

Note

Fluorination of methyl 6-deoxy-1-thiohexopyranosides with diethylaminosulfur trifluoride (DAST), and the formation of unexpected products

Takashi Ando, Hideharu Ishida, Makoto Kiso and Akira Hasegawa *

Department of Applied Bioorganic Chemistry, Gifu University, Gifu 501-11 (Japan)

(Received April 21st, 1993; accepted June 6th, 1993)

The selectin family, such as E-selectin (endothelial-leukocyte adhesion molecule-1), P-selectin (GMP-1), and L-selectin (leucocyte adhesion molecule-1), recognizes the sialyl Le^x determinant¹, α -Neu5Ac-(2 → 3)- β -D-Gal-(1 → 4)-[α -L-Fuc-(1 → 3)]- β -D-GlcNAc, which is found as the terminal carbohydrate structure in both glycolipids and glycoproteins. In the course of an investigation^{2–4} on the structural requirements for the carbohydrate ligand of selectin recognition, we have demonstrated that both the fucose and sialic acid residues are required for full recognition. In order to clarify the structural requirement for the L-fucose moiety of the selectin recognition, we have designed a synthesis of the fluorinated 6-deoxyhexopyranose-containing the sialyl Le^x epitope. We report here fluorination of some methyl 6-deoxy-1-thiohexopyranoside derivatives.

We chose the readily available methyl 2,3,4-tri-*O*-acetyl-1-thio- β -L-fucopyranoside^{3b} (**1**) and methyl 1-thio- α -L-rhamnopyranoside⁵ (**5**) as the starting materials, and diethylaminosulfur trifluoride (DAST) as the fluorinating reagent. Although DAST is widely used to produce fluorinated carbohydrates in high yield, unpredictable products^{6–8} are occasionally formed. Treatment of methyl 1-thio- β -L-fucopyranoside (**2**), prepared by *O*-deacetylation of methyl 2,3,4-tri-*O*-acetyl-1-thio- β -L-fucopyranoside^{3b} (**1**), with 2,2-dimethoxypropane in *N,N*-dimethylformamide (DMF) in the presence of *p*-toluenesulfonic acid monohydrate gave methyl 3,4-*O*-isopropylidene-1-thio- β -L-fucopyranoside (**3**) in 90% yield **. The ¹H NMR spectrum revealed the presence of one isopropylidene and two methyl groups, H-2 as a doublet of doublets, and H-1 as a doublet, confirming the structure assigned. On treatment with DAST in dichloromethane for 1 h at 0°C, compound **3** afforded

* Corresponding author.

** Compounds **2** and **3** have been previously reported⁹, although they were not fully characterized.

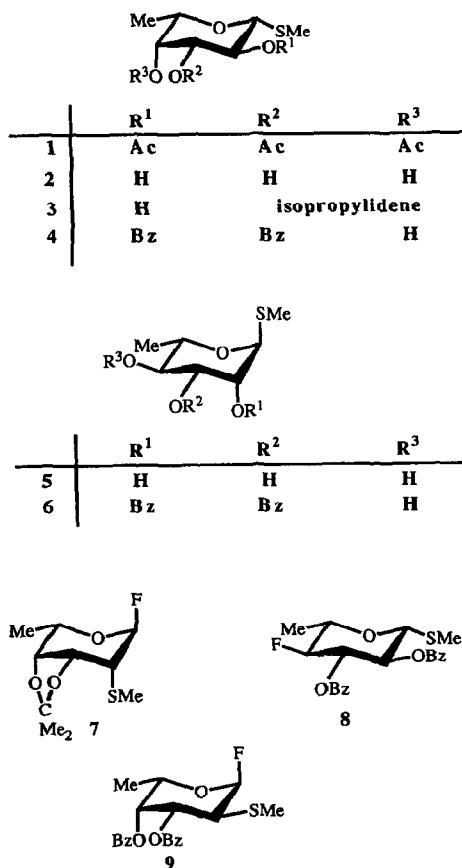


Fig. 1.

an unexpected glycosyl fluoride, 6-deoxy-3,4-*O*-isopropylidene-2-*S*-methyl-2-thio- α -*L*-talopyranoside (**7**) in 66% yield; the methylthio group at C-1 in **3** had migrated to C-2 and a fluorine atom became attached to C-1, analogous to behavior observed previously^{6–8a}. The ¹H NMR spectrum of **7** exhibited the presence of one C-methyl group, one *S*-methyl group, an isopropylidene group, H-1 as a doublet of doublets at δ 5.68 ($J_{1,2}$ 6.6, $J_{1,F}$ 62.1 Hz), and H-2 as a multiplet, confirming the structure assigned.

