



Note

Synthesis of 1-C-(tetra-O-acetyl- β -D-galactopyranosyl)-2,3-diiodo-1-propene and its reaction with primary amines [☆]

Uthai Sakee ^{a,*}, Chiradet Nasuk ^a, Ronald Grigg ^{b,*}

^a Center of Excellence for Innovation in Chemistry (PERCH-CIC), Department of Chemistry, Faculty of Science, Mahasarakham University, Mahasarakham 44150, Thailand

^b Molecular Innovation, Diversity and Automated Synthesis (MIDAS) Centre, School of Chemistry, University of Leeds, Leeds LS2 9JT, United Kingdom

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ABSTRACT

Iodination of 1-C-(tetra-O-acetyl- β -D-galactopyranosyl)allene affords an *E/Z*-mixture of 1-C-(tetra-O-acetyl- β -D-galactopyranosyl)-2,3-diiodo-1-propene. *S_N2* displacement of the allylic iodide with primary amines affords an *E/Z*-mixture of allylic amines under a variety of conditions. Due to its experimental simplicity and low cost of reagents, this procedure may find wide use in the laboratory for functionalization of sugar allenes.

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Carbohydrate-based starting materials are widely used in target-oriented syntheses of optically active compounds.¹ C-Glycosyl compounds ('C-glycosides') with unsaturated aglycones are useful for further functionalization purposes because of their potential for the introduction of new stereogenic centers in the side chain.² The addition of iodine to allene is well established;^{3–5} however, this type of reaction has not been reported for sugar allenes. In this communication, we report the addition of iodine to 1-C-(tetra-O-acetyl- β -D-galactopyranosyl)allene (**1**)⁶ in a variety of organic solvents and the subsequent *S_N2* displacement of the allylic iodide in the diiodo addition products with primary amines. We found that in all cases an *E/Z*-mixture of *E*-**2** and *Z*-**2** was formed and that the *E/Z* ratio was influenced by the solvent, time, and temperature⁷ (Table 1).

In Table 1 EtOH is identified as the best solvent for the reactions studied. The products were formed by addition of iodine to the terminal π -bond of the allene moiety. The other regioisomer resulting from *I*₂ addition to the internal π -bond of allene (**1**) was not observed. Selectivity for (*Z*)-**2** increased with increasing reaction time and temperature (Table 1, entries 5, 7, and 8), and further studies of this trend are needed. The *E/Z*-isomerization of the diiodides is presumed to proceed via a radical mechanism⁷ involving excess *I*₂ (Scheme 1).

The olefin geometry was assigned on the basis of the chemical shift of the vinylic protons in the ¹H NMR spectrum of the diiodide

reaction mixture. The vinyl proton resonance for the major *Z*-**2** (δ 6.36) is 0.04 ppm upfield from the corresponding resonance for the *E*-isomer *E*-**2** (δ 6.40). This assignment was also confirmed by appropriate NOE experiments (Fig. 1).

Irradiation at the resonance frequency of H₃ in the *Z*-isomer *Z*-**2** (δ 6.36) resulted in an enhancement of the H₄ and H_{1a} resonances of 8.5 and 6.3%, respectively, whilst irradiation at the analogous resonance frequency of the *E*-isomer *E*-**2** (δ 6.40) did not result in any enhancement of H_{1a} or H_{1b} but gave a 5.1% enhancement of H₄.

The crude (1:1.9-*E/Z*)-mixture of allylic iodides **2** was subjected to *S_N2* displacement with benzylamine nucleophiles **4a**, **b** varying both solvent (CH₂Cl₂, THF) and base (Et₃N, K₂CO₃). Part of this study is summarized in Table 2.

The geometry of *E*-**5a** and *Z*-**5a** was determined by NOE experiments as shown in Figure 2.

In conclusion, we have established a simple route from 1-C-(tetra-O-acetyl- β -D-galactopyranosyl)allene (**1**) to (*E/Z*)-1-C-(tetra-O-acetyl- β -D-galactopyranosyl)-2,3-diiodo-1-propene (**2**) in >95% yield. Regioselective substitution of the allylic diiodide by benzylamines can be achieved in moderate to good yield (Table 2, entries 1 and 6).

