded **12** in 80% yield. Regioselective acetal ring opening of this intermediate with sodium cyanoborohydride and trifluoroacetic acid in DMF²⁰ gave methyl 2,3,6-tri-*O*-(4-methoxybenzyl)- α -D-glucopyranoside (**13a**) in 88% together with 5% of its regioisomer **13b**. Triflation of compound **13a** with triflic anhydride and pyridine in CH₂Cl₂ afforded triflate **14** in 89% yield. Benzylation of methyl 4,6-*O*-benzylidene-2-*O*-[(trifluoromethyl)sulfonyl]- α -D-glucopyranoside²¹ (**15**) with benzyltrichloroacetimidate and a catalytic amount of triflic acid²² furnished the known triflate **16** in improved yield compared to the literature.²³ The synthesis of triflates **18**, **20**, **22** and **24** was achieved starting from the corresponding alcohols **17**²⁴, **19**²⁵, **21**²⁶ and **23**²⁷ by treatment with triflic anhydride and pyridine in CH₂Cl₂ in 86%, 81%, 89% and 93% yield, respectively. Nucleophilic substitution of triflate **24** with tetraacetylammonium acetate at room temperature in DMF led to intermediate **25** in 94% yield. Zemplen deprotection of the acetate in quantitative yield followed by triflation with triflic anhydride and pyridine in CH₂Cl₂ furnished the particularly unstable compound **27** in 51% yield. Regioselective reductive ring opening of benzyl 2,3-di-*O*-benzyl-4,6-*O*-benzylidene- β -D-galactopyranoside²⁸ (**28**) with boron-trimethylamine complex and aluminium chloride in THF²⁹ afforded benzyl 2,3,6-tri-*O*-benzyl- β -D-galactopyranoside³⁰ (**29**) as a sole product in 76% yield. Treatment of compound **29** with triflic anhydride and pyridine in CH₂Cl₂ furnished compound **30** in 95% yield.



	R ₁	R ₂	R ₃	R ₄	R ₅
7	Ac	OAc	H	Ac	H
8	Ac	OAc	H	Ac	Tf
11	H	OH	H	4-MeOPh-CH	
12	MPM	OMPM	H	4-MeOPh-CH	
13a	MPM	OMPM	H	OH	OMPM
13b	MPM	OMPM	H	OMPM	OH
14	MPM	OMPM	H	OTf	OMPM
15	Tf	OH	H	Ph-CH	
16	Tf	OBn	H	Ph-CH	
17	Bn	OH	H	Ph-CH	
18	Bn	OTf	H	Ph-CH	
19	Bn	H	OH	Ph-CH	
20	Bn	H	OTf	Ph-CH	

	R ₁	R ₂	R ₃
21	H	OBn	H
22	Tf	OBn	H
23	Bn	OH	H
24	Bn	OTf	H
25	Bn	H	OAc
26	Bn	H	OH
27	Bn	H	OTf



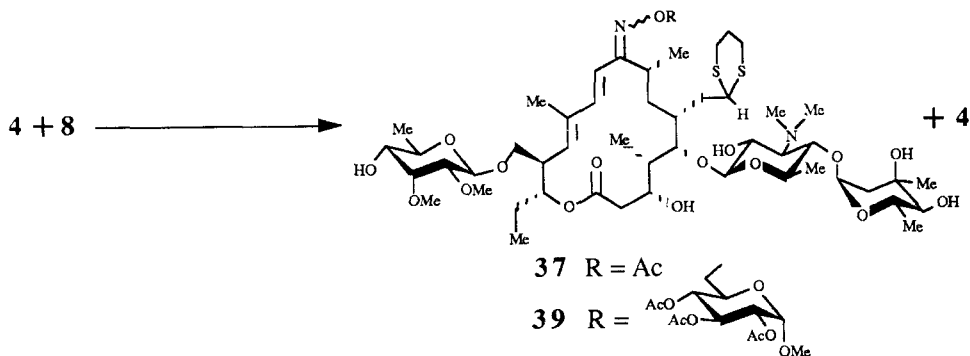
	R ₁	R ₂	R ₃	R ₄	R ₅
9	H	OMe	Bz	H	Bz
10	H	OMe	Bz	Tf	Bz
28	OBn	H	Bn	Ph-CH	
29	OBn	H	Bn	H	Bn
30	OBn	H	Bn	Tf	Bn

	R ₁	R ₂	R ₃	R ₄	R ₅
31	H,OH	OH,H	H	H	H
33	OMe	H	Ac	Ac	Ac
34	OMe	H	Bn	Ph-CH	
35	OMe	H	Bn	H	Bn
36	OMe	H	Bn	Tf	Bn

(MPM = 4-Methoxyphenylmethyl)

Scheme 3. Preparation of carbohydrate triflates.

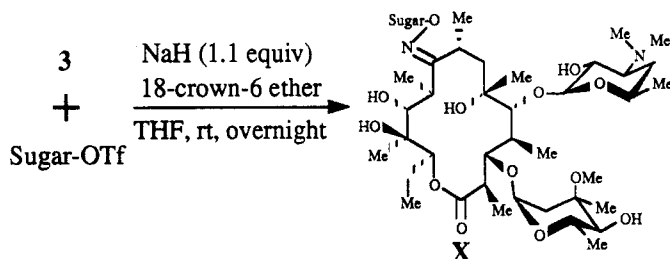
Treatment of maltose monohydrate (**31**) with acetic anhydride and a catalytic amount of perchloric acid in glacial acetic acid followed by addition of 30% hydrobromic acid in glacial acetic acid afforded 2,3,6,2',3',4',6'-hepta-*O*-acetyl- α -D-maltosyl bromide (**32**), which was further transformed to the corresponding methyl- β -D-glycoside **33** via a Koenigs-Knorr reaction in the presence of mercuric (II) cyanide in MeOH in 61% overall yield. Deesterification of **33** was carried out using the Zemplen procedure; the crude

Table 1. Optimization of the coupling reaction conditions.

Entry	Experimental conditions	Yields (%)		
		37	39	4
1	THF, rt triflate added to anion	14	25	50
2	THF, rt anion added to triflate	traces	44	37
3	THF, rt 18-crown-6 ether anion added to triflate	traces	53	25
4	THF/DMPU 5:2, rt anion added to triflate	traces	48	40

product was treated with dimethoxytoluene in the presence of a catalytic amount of *p*-toluenesulfonic acid in DMF.³² After removal of the excess reagent and solvent, the residue was treated with sodium hydride and benzyl bromide in DMF to give **34** in 68% overall yield. Regioselective reductive acetal ring opening of compound **34** with boron-trimethylamine complex and aluminium chloride in THF²⁹ afforded alcohol **35** as a sole product in 70% yield. Treatment of **35** with triflic anhydride and pyridine in CH₂Cl₂ furnished triflate **36** in 84% yield.

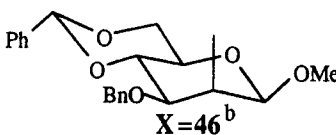
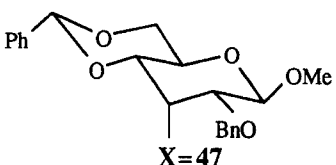
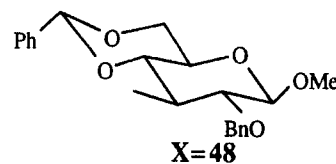
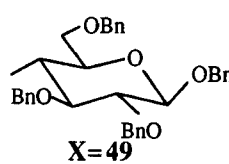
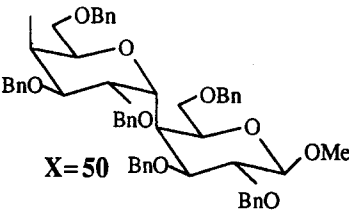
Having prepared these triflates, their nucleophilic substitution with compounds **3** and **4** was undertaken. Direct addition of triflate **8** in solution in THF into the oximate ion of compound **4** in THF at room temperature led mainly to the formation of undesired products [e.g., **37** or methyl 2,4-di-*O*-acetyl-3,6-anhydro- α -D-glucopyranoside (**38**)] together with recovered starting material rather than to compound **39** (Table 1, entry 1).

Table 2. *O*-Glycosyloxime derivatives of erythromycin A.

Entry	Sugar-OTf	Temperature	Sugar Compound	Yield (%)
1	8	rt	<p style="text-align: center;">X=42</p>	61
2	10	rt	<p style="text-align: center;">X=43</p>	65
3	14	rt	<p style="text-align: center;">X=44</p>	82
4	16	reflux	no reaction	-
5	18^a	rt	no reaction	-
6	18	reflux	<p style="text-align: center;">45 elimination</p>	-

(continued)

Table 2. (continued)

Entry	Sugar-OTf	Temperature	Sugar Compound	Yield (%)
7	20 ^a	rt	elimination 45	
8	22 ^a	reflux	 X=46^b	60
9	24 ^a	rt	 X=47	70
10	27	0°C	 X=48	47
11	30	rt	 X=49	90
12	36	rt	 X=50	65

a. See note 33.

b. Obtained as a *Z* and *E* mixture of oximes.

The importance of these side reactions was, however, controlled using an inverse procedure (i.e., dropwise addition of the oximate ion into the triflate in solution in THF) (Table 1, entry 2) and almost avoided when 18-crown-6 ether or a cosolvent was used (Table 1, entries 3 and 4). Reproducing the above optimum conditions, 9-[*O*-(methyl 2,3,6-tri-*O*-benzoyl- α -D-glucopyranosid-4-yl)oxime] of tylosin 20-(1,3-dithiane) (**40**) and 9-{*O*-[methyl 2,3,6-tri-*O*-(4-methoxybenzyl)- α -D-glucopyranosid-4-yl]oxime} of tylosin 20-(1,3-dithiane) (**41**) were obtained in 63 and 72% yield starting from triflates **10** and **14**, respectively. These substantially improved yields may reflect the greater stability of triflates **10** and **14** towards the applied basic conditions.

In the same manner, *O*-glycosyloxime derivatives of erythromycin A (**1**) were prepared using compound **3** as nucleophile (Table 2). Results were strongly dependent on the nature of the triflates and particularly on their anomeric configuration. Methyl 2-*O*- or 3-*O*-[(trifluoromethyl)sulfonyl]- α -D-glycopyranoside derivatives did not react or led to elimination products. Triflate **16** (Table 2, entry 4) was recovered unchanged whereas the isomer **22** led to a mixture of oximes (*Z* and *E*) **46**,³⁴ when the reaction was run at reflux, in 70% yield (Table 2, entry 8). The difference of reactivity of triflates **16** and **22** is surprising and may be explained by more unfavorable dipolar interactions at the transition state for the former compound.³⁵ Triflate **24** and its unstable stereomer **27** smoothly afforded the corresponding *O*-glycosyloxime derivatives **47** and **48** at room temperature or 0 °C, respectively (Table 2, entries 9 and 10). In contrast, the lack of reactivity of triflate **18** (Table 2, entry 5) may be attributed to 1,3-dipolar interactions with the anomeric methoxy group. Under forcing conditions (Table 2, entry 6), this substrate reacted preferentially to give methyl 2-*O*-benzyl-4,6-*O*-benzylidene-3-deoxy- α -D-*erythro*-hex-2-enopyranoside (**45**). Starting from isomer **20**, the same unsaturated carbohydrate was obtained from room temperature (Table 2, entry 7). The formation of this elimination product was in agreement with recent observations.^{33b} Reasonable yields of *O*-glycosyloxime derivatives were obtained starting from primary triflates (Table 2, entry 1 and Scheme 2) or from axial (Table 2, entries 2 and 11) or equatorial (Table 2, entries 3 and 12) 4-*O*-triflated derivatives regardless of the anomeric configuration.