Treatment of methyl 2,3-di-*O*-benzoyl-1-thio- β -*L*-fucopyranoside (**4**, prepared by 2,3-di-*O*-benzoylation of compound **2**) with DAST gave the desired 4-fluoro compound, methyl 2,3-di-*O*-benzoyl-4,6-dideoxy-4-fluoro-1-thio- β -*L*-glucopyranoside (**8**) in 45% yield. The ¹H NMR spectrum of **8** exhibited signals for H-5 at δ 3.81 ($J_{4,5}$ 9.4, $J_{5,6}$ 6.0, and $J_{5,F}$ 2.7 Hz), H-4 at δ 4.35 ($J_{3,4}$ = $J_{4,5}$ = 9.4, $J_{4,F}$ 50.3 Hz), H-3 at δ 5.76 ($J_{2,3}$ = $J_{3,4}$ = 9.4, $J_{3,F}$ 13.3 Hz), H-2 at δ 5.44 ($J_{1,2}$ 9.8 Hz), and H-1 at δ 4.61, consistent with the assignment of the 4-fluoro- β -*L*-glucopyranose

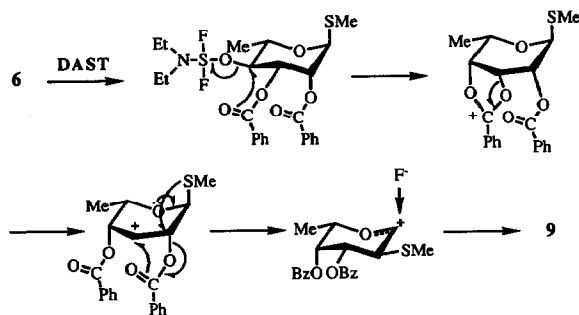


Fig. 2.

configuration. In contrast, when treated with DAST under the same conditions, methyl 2,3-di-*O*-benzoyl-1-thio- α -L-rhamnopyranoside [derived from methyl 1-thio- α -L-rhamnopyranoside⁵ (5) by selective benzylation] gave the unpredicted 1-fluoro compound **9** in 42% yield. In the ¹H NMR spectrum of **9**, H-1 appeared as a doublet of doublets at δ 5.94 ($J_{1,F}$ 51.3, $J_{1,2}$, 2.4 Hz) and H-2 as a multiplet at δ 3.29 ($J_{2,3}$ 11.7, $J_{2,F}$ 30.2 Hz), indicating the 1-fluoro substituent to be *cis* to the *S*-methyl group on C-2.

As regards the mechanism of formation of **9**, as proposed in the scheme, the oxygen atom of the hydroxyl group at C-4 replaces a fluorine atom of DAST, with loss of hydrogen fluoride, to form the sulfoxo group, and the intermediate may then be converted via orthoester formation with inversion of configuration at the C-4. Successive, intramolecular rearrangements of the benzoyl and *S*-methyl groups, and then 1-fluorination from the β -side, would give methyl 3,4-di-*O*-benzoyl-2-*S*-methyl-2-thio- α -L-fucopyranosyl fluoride (**9**).

EXPERIMENTAL

General methods.—Melting points are uncorrected. Optical rotations were determined with a Union PM-201 polarimeter at 25°C. ¹H NMR spectra were recorded with a Jeol JNM-GX 270 spectrometer. Preparative chromatography was performed on silica gel (Wako Co., 200 mesh) with the solvents specified. Concentrations were conducted in vacuo.

Methyl 1-thio- β -L-fucopyranoside (2).—To a solution of methyl 2,3,4-tri-*O*-acetyl-1-thio- β -L-fucopyranoside^{3b} (**1**; 2.4 g, 7.5 mmol) in MeOH (15 mL) was added NaOMe (30 mg), and the mixture was treated with Amberlite IR-120 (H⁺) resin to remove the base. The solution was concentrated to give a crystalline product. Recrystallization from EtOH–Et₂O gave **2** (1.4 g, 96%) as needles; mp 103–105; $[\alpha]_D^{20} +21.0^\circ$ (*c* 1.0, CHCl₃); ¹H NMR (CD₃OD); δ 1.34 (d, 3 H, $J_{5,6}$ 6.4 Hz, MeCH), 2.26 (s, 3 H, MeS), 3.56 (dd, 1 H, $J_{2,3}$ 9.2, $J_{3,4}$ 2.4 Hz, H-3), 3.63 (t, 1 H, $J_{1,2} = J_{2,3} = 9.2$ Hz, H-2), 3.74 (q, 1 H, H-5), 3.75 (d, 1 H, $J_{3,4}$ 2.6 Hz, H-4), and

4.28 (d, 1 H, H-1). Anal. Calcd for $C_7H_{14}O_4S$ (194.3): C, 43.28; H, 7.26. Found: C, 43.26; H, 7.47.

