

Synthetic Communications: An International Journal for Rapid Communication of Synthetic Organic Chemistry

Publication details, including instructions for
authors and subscription information:

<http://www.tandfonline.com/loi/lcyc20>

An Improved Preparation of 4-Hydroxymethyl-L- Phenylalanine

Enrico Morera ^a, Giorgio Ortar ^a & Aurelio Varani ^a

^a Dipartimento di Studi Farmaceutici e Centro di
Studio per la Chimica del Farmaco del C.N.R.,
Università "La Sapienza", 00185, Roma, Italy
Version of record first published: 11 Mar 2009.

To cite this article: Enrico Morera , Giorgio Ortar & Aurelio Varani (1998):
An Improved Preparation of 4-Hydroxymethyl-L-Phenylalanine, Synthetic
Communications: An International Journal for Rapid Communication of Synthetic
Organic Chemistry, 28:22, 4279-4285

To link to this article: <http://dx.doi.org/10.1080/00397919809458709>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.tandfonline.com/page/terms-and-conditions>

This article may be used for research, teaching, and private study
purposes. Any substantial or systematic reproduction, redistribution,
reselling, loan, sub-licensing, systematic supply, or distribution in any
form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

AN IMPROVED PREPARATION OF 4-HYDROXYMETHYL-L-PHENYLALANINE

Enrico Morera, Giorgio Ortar,* and Aurelio Varani

Dipartimento di Studi Farmaceutici e Centro di Studio per la Chimica del Farmaco
del C.N.R., Università "La Sapienza", 00185 Roma, Italy

Abstract: Palladium-catalyzed hydroformylation of methyl esters of *N*-Boc-L-tyrosine triflate (**1a**) and *N*-Boc-4-iodo-L-phenylalanine (**1b**) followed by reduction with sodium borohydride and standard deprotection procedures affords the title compound in 77 and 83% overall yield, respectively.

Tyrosine phosphorylation by protein tyrosine kinases (PTK) is of central importance in a broad range of cellular processes including cell growth, differentiation, and death.¹ Alterations in the activity of PTKs are associated with a number of human diseases like cancer, diabetes, and immune dysfunctions.²

In addition, the presence of a sulphated tyrosine residue is essential for the expression of the biological activity of a series of secretory and neuropeptides such as gastrin,³ cholecystokinin,⁴ fibronectin,⁵ and leucosulfakinin.⁶

The preparation of amino acids mimicking tyrosine phosphate or sulphate residues for the development of PTK inhibitors or of peptide analogues is therefore of widespread interest.⁷

*To whom correspondence should be addressed.

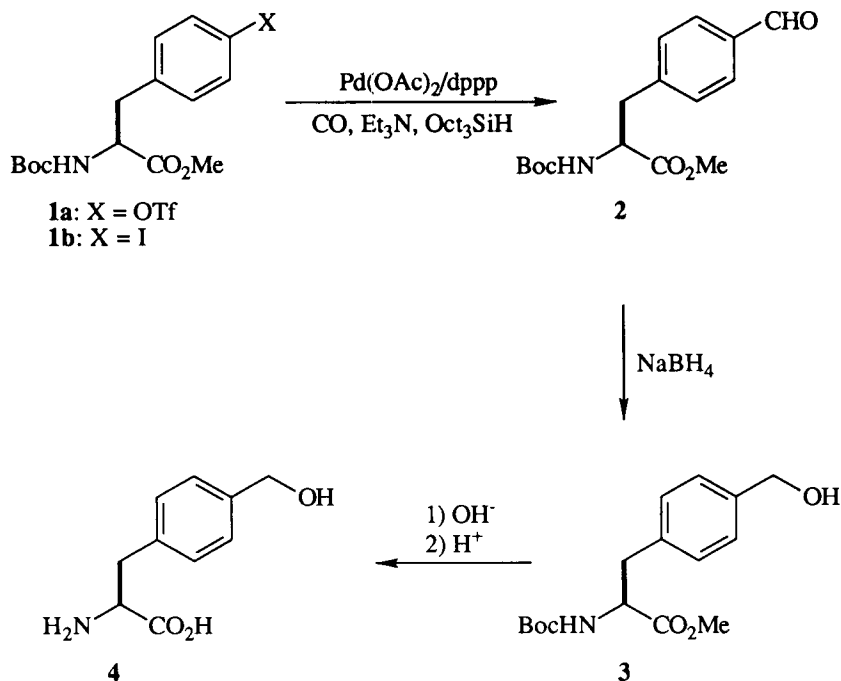
4-Hydroxymethyl-L-phenylalanine (**4**) is a versatile unnatural amino acid which has been employed to this end by different authors both in linear strategies^{7b} and in divergent ones.⁸ Compound **4** was also found of interest in metabolic studies.⁹

Previous reports on the synthesis of 4-hydroxymethyl-L-phenylalanine have involved the nonselective preparation of the D,L-form followed by an enzymatic resolution,^{7b,9a} or have exploited the chiral phase-transfer catalysis methodology of O'Donnell.⁸ The first approach appears to be inefficient and inconvenient for large scale preparations, while the latter produces a 4/1 ratio of L and D enantiomers.