CONCLUSION

In this paper, the first examples of substitution of carbohydrate triflates by oximate ions have been described. Given the poor nucleophilicity compared to the basicity of these ions, the issue of the coupling reaction is strongly dependent on the nature of the triflate.

Application of this reaction allowed the entry to a new class of macrolides, the 9-*O*-glycosyloximes of erythromycin A and tylosin. The antibacterial activity of the described compounds after liberation of all the hydroxyl groups from the respective esters, ethers and acetals will be reported upon permission of our industrial partner.

EXPERIMENTAL

General methods. Meltings points were determined with a Reichert-Jung apparatus and are not corrected. Optical rotations were measured with a Perkin-Elmer Model 241 polarimeter. Thin-layer chromatography (TLC) was performed using E. Merck plates of silica gel 60 with fluorescent indicator. Visualization was effected by spraying plates with 5% H₂SO₄ in ethanol followed by heating at 120-140 °C. Flash chromatography was conducted with silica gel (0.040-0.063 mm, E. Merck). THF was distilled over sodium/benzophenone, DMF, DMSO and pyridine were distilled over CaH₂. CH₂Cl₂ was distilled over P₂O₅ and MeOH over magnesium. Nuclear magnetic resonance (¹H and ¹³C NMR) spectra were recorded in CDCl₃ on Bruker WP 200, Bruker WP 300 and Bruker WP 300 spectrometers. Chemical shifts are recorded in ppm (δ) relative to tetramethylsilane as internal standard. For clarity, chemical shifts of aromatic hydrogens and carbons have been omitted. CIMS and FABMS spectra were obtained on AEI-SM-9 and SM-80 spectrometers, respectively. High resolution mass spectra (HRMS) were run on a VG-ZAB-SEQ spectrometer by the Service Central d'Analyse, Vernaison. Elemental analyses were performed by the microanalytical laboratory at the ICSN, Gif sur Yvette. In NMR data, "" and "" for compounds **6**, **42**, **43**, **44**, **46**, **47**, **48**, **49** and **39**, **40**, **41**, respectively, refer to the newly introduced carbohydrate moiety. "" and "" for compound **50** refer to the galactose and the glucose moiety of the disaccharide, respectively.

General procedure A for trifluoromethanesulfonylation of carbohydrates (13a, 17, 19, 21, 23, 26, 29, 35). A mixture of the carbohydrate substrate (1 equiv) and pyridine (2 equiv) in CH₂Cl₂ (3 mL/mmol of carbohydrate) was treated with trifluoromethanesulfonic anhydride (1.15 to 1.8 equiv) at -20 °C under argon. The reaction was stirred for 2 hours, then warmed up to room temperature. After completion (monitored by TLC), the reaction was quenched with water. The mixture was then extracted with CH₂Cl₂. The organic layer was dried over Na₂SO₄, filtered and concentrated under reduced pressure to furnish a residue which was further purified by flash chromatography on silica gel.

General procedure B for the coupling reactions between trifluoromethanesulfonates (8, 10, 14, 24, 30, 36) and oximes (3) and (4).

A mixture of oxime (1 equiv) and 18-crown-6 ether (0.3 to 1 equiv) in THF (5 mL/mmol of oxime) was treated with sodium hydride (1.1 equiv) [sodium hydride (60% dispersion in mineral oil) was washed with heptane] at room temperature, under argon. After stirring during 20 minutes, the mixture was slowly transferred into a solution of triflate (1 equiv) in THF (5 mL/mmol of oxime). The reaction was stirred at room temperature overnight then quenched by adding Florisil® and concentrated under reduced pressure. The residue was purified by flash chromatography on silica gel.

(E) -9-[O -(1,2;3,4-di- O -isopropylidene- α -D-galactopyranosid-6-yl) oxime] of erythromycin A (6). To a mixture of (E)-9-oxime of erythromycin A **3** (618 mg, 0.83 mmol) and 18-crown-6 ether (304 mg, 0.83 mmol) in THF (4 mL) was added, under argon, sodium hydride (20 mg) [sodium hydride (60% dispersion in mineral oil, 33 mg) was washed with heptane]. The mixture was then stirred for 20 min at room temperature. A solution of triflate **5** (426 mg, 1.08 mmol) in THF (5 mL) was then added dropwise *via* a syringe into the solution. The reaction mixture was stirred overnight. The solvent was removed under reduced pressure and the residue purified by flash chromatography (eluent dichloromethane/methanol/concd ammonia 10:1:0.05 v/v/v) to afford compound **6** (625 mg, 75%) as a foam: R_f 0.46 (dichloromethane/methanol/concd ammonia 15:1:0.05 v/v/v); [α]_D -58 (c 0.8, chloroform); ¹H NMR (200 MHz) δ 1.00 (d, 3H, J_{8,8Me} = 7 Hz, 8-CH₃), 1.30, 1.35, 1.43 and 1.50 (4s, 12H, 2 C(CH₃)₂), 4.33 (dd, 1H, J_{1'',2''} = 5.1 Hz, J_{2'',3''} = 2.5 Hz, H-2''), 4.42 (d, 1H, J_{1',2'} = 7.2 Hz, H-1'), 4.62 (dd, 1H, J_{3'',4''} = 7.6 Hz, H-3''), 4.86 (d, 1H, J_{1'',2''} = 5 Hz, H-1''), 4.98 (dd, 1H, J_{13,14ax} = 2 Hz, J_{13,14eq} = 11 Hz, H-13), 5.55 (d, 1H, H-1''); ¹³C NMR (50 MHz) δ 15.1 (10-OCH₃), 18.6 (8-CH₃), 24.5, 24.6, 26.0 and 26.3 (4 CH₃), 25.8 (C-8), 33.0 (C-10), 65.9 (C-5''), 72.8 (C-6''), 71.0, 71.1 and 71.2 (C-2'', C-3'' and C-4''), 96.5 (C-1''), 108.4 and 109.4 (2 C(Me)₂), 171.0 (C-9); FABMS (positive) *m/z* 1013 (M+Na)⁺, 991 (M+H)⁺, 833 (M+H-(Cladinose-H))⁺. HRMS Calcd for C₄₉H₈₇N₂O₁₈ (991.5954). Found: 991.5957.

Methyl 2,3,4-tri-O-acetyl-6-O-[(trifluoromethyl)sulfonyl]- α -D-glucopyranoside (8). A solution of methyl 2,3,4-tri-O-acetyl- α -D-glucopyranoside¹⁵ **7** (1.51 g, 4.71 mmol) was converted to the triflate **8** following a literature procedure.¹⁶ Compound **8** (1.84 g, 84%) was obtained after flash chromatography (eluent ether/pentane 3:2 v/v) as an unstable syrup; R_f 0.43 (ethyl acetate/heptane 1:1 v/v); ¹H NMR (200 MHz) δ 2.02, 2.07 and 2.09 (3s, 9H, 3 CH₃), 3.45 (s, 3H, OCH₃), 4.12 (ddd, 1H, J_{4,5} = 3 Hz, J_{5,6a} = 9.8 Hz, J_{5,6b} = 5.5 Hz, H-5), 4.50-4.58 (m, 2H, H-6a and H-6b), 4.88 (dd, 1H, J_{1,2} = 3.9 Hz, J_{2,3} = 10.1 Hz, H-2), 4.98 (d, 1H, H-1), 5.00 (dd, 1H, J_{3,4} = 10.1 Hz, H-4), 5.50 (t, 1H, H-3); ¹³C NMR (50 MHz) δ 20.2, 20.5 and 20.5 (3 CH₃), 55.7 (OCH₃),

67.0 (C-5), 68.6 (C-4), 69.7 and 70.5 (C-2 and C-3), 73.9 (C-6), 96.8 (C-1), 118.6 (q, $J = 320.4$ Hz, CF_3), 169.6 and 170.0 (3 OCOCH_3); FABMS (positive) m/z 475 ($\text{M}+\text{Na}$)⁺.

Methyl 2,3,6-tri-*O*-benzoyl-4-*O*-(trifluoromethyl sulfonyl)- α -D-galactopyranoside (10). Methyl 2,3,6-tri-*O*-benzoyl- α -D-galactopyranoside¹⁷ **9** (1.52 g, 3 mmol) was reacted with trifluoromethanesulfonic anhydride (0.76 mL, 4.5 mmol) in the presence of pyridine (0.47 mL, 6 mmol) following general procedure A. Compound **10** (1.79 g, 84%) was obtained after flash chromatography (eluent ethyl acetate/heptane 3:7 v/v) followed by crystallization from EtOH: Rf 0.53 (ethyl acetate/heptane 1:1 v/v); mp 136 °C (dec) (lit.¹⁷ mp 137-138 °C); $[\alpha]_{\text{D}}^{+101}$ (c 1, chloroform) [lit.¹⁷ $[\alpha]_{\text{D}}^{+103.6}$ (c 1, chloroform)]; ¹H NMR (200 MHz): identical to the literature; ¹³C NMR (50 MHz) δ 56.2 (OCH_3), 61.6 (C-6), 66.0, 67.7 and 68.4 (C-2, C-3 and C-5), 83.1 (C-4), 97.7 (C-1), 118.5 (q, $J = 321$ Hz, CF_3), 165.9 (3 OCOCH_3); FABMS (positive) m/z 661 ($\text{M}+\text{Na}$)⁺.

Anal. Calcd for $\text{C}_{29}\text{H}_{25}\text{F}_3\text{O}_{11}\text{S}$ (638.58): C, 54.54; H, 3.95; S, 5.02. Found: C, 54.69; H, 4.02; S, 4.80.