Methyl 3,4-O-isopropylidene-1-thio- β -L-fucopyranoside (3).—To a solution of **2** (2.0 g, 10.3 mmol) in DMF (10 mL) were added 2,2-dimethoxypropane (2.5 mL) and *p*-toluenesulfonic acid monohydrate (50 mg), and the mixture was stirred for 5 h at 60°C, and then neutralized with Amberlite IR-410 (HO[−]) resin. The solution was concentrated. Column chromatography (1:1 EtOAc–hexane) of the residue on silica gel (100 g) afforded **3** (2.2 g, 90%). Recrystallization from ether gave needles; mp 63–65°C; $[\alpha]_D -47.5^\circ$ (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃): δ 1.37, 1.54 (2 s, 6 H, Me₂C), 1.41 (d, 3 H, *J*_{5,6} 6.6 Hz, MeC), 2.21 (s, 3 H, MeS), 3.57 (dd, 1 H, *J*_{1,2} 10.2, *J*_{2,3} 6.7 Hz, H-2), and 4.14 (d, 1 H, H-1). Anal. Calcd for $C_{10}H_{18}O_4S$ (234.3): C, 51.26; H, 7.74. Found: C, 51.23; H, 7.49.

Methyl 2,3-di-O-benzoyl-1-thio- β -L-fucopyranoside (4).—To a solution of **2** (2.5 g, 12.9 mmol) in dry pyridine (9 mL) and dry CH₂Cl₂ (18 mL) was added dropwise a solution of BzCl (3.1 mL, 27 mmol) in dry CH₂Cl₂ (7.5 mL) at −40°C, and the mixture was stirred for 30 min at −40°C. Methanol (1 mL) was added and the mixture was concentrated, then extracted with CH₂Cl₂. The extract was successively washed with 2 M HCl and H₂O, dried (Na₂SO₄), and concentrated. Column chromatography (1:3 EtOAc–hexane) of the residue on silica gel (50 g) gave **4** (3.4 g, 66%) as a syrup; $[\alpha]_D -96.2^\circ$ (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃): δ 1.41 (d, 3 H, *J*_{5,6} 6.4 Hz, MeC), 2.24 (s, 3 H, MeS), 3.93 (q, 1 H, H-5), 4.14 (br d, 1 H, *J*_{3,4} 3.1 Hz, H-4), 4.59 (d, 1 H, *J*_{1,2} 9.9 Hz, H-1), 5.36 (dd, 1 H, *J*_{2,3} 10.1 Hz, H-3), 5.80 (t, 1 H, *J*_{1,2} = *J*_{2,3} = 10.0 Hz, H-2), and 7.31–8.07 (m, 10 H, 2 Ph). Anal. Calcd for $C_{21}H_{22}O_6S$ (402.5): C, 62.67; H, 5.51. Found: C, 62.53; H, 5.68.

Methyl 2,3-di-O-benzoyl-1-thio- α -L-rhamnopyranoside (6).—To a solution of methyl 1-thio- α -L-rhamnopyranoside⁵ (**5**; 900 mg, 4.6 mmol) in dry pyridine (3.2 mL) and CH₂Cl₂ (6.4 mL) was added dropwise a solution of BzCl (1.1 mL, 92 mmol) in dry CH₂Cl₂ (2.6 mL) at −40°C, and the mixture was stirred for 30 min at −40°C. Methanol (3 mL) was added to the mixture, which was concentrated and then extracted with CH₂Cl₂. The extract was successively washed with 2 M HCl and H₂O, dried (Na₂SO₄), and then concentrated. Column chromatography (1:3 EtOAc–hexane) of the residue on silica gel (100 g) gave **6** (1.4 g, 75%) as an amorphous mass; $[\alpha]_D -5.9^\circ$ (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃): δ 1.46 (d, 3 H, *J*_{5,6} 6.2 Hz, MeC), 2.21 (s, 3 H, MeS), 3.96 (t, 1 H, *J*_{3,4} = *J*_{4,5} = 9.6 Hz, H-4), 4.22 (q, 1 H, H-5), 5.29 (d, 1 H, *J*_{1,2} 1.6 Hz, H-1), 5.48 (dd, 1 H, *J*_{2,3} 3.4 Hz, H-3), 5.70 (dd, 1 H, H-2), and 7.26–8.09 (m, 10 H, 2 Ph). Anal. Calcd for $C_{21}H_{22}O_6S$ (402.5): C, 62.67; H, 5.51. Found: C, 62.68; H, 5.62.