As part of our interest in the palladium-catalyzed elaboration of α -amino acids,¹⁰ we report here an alternative and convenient procedure for the preparation of **4** which utilizes as the key step a recently reported procedure for the palladium-catalyzed hydroformylation of aryl triflates (Scheme).¹¹

The process starts with the tyrosine triflate or the 4-iodophenylalanine derivatives **1a,b**, both readily available from the corresponding protected tyrosine¹² or from L-phenylalanine,¹³ respectively. Reaction of **1a** and **1b** with carbon monoxide and trioctylsilane in the presence of Pd(OAc)₂/dppp¹⁴ as the catalyst following a procedure similar to that introduced by Kotsuki¹¹ afforded the aldehyde **2** in 85 and 89% yield, respectively. Faster conversion was observed with **1a**. In fact, when 3 mol% of the catalyst were employed, the reaction with **1a** went to completion in 4 h, while 12 h were required for the complete consumption of **1b**. To the best of our knowledge, this is the first application of the Kotsuki's method to an aryl iodide.

Hartman and Halczenko¹⁵ have already reported a palladium-catalyzed carbonylative approach to *N*-Boc-4-formyl-L-phenylalanine from *N*-Boc-4-iodo-L-phenylalanine by the use of tributyltin hydride as hydrogen donor. A paper describing an application of this procedure to the preparation of the pentapeptide Ac-Ile-Phe(4-formyl)-Gly-Glu(O-tBu)-Phe-NH₂, key intermediate for divergent PTK pp60^{c-src} inhibitor synthesis, has also appeared during completion of our work.^{7a} The formylation of organic halides or triflates with carbon monoxide and Bu₃SnH presents however the disadvantage that it requires the slow addition of the reagent (≥ 2.5 h), in order to minimize the direct reduction of the halide or triflate.¹⁶



Scheme

Reduction of **2** with sodium borohydride at room temperature afforded the hydroxymethyl derivative **3** in 91% yield. The methyl ester function remained substantially unaffected under the conditions described in the Experimental Section. If, however, a 1.5 molar excess of the hydride was employed and the reaction time was increased to 1 h, the yield of **3** diminished to 75% and a 21% of the bis-alcohol was isolated. It is well known that the presence of suitable neighbouring groups enhances the reactivity of NaBH_4 toward esters.¹⁷

The two-step reaction sequence leading to **3** can be conveniently conducted as a one-pot operation that omits the isolation of the aldehyde. The alcohol **3** was isolated in this case in 79 and 85% yield from **1a** and **1b**, respectively.¹⁸

The free amino acid **4** was obtained almost quantitatively by saponification of **3** with a $\text{NaOH}/\text{MeOH}/\text{H}_2\text{O}$ mixture at room temperature followed by removal of

the Boc protecting group using a solution of dry HCl in AcOEt. Physical data for **4**, in particular the optical rotation, were in good agreement with the values reported in literature.^{9a}

EXPERIMENTAL SECTION

Melting points were determined on a Kofler hot-stage apparatus and are uncorrected. Optical rotations were measured at 22 °C with a Schmidt-Haensch Polartronic D polarimeter (1 dm-cell) in 2% CHCl₃ (**1-3**) or H₂O (**4**) solutions. IR spectra were recorded on a Perkin-Elmer 983 spectrophotometer as KBr disks. ¹H and ¹³C NMR spectra were obtained on a Varian XL-300 spectrometer using CDCl₃ (**1-3**) or D₂O (**4**) as solvent and Me₄Si or 3-(trimethylsilyl)propionic acid-d₄ sodium salt as internal standard, respectively. Elemental analyses were performed by Servizio Microanalisi del C.N.R., Area della Ricerca di Roma, 00016 Monterotondo Stazione, Roma, Italy.