Methyl 2,3-di-*O*-(4-methoxybenzyl)-4,6-*O*-(4-methoxybenzylidene)- α -D-glucopyranoside (12). Methyl 4,6-*O*-(4-methoxybenzylidene)- α -D-glucopyranoside²⁰ **11** (2.6 g, 8.32 mmol) was dissolved in DMSO (40 mL) and added dropwise, under argon, to a suspension of sodium hydride (519 mg, 21.63 mmol) [sodium hydride (60% dispersion in mineral oil, 865 mg) was washed with heptane] in DMSO (35 mL) and stirred at room temperature for 30 min. 4-Methoxybenzyl chloride (3.38 mL, 24.96 mmol) was then added dropwise and the reaction mixture was further stirred overnight at room temperature. Crude **12** was precipitated by addition of a few drops of cold water. The solid was filtered and washed with water and heptane. Finally, crystallization from EtOH furnished compound **12** as a white solid (3.7 g, 80%): Rf 0.38 (ethyl acetate/heptane 1:1 v/v); mp 123-125 °C; $[\alpha]_{\text{D}}^{+46}$ (c 1, chloroform); ¹H NMR (200 MHz) δ 3.40 (s, 3H, 1- OCH_3), 3.81 and 3.82 (2s, 3 and 6H, 3 OCH_3), 3.99 (dd, 1H, $J_{2,3} = 8.4$ Hz, $J_{3,4} = 8.3$ Hz, H-3), 4.22 (dd, 1H, $J_{1,2} = 3.6$ Hz, H-2), 4.52 (d, 1H, H-1), 4.58-4.81 (m, 4H, 2 CH_2Ar), 5.42 (s, 1H, H-7); ¹³C NMR (50 MHz) δ 55.4 (4 OCH_3), 62.5 (C-5), 69.2 (C-6), 73.5 and 75.0 (2 CH_2Ar), 78.4, 79.0 and 82.3 (C-2, C-3 and C-4), 99.5 (C-1), 101.4 (C-7); CIMS m/z 553 ($\text{M}+\text{H}$)⁺.

Anal. Calcd for $\text{C}_{31}\text{H}_{36}\text{O}_9$ (503.38): C, 67.38; H, 6.57. Found: C, 67.43; H, 6.65.

Methyl 2,3,6-tri-*O*-(4-methoxybenzyl)- α -D-glucopyranoside (13a). A solution of trifluoroacetic acid (2.62 mL, 25.5 mmol) in DMF, precooled at 0 °C, was added dropwise to a stirred mixture containing compound **12** (933 mg, 1.7 mmol), sodium cyanoborohydride (1.06 g, 17 mmol) and 3 Å molecular sieves in DMF (12 mL). The reaction mixture was kept at room temperature during 24 hours and then, filtered through Celite®. The filtrate was poured into ice-cold saturated aqueous sodium hydrogen

carbonate. The aqueous layer was extracted three times with CH₂Cl₂. The combined extracts were washed with brine, dried (Na₂SO₄), filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (eluent ethyl acetate/heptane 4:6 v/v) to furnish compound **13a** (830 mg, 88%) as a colorless syrup and its regioisomer **13b** (46 mg, 5%), which was crystallized from EtOH.

Description of compound **13a**: R_f 0.43 (ethyl acetate/heptane 1:1 v/v); [α]_D +2 (c 1, chloroform); ¹H NMR (200 MHz) δ 2.2 (br s, 1H, 4-OH), 3.32 (s, 3H, 1-OCH₃), 3.42 (dd, 1H, J_{5,6a} = 3.4 Hz, J_{6a,6b} = 9.4 Hz, H-6a), 3.49 (br d, 1H, H-6b), 3.75 (s, 9H, 3 OCH₃), 4.42 (d, 1H, J_{1,2} = 3.6 Hz, H-1), 4.48-4.63 (m, 5H, CH₂Ar), 4.84 (d, 1H, J = 11.1 Hz, CHHAr); ¹³C NMR (50 MHz) δ 54.5 (4 OCH₃), 68.8 (C-6), 69.9 and 70.2 (C-4 and C-5), 72.0, 72.5 and 74.3 (3 CH₂Ar), 78.9 and 80.7 (C-2 and C-3), 97.8 (C-1); FABMS (positive) *m/z* 561 (M+Li)⁺.

Anal. Calcd for C₃₁H₃₈O₉ (554.64): C, 67.13; H, 6.91. Found: C, 67.06; H, 6.95.

Description of regioisomer **13b**: R_f 0.15 (ethyl acetate/heptane 1:1 v/v); mp 79-80 °C; [α]_D +1 (c 1, chloroform); ¹H NMR (200 MHz) δ 1.77 (br s, 1H, 6-OH), 3.35 (s, 3H, 1-OCH₃), 3.78 and 3.82 (2s, 3 and 6H, 3 OCH₃), 3.96 (t, 1H, J_{2,3} = J_{3,4} = 9 Hz, H-3), 4.50 (d, 1H, J_{1,2} = 3.4 Hz, H-1), 4.52-4.96 (m, 6H, 3 CH₂Ar); ¹³C NMR (50 MHz) δ 55.2 and 55.3 (4 OCH₃), 62.0 (C-6), 70.7 (C-5), 73.1, 74.7 and 75.4 (3 CH₂Ar), 77.3 (C-4), 79.8 and 81.8 (C-2 and C-3), 98.3 (C-1); CIMS *m/z* 555 (M+H)⁺.

Anal. Calcd for C₃₁H₃₈O₉ (554.64): C, 67.13; H, 6.91. Found: C, 67.25; H, 6.95.

Methyl 2,3,6-tri-O-(4-methoxybenzyl)-4-O-[(trifluoromethyl)sulfonyl]- α -D-glucopyranoside (14). Compound **13a** (815 mg, 1.47 mmol) was reacted with trifluoromethanesulfonic anhydride (0.45 mL, 2.65 mmol) in the presence of pyridine (0.41 mL, 2.94 mmol) following general procedure A. Compound **14** (898 mg, 89%) was obtained after flash chromatography (eluent diethylether/pentane 3:2 v/v) as an unstable syrup: R_f 0.54 (ethyl acetate/heptane 1:1 v/v); [α]_D +31 (c 0.43, chloroform); ¹H NMR (200 MHz) δ 3.30 (s, 3H, 1-OCH₃), 3.78 (s, 9H, 3 OCH₃), 4.31-4.52 (m, 4H, H-1, H-3, H-4 and CHHAr), 4.52-4.92 (m, 5H, CH₂Ar); ¹³C NMR (50 MHz) δ 55.0 (3 OCH₃), 55.5 (1-OCH₃), 67.2 (C-6), 67.6 (C-5), 73.1, 73.1 and 75.0 (3 CH₂Ar), 77.5 (C-3), 79.8 and 81.6 (C-2 and C-4), 97.8 (C-1), 118.5 (q, J = 320.4 Hz, CF₃); FAB-MS *m/z* 709 (M+Na)⁺.

Methyl 3-O-benzyl-4,6-O-benzylidene-2-O-[(trifluoromethyl)sulfonyl]- α -D-glucopyranoside (16). To a stirred solution of methyl 4,6-O-benzylidene-2-O-[(trifluoromethyl)sulfonyl]- α -D-glucopyranoside²¹ **15** (1.75 g, 4.23 mmol) and benzyl-2,2,2-trichloroacetimidate (1.18 mL, 6.34 mmol) in a mixture of cyclohexane/CH₂Cl₂ 2:1 (v/v) (42 mL) was added, a catalytic amount of trifluoromethanesulfonic acid (211 μ L) at room temperature, under argon. The mixture was stirred until the starting material had

completely reacted as monitored by TLC. The crystalline trichloroacetamidate was removed by filtration through Celite® and the filtrate washed with aqueous saturated sodium hydrogen carbonate and water. The organic layer was dried (Na₂SO₄), filtered and concentrated under reduced pressure. Compound **16** (1.28 g, 60%) was obtained after flash chromatography (eluent ethyl acetate/heptane 2:8 v/v) followed by crystallization from Et₂O/heptane as a white solid: R_f 0.36 (ethyl acetate/heptane 2:8 v/v); mp 93–94°C (dec); [α]_D +27 (c 1, chloroform); ¹H NMR (200 MHz) δ 3.48 (s, 3H, 1-OCH₃), 3.70 (t, 1H, J_{3,4} = J_{4,5} = 9.3 Hz, H-4), 3.73 (dd, 1H, J_{5,6ax} = 12 Hz, J_{6ax,6eq} = 9.7 Hz, H-6ax), 3.88 (ddd, 1H, J_{5,6eq} = 4.1 Hz, H-5), 4.13 (t, 1H, J_{2,3} = 9.3 Hz, H-3), 4.32 (dd, 1H, H-6eq), 4.75 (dd, 1H, J_{1,2} = 3.6 Hz, H-2), 4.76 (d, 1H, J = 10.9 Hz, CHHPh), 4.87 (d, 1H, CHHPh), 4.98 (d, 1H, H-1), 5.56 (s, 1H, H-7); ¹³C NMR (50 MHz) δ 55.7 (1-OCH₃), 62.3 (C-5), 68.7 (C-6), 75.1 (CH₂Ph), 75.2 (C-3), 82.0 (C-4), 83.7 (C-2), 97.7 (C-1), 101.6 (C-7), 118.6 (q, J = 320.4 Hz, CF₃); CISM *m/z* 505 (M+H)⁺.

Methyl 2-*O*-benzyl-4,6-*O*-benzylidene-3-*O*-[(trifluoromethyl)sulfonyl]-α-D-glucopyranoside (18). Methyl 2-*O*-benzyl-4,6-*O*-benzylidene-α-D-glucopyranoside²⁴ **17** (1.12 g, 3 mmol) was reacted with trifluoromethanesulfonic anhydride (0.757 mL, 4.5 mmol) in the presence of pyridine (0.485 mL, 6 mmol) following general procedure A. Compound **18** (1.30 g, 86%) was obtained after flash chromatography (eluent diethylether/heptane 4:6 v/v) followed by crystallization from Et₂O-heptane: R_f 0.18 (diethylether/heptane 1:1 v/v); mp 91–92 °C (dec); [α]_D +9 (c 1, chloroform); ¹H NMR (200 MHz) δ 3.38 (s, 3H, OCH₃), 4.26 (dd, 1H, J_{5,6eq} = 4.1 Hz, J_{6ax,6eq} = 9.6 Hz, H-6eq), 4.50 (d, 1H, J_{1,2} = 3.5 Hz, H-1), 4.53 (d, 1H, J = 12.2 Hz, CHHPh), 4.80 (d, 1H, CHHPh), 5.17 (t, 1H, J_{2,3} = J_{3,4} = 9.3 Hz, H-3), 5.55 (s, 1H, H-7); ¹³C NMR (50 MHz) δ 55.7 (OCH₃), 62.6 (C-5), 68.8 (C-6), 73.8 (CH₂Ph), 77.3 and 78.7 (C-2 and C-4), 84.7 (C-3), 99.0 (C-1), 101.6 (C-7), 118.6 (q, J = 320.5 Hz, CF₃); CIMS *m/z* 505 (M+H)⁺.

Anal. Calcd for C₂₂H₂₃F₃O₈S (504.48): C, 52.38; H, 4.59; S, 6.36. Found: C, 52.19; H, 4.58; S, 6.41.

