Regioselective Reductive Ring Opening of Benzylidene Acetals Using Triethylsilane and Iodine

Rajib Panchadhayee, Anup Kumar Misra*

Division of Molecular Medicine, Bose Institute, P-1/12, C. I. T. Scheme VII M, Kolkata 700054, India Fax +91(33)23553886; E-mail: akmisra69@gmail.com *Received 22 January 2010*

Abstract: Novel reaction conditions have been developed for the regioselective reductive ring opening of benzylidene acetals in carbohydrate derivatives using triethylsilane and molecular iodine. The reaction is fast, compatible with most of the functional groups encountered in the oligosaccharide synthesis, and yields were excellent. The reaction conditions are equally effective in thioglycosides.

Key words: carbohydrate, benzylidene acetal, regioselective, iodine, triethylsilane

Judicious protection of hydroxyl groups in the polyhydroxylated carbohydrate skeleton plays an important role in the synthesis of complex oligosaccharides and natural products.¹ Although chemistry for the selective protection and deprotection of carbohydrates has been extensively studied, there is still a need for improved methodology. Benzylidene acetals have found extensive application in oligosaccharide synthesis because of the group's utility for the simultaneous protection of two hydroxyl groups and its removal under acidic or neutral conditions.² Transformation of a benzylidene acetal to a benzyl or benzoyl group are useful conversions in the synthesis of oligosaccharides. The 4,6-O-benzylidene acetal in carbohydrate derivatives can be regioselectively opened to give either 6-O-benzylated or 4-O-benzylated derivatives depending on the reducing agent used.² For the selective removal of benzylidene acetasl a number of methods are available in the literature, including NaBH₃CN-HCl,³ NaBH₃CN-TFA,⁴ Et₃SiH–TFA,⁵ Et₃SiH–BF₃·OEt₂⁶ for the preparation of 6-O-benzylated derivatives and AlCl₃-LiAlH₄,⁷ DIBAL-H,⁸ Me₃NBH₃-AlCl₃,⁴ BH₃·THF-TMSOTf,⁹ Bu₂BOTf-BH₃·THF,¹⁰ CoCl₂-BH₃·THF¹¹ and others¹² for the preparation of 4-O-benzylated derivatives. Nevertheless, many of these methods for the preparation of 6-Obenzylated carbohydrate derivatives have shortcomings such as formation of byproducts, use of expensive reagents, incompatibility with other functional groups, and harsh reaction conditions. Therefore, development of mild, efficient, and metal-free methodology for the regioselective ring opening of benzylidene acetals is still During attractive. the synthesis of complex oligosaccharides¹³ we applied Et₃SiH-TFA⁵ or Et₃SiH- $BF_3 \cdot OEt_2^6$ to the regioselective opening of 4,6-O-benzylidene acetals in sugar derivatives to obtain 6-O-benzylated derivatives. However, in some cases we obtained poor yields because of the instability of some functional groups under the reaction conditions. Alternative use of NaBH₃CN–HCl·Et₂O requires repeated purification of the products to remove borate salts, and the Et₃SiH-TFA system did not work in D-galactose derivatives. Thus we sought to explore other possible reagent combinations for the reductive ring opening of benzylidene acetals of Dglucose and D-galactose derivatives. We envisaged that use of a Et₃SiH and molecular iodine combination could serve to improve the yield of the products. We disclose herein convenient reaction conditions for the regioselective ring opening of 4,6-O-benzylidene acetals of carbohydrate derivatives into 6-O-benzylated derivatives using combination of triethylsilane-molecular iodine (Scheme 1).



Scheme 1 Regioselective ring opening of benzylidene acetal using triethylsilane and iodine.

In a set of initial experiments, methyl 2,3-di-O-acetyl-4,6-*O*-benzylidene- α -D-glucopyranoside (1) was allowed to stir with triethylsilane and iodine in MeCN at room temperature, varying the quantity of reagents. It was observed that treatment of compound 1 with 1.5 equiv. of Et₃SiH and 0.2 equiv. of iodine in MeCN could efficiently produce methyl 2,3-di-O-acetyl-6-O-benzyl-a-D-glucopyranoside (14) in 10 min at room temperature. Reducing the quantity of reagents resulted in an incomplete transformation even after 5 h. Use of other organic solvents such as CH₂Cl₂, THF produced a similar result in the formation of compound 14. Following these reaction condition, a series of 4,6-O-benzylidene acetal derivatives derived from Dglucose and D-galactose was transformed into the corresponding 6-O-benzylated derivatives (Scheme 1, Table 1). Noteworthy features are the absence of formation of 4-O-benzylated derivatives and stability of the majority of the functional groups used for the protection of hydroxy and amino groups in carbohydrates. Furthermore benzylidene acetals of thio- and selenoglycosides can also be regioselectively opened without any side reaction (Table 1, entries 8, 10 and 12); despite the fact that iodine has been used for thioglycoside activation.¹⁴ The reaction conditions were equally effective in D-glucose and D-