1. Experimental

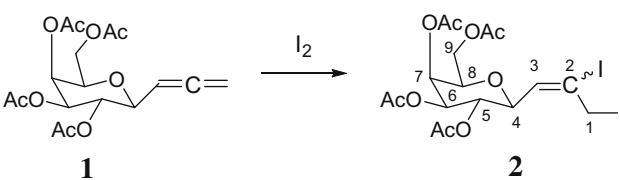
1.1. General methods

All reagents were commercially available and used without purification. Flash column chromatography was performed using

[☆] IUPAC nomenclature: (2*E*/*Z*)-5*S*,6*R*,7*S*,9-tetra-O-acetyl-4,8-anhydro-1,2,3-trideoxy-1,2-diiodonon-2-enitol.

* Corresponding authors. Tel.: +66 43 754246; fax: +66 43 754246 (U.S.).

E-mail addresses: uthai.s@msu.ac.th (U. Sakee), R.Grigg@leeds.ac.uk (R. Grigg).

Table 1The reaction of β -C-galactose allene (**1**) with iodine^a


Entry	Solvent	Temperature ^b (°C)	Time ^c (h)	Yield ^d (%)	E/Z ^e
1	CH ₂ Cl ₂	rt	48	20	1:2.3
2	CH ₂ Cl ₂	40	24	25	1:1.7
3	MeOH	rt	24	30	1:1.9
4	MeOH	70	3	40	1:2.3
5	EtOH	80	3	92	1:1.9
6	MeC ₆ H ₅	90	3	22	1:1.5
7	EtOH	Reflux	3	96	1:3.2
8	EtOH	Reflux	1.5	97	1:1.7
9	CH ₃ CN	Reflux	1.5	81	1:4.1

^a Compound **1** (0.540 mmol), I₂ (0.594 mmol), and 20 mL of solvent were used.^b Oil bath temperature except where noted.^c TLC monitoring to 100% completion.^d Isolated yield.^e Ratios were determined by ¹H NMR.

Silica Gel 60 (230–400 mesh). Optical rotations were measured on an AA-100 polarimeter at room temperature and are given in 10⁻¹ deg cm² g⁻¹. Accurate molecular masses were recorded on a Bruker micrOTOF instrument. Infrared spectra were recorded using a Perkin–Elmer FTIR spectrometer. The IR samples were prepared as thin films by evaporation of a solution of the compound in CH₂Cl₂ onto a germanium plate. Nuclear magnetic resonance spectra were determined at 500 MHz (¹H) and at 75 MHz (¹³C) on Bruker spectrometers. Chemical shift values are quoted in parts per million (ppm) downfield from TMS, and coupling constants are in hertz. Chemical shift multiplicities are reported as s = singlet, d = doublet, dd = doublet of doublets, t = triplet, q = quartet, m = multiplet, br = broad, and br s = broad singlet, app. t = apparent triplet. 1-C-(tetra-O-acetyl- β -D-galactopyranosyl)allene (**1**) was prepared according to a literature method.⁶

1.2. (2E)-5S,6R,7S,9-Tetra-O-acetyl-4,8-anhydro-1,2,3-trideoxy-1,2-diiodonon-2-enitol (E-2) and (2Z)-5S,6R,7S,9-Tetra-O-acetyl-4,8-anhydro-1,2,3-trideoxy-1,2-diiodonon-2-enitol (Z-2)

Iodine (0.754 g, 2.970 mmol) was added to a stirred solution of the 1-C-(tetra-O-acetyl- β -D-galactopyranosyl)allene (**1**) (1.0 g, 2.7 mmol) in EtOH (200 mL). The resulting mixture was stirred and heated under reflux for 3 h. The solvent was then removed, the residue was dissolved in Et₂O (100 mL), and the solution was sequentially washed with satd sodium thiosulfate (100 mL), satd NaHCO₃ (100 mL), and satd brine (100 mL). The organic layer was separated, dried (Na₂SO₄), and filtered, and the filtrate evaporated under reduced pressure. Purification of the residue by flash column chromatography, eluting with 2:1 hexane–Et₂O, gave a 1:3.2 mixture of E- and Z-2 as a pale-yellow gum (1.61 g, 2.59 mmol, 96%).

Gradient flash column chromatography of the mixture of E- and Z-2 with 3:1 to 2:1 hexane–Et₂O afforded partial separation of E- and Z-2 with some fractions containing mixtures of E- and Z-2.