***N*-Boc-4-formyl-L-phenylalanine Methyl Ester (2).** A mixture of *N*-Boc-L-tyrosine triflate methyl ester (**1a**, 427 mg, 1 mmol), Pd(OAc)₂ (7 mg, 0.03 mmol), dppp (12 mg, 0.03 mmol), and Et₃N (0.35 mL, 2.5 mmol) in DMF (5 mL) was purged with carbon monoxide for 10 min. Then, trioctylsilane (0.9 mL, 2 mmol) was introduced in one portion and the mixture was stirred under a CO balloon at 70 °C for 4 h. After dilution with water, the mixture was extracted with ether. The organic phase was washed with water, saturated NaHCO₃, and water, dried (Na₂SO₄), and evaporated. The residue (964 mg) was chromatographed on silica gel (30 g) using hexane/AcOEt = 75/25 as eluent to give 261 mg (85%) of **2**: mp 91-92 °C (CH₂Cl₂/hexane); [α]_D +64°; IR 3377, 1759, 1689, 1527, 1351, 1167 cm⁻¹; ¹H NMR δ 1.41 (9H, s, *t*-Bu), 3.11 (1H, dd, *J* = 13.9, 6.6 Hz, CHH), 3.24 (1H, dd, *J* = 13.9, 5.5 Hz, CHH), 3.73 (3H, s, CO₂Me), 4.64 (1H, m, α-CH), 5.05 (1H, d, *J* = 7.7 Hz, NH), 7.32 (2H, d, *J* = 8.0 Hz, ArH), 7.82 (2H, d, *J* = 8.0 Hz, ArH), 9.99 (1H, s, CHO); ¹³C δ 28.27, 38.70, 52.40, 54.20, 80.18, 129.93, 130.06, 135.34, 143.45, 169.81, 171.88, 191.83. Anal. Calcd for

C₁₆H₂₁NO₅ (307.4): C, 62.52; H, 6.88; N, 4.56. Found: C, 62.55; H, 7.08; N, 4.66.

The aldehyde **2** could be prepared and isolated in a similar manner using *N*-Boc-4-iodo-L-phenylalanine methyl ester (**1b**, 405 mg, 1 mmol) as the starting material. In this case, 5 mol% of Pd(OAc)₂ and dppp were employed. The reaction time was 4 h and the yield of **2** was 89%.

***N*-Boc-4-hydroxymethyl-L-phenylalanine Methyl Ester (3).** To a stirred solution of **2** (307 mg, 1 mmol) in CH₂Cl₂ (5 mL) NaBH₄ (45 mg, 1.2 mmol) was added in one portion, and MeOH (1 mL) was then added dropwise. The mixture was stirred at room temperature for 0.5 h, then treated with 10% aqueous acetic acid, and extracted with AcOEt. The organic phase was washed twice with water, dried (Na₂SO₄), and evaporated. The residue (328 mg) was chromatographed on silica gel (10 g) using hexane/AcOEt = 65/35 as eluent to give 280 mg (91%) of **3**: mp 82–83 °C (CH₂Cl₂/hexane); [α]_D +49°; IR 3491, 3445, 1748, 1702, 1512, 1358, 1170 cm⁻¹; ¹H NMR δ 1.42 (9H, s, *t*-Bu), 2.60 (1H, br, OH), 3.05 (1H, dd, *J* = 13.7, 5.9 Hz, CHH), 3.12 (1H, dd, *J* = 13.7, 5.9 Hz, CHH), 3.72 (3H, s, CO₂Me), 4.58 (1H, m, α-CH), 4.67 (2H, s, CH₂OH), 4.98 (1H, d, *J* = 7.9 Hz, NH), 7.12 (2H, d, *J* = 8.0 Hz, ArH), 7.30 (2H, d, *J* = 8.0 Hz, ArH); ¹³C δ 28.29, 38.00, 52.24, 54.40, 65.04, 79.97, 127.24, 129.49, 135.42, 139.64, 155.08, 172.30. Anal. Calcd for C₁₆H₂₃NO₅ (309.4): C, 62.12; H, 7.49; N, 4.53. Found: C, 62.11; H, 7.69; N, 4.61.

One-Pot Preparation of 3. After completion of the carbonylation reaction on either **1a** or **1b**, the mixture was flushed with N₂ for 5 min and cooled at 0 °C. NaBH₄ (57 mg, 1.5 mmol) was added in one portion, and MeOH (1 mL) was then added dropwise. The mixture was stirred at room temperature for 1 h, then worked-up as described above to give, after chromatography, 243 mg (79%) or 262 mg (85%) of **3**, respectively.

4-Hydroxymethyl-L-phenylalanine (4). A solution of **3** (309 mg, 1 mmol) in MeOH (3 mL) and 2N NaOH (1 mL) was stirred at room temperature for 4 h, concentrated, acidified with a slight excess of 2N HCl, and extracted with AcOEt. The organic phase was washed twice with water, dried (Na₂SO₄), and