6-Deoxy-3,4-O-isopropylidene-2-S-methyl-2-thio- α -L-talopyranosyl fluoride (7).—To a solution of **3** (50 mg, 0.21 mmol) in dry CH₂Cl₂ (2 mL) was added DAST (70 μ L), and the mixture was stirred for 1 h at 0°C. Methanol (0.5 mL) was added to the mixture, which was then concentrated. Column chromatography (1:5 EtOAc–hexane) of the residue on silica gel (10 g) gave **7** (33.5 mg, 66%) as a syrup; $[\alpha]_D -44.5^\circ$ (*c* 0.7, CHCl₃); ¹H NMR (CDCl₃): δ 1.30 (d, 3 H, *J*_{5,6} 6.4 Hz, MeC), 1.35,

1.46 (2 s, 6 H, Me₂C), 2.29 (s, 3 H, MeS), 2.81 (ddd, 1 H, $J_{1,2}$ 6.6, $J_{2,3}$ 2.5 $J_{2,F}$ 18.2 Hz, H-2), 3.94 (br q, 1 H, H-5), 4.15 (d, 1 H, $J_{3,4}$ 7.7 Hz, H-4), 4.61 (ddd, 1 H, $J_{3,F}$ 4.2 Hz, H-3), and 5.68 (dd, 1 H, $J_{1,F}$ 62.1 Hz, H-1). Anal. Calcd for C₁₀H₁₇FO₃S (236.3): C, 50.83; H, 7.25. Found: C, 50.98; H, 7.36.

Methyl 2,3-di-O-benzoyl-4,6-dideoxy-4-fluoro-1-thio-β-L-glucopyranoside (8).—To a solution of **4** (100 mg, 0.25 mmol) in dry CH₂Cl₂ (2 mL) was added DAST (82 μL, 0.63 mmol) at 0°C, and the mixture was stirred for 2 h at 0°C. Methanol (0.5 mL) was added to the mixture which was then concentrated. Column chromatography (1:6 EtOAc–hexane) of the residue on silica gel (20 g) afforded **8** (45 mg, 46%) as an amorphous mass; $[\alpha]_D -50.8^\circ$ (c 0.9, CHCl₃); ¹H NMR (CDCl₃): δ 1.47 (dd, 3 H, $J_{5,6}$ 6.0, $J_{F,6}$ 1.3 Hz, MeC), 2.22 (s, 3 H, MeS), 3.81 (ddd, 1 H, $J_{4,5}$ 9.4, $J_{5,F}$ 2.7 Hz, H-5), 4.35 (dt, 1 H, $J_{3,4} = J_{4,5} = 9.4$, $J_{4,F}$ 50.3 Hz, H-4), 4.61 (d, 1 H, $J_{1,2}$ 9.8 Hz, H-1), and 5.44 (near t, 1 H, $J_{1,2} = J_{2,3} = 9.8$ Hz, H-2), 5.76 (dt, 1 H, $J_{3,F}$ 13.3 Hz, H-3), and 7.38–8.01 (m, 10 H, 2 Ph). Anal. Calcd for C₂₁H₂₁FO₅S (404.5): C, 62.36; H, 5.23. Found: C, 62.33; H, 5.05.