Methyl 2-*O*-benzyl-4,6-*O*-benzylidene-3-*O*-[(trifluoromethyl)sulfonyl]-α-D-allopyranoside (20). Methyl 2-*O*-benzyl-4,6-*O*-benzylidene-α-D-allopyranoside²⁵ **19** (40 mg, 0.11 mmol) was reacted with trifluoromethanesulfonic anhydride (27.12 μL, 0.17 mmol) in the presence of pyridine (86.08 μL, 0.22 mmol) following general procedure A. Compound **20** (44 mg, 82%) was obtained after flash chromatography (eluent diethylether/pentane 1:2 v/v) as a syrup: R_f 0.45 (ethyl acetate/heptane 1:1 v/v); [α]_D -5 (c 1, chloroform); ¹H NMR (200 MHz) δ 3.46 (s, 3H, OCH₃), 3.60 (t, 1H, J_{1,2} = J_{2,3} = 4 Hz, H-2), 3.63 (d, 1H, J_{4,5} = 10 Hz, H-4), 3.69 (t, 1H, J_{5,6ax} = J_{6ax,6eq} = 10 Hz, H-6ax), 4.20 (dt, 1H, J_{5,6eq} = 5.1 Hz, 5-H), 4.35 (dd, 1H,

H-6eq), 4.62 (d, 1H, *J* = 12.7 Hz, *CHHPh*), 4.72 (d, 1H, H-1), 4.84 (d, 1H, *CHHPh*), 5.45 (br s, 1H, H-3), 5.54 (s, 1H, H-7); ¹³C NMR (50 MHz) δ 56.3 (OCH₃), 58.1 (C-5), 69.2 (C-6), 71.6 (CH₂Ph), 72.2 (C-2), 75.5 (C-4), 80.7 (C-3), 98.5 (C-1), 102.4 (C-7), 118.6 (q, *J* = 320.5 Hz, CF₃); FABMS (positive) *m/z* 505 (M+H)⁺.

Methyl 3-*O*-benzyl-4,6-*O*-benzylidene-2-*O*-[(trifluoromethyl)sulfonyl]-β-D-glucopyranoside (22). Methyl 3-*O*-benzyl-4,6-*O*-benzylidene-β-D-glucopyranoside²⁶ **21** (405 mg, 1.09 mmol) was reacted with trifluoromethanesulfonic anhydride (0.275 mL, 1.64 mmol) in the presence of pyridine (0.176 mL, 2.18 mmol) following general procedure A. Compound **22** (490 mg, 89%) was obtained as a white solid after flash chromatography (eluent ethyl acetate/pentane 3:7 v/v) followed by crystallization from a mixture of Et₂O-heptane: R_f 0.42 (ethyl acetate/heptane 3:7 v/v); mp 104 °C (dec); [α]_D -51 (*c* 1, chloroform); ¹H NMR (200 MHz) δ 3.58 (s, 3H, OCH₃), 3.77 (t, 1H, *J*_{5,6ax} = *J*_{6ax,6eq} = 10.3 Hz, H-6ax), 3.88 (dd, 1H, *J*_{2,3} = 7.8 Hz, *J*_{3,4} = 8.4 Hz, H-3), 4.41 (dd, 1H, *J*_{5,6eq} = 4 Hz, H-6eq), 4.52 (d, 1H, *J*_{1,2} = 7.9 Hz, H-1), 4.61 (dd, 1H, H-2), 4.80 (d, 1H, *J* = 10.6 Hz, *CHHPh*), 4.93 (d, 1H, *CHHPh*), 5.59 (1H, s, H-7); ¹³C NMR (CDCl₃, 50 MHz): δ 57.6 (OCH₃), 66.4 (C-5), 68.5 (C-6), 75.0 (CH₂Ph), 77.7 (C-3), 81.7 (C-4), 84.8 (C-2), 101.5 (C-1 and C-7), 118.6 (q, *J* = 320.4 Hz, CF₃); CIMS *m/z* 505 (M+H)⁺.

Anal. Calcd for C₂₂H₂₃F₃O₈S (504.48): C, 52.38; H, 4.59; S, 6.36. Found: C, 52.61; H, 4.38; S, 6.28.

Methyl 2-*O*-benzyl-4,6-*O*-benzylidene-3-*O*-[(trifluoromethyl)sulfonyl]-β-D-glucopyranoside (24). Methyl 2-*O*-benzyl-4,6-*O*-benzylidene-β-D-glucopyranoside²⁷ **23** (1.2 g, 3.23 mmol) was reacted with trifluoromethanesulfonic anhydride (0.81 mL, 4.84 mmol) in the presence of pyridine (0.53 mL, 6.45 mmol) following general procedure A. Compound **24** (1.5 g, 93%) was obtained as white solid after flash chromatography (eluent diethylether/pentane 1:3 v/v), followed by crystallization from Et₂O: R_f 0.57 (ethyl acetate/heptane 1:1 v/v); mp 85-87 °C (dec); [α]_D -36 (*c* 1, chloroform); ¹H NMR (200 MHz) δ 3.58 (s, 3H, OCH₃), 3.80 (t, 1H, *J*_{3,4} = *J*_{4,5} = 9.4 Hz, H-4), 3.82 (t, 1H, *J*_{5,6ax} = *J*_{6ax,6eq} = 10.6 Hz, H-6ax), 4.43 (dd, 1H, *J*_{5,6eq} = 5.1 Hz, H-6eq), 4.58 (d, 1H, *J*_{1,2} = 9.4 Hz, H-1), 4.76 (d, 1H, *J* = 10.4 Hz, *CHHPh*), 4.89 (d, 1H, *CHHPh*), 4.96 (t, 1H, *J*_{2,3} = 9.4 Hz, H-3), 5.56 (s, 1H, H-7); ¹³C NMR (50 MHz) δ 57.7 (OCH₃), 65.7 (C-5), 68.5 (C-6), 75.2 (CH₂Ph), 77.9 (C-2), 79.2 (C-4), 85.9 (C-3), 101.4 (C-7), 105.1 (C-1), 118.5 (q, *J* = 320.6 Hz, CF₃); FABMS (positive): *m/z* 505 (M+H)⁺.

Methyl 3-*O*-acetyl-2-*O*-benzyl-4,6-*O*-benzylidene-β-D-allopyranoside (25). To a stirred solution of triflate **24** (800 mg, 1.59 mmol) in DMF (10 mL) was added, under argon, anhydrous tetraethylammonium acetate (1.24 g, 4.77 mmol) in one portion at

room temperature. The mixture was stirred until the starting material had completely reacted as monitored by TLC. The crude mixture was then diluted in Et₂O and washed with water. The organic layer was dried (Na₂SO₄), filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (eluent ethyl acetate/heptane 3:7 v/v) to afford compound **25** (600 mg, 94%) as a colorless syrup: R_f 0.37 (ethyl acetate/heptane 3:7 v/v); [α]_D -38 (c 1, chloroform); ¹H NMR (200 MHz) δ 2.21 (s, 3H, CH₃), 3.42 (dd, 1H, J_{1,2} = 7.9 Hz, J_{2,3} = 2.8 Hz, H-2), 3.60 (s, 3H, OCH₃), 3.60 (dd, 1H, J_{3,4} = 2.8 Hz, J_{4,5} = 10.1 Hz, H-4), 3.74 (t, 1H, J_{5,6ax} = J_{6ax,6eq} = 10.1 Hz, H-6ax), 3.95 (dt, 1H, J_{5,6eq} = 4.7 Hz, H-5), 4.39 (dd, 1H, H-6eq), 4.67 (d, 1H, J = 11.8 Hz, CHHPh), 4.72 (d, 1H, H-1), 4.74 (d, 1H, CHHPh), 5.52 (s, 1H, H-7), 5.85 (t, 1H, H-3); ¹³C NMR (50 MHz) δ 21.0 (CH₃), 57.3 (OCH₃), 63.9 (C-5), 68.1 (C-3), 69.2 (C-6), 72.4 (CH₂Ph), 76.1 and 76.9 (C-2 and C-4), 101.4 (C-7), 102.5 (C-1), 170.1 (OCOCH₃); CIMS *m/z* 415 (M+H)⁺, 383 (M+H-H₂O)⁺.

Anal. Calcd for C₂₃H₂₆O₇ (414.46): C, 66.65; H, 6.33. Found: C, 66.72; H, 6.21.

Methyl 2-O-benzyl-4,6-O-benzylidene-β-D-allopyranoside (26). To a stirred solution of compound **25** (550 mg, 1.37 mmol) in MeOH (7 mL) was added sodium methylate (74 mg, 1.37 mmol) at room temperature. The mixture was stirred at room temperature until the starting material had completely reacted as monitored by TLC. The solvent was then evaporated under reduced pressure and the residue triturated with Et₂O to furnish compound **26** (485 mg, 95%) as a white solid: R_f 0.20 (ethyl acetate/heptane 3:7 v/v); mp 122-123 °C, [α]_D -45 (c 1, chloroform), ¹H NMR (200 MHz) δ 3.28 (dd, 1H, J_{1,2} = 8 Hz, J_{2,3} = 3.8 Hz, H-2), 3.48 (dd, 1H, J_{3,4} = 2 Hz, J_{4,5} = 10.1 Hz, H-4), 3.53 (s, 3H, OCH₃), 3.67 (t, 1H, J_{5,6ax} = J_{6ax,6eq} = 10.1 Hz, H-6ax), 3.99 (dt, 1H, J_{5,6eq} = 5 Hz, H-5), 4.27 (br s, 1H, H-3), 4.35 (dd, 1H, H-6eq), 4.66 (d, 1H, J = 12.1 Hz, CHHPh), 4.72 (d, 1H, H-1), 4.81 (d, 1H, CHHPh), 5.55 (s, 1H, H-7); ¹³C NMR (50 MHz) δ 57.1 (OCH₃), 62.5 (C-5), 68.1 (C-3), 69.0 (C-6), 72.5 (CH₂Ph), 77.3 and 78.6 (C-2 and C-4), 101.7 (C-7), 102.2 (C-1); CIMS *m/z* 373 (M+H)⁺.

Anal. Calcd for C₂₁H₂₄O₆ (372.42): C, 67.73; H, 6.49. Found: C, 67.90; H, 6.63.