evaporated. To a solution of the residue (300 mg) in AcOEt (2 mL) was added a saturated solution of anhydrous HCl in AcOEt (4.5 mL). The solution was stirred at room temperature for 3 h and evaporated to obtain a white solid (245 mg). The crude hydrochloride was dissolved in water (20 mL) and passed through a column of the anion exchange resin Amberlite IRA-93 (6 g). The effluent was evaporated to dryness (after decolourization with activated charcoal) to give 191 mg (98%) of **4**: mp 205-207 °C (EtOH/H₂O); [α]_D -32° (lit.^{9a} 213-215 °C; [α]_D -32.5°); IR 3291, 1601, 1485, 1402, 1325 cm⁻¹; ¹H NMR δ 3.11 (1H, dd, J = 14.4, 7.9 Hz, CHH), 3.27 (1H, dd, J = 14.4, 5.1 Hz, CHH), 3.98 (1H, m, α -CH), 4.62 (2H, s, CH₂OH), 7.31 (2H, d, J = 7.8 Hz, ArH), 7.39 (2H, d, J = 7.8 Hz, ArH); ¹³C NMR δ 38.74, 58.66, 66.26, 130.82, 132.30, 137.21, 142.17, 176.56.

REFERENCES AND NOTES

1. Groundwater, P. W., Solomons, K. R. H., Drewe, J. A., and Munawar, M. A. in "Progress in Medicinal Chemistry", Ellis, G. P. and Luscombe, D. K. Eds., Elsevier, Amsterdam, **1996**, Vol 33, pp. 233-329. Fry, D. W. in "Annual Reports in Medicinal Chemistry", Bristol, J. A. Editor-in-Chief, Academic Press, San Diego, **1996**, Vol 31, pp. 151-160.
2. Gishizky, M. L. in "Annual Reports in Medicinal Chemistry", Bristol, J. A. Editor-in-Chief, Academic Press, San Diego, **1995**, Vol 30, pp. 247-253 and references therein.
3. Gregory, H., Hardy, P. M., Jones, D. S., Kenner, G. W., and Sheppard, R. C. *Nature* **1964**, 204, 931.
4. Yajima, H., Mori, Y., Koyama, K., Tobe, T., Setoyama, M., Adachi, H., Kanno, T., and Saito, A. *Chem. Pharm. Bull.* **1976**, 24, 1110. Mutt, V. and Jorpes, J. E. *Eur. J. Biochem.* **1968**, 6, 156.
5. Liu, M.-C. and Lipmann, F. *Proc. Natl. Acad. Sci. U.S.A.* **1985**, 82, 34.
6. Nachman, R. J., Holman, G. M., Haddon, W. F., and Hayes, T. K. *Pept. Res.* **1989**, 2, 171. Nachman, R. J., Holman, G. M., Cook, B. J., Haddon,

- W. R., and Ling, N. *Biochem. Biophys. Res. Commun.* **1986**, *140*, 357.
- Nachman, R. J., Holman, G. M., Haddon, W. F., and Ling, N. *Science* **1986**, *234*, 71.
7. (a) Lai, J. H., Marsilje, T. H., Choi, S., Nair, S. A., and Hangauer, D. G. *J. Pept. Res.* **1998**, *51*, 271 and references therein. (b) Gonzalez-Muniz, R., Cornille, F., Bergeron, F., Ficheux, D., Pothier, J., Durieux, C., and Roques, B. P. *Int. J. Pept. Protein Res.* **1991**, *37*, 331. (c) Tilley, J. W., Ramakanth, S., Wagner, R., and Mulkerins, K. *J. Org. Chem.* **1990**, *55*, 906. (d) Marseigne, I. and Roques, B. P. *J. Org. Chem.* **1988**, *53*, 3621.
8. Kim, M. H., Lai, A. H., and Hangauer, D. G. *Int. J. Pept. Protein Res.* **1994**, *44*, 457.
9. (a) Smith, S. C. and Sloane, N. H. *Biochim. Biophys. Acta* **1967**, *148*, 414. (b) Plaut, G. W. E. *J. Med. Chem.* **1965**, *8*, 554.
10. Morera, E. and Ortar, G. *Synlett* **1997**, 1403. Ciattini, P. G., Morera, E., and Ortar, G. *Tetrahedron Lett.* **1995**, *36*, 4133.
11. Kotsuki, H., Datta, F. K., and Suenaga, H. *Synthesis* **1996**, 470.
12. Shieh, W.-C. and Carlson, J. A. *J. Org. Chem.* **1992**, *57*, 379.
13. Lei, H., Stoakes, M. S., Herath, K. P. B., Lee, J., and Schwabacher, A. W. *J. Org. Chem.* **1994**, *59*, 4206.
14. Dppp refers to 1,3-bis(diphenylphosphino)propane.
15. Hartman, G. D. and Halczenko, W. *Synth. Commun.* **1991**, *21*, 2103.
16. Baillargeon, V. P. and Stille, J. K. *J. Am. Chem. Soc.* **1986**, *108*, 452.
17. Sasaki, N. A., Hashimoto, C., and Potier, P. *Tetrahedron Lett.* **1987**, *28*, 6069. Brown, M