3,4-Di-O-benzoyl-2-S-methyl-2-thio-α-L-fucopyranosyl fluoride (9).—To a solution of **6** (60 mg, 0.14 mmol) in dry CH₂Cl₂ (1.4 mL) was added DAST (50 μL, 0.38 mmol) at 0°C, and the mixture was stirred for 1 h at 0°C. A similar work up as described for **7** gave **9** (25 mg, 42%) as an amorphous mass; $[\alpha]_D -241.1^\circ$ (c 0.84, CHCl₃); ¹H NMR (CDCl₃): δ 1.29 (d, 3 H, $J_{5,6}$ 6.7 Hz, MeC), 2.16 (s, 3 H, MeS), 3.29 (ddd, 1 H, $J_{1,2}$ 2.4, $J_{2,3}$ 11.7, $J_{2,F}$ 30.2 Hz, H-2), 4.53 (q, 1 H, H-5), 5.71 (m, 2 H, H-3,4), 5.94 (dd, 1 H, $J_{1,F}$ 51.3 Hz, H-1), and 7.30–8.06 (m, 10 H, 2 Ph). Anal. Calcd for C₂₁H₂₁FO₅S (404.5): C, 62.36; H, 5.23. Found: C, 62.49; H, 5.18.

ACKNOWLEDGMENT

This work was supported in part by a Grant-in-Aid (No. 03660132) for Scientific Research from the Ministry of Education, Science and Culture of Japan.

REFERENCES

- (a) G. Walz, A. Aruffo, W. Kolanus, M. Bevilacqua, and B. Seed, *Science*, 250 (1990) 1132–1135; (b) M.J. Polley, M.L. Philips, E. Wayner, E. Nudelman, A.K. Singhal, S. Hakomori, and J.C. Paulson, *Proc. Natl. Acad. Sci. USA*, 88 (1991) 6224–6228; (c) M. Tiemeyer, S.J. Swiedler, M. Ishihara, M. Moleland, H. Schweingruber, P. Hirtzer, and B.K. Brandley, *ibid.*, 88 (1991) 1138–1142; (d) Y. Imai, M.S. Singer, C. Fennie, L.A. Lasky, and S.D. Rosen, *J. Cell Biol.*, 113 (1991) 1213–1221.
- (a) C. Foxall, S.R. Watson, D. Dowbenko, C. Fennie, L.A. Lasky, M. Kiso, A. Hasegawa, D. Asa, and B.K. Brandley, *J. Cell Biol.*, 117 (1992) 895–902; (b) D. Tyrrell, P. James, N. Rao, C. Foxall, S. Abbas, F. Dasgupta, M. Nashed, A. Hasegawa, M. Kiso, D. Asa, J. Kidd, and B.K. Brandley, *Proc. Natl. Acad. Sci. USA*, 88 (1991) 10372–10376; (c) M. Larkin, T.J. Ahern, M.S. Stoll, M. Shaffer, D. Sako, J. O'Brien, C.T. Yuen, A.M. Lawson, R.A. Childs, K.M. Barone, P.R. Langer-Safer, A. Hasegawa, M. Kiso, G.R. Larsen, and T. Feizi, *J. Biol. Chem.*, 267 (1992) 13661–13668.
- (a) A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, *Carbohydr. Res.*, 209 (1991) c1–c4; (b) and *J. Carbohydr. Chem.*, 10 (1991) 549–560; (c) A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, *J. Carbohydr. Chem.*, 10 (1991) 729–738; (d) A. Hasegawa, T. Ando, A. Kameyama, and M. Kiso, *Carbohydr. Res.*, 230 (1992) c1–c5; (e) and *J. Carbohydr. Chem.*, 11 (1992) 645–658.

- 4 (a) M. Yoshida, A. Uchimura, M. Kiso, and A. Hasegawa, *Glycoconjugate J.*, in press; (b) T. Ando, M. Kato, M. Kiso, and A. Hasegawa, *Nippon Nogeikagaku kaishi*, 67 (1993) 417.
- 5 V. Pozsgay and H.J. Jennings, *J. Org. Chem.*, 53 (1988) 4042–4052.
- 6 A. Hasegawa, M. Goto, and M. Kiso, *J. Carbohydr. Chem.*, 4 (1985) 627–638.
- 7 K.C. Nicolaou, T. Ladduwahetty, J.L. Randall, and A. Chucholowski, *J. Am. Chem. Soc.*, 108 (1986) 2466–2467.
- 8 (a) P. Kovac, H.J.C. Yeh, G.L. Jung, and C.P.J. Glaudemans, *J. Carbohydr. Chem.*, 5 (1986) 497–512; (b) P. Kovac, H.J.C. Yeh, and G.L. Jung, *J. Carbohydr. Chem.*, 6 (1987) 423–439; (c) E. Petrákova, H.J.C. Yeh, P. Kovac, and C.P.J. Glaudemans, *J. Carbohydr. Chem.*, 11 (1992) 407–412.
- 9 R.K. Jain and K.L. Matta, *Tetrahedron Lett.*, 31 (1990) 4325–4328.