Methyl 2-O-benzyl-4,6-O-benzylidene-3-O-[(trifluoromethyl)sulfonyl]-β-D-allopyranoside (27). Alcohol **26** (358 mg, 0.96 mmol) was reacted with trifluoromethanesulfonic anhydride (0.25 mL, 1.48 mmol) in the presence of pyridine (0.155 mL, 1.92 mmol) following general procedure A. Unstable triflate **27** (247 mg, 51%) was obtained after flash chromatography (eluent ethyl acetate/pentane 3:7 v/v) followed by crystallization from Et₂O-heptane: R_f 0.33 (ethyl acetate/heptane 3:7 v/v); mp (dec); [α]_D -32 (c 1, chloroform); ¹H NMR (200 MHz) δ 3.48 (dd, 1H, J_{1,2} = 7.5 Hz, J_{2,3} = 3 Hz, H-2), 3.62 (3H, s, OCH₃), 3.68 (dd, 1H, J_{3,4} = 2 Hz, J_{4,5} = 10.2 Hz, H-4), 3.77

(t, 1H, $J_{5,6ax} = J_{6ax,6eq} = 10.2$ Hz, H-6ax), 3.97 (dt, 1H, $J_{5,6eq} = 4.6$ Hz, H-5), 4.44 (dd, 1H, H-6eq), 4.77 (d, 1H, H-1), 4.84 (s, 2H, CH₂Ph), 5.36 (br s, 1H, H-3), 5.55 (s, 1H, H-7); ¹³C NMR (50 MHz) δ 57.6 (OCH₃), 63.6 (C-5), 69.1 (C-6), 73.5 (CH₂Ph), 74.6 and 75.8 (C-2 and C-4), 84.1 (C-3), 102.4 and 102.5 (C-1 and C-7), 118.6 (q, $J = 320$ Hz, CF₃); CIMS m/z 505 (M+H)⁺.

Benzyl 2,3,6-tri-*O*-benzyl-4-*O*-[(trifluoromethyl) sulfonyl]- β -D-galactopyranoside (30). Benzyl 2,3,6-tri-*O*-benzyl- β -D-galactopyranoside **29** (400 mg, 0.74 mmol) was reacted with trifluoromethanesulfonic anhydride (231 μ L, 1.33 mmol) in the presence of pyridine (122 μ L, 1.48 mmol) following general procedure A. Compound **30** (473 mg, 95%) was obtained after flash chromatography (eluent diethylether/pentane 1:4 v/v) as a syrup: R_f 0.27 (ethyl acetate/heptane 1:4 v/v); $[\alpha]_D +1$ (c 1, chloroform); ¹H NMR (200 MHz) δ 3.61 (dd, 1H, $J_{2,3} = 9.9$ Hz, $J_{3,4} = 2.7$ Hz, H-3), 4.52 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1), 4.44-5.02 (m, 8H, 4 CH₂Ph), 5.42 (d, 1H, H-4); ¹³C NMR (50 MHz) δ 67.4 (C-6), 71.4 (C-5), 71.4, 73.2, 73.9 and 75.8 (4 CH₂Ph), 78.2 and 78.6 (C-2 and C-3), 82.0 (C-4), 102.7 (C-1), 118.6 (q, $J = 320.1$ Hz, CF₃); FABMS (positive) m/z 673 (M+H)⁺.

Methyl 2,3,6,2',3',6'-hexa-*O*-benzyl- β -D-maltoside (35). A mixture of methyl 2,3,6,2',3'-penta-*O*-benzyl-4',6'-*O*-benzylidene- β -D-maltoside **34** (2.85 g, 3.18 mmol), borane-trimethylamine complex (1.4 g, 19.1 mmol) and 4Å molecular sieves in THF (50 mL) was stirred, under argon, for 30 minutes at room temperature. Aluminium chloride (2.65 g, 19.1 mmol) was then added in several portions and the reaction monitored by TLC. After completion (3 hours), the molecular sieves were filtered on Celite® and the filtrate treated with Dowex 50 (H⁺) resin. The mixture was filtered and coconcentrated three times with MeOH under reduced pressure. The residue was finally purified by flash chromatography (eluent ethyl acetate/heptane 1:4 v/v) to afford compound **35** (2 g, 70%) as a syrup: R_f 0.55 (ethyl acetate/heptane 1:1 v/v); $[\alpha]_D +27$ (c 1, chloroform); ¹H NMR (200 MHz) δ 3.60 (s, 3H, OCH₃), 4.08 (br t, 1H, $J_{5,6} = 9$ Hz, H-5), 4.35 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1), 4.35-5.10 (m, 12H, 6 CH₂Ph), 5.69 (d, 1H, $J_{1',2'} = 3.6$ Hz, H-1'); ¹³C NMR (50 MHz) δ 56.9 (OCH₃), 69.5 and 70.0 (C-6 and C-6'), 70.9 and 71.6 (C-4' and C-5'), 73.1 (C-5), 74.5 (C-4), 73.0, 73.5, 73.7, 73.9, 74.8 and 75.3 (6 CH₂Ph), 79.2 (C-2'), 81.4 (C-3'), 82.3 (C-2), 84.8 (C-3), 96.7 (C-1'), 104.7 (C-1); FABMS (positive) m/z 897 (M+H)⁺.

Anal. Calcd for C₅₅H₆₀O₁₁ (897.08): C, 73.64; H, 6.74. Found: C, 73.36; H, 6.98.

Methyl 2,3,6,2',3',6'-hexa-*O*-benzyl-4'-*O*-[(trifluoromethyl)sulfonyl]- β -D-maltoside (36). Alcohol **35** (705 mg, 0.79 mmol) was reacted with

trifluoromethanesulfonic anhydride (239 μ L, 1.42 mmol) in the presence of pyridine (128 μ L, 1.58 mmol) following general procedure A. Compound **36** (676 mg, 84%) was obtained after flash chromatography (eluent ethyl acetate/pentane 3:7 v/v) as an unstable syrup: Rf 0.62 (ethyl acetate/heptane 1:1 v/v); $[\alpha]_D^{+41}$ (*c* 1, chloroform); ^1H NMR (200 MHz) δ 3.61 (3H, OCH₃), 5.69 (d, 1H, $J_{1',2'} = 3.4$ Hz, H-1'); ^{13}C NMR (50 MHz) δ 56.8 (OCH₃), 67.4 (C-6'), 68.7 (C-5'), 69.0 (C-6), 72.7 (C-5), 74.4 (C-4), 72.6, 73.2, 73.6, 74.4 and 75.0 (6 CH₂Ph), 77.7 (C-3'), 79.6 (C-4'), 81.8 (C-2'), 82.1 (C-2), 84.5 (C-3), 95.8 (C-1'), 104.7 (C-1), 118.6 (q, $J = 320$ Hz, CF₃); FABMS (positive) m/z 1029 (M+H)⁺.

9-*O*-Acetyloxime of tylosin 20-(1,3-dithiane) (37). Rf 0.53 (dichloromethane/methanol/concd ammonia 20:1:0.05 v/v/v); ^1H NMR (200 MHz) δ 2.01 and 2.08 (OCOCH₃); ^{13}C NMR (50 MHz) δ 12.5 (8-CH₃), 20.7 and 20.9 (OCOCH₃), 26.1 (SCH₂CH₂), 26.1 (2 SCH₂), 35.1 and 40.2 (C-8), 44.6 (C-20), 138.4 (C-13), 160.2 (C-9), 169.8 and 170.1 (OCOCH₃); FABMS (positive) m/z 1085 (M+Na)⁺, 1063 (M+H)⁺.

Methyl 2,4-di-*O*-acetyl-3,6-anhydro- α -D-glucopyranoside (38). Rf 0.1 (ethyl acetate 1:1 v/v); ^1H NMR (300 MHz) δ 2.01 and 2.13 (2s, 6H, 2 OCOCH₃), 3.50 (s, 3H, OCH₃), 3.92 (dd, 1H, $J_{5,6a} = 1.6$ Hz, $J_{6a,6b} = 8.6$ Hz, H-6a), 4.13 (d, 1H, H-6b), 4.48 (t, 1H, $J_{4,5} = 1.6$ Hz, H-5), 4.56 (t, 1H, $J_{2,3} = J_{3,4} = 3.6$ Hz, H-3), 4.66 (dd, 1H, H-4), 4.93 (d, 1H, $J_{1,2} = 2.6$ Hz, H-1), 5.03 (dd, 1H, H-2); ^{13}C NMR (62.5 MHz) δ 20.7 and 20.8 (2 OCOCH₃), 58.0 (OCH₃), 67.9 (C-2), 68.2 (C-6), 69.7 (C-3), 71.0 (C-4), 73.4 (C-5), 96.7 (C-1), 169.9 (2 OCOCH₃). CIMS m/z 261 (M+H)⁺, 229 (M+H-MeOH)⁺.

9-[*O*-(Methyl 2,3,4-tri-*O*-acetyl- α -D-glucopyranosid-6-yl)oxime] of tylosin 20-(1,3-dithiane) (39). 9-Oxime of tylosin 20-(1,3-dithiane) **4** (500 mg, 0.42 mmol) was reacted with triflate **8** (189 mg, 0.42 mmol) in the presence of 18-crown-6 ether (155 mg, 0.42 mmol) following general procedure B. Compound **39** (292 mg, 53%) was obtained after flash chromatography (eluent dichloromethane/methanol/concd ammonia 20:1:0.05 v/v/v) as a white foam: Rf 0.59 (dichloromethane/methanol/concd ammonia 20:1:0.05 v/v/v); ^1H NMR (200 MHz) δ 3.22 (s, 3H, 1'''-OCH₃), 5.02 (d, 1H, $J_{1''',2'''} = 4.7$ Hz, H-1'''); ^{13}C NMR (50 MHz) δ 12.4 (8-CH₃), 20.7 and 20.8 (3 OCOCH₃), 26.2 (SCH₂CH₂), 26.2 (2 SCH₂), 35.1 and 40.3 (C-8), 44.5 (C-20), 54.9 (1'''-OCH₃), 67.2 (C-5'''), 71.0, 71.0 and 71.1 (C-2''', C-3''' and C-4'''), 72.8 (C-6'''), 96.4 (C-1'''), 138.3 (C-13), 160.7 (C-9), 169.7 and 170.0 (3 OCOCH₃); FABMS (positive) m/z 1345 (M+Na)⁺, 1323 (M+H)⁺. HRMS Calcd for C₆₂H₁₀₃N₂O₂₄S₂ (1323.6342). Found: 1323.6315.

9-[*O*-(Methyl 2,3,6-tri-*O*-benzoyl- α -D-glucopyranosid-4-yl)oxime] of tylosin 20-(1,3-dithiane) (40). Compound **4** (1g, 1 mmol) was reacted with triflate **10** (625 mg, 1 mmol) in the presence of 18-crown-6 ether (364 mg, 1 mmol) following general procedure B. Product **40** (926 mg, 63%) was obtained after flash chromatography (eluent dichloromethane/methanol/concd ammonia 20:1:0.05 v/v/v) as a foam: R_f (dichloromethane/methanol/concd ammonia 10:1:0.05 v/v/v); ¹H NMR (200 MHz) δ 3.46 (1''''-OCH₃), 5.14 (d, 1H, J_{1''''',2'''''} = 3.1 Hz, H-1'''''), 6.08 (t, 1H, J_{2''''',3'''''} = J_{3''''',4'''''} = 9.6 Hz, H-3'''''); ¹³C NMR (50 MHz) δ 12.1 (8-CH₃), 25.8 (SCH₂CH₂), 26.0 (2 SCH₂), 35.1 and 41.0 (C-8), 44.1 (C-20), 55.0 (1''''-OCH₃), 63.5 (C-6'''''), 67.7 (C-5'''''), 69.9 (C-3'''''), 72.5 (C-2'''''), 77.4 (C-4'''''), 95.7 (C-1'''''), 162.8 (C-9), 165.0, 165.8 and 165.8 (3 OCOBz); FABMS (positive) *m/z* 1509 (M+H)⁺. HRMS Calcd for C₇₇H₁₀₉N₂O₂₄S₂ (1509.6812). Found: 1509.6873.