1.2.1. Data for E-2

Pale-yellow gum; [α]_D²⁴ +238.5 (c 0.218, CHCl₃); IR (film): 2971, 2121, 1737, 1615, 1423, 1366, 753, 600, 542 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 6.4 (1H, d, J 6.4, 3-H), 5.42–5.44 (1H, m, 7-H), 5.3 (1H, dd, J 10.7 and 6.0, 5-H), 5.17 (1H, dd, J 10.7 and 3.0, 6-H), 4.8 (1H, app t, J 6.0 and 5.95, 4-H), 4.56 and 4.37 (2H, 2 \times d, J 10.25, 1-H), 4.0–4.2 (3H, m, 8-H and 2 \times 9-H), 2.1, 2.08, 2.03 and 2.01 (12H, 4 \times s, 4 \times OCOMe); ¹³C NMR (75 MHz, CDCl₃): δ 169.2, 169.1, 169.0, 168.8, 134.2, 129.2, 107.4, 69.7, 68.0, 67.0, 66.4, 65.7, 60.2, 20.9, 19.8, 19.7, 19.6; ESIMS (positive-ion): m/z 625 (MH⁺, 100); HRMS: calcd for C₁₇H₂₂I₂NaO₉ (M + Na): 646.9245; found: 646.9255.

1.2.2. Data for Z-2

Pale-yellow gum; [α]_D²⁴ +253.5 (c 0.493, CHCl₃); IR (film): 2971, 2121, 1737, 1615, 1423, 1366, 753, 600, 542 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 6.36 (1H, d, J 7.7, 3-H), 5.42–5.46 (1H, m, 7-H), 5.36 (1H, dd, J 10.25 and 6.0, 5-H), 5.18 (1H, dd, J 10.25 and 3.0, 6-H), 4.78 (1H, app. t, J 6.85 and 6.4, 4-H), 4.50 and 4.45 (2H,

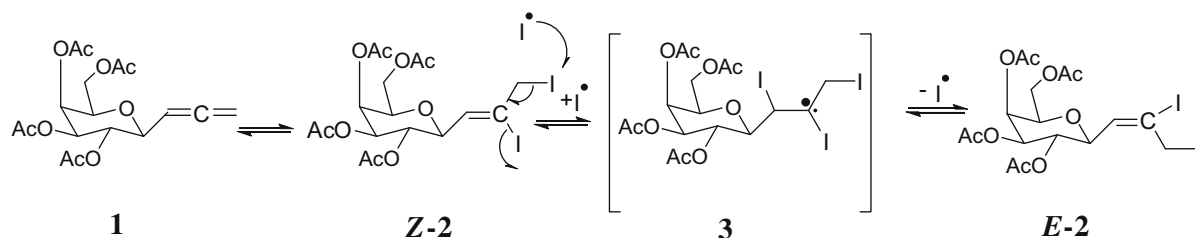
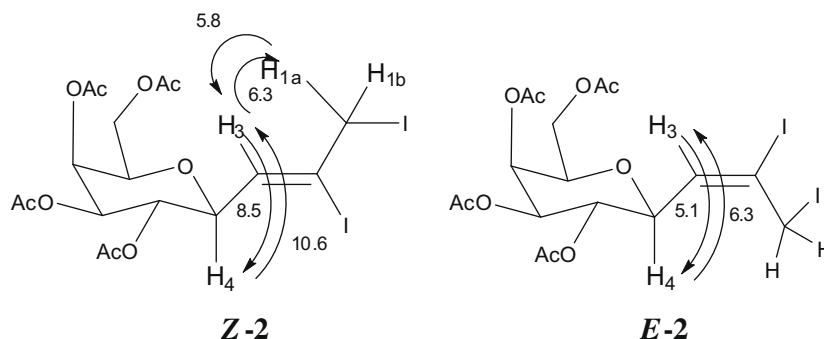
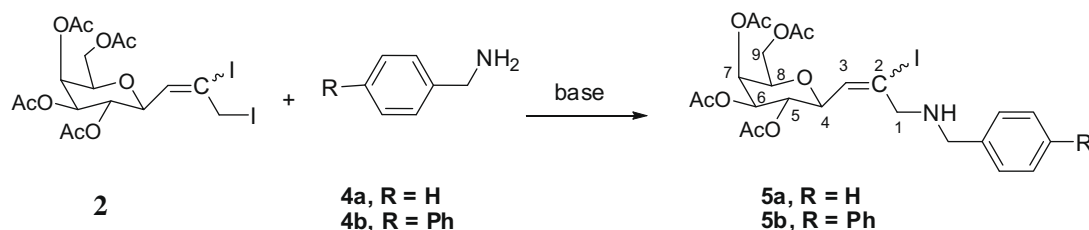
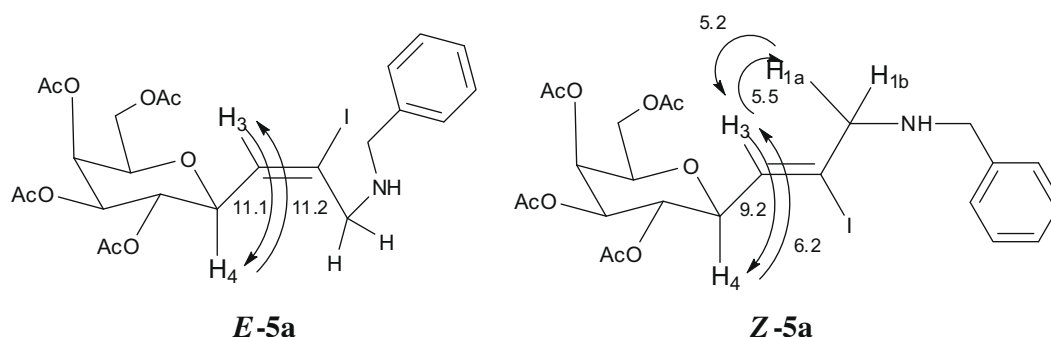
**Scheme 1.** Proposed E/Z isomerization mechanism.**Figure 1.** NOE data for E-2 and Z-2.