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galactose derivatives unlike the Et₃SiH–TFA combination,⁵ which can be used in D-glucose derivatives only. Finally, stereochemistry at the anomeric center does not influence the product formation. In order to establish the significant advantages of the present protocol, a comparative study was also carried out with the existing literature reported reaction conditions (Table 2) from which it is clear that the present reaction condition produces higher yields of the product in shorter time. In summary, a novel, mild reaction condition has been developed for the regioselective reductive ring opening of benzylidene acetals in the carbohydrate backbone using triethylsilane and molecular iodine. This straightforward reaction is rapid and high yielding and can be scaled-up. Use of readily available reagents, without requirement of heavy metal salts, high boiling solvents or Lewis acids makes this reaction protocol an attractive alternative to existing methodology.

Table 1	Regioselective Rin	g Opening of Benzylide	ne Acetal of Carbohydrate Derivatives	Using Et ₃ SiH and I ₂
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Entry	Substrates ^a	Products ^a	Time (min)	Yield (%)
1	Ph TO O AcO AcO OMe	HO AcO AcO OMe	10	95 ¹⁵
2	1 Ph TO TO BzO BzO OMe	14 HO OBn BzO BzO OMe	10	92 ^{12b}
3	2 Ph TO O BnO BnO OMe 3	15 HO BNO BNO OME	10	92 ¹⁶
4	Ph TO O Aco OMp NPhth	HO AcO NPhth	15	87 ¹⁷
5	Ph TO OBn BnO OMe	BnO OBn HO -O BnO OMe	10	90 ⁷
6	Ph TO AcO BnO OMe	HO Aco Bno OMe	10	92 ¹⁸
7		19 HO Allo BnO OMe	10	88 ¹⁹
8	Ph TO O BZO BZO SPh 8	$\begin{array}{c} 20 \\ HO \\ BZO \\ BZO \\ BZO \\ SPh \\ \end{array}$	10	82 ²⁰
9	Ph CO AcO AcO AcO OMp	$\begin{array}{c} HO \\ AcO \\ AcO \\ AcO \\ AcO \\ AcO \\ OMp \\ 22 \end{array}$	25	90
	9			

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Entry	Substrates ^a	Products ^a	Time (min)	Yield (%)
10	AcO AcO SEt	$\begin{array}{c} HO \qquad OBn \\ AcO \qquad AcO \qquad SEt \\ 23 \end{array}$	20	80 ²¹
11	Ph b BzO BzO BzO OMp	HO OBn BzO OMp 24	25	88 ²²
12	$\frac{11}{AcO}$	HO AcO 25		
13	Ph to to Aco Aco Aco Aco OMe	HO ACO ACO ACO ACO ACO ACO ACO ACO	30	85
	13			

Table 1Regioselective Ring Opening of Benzylidene Acetal of Carbohydrate Derivatives Using Et_3SiH and I_2 (continued)

^a Mp: 4-methoxyphenyl, NPhth: N-phthalimido, All: allyl, Bn: benzyl, Bz: benzoyl. Known compounds gave ¹H NMR and ¹³C NMR spectra that matched the data reported in the cited references.

 Table 2
 Comparison of Reaction Conditions for the Preparation of 6-O-Benzyl Carbohydrate Derivatives

Substrate	Product	Reagent	Time (min)	Yield (%)
		NaBH ₃ CN–HCl·Et ₂ O	5	87 ³
PhTO 070	OBn	Et ₃ SiH–TFA	180	80 ⁵
BnO OMe BnO	BnO OMe BnO	$Et_3SiH-BF_3\cdot OEt_2$	240	84 ^a
3		Et ₃ SiH–I ₂	10	92 ^a

^a This study.

Typical Experimental Conditions

To a solution of compound 1 (1 mmol) in MeCN (5 mL) were added triethylsilane (1.5 mmol) and iodine (0.2 mmol) at 0–5 °C, and the reaction mixture was allowed to stir at the same temperature for the appropriate time (Table 1). After completion of reaction (TLC), the reaction mixture was diluted with CH_2Cl_2 (20 mL). The organic layer was successively washed with sat. NaHCO₃ and H₂O, dried (Na₂SO₄), and concentrated under reduced pressure. The crude product was purified using hexane–EtOAc (4:1) as eluent to furnish pure compound 14 (95%).²³

Supporting Information for this article is available online at http://www.thieme-connect.com/ejournals/toc/synlett.

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- (23) Spectroscopic data for novel products are presented below. To aid assignments, data were taken after acetylation of the products.