9-{*O*-[Methyl 2,3,6-tri-*O*-(4-methoxybenzyl)- α -D-galactopyranosid-4-yl]oxime} of tylosin 20-(1,3-dithiane) (41). Compound **4** (642 mg, 0.62 mmol) was reacted with triflate **14** (475 mg, 0.62 mmol) in the presence of 18-crown-6 ether (70 mg, 0.20 mmol) following general procedure B. Compound **41** (694 mg, 72%) was obtained after flash chromatography (eluent dichloromethane/methanol/concd ammonia 20:1:0.05 v/v/v) as a foam: R_f 0.58 (dichloromethane/methanol/concd ammonia 10:1:0.05 v/v/v); ¹H NMR (200 MHz) δ 3.38 (s, 3H, 1''''-OCH₃), 4.51 (d, 1H, J_{1''''',2'''''} = 2.4 Hz, H-1'''''); ¹³C NMR (50 MHz) δ 12.7 and 12.9 (8-CH₃), 26.2 (SCH₂CH₂), 26.2 (2 SCH₂), 36.1 and 40.6 (C-8), 45.6 (C-20), 55.3 (1''''-OCH₃ and 3 OCH₃), 68.9 and 69.2 (C-5'''''), 69.5 (C-6'''''), 71.0, 73.2 and 73.6 (C-4'''''), 99.1 (C-1'''''), 137.4 (C-13), 160.5 (C-9); FABMS (positive) *m/z* 1557 (M+H)⁺. HRMS Calcd for C₈₀H₁₂₁N₂O₂₄S₂ (1557.7751). Found: 1557.7712.

(*E*)-9-[*O*-(Methyl 2,3,4-tri-*O*-acetyl- α -D-glucopyranosid-6-yl)oxime] of erythromycin A (42). (*E*)-9-oxime of erythromycin A **3** (400 mg, 0.53 mmol) was reacted with triflate **8** (242 mg, 0.53 mmol) in the presence of 18-crown-6 ether (200 mg, 0.53 mmol) following general procedure B. Compound **42** (243 mg, 61%) was obtained after flash chromatography (eluent dichloromethane/methanol/concd ammonia 15:1:0.05 v/v/v) as a foam: R_f 0.40 (dichloromethane/methanol/concd ammonia 10:1:0.05); [α]_D -3 (c 0.34, chloroform); ¹H NMR (300 MHz) δ 1.00 (d, 3H, J_{8,8Me} = 7 Hz, 8-CH₃), 1.96 and 2.02 (2s, 3 and 6H, 3 CH₃), 2.66 (br q, 1H, J_{10,10Me} = 7 Hz, H-10), 2.86 (dd, 1H, J_{2,3} = 9.5 Hz, J_{2,2Me} = 7.5 Hz, H-2), 3.22 (s, 3H, 1''''-OCH₃), 3.38 (s, 3H, 3''''-OCH₃), 3.52 (d, 1H, J_{4,5} = 7.8 Hz, H-5), 3.61 (br s, 1H, H-11), 4.15 (dd, 1H, J_{5,6''''b} = 1.4 Hz, J_{6''''a,6''''b} = 13 Hz, H-6''''b), 4.32 (br s, 1H, OH), 4.43 (d, 1H, J_{1',2'} = 7.2 Hz, H-1'), 4.88 (dd, 1H, J_{1''',2'''} = 4.7 Hz, J_{2''',3'''} = 10.3 Hz, H-2'''), 4.94 (d, 1H, J_{1'',2''} = 5 Hz, H-1''), 5.02 (d,

1H, H-1'''), 5.07 (t, 1H, $J_{3,4} = J_{4,5} = 10.3$ Hz, H-4'''), 5.13 (br d, 1H, $J_{13,14ax} = 10$ Hz, H-13), 5.48 (t, 1H, H-3'''); ^{13}C NMR (62.5 MHz) δ 14.6 (10-CH₃), 19.0 (8-CH₃), 20.8 and 20.9 (3 CH₃), 26.6 (C-8), 32.2 (C-10), 56.0 (1'''-OCH₃), 67.8 (C-5'''), 69.3 (C-4'''), 70.8 and 71.1 (C-2''' and C-3'''), 71.9 (C-6'''), 97.0 (C-1'''), 169.7 and 170.0 (3 OCOCH₃), 171.8 (C-9); FABMS (positive) m/z 1073 (M+Na)⁺, 1051 (M+H)⁺, 893 [M+H-(Cladinose-H)]⁺. HRMS Calcd for C₅₀H₈₇N₂O₂₁ (1051.5801). Found: 1051.5837.

(E)-9-[O-(Methyl 2,3,6-tri-O-benzoyl- α -D-glucopyranosid-4-yl)oxime] of erythromycin A (43). (E)-9-oxime of erythromycin A **3** (543 mg, 0.73 mmol) was treated with triflate **10** (328 mg, 0.73 mmol) in the presence of 18-crown-6 ether (270 mg, 0.73 mmol) following general procedure B. Compound **43** (650 mg, 65%) was obtained after flash chromatography (eluent dichloromethane/methanol/concd ammonia 10:1:0.05 v/v/v) as a foam: Rf 0.39 (dichloromethane/methanol/concd ammonia 10:1:0.05); $[\alpha]_D^{+5}$ (c 0.46, chloroform); ^1H NMR (250 MHz) δ 1.00 (d, 3H, $J_{8,8Me} = 7$ Hz, 8-CH₃), 2.66 (br d, 1H, $J_{10,10Me} = 6.9$ Hz, H-10), 3.21 (s, 3H, 3''-OCH₃), 3.28 (dd, 1H, $J_{1',2'} = 7.2$ Hz, $J_{2',3'} = 10.3$ Hz, H-2'), 3.38 (s, 3H, 1'''-OCH₃), 3.63 (br s, 1H, H-11), 4.73 (d, 1H, $J_{1'',2''} = 4.7$ Hz, H-1''), 4.92 (dd, 1H, $J_{1''',2'''} = 3.5$ Hz, $J_{2''',3'''} = 9.6$ Hz, H-2'''), 5.11 (d, 1H, H-1'''), 5.17 (br d, 1H, $J_{13,14ax} = 10.6$ Hz, H-13), 6.08 (t, 1H, $J_{3'',4''} = 9.6$ Hz, H-3'''); ^{13}C NMR (50 MHz) δ 14.6 (10-CH₃), 18.5 (8-CH₃), 26.6 (C-8), 34.4 (C-10), 55.6 (1'''-OCH₃), 62.9 (C-6'''), 67.7 (C-5'''), 69.9 (C-3'''), 72.5 (C-2'''), 78.8 (C-4'''), 97.1 (C-1'''), 165.9 and 166.4 (3 OCOBz), 172.5 (C-9); FABMS (positive) m/z 1237 (M+H)⁺ and 1079 [M+H-(Cladinose-H)]⁺. HRMS Calcd for C₆₅H₉₃N₂O₂₁ (1237.6271). Found: 1237.6279.

(E)-9-{O-[Methyl 2,3,4-tri-O-(4-methoxybenzyl)- α -D-galactopyranosid-4-yl]oxime} of erythromycin A (44). (E)-9-oxime of erythromycin A **3** (373 mg, 0.50 mmol) was treated with triflate **14** (342 mg, 0.50 mmol) in the presence of 18-crown-6 ether (56 mg, 0.15 mmol) following general procedure B. Compound **44** (550 mg, 82%) was obtained after flash chromatography (dichloromethane/methanol/concd ammonia 20:1:0.05 v/v/v) as a foam: Rf 0.46 (dichloromethane/methanol/concd ammonia 20:1:0.05 v/v/v); $[\alpha]_D^{-19}$ (c 1, chloroform); ^1H NMR (300 MHz) δ 1.00 (d, 3H, $J_{8,8Me} = 7$ Hz, 8-CH₃), 2.92 (d, 1H, $J_{4'',5''} = 9.7$ Hz, H-4''), 3.32 (s, 3H, 3''-OCH₃), 3.37 (s, 3H, 1'''-OCH₃), 3.70 (s, 9H, 3 OCH₃), 3.88 (d, 1H, $J_{4,5} = 8$ Hz, H-5), 4.41 (d, 1H, $J_{1',2'} = 7.2$ Hz, H-1'), 4.51 (d, 1H, $J_{1''',2'''} = 3.4$ Hz, H-1'''), 4.43-4.63 (m, 6H, H-4'', CHHAr and 2 CH₂Ar), 4.72 (d, 1H, $J = 13$ Hz, CHHAr), 4.76 (d, 1H, $J_{1'',2''} = 4.6$ Hz, H-1''), 5.12 (dd, 1H, $J_{13,14ax} = 10.6$ Hz, $J_{13,14eq} = 2.5$ Hz, 13-H); ^{13}C NMR (62.5 MHz) δ 14.5 (10-CH₃), 18.8 (8-CH₃), 26.5 (C-8), 32.8 (C-10), 55.2 (3 OCH₃), 55.3 (1'''-OCH₃),

68.3 (C-6'''), 68.8 (C-5'''), 71.0, 72.8 and 73.1 (3 CH₂Ar), 75.3 (C-2'''), 76.8 (C-3'''), 77.7 (C-4'''), 98.8 (C-1'''), 171.0 (C-9); FABMS (positive) *m/z* 1285 (M+H)⁺. HRMS Calcd for C₆₈H₁₀₅N₂O₂₁ (1285.7210). Found: 1285.7251.

Methyl 2-*O*-benzyl-4,6-*O*-(4-methoxybenzyl)-3-deoxy- α -D-erythro-hex-2-enopyranoside (45). Rf 0.34 (ethyl acetate/heptane 3:7 v/v); [α]_D +23 (*c* 1, chloroform); ¹H NMR (200 MHz) δ 3.48 (s, 3H, OCH₃), 3.79 (t, 1H, J_{5,6ax} = J_{6ax,6eq} = 9.5 Hz, H-6ax), 3.97 (dt, 1H, J_{5,6eq} = 4.1 Hz, H-6eq), 4.81 (s, 1H, H-1), 4.78 (d, 1H, J = 14.5 Hz, CHHPh), 4.82 (d, 1H, CHHPh), 5.05 (br s, 1H, H-3), 5.55 (s, 1H, H-7); ¹³C NMR (50 MHz) δ 56.2 (OCH₃), 65.2 (C-5), 69.2 and 69.8 (C-6 and CH₂Ph), 76.2 (C-4), 96.9 (C-3), 98.5 (C-1), 102.0 (C-7), 153.1 (C-2); CIMS *m/z* 355 (M+H)⁺, 323, 249 and 247.