Table 2The reaction of diiodide **2** with primary amines under different reaction conditions^a

Entry	Nucleophile	Base (mol equiv)	Solvent	Temperature (°C)	Time ^b (h)	Product, yield ^c (%)	E:Z
1	4a	Et ₃ N (1.1)	CH ₂ Cl ₂	rt	24	5a (45)	1:1.25
2	4a	Et ₃ N (2)	CH ₂ Cl ₂	rt	24	5a (27)	1:1.25
3	4a	Et ₃ N (1.1)	CH ₂ Cl ₂	40	12	5a (22)	1:1.2
4	4b	Et ₃ N (1.1)	CH ₂ Cl ₂	rt	24	5b (53)	1:1.3
5	4b	Et ₃ N (1.1)	THF	rt	24	5b (25)	1:1.5
6	4b	K ₂ CO ₃ (5)	CH ₂ Cl ₂	Reflux	24	5b (79)	1:2.16
7	4a	K ₂ CO ₃ (5)	CH ₂ Cl ₂	Reflux	24	5a (25)	1:1.5

^a Compound **2** (1: 1.9-*E/Z*-mixture) (0.33 mmol), nucleophile (1.1 mol equiv), and 5 mL of solvent were used.^b Reaction monitored by TLC.^c Isolated yield.**Figure 2.** NOE data for **E-5a** and **Z-5a**.

2 × d, *J* 10.25, 1-H), 4.05–4.15 (2H, m, 2 × 9-H), 4.02 (1H, app t, *J* 6.0 and 5.1), 2.1, 2.06 and 2.03 (12H, 4 × s, 4 × OCOMe); ¹³C NMR (75 MHz, CDCl₃): δ 170.9, 170.56, 170.52, 170.2, 130.6, 76.6, 69.9, 69.0, 67.8, 67.5, 62.0, 21.2, 21.1, 21.0; ESIMS (positive-ion): *m/z* 625 (MH⁺, 100); HRMS: calcd for C₁₇H₂₂I₂NaO₉ (M+Na): 646.9245; found: 646.9255.

1.3. General procedure for the reaction of **2** with primary amines

A 1:1.9 mixture of *E*- and *Z*-**2** (0.33 mmol) was dissolved in CH₂Cl₂ (5 mL), and K₂CO₃ (5.0 mol equiv) or Et₃N (1.1 mol equiv), and amine (1.1 mol equiv) were added. The resulting mixture was stirred and heated under reflux for 24 h, cooled, poured into water, and extracted with CH₂Cl₂ (2 × 20 mL). The organic layer was separated and washed with water (2 × 50 mL), dried (Na₂SO₄), and filtered, and the filtrate was evaporated. Purification of the residue by flash column chromatography afforded the product.