4-Methoxyphenyl 2,3,4-Tri-*O*-acetyl-6-*O*-benzyl-β-Dgalactopyranoside (Acetylated 22)

¹H NMR (500 MHz, CDCl₃): δ = 7.25–7.17 (m, 5 H, ArH), 6.87 (d, *J* = 9.0 Hz, 2 H, ArH), 6.70 (d, *J* = 9.0 Hz, 2 H, ArH), 5.40 (d, *J* = 3.3 Hz, 1 H, H-4), 5.31 (dd, *J* = 7.9 Hz each, 1 H, H-2), 4.99 (dd, *J* = 10.4, 3.4 Hz, 1 H, H-3), 4.83 (d, *J* = 7.9 Hz, 1 H, H-1), 4.48 (d, *J* = 11.9 Hz, 1 H, PhCH_{2a}), 4.36 (d, *J* = 11.9 Hz, 1 H, PhCH_{2b}), 3.87–3.84 (m, 1 H, H-5), 3.68 (s, 3 H, OCH₃), 3.52–3.44 (m, 2 H, H-6_{a,b}), 2.01, 1.99, 1.93 (3 s, 9 H, 3 COCH₃). ¹³C NMR (125 MHz, CDCl₃): δ = 170.3, 170.1, 169.5 (3 *C*OCH₃), 156.0–114.9 (ArC), 101.1 (C-1), 73.9, 72.9, 71.5, 69.4, 67.9, 67.8, 55.8, 21.1, 21.0, 20.9 (3 COCH₃). ESI-MS (C₂₆H₃₀O₁₀): *m*/*z* = 525.1 [M + Na]⁺.

Phenyl 2,3,4-Tri-O-acetyl-6-O-benzyl-1-seleno-β-Dglucopyranoside (Acetylated 25)

¹H NMR (500 MHz, CDCl₃): δ = 7.59–7.21 (m, 10 H, ArH), 5.15 (t, J = 9.2 Hz, 1 H, H-3), 5.01 (t, J = 9.8 Hz, 1 H, H-2), 4.97 (t, J = 9.2 Hz, 1 H, H-4), 4.88 (d, J = 9.9 Hz, 1 H, H-1), 4.53–4.46 (2 d, J = 11.8 Hz, 2 H, PhCH₂), 3.67–3.63 (m, 1 H, H-5), 3.56–3.54 (m, 2 H, H-6_{a,b}), 2.14, 2.05, 1.96 (3 s, 9 H, 3 COCH₃). ¹³C NMR (125 MHz, CDCl₃): δ = 170.4, 169.6, 169.4 (3 COCH₃), 138.2–127.6 (ArC), 81.4 (C-1), 78.9, 74.4, 73.9, 71.3, 69.4 (2 C), 21.1, 20.9 (2 C) (3 COCH₃). ESI-MS (C₂₅H₂₈O₈Se): m/z = 559.1 [M + Na]⁺. Methyl (2,3,4-Tri-*O*-acetyl-6-*O*-benzyl-β-D-

glucopyranosyl)-(1→6)-2,3,4-tri-*O*-acetyl-α-Dglucopyranoside (Acetylated 26)

¹H NMR (500 MHz, CDCl₃): $\delta = 7.33-7.24$ (m, 5 H, ArH), 5.43 (t, J = 10.0 Hz each, 1 H, H-3_A), 5.16 (t, J = 9.4 Hz each, 1 H, H-4_A), 5.03 (t, J = 9.5 Hz each, 1 H, H-3_B), 4.96 (dd, J = 7.9 Hz each, 1 H, H-2_B), 4.90 (t, J = 9.4 Hz each, 1 H, H-4_B), 4.88 (d, J = 3.6 Hz, 1 H, H-1_A), 4.82 (dd, J = 10.1, 3.6 Hz, 1 H, H-2_A), 4.55 (d, J = 11.9 Hz, 1 H, PhCH_{2a}), 4.52 (d, J = 7.9 Hz, 1 H, H-1_B), 4.48 (d, J = 11.9 Hz, 1 H, PhCH_{2b}), 3.94–3.89 (m, 2 H, H-6_{abA}), 3.65–3.61 (m, 1 H, H-5_B), 3.55–3.51 (m, 3 H, H-5_A, H-6_{abB}), 3.37 (s, 3 H, OCH₃), 2.06, 2.03, 1.98, 1.89 (4 s, 18 H, 6 COCH₃). ¹³C NMR (125 MHz, CDCl₃): $\delta = 170.4$, 170.1, 170.0, 169.7, 169.6, 169.4 (6 COCH₃), 137.9–128.1 (ArC), 101.2 (C-1_B), 96.8 (C-1_A), 73.9, 73.7, 73.3, 71.6, 71.2, 70.6, 69.7, 69.4, 69.2, 68.5, 68.3, 55.5 (OCH₃), 21.1 (2 C), 21.0 (2 C), 20.9 (2 C). ESI-MS (C₃₀H₄₀O₁₆): m/z = 679.2 [M + Na]⁺. Copyright of Synlett is the property of Georg Thieme Verlag Stuttgart and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.