9-[*O*-(Methyl 3-*O*-benzyl-4,6-*O*-benzylidene- β -D-mannopyranosid-2-yl)oxime] of erythromycin A (46). To a mixture of (*E*)-9-oxime of erythromycin A **3** (300 mg, 0.40 mmol) and 18-crown-6 ether (45 mg, 0.12 mmol) in THF (2 mL) was added, under argon, sodium hydride (11 mg, 0.44 mmol) [sodium hydride (60% dispersion in mineral oil, 19 mg) was washed with heptane]. The mixture was stirred for 20 minutes at room temperature and then transferred via a syringe to a mixture of triflate **22** (202 mg, 0.44 mmol) in THF (2 mL). The reaction mixture was refluxed for 3 hours. After cooling, the reaction was quenched with Florisil[®] and concentrated under reduced pressure. The residue was purified by flash chromatography (eluent dichloromethane/methanol/concd ammonia 10:1:0.05 v/v/v) to furnish a 70:30 mixture (as determined by NMR) of *Z* and *E* oximes **46** (260 mg, 60%) as a foam: Rf 0.36 (dichloromethane/methanol/concd ammonia 10:1:0.05); ¹H NMR (300 MHz) δ 4.30-4.50 (m, 4H, H-1', H-1'', H-2'''(*E*) and H-6'''eq), 4.68-4.80 (m, 1H, H-2'''(*Z*)), 4.82 (d, 1H, J_{1'',2''} = 4.3 Hz, H-1''(*E*)), 4.87 (d, 1H, J_{1'',2''} = 4.3 Hz, H-1''(*Z*)), 5.22 (d, 1H, J_{13,14ax} = 10.7 Hz, H-13(*E*)), 5.29 (d, 1H, J_{13,14ax} = 10.7 Hz, H-13(*Z*)), 5.54 (s, 1H, H-7'''(*E*)), 5.64 (s, 1H, H-7'''(*Z*)); ¹³C NMR (62.5 MHz) (*Z* isomer) δ 14.9 (10-CH₃), 19.1 (8-CH₃), 26.5 (C-8), 33.1 (C-10), 57.3 (1'''-OCH₃), 67.2 (C-5'''), 69.8 (C-6'''), 72.1 (CH₂Ph), 75.2 (C-3'''), 79.1 (C-4'''), 80.6 (C-2'''), 101.7 (C-1''' and C-7'''), 170.2 (C-9); (*E* isomer) δ 11.3 (10-CH₃), 19.1 (8-CH₃), 34.4 (C-10), 35.5 (C-8), 57.3 (1'''-OCH₃), 67.4 (C'''-5), 68.9 (C-6'''), 72.1 (CH₂Ph), 75.8 (C-3'''), 79.7 (C-4'''), 80.6 (C-2'''), 101.7 (C-1''' and C-7'''), 168.3 (C-9); FABMS (positive) *m/z* 1125 (M+Na)⁺, 1103 (M+H)⁺, 945 (M+H-(Cladinose-H))⁺. HRMS Calcd for C₅₈H₉₁N₂O₁₈ (1103.6267). Found: 1103.6260.

(*E*)-9-[*O*-(Methyl 2-*O*-benzyl-4,6-*O*-benzylidene- β -D-allopyranosid-3-yl)oxime] of erythromycin A (47). (*E*)-9-oxime of erythromycin A **3** (1.2 g, 1.60 mmol) was treated with triflate **24** (807 mg, 1.60 mmol) in the presence of 18-crown-6

ether (180 mg, 0.48 mmol) following general procedure B. Compound **47** (1.23 g, 70%) was obtained after flash chromatography (eluent dichloromethane/methanol/concd ammonia 10:1:0.05 v/v/v) and crystallization from diethylether/heptane as a white solid: Rf 0.38 (dichloromethane/methanol/concd ammonia 10:1:0.05 v/v/v); mp 153-155 °C; $[\alpha]_D -72$ (c 1, chloroform); ^1H NMR (300 MHz) δ 1.00 (d, 3H, $J_{8,\text{Me}} = 7$ Hz, 8-CH₃), 2.59 (br d, 1H, $J_{10,10\text{Me}} = 7$ Hz, H-10), 3.21 (s, 3H, 3''-OCH₃), 3.22-3.32 (2H, m, 2'-H and 2'''-H), 3.33 (1H, dd, $J_{3'',4''} = 2.5$ Hz, $J_{4'',5''} = 9$ Hz, H-4''), 3.53 (s, 3H, 1'''-OCH₃), 3.78 (br s, 1H, H-11), 3.95 (m, 1H, H-5''), 4.08 (d, 1H, $J_{2,3} = 9.8$ Hz, H-3), 4.25 (dd, 1H, $J_{5'',6''\text{eq}} = 3.8$ Hz, $J_{6''\text{ax},6''\text{eq}} = 10$ Hz, H-6''eq), 4.32 (t, 1H, $J_{2'',3''} = 2.5$ Hz, H-3''), 4.42 (d, 1H, $J_{1',2'} = 6.7$ Hz, H-1'), 4.53 (1H, d, $J = 12.4$ Hz, CHHPh), 4.62 (d, 1H, $J_{1'',2''} = 8.2$ Hz, H-1''), 4.75 (d, 1H, $J_{1'',2''} = 4.9$ Hz, H-1''), 4.78 (d, 1H, CHHPh), 5.18 (br d, 1H, $J_{13,14\text{ax}} = 10.4$ Hz, H-13), 5.36 (s, 1H, H-7'''); ^{13}C NMR (50 MHz) δ 14.4 (10-CH₃), 18.6 (8-CH₃), 26.7 (C-8), 33.0 (C-10), 57.5 (1'''-OCH₃), 64.0 (C-5''), 69.4 (C-6''), 72.9 (CH₂Ph), 75.1 (C-2''), 77.8 (C-4''), 80.6 (C-3''), 101.2 (C-7''), 103.4 (C-1''), 170.0 (C-9); FABMS (positive) m/z 1125 (M+Na)⁺, 1103 (M+H)⁺. HRMS Calcd for C₃₆H₆₇N₂O₁₅ (1103.6266). Found: 1103.6320.

(E)-9-[O-(Methyl 2-O-benzyl-4,6-O-benzylidene-β-D-glucopyranosid-3-yl)oxime] of erythromycin A (48). To a mixture of (E)-9-oxime of erythromycin A **3** (225 mg, 0.30 mmol) and 18-crown-6 ether (33 mg, 0.09 mmol) in THF (1.5 mL) was added, under argon, sodium hydride (8 mg, 0.33 mmol) [sodium hydride (60% dispersion in mineral oil, 14 mg) was washed with heptane]. The mixture was then stirred for 10 minutes at room temperature and then added dropwise via a syringe into a mixture of triflate **27** (151 mg, 0.3 mmol) in THF (1.5 mL) at 0 °C. The reaction mixture was then stirred for 10 hours at 0 °C before being quenched with Florisil®. The solvent was removed under reduced pressure and the residue purified by flash chromatography (eluent dichloromethane/methanol/concd ammonia 10:1:0.05 v/v/v) to afford compound **48** (155 mg, 47%) as a foam: Rf 0.37 (dichloromethane/methanol/concd ammonia 10:1:0.05); $[\alpha]_D -52$ (c 0.93, chloroform); ^1H NMR (300 MHz) δ 1.00 (d, 3H, $J_{8,\text{Me}} = 7$ Hz, 8-CH₃), 2.48 (ddd, 1H, $J_{2',3'} = 10.1$ Hz, $J_{3',4'\text{ax}} = 12$ Hz, $J_{3',4'\text{eq}} = 2$ Hz, H-3'), 2.52 (br q, 1H, $J_{10,10\text{Me}} = 6.9$ Hz, H-10), 2.96 (d, 1H, $J_{4'',5''} = 8.4$ Hz, H-4''), 3.16 (dd, 1H, $J_{1',2'} = 7.2$ Hz, H-2'), 3.22 (s, 3H, 3-OCH₃), 3.51 (s, 3H, 1'''-OCH₃), 3.52 (br s, 1H, H-11), 3.52 (t, 1H, $J_{1'',2''} = J_{2'',3''} = 8.8$ Hz, H-2''), 3.85 (d, 1H, $J_{2,3} = 11.1$ Hz, H-3), 3.92 (dq, 1H, $J_{5'',5''\text{Me}} = 6.2$ Hz, $J_{4'',5''} = 9.7$ Hz, H-5''), 4.26 (d, 1H, H-1'), 4.27 (dd, 1H, $J_{3'',4''} = 9.4$ Hz, H-3''), 4.42 (d, 1H, H-1''), 4.61 (d, 1H, $J = 10.3$ Hz, CHHPh), 4.71 (d, 1H, $J_{1'',2''} = 4.7$ Hz, H-1''), 4.78 (d, 1H, CHHPh), 4.92 (dd, 1H, $J_{13,14\text{ax}} = 10.1$ Hz, $J_{13,14\text{eq}} = 2.3$ Hz, H-13), 5.46 (s, 1H, H-7'''); ^{13}C NMR (50 MHz) δ 14.2 (10-CH₃), 18.4 (8-

CH₃), 26.4 (C-8), 32.9 (C-10), 56.9 (1''''-OCH₃), 65.7 (C-5'''), 68.4 (C-6'''), 74.5 (CH₂Ph), 78.3 and 78.7 (C-2''' and C-4'''), 82.7 (C-3'''), 101.3 (C-7'''), 102.2 (C-1'''), 171.6 (C-9); FABMS (positive) *m/z* 1125 (M+Na)⁺, 1103 (M+H)⁺. HRMS Calcd for C₅₈H₉₁N₂O₁₈ (1103.6267). Found: 1103.6293.