1.3.1. (2*E*)-5*S*,6*R*,7*S*,9-Tetra-*O*-acetyl-4,8-anhydro-1-(benzylamino)-1,2,3-trideoxy-2-iodonon-2-enitol (**E-5a**) and (2*Z*)-5*S*,6*R*,7*S*,9-tetra-*O*-acetyl-4,8-anhydro-1-(benzylamino)-1,2,3-trideoxy-2-iodonon-2-enitol (**Z-5a**)

Compounds **E-5a** and **Z-5a** were prepared by the general procedure from 1:1.9-*E/Z*-**2** (0.400 g, 0.641 mmol) in CH₂Cl₂ (7 mL), K₂CO₃ (0.442 g, 3.205 mmol), and benzylamine (**4a**) (0.075 g,

0.705 mmol) under reflux for 24 h. Purification of the residue by gradient elution flash column chromatography with 1:1 to 1:2 hexane–EtOAc afforded **E-5a** (0.096 g, 0.160 mmol, 25%) and **Z-5a** (0.208 g, 0.346 mmol, 54%) as pale-yellow gums.

1.3.1.1. Data for E-5a. [α]_D²⁴ –190.7 (c 1.206, CHCl₃); IR (film): 3473, 2970, 1739, 1624, 1489, 1434, 1365, 1217, 701, 538 cm^{–1}; ¹H NMR (500 MHz, CDCl₃): δ 7.32–7.33 (5H, m, Ar-H), 6.6 (1H, d, *J* 6.8, 3-H), 5.41–5.42 (1H, m, 7-H), 5.2–5.26 (2H, m, 5-H and 6-H), 4.9 (1H, app t, *J* 6.3 and 6.2, 4-H), 4.40–4.1 (3H, m, 8-H and 2 × 9-H), 3.7 and 3.6 (2H, 2d, *J* 12.9, NCH₂Ph), 3.5 and 3.4 (2H, 2d, *J* 14.8, 2 × 1-H), 2.1, 2.06, 2.01 and 1.96 (12H, 4 × s, 4 × OCOMe); ¹³C NMR (75 MHz, CDCl₃): δ 170.7, 170.6, 170.5, 170.2, 139.2, 135.4, 128.9, 128.7, 127.1, 117.5, 70.9, 69.2, 68.4, 68.1, 67.5, 62.1, 54.2, 52.1, 21.6, 21.2; ESIMS (positive-ion): *m/z* 604 (MH⁺, 100); HRMS: calcd for C₂₄H₃₁INO₉ (M+H): 604.1049; found: 604.1070.

1.3.1.2. Data for Z-5a. [α]_D²⁴ +107.2 (c 1.212, CHCl₃); IR (film): 3473, 2970, 1739, 1624, 1489, 1434, 1365, 1217, 701, 538 cm^{–1}; ¹H NMR (500 MHz, CDCl₃): δ 7.34–7.35 (5H, m, Ar-H), 6.2 (1H, d, *J* 7.7, 3-H), 5.44 (1H, dd, *J* 3.3 and 1.6, 7-H), 5.41 (1H, dd, *J* 10.3 and 5.9, 5-H), 5.2 (1H, dd, *J* 10.3 and 3.3, 6-H), 4.78 (1H, dd, *J* 7.7 and 5.9, 4-H), 4.0–4.2 (3H, m, 6-H and 2 × 9-H), 3.7 and 3.68 (2H, 2d, *J* 13.0, 2 × 1-H), 3.57 and 3.53 (2H, 2d, *J* 15.5, NCH₂Ph), 2.1, 2.04 and 2.02 (12H, 4 × s, 4 × OCOMe); ¹³C NMR (75 MHz, CDCl₃): δ 170.9, 170.6, 170.5, 170.2, 139.7, 129.0, 128.7, 128.2,

127.7, 119.5, 76.5, 69.7, 69.0, 67.9, 67.6, 62.1, 61.0, 51.4, 21.3, 21.2, 21.1, 21.0; ESIMS (positive-ion): m/z 604 (MH^+ , 100); HRMS: calcd for $C_{24}H_{31}INO_9$ ($M+H$): 604.1049; found: 604.1070.

1.3.2. (2E)-5S,6R,7S,9-Tetra-O-acetyl-4,8-anhydro-1-[(biphenyl-4-ylmethyl)amino]-1,2,3-trideoxy-2-iodonon-2-enitol (E-5b) and (2Z)-5S,6R,7S,9-tetra-O-acetyl-4,8-anhydro-1-[(biphenyl-4-ylmethyl)amino]-1,2,3-trideoxy-2-iodonon-2-enitol (Z-5b)

Compounds *E/Z* **5b** were prepared by the general procedure from 1:1.9-*E/Z*-**2** (0.400 g, 0.641 mmol) in CH_2Cl_2 (7 mL), Et_3N (0.071 g, 0.705 mmol) and 1-biphenyl-4-ylmethanamine (**4b**) (0.128 g, 0.705 mmol) at room temperature for 24 h. Workup, followed by gradient elution flash column chromatography with 1:1 to 1:2 hexane- $EtOAc$, afforded **E-5b** (0.088 g, 0.147 mmol, 23%) and **Z-5b** (0.116 g, 0.192 mmol, 30%) as pale-yellow gums.

1.3.2.1. Data for E-5b. $[\alpha]_D^{24}$ –110.7 (c 0.659, $CHCl_3$); IR (film): 3474, 3003, 2970, 1742, 1488, 1435, 1366, 1228, 1217, 1084, 1053, 911, 765, 700, 530 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$): δ 7.5 and 7.6 (4H, 2d, J 7.7, Ar-H), 7.3–7.45 (5H, m, Ar-H), 6.6 (1H, d, J 6.8, 3-H), 5.4–5.45 (1H, m, 7-H), 5.26 (1H, dd, J 10.6 and 5.6, 5-H), 5.2 (1H, dd, J 10.3 and 3, 6-H), 4.95 (1H, app t, J 6.4 and 6.0, 4-H), 4.0–4.1 (3H, m, 8-H and $2 \times$ 9-H), 3.76 and 3.68 (2H, 2d, J 13.25, $2 \times$ 1-H), 3.5 and 3.4 (2H, 2d, J 14.6, NCH_2Ar), 2.1, 2.07, 2.0 and 1.97 (12H, $4 \times$ s, $4 \times$ $OCOMe$); ^{13}C NMR (75 MHz, $CDCl_3$): δ 170.7, 170.6, 170.5, 170.2, 141.2, 140.6, 138.9, 135.4, 129.2, 129.1, 127.7, 127.6, 127.4, 117.8, 71.0, 69.1, 68.4, 68.1, 67.5, 62.1, 54.3, 51.8, 21.3, 21.2, 21.0; ESIMS (positive-ion): m/z 680 (MH^+ , 100); HRMS: calcd for $C_{30}H_{35}INO_9$ ($M+H$): 680.1351; found: 680.1365.

1.3.2.2. Data for (Z-5b). $[\alpha]_D^{24}$ +234.9 (c 0.166, $CHCl_3$); IR (film): 3474, 3003, 2970, 1742, 1488, 1435, 1366, 1228, 1217, 1084, 1053, 911, 765, 700, 530 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$): δ 7.6

(4H, app t, J 7.7 and 6.8, Ar-H), 7.4–7.45 (4H, m, $4 \times$ Ar-H), 7.34 (1H, t, J 7.25, Ar-H), 6.25 (1H, d, J 7.7, 3-H), 5.43–5.45 (1H, m, 7-H), 5.41 (1H, dd, J 10.3 and 5.95, 5-H), 5.2 (1H, dd, J 10.3 and 3.0, 6-H), 4.95 (1H, app t, J 6.8 and 6.4, 4-H), 4.05–4.17 (3H, m, 8-H and $2 \times$ 9-H), 3.75 and 3.72 (2H, 2d, J 13.25, $2 \times$ 1-H), 3.59 and 3.55 (2H, 2d, J 15.8, NCH_2Ar), 2.17, 2.15, 2.03 and 2.02 (12H, $4 \times$ s, $4 \times$ $OCOMe$); ^{13}C NMR (75 MHz, $CDCl_3$): δ 170.4, 170.1, 170.0, 169.7, 140.8, 140.2, 138.4, 128.7, 127.9, 127.3, 127.2, 127.0, 119.1, 76.1, 69.3, 68.5, 68.3, 67.5, 67.2, 61.7, 60.5, 50.7, 20.9, 20.8, 20.7, 20.6; ESIMS (positive-ion): m/z 680 (MH^+ , 100); HRMS: calcd for $C_{30}H_{35}INO_9$ ($M+H$): 680.1351; found: 680.1365.

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