(*E*)-9-[*O*-(Benzyl 2,3,6-tri-*O*-benzyl-β-D-glucopyranosid-4-yl)oxime] of erythromycin A (49). (*E*)-9-oxime of erythromycin A **3** (403 mg, 0.54 mmol) was treated with triflate **30** in the presence of 18-crown-6 ether (80 mg, 0.12 mmol) following general procedure B. Compound **49** (615 mg, 90%) was obtained after flash chromatography (eluent dichloromethane/methanol/concd ammonia 15:1:0.05 v/v/v) as a foam: R_f 0.32 (dichloromethane/methanol/concd ammonia 15:1:0.05); [α]_D -73 (c 1, chloroform); ¹H NMR (300 MHz) δ 1.00 (d, 3H, J_{8,8Me} = 7 Hz, 8-CH₃), 2.61 (br q, 1H, J_{10,10Me} = 7 Hz, 10-H), 2.90 (d, 1H, J_{4'',5''} = 9.7 Hz, H-4''), 3.18 (dd, 1H, J_{1',2'} = 7.1 Hz, J_{2',3'} = 10.2 Hz, H-2'), 3.22 (s, 3H, 3''-OCH₃), 3.49 (d, 1H, J_{4,5} = 7.3 Hz, H-5), 3.57 (dd, 1H, J_{5,6'''a} = 3.9 Hz, J_{6'''a,6'''b} = 10.9 Hz, H-6'''a), 3.73 (s, 1H, H-11), 3.91 (t, 1H, J_{2'',3''} = J_{3'',4''} = 9.2 Hz, H-3'''), 4.09 (t, 1H, J_{4'',5''} = 9.2 Hz, H-4'''), 4.34 (d, 1H, H-1'), 4.47 (d, 1H, J_{1'',2''} = 7.5 Hz, H-1'''), 4.51-4.69 (m, 6H, 3 CH₂Ph), 4.72 (d, 1H, J_{1',2'} = 4.5 Hz, H-1''), 4.82 (d, 1H, J = 12 Hz, CHHPh), 4.84 (d, 1H, CHHPh), 5.04 (dd, 1H, J_{13,14ax} = 9.3 Hz, J_{13,14eq} = 2.3 Hz, H-13); ¹³C NMR (62.5 MHz) δ 14.9 (10-OCH₃), 18.8 (8-CH₃), 26.8 (C-8), 33.0 (C-10), 69.6 (C-6'''), 73.3 (C-5'''), 73.3 and 75.0 (4 CH₂Ph), 80.7 (C-3'''), 81.4 (C-2'''), 82.3 (C-2'''), 102.7 (C-1'''), 172.1 (C-9); FABMS (positive) *m/z* (M+Na)⁺ and 1271 (M+H)⁺. HRMS Calcd for C₇₁H₁₀₃N₂O₁₈ (1271.7206). Found: 1271.7269.

(*E*)-9-{*O*-[2,3,6-Tri-*O*-benzyl-1-*O*-(methyl 2,3,6-tri-*O*-benzyl-β-D-glucopyranosid-4-yl)-α-D-galactopyranosyl-4-yl]oxime} of erythromycin A (50). (*E*)-9-oxime of erythromycin A **3** (473 mg, 0.63 mmol) was reacted with triflate **36** (650 mg, 0.63 mmol) in the presence of 18-crown-6 ether (70 mg, 0.19 mmol) following general procedure B. Compound **50** (770 mg, 75%) was obtained after flash chromatography (eluent dichloromethane/methanol/concd ammonia 10:1:0.05) as a foam: R_f 0.43 (dichloromethane/methanol/concd ammonia 10:1:0.05); [α]_D +4 (c 0.2, chloroform); ¹H NMR (300 MHz) δ 1.00 (d, 3H, J_{8,8Me} = 7 Hz, 8-CH₃), 2.53 (br d, 1H, J_{10,10Me} = 7.1 Hz, H-10), 3.51 (dd, 1H, J_{5'',6'''a} = 2.1 Hz, J_{6'''a,6'''b} = 7.4 Hz, H-6'''a), 3.58 (s, 3H, 1'''-OCH₃), 3.84 (dd, 1H, J_{2'',3''} = 10.7 Hz, J_{3'',4''} = 3.1 Hz, H-3'''), 4.32 (d, 1H, J_{1'',2''} = 7.7 Hz, H-1'''), 4.45 (d, 1H, J_{1',2'} = 7.4 Hz, H-1'), 4.67 (d, 1H, H-4'''), 4.39-4.92 (m, 12H, 6 CH₂Ph), 4.86 (d, 1H, J_{1',2'} = 4.8 Hz, H-1''), 5.18 (dd, 1H, J_{13,14ax} = 11 Hz, J_{13,14eq} = 2.1 Hz, H-13), 5.71 (d, 1H, J_{1'',2''} = 3.7 Hz, H-1'''); ¹³C NMR (62.5 MHz) δ 14.5 (10-CH₃), 18.8 (8-CH₃), 26.6 (C-8), 33.0 (C-10), 56.9 (1'''-OCH₃), 68.4

(C-6'''), 70.0 (C-6'''), 70.8 (C-5'''), 71.1, 73.4, 73.6 and 73.7 (4 CH₂Ph), 73.7 (C-5'''), 73.9 and 74.5 (2 CH₂Ph), 74.6 (C-4'''), 75.3 (C-2'''), 77.4 (C-3'''), 77.5 (C-4'''), 82.5 (C-2'''), 84.6 (C-3'''), 97.5 (C-1'''), 104.5 (C-1'''), 171.1 (C-9); FABMS (positive) *m/z* 1628 (M+H)⁺. HRMS Calcd for C₉₂H₁₂₇N₂O₂₃ (1267.8830). Found: 1267.8892.

ACKNOWLEDGMENTS

This work was supported by a grant awarded to C. G. by the Centre National de la Recherche Scientifique and A.D.I.R.

REFERENCES AND NOTES

1. S. Omura in *Macrolide Antibiotics. Chemistry, Biology and Practice*, S. Omura, Ed.; Academic Press: New York, 1984.
2. a) G. Babany, D. Larrey and D. Pessayre, *Prog. Drug Metab.*, **11**, 61 (1988); b) T.M. Ludden, *Clin. Pharmacokin.*, **10**, 63 (1985).
3. H. Sakakibara and S. Omura in *Macrolide Antibiotics. Chemistry, Biology and Practice*, S. Omura Ed.; Academic Press: New York, 1984, p 85.
4. M. Delaforge, P. Ladam, G. Bouillé, J. Gharbi-Benarous, M. Jaouen and J.-P. Girault, *Chem.-Biol. Interact.*, **85**, 215 (1992).
5. a) M. Delaforge, M. Jaouen and D. Mansuy, *Biochem. Pharmacol.*, **32**, 2309 (1983); b) J. Gharbi-Benarous, P. Ladam, M. Delaforge and J.-P. Girault, *J. Chem. Soc., Perkin Trans. I*, 2303 (1993).
6. G. Lukacs, C. Ruggeri-Duchatelle, A. Dessinges, A. Olesker, M. Laborde and L. Ming, *Eur. Pat. Appl.* 366 560, 1990.
7. J.-C. Gasc, S. Gouin d'Ambrières, A. Lutz and J.-F. Chantot, *J. Antibiot.*, **44**, 313 (1991).
8. P.A. Larrey and R. Faghih in *Recent Progress in the Chemical Synthesis of Antibiotics and Related Microbial Products*, G. Lukacs Ed.; Springer-Verlag: Berlin, 1993, p 121.
9. M. Pozo and V. Gotor, *J. Chem. Soc., Perkin Trans. I*, 1001 (1993).
10. a) J.M.J. Tronchet, D. Schazzenbach, E. Winter-Mihaly, C. Diamantides, U. Likic, G. Galland-Barrera, C. Jurrand, K. Deen Pallie, J. Ohja-Poncet, J. Rupp and G. Moret, *Helv. Chim. Acta*, **65**, 1404 (1982); b) E.M. Nashed, E. Grochowski and E. Czyzewska, *Carbohydr. Res.*, **196**, 184 (1990).
11. a) J.M.J. Tronchet, G. Zosimo-Landolfo, G. Galland-Barrera and N. Dolahatshi, *Carbohydr. Res.*, **204**, 145 (1990); b) K.C. Nicolaou and R.C. Groneberg, *J. Am. Chem. Soc.*, **112**, 4085 (1990).
12. Gasc *et al*⁷ have reported the synthesis of (*E*)-9-*O*-erythromycin A oxime in 74% yield whereas the corresponding (*E*)-9-*O*-methyloxime of erythromycin A was obtained in only 20% yield in the same conditions.
13. C. Grandjean, *Thesis*, University of Paris XI, 1994.
14. R.W. Binkley, M.G. Ambrose and D.G. Hehemann, *J. Org. Chem.*, **45**, 4387 (1980).
15. K. Dax, W. Wolflehner and H. Weidmann, *Carbohydr. Res.*, **65**, 132 (1978).
16. M.G. Ambrose and R.W. Binkley, *J. Org. Chem.*, **48**, 674 (1983).
17. L.A. Reed, III and L. Goodman, *Carbohydr. Res.*, **94**, 91 (1981).

18. R.W. Binkley, *J. Org. Chem.*, **56**, 3892 (1991).
19. T. Iwashige and H. Saeki, *Chem. Phar. Bull.*, **15**, 1803 (1967).
20. R. Johansson and B. Samuelsson, *J. Chem. Soc., Perkin Trans. I*, 2371 (1984).
21. S. Knapp, A.B.J. Naughton, C. Jaramillo and B. Pipik, *J. Org. Chem.*, **57**, 7328 (1992).
22. a) T. Iversen and D.R. Bundle, *J. Chem. Soc., Chem. Commun.*, 1240 (1981); b) U. Widmer, *Synthesis*, 568 (1987).
23. H.H. Baer and B. Radatus, *Carbohydr. Res.*, **128**, 165 (1984).
24. T. Ogawa and T. Kaburagi, *Carbohydr. Res.*, **103**, 53 (1982).
25. Y. Kondo, *Carbohydr. Res.*, **30**, 386 (1973).
26. H.B. Borén, P.J. Garegg, L. Kenne, L. Maron and S. Svensson, *Acta Chem. Scand.*, **26**, 644 (1972).
27. P.J. Garegg, T. Iversen and S. Oscarson, *Carbohydr. Res.*, **50**, C-12 (1976).
28. J.R. Turvey and T.P. Williams, *J. Chem. Soc.*, 2119 (1962).
29. M. Ek, P.J. Garegg, H. Hultberg and S. Oscarson, *J. Carbohydr. Chem.*, **2**, 305 (1983).
30. A. Liptak, I. Jodal and P. Manasi, *Carbohydr. Res.*, **44**, 1 (1975).
31. F.H. Newth, S.D. Nicholas, F. Smith and L.F. Wiggins, *J. Chem. Soc.*, 2550 (1949).
32. M.E. Evans, *Carbohydr. Res.*, **21**, 473 (1972).
33. During the preparation of this manuscript, synthesis of these triflates has been mentioned but their precise description not effected (a) or not given (b): a) M. Kassou and S. Castillon, *J. Org. Chem.*, **60**, 4353 (1995); b) A. El Nemr and T. Tsuchiya, *Tetrahedron Lett.*, **36**, 7665 (1995).
34. Partial isomerisation of (*E*)-9-erythromycin A oxime into the *Z* isomer in basic media has already been observed: see reference 7 and also R.R. Wilkening, R.W. Ratcliffe, G.A. Doss, K.F. Bartizal, A.C. Graham and C.M. Herbert, *Bioorg. Med. Chem. Lett.*, **3**, 1287 (1993).
35. A.C. Richardson, *Carbohydr. Res.*, **10**, 395 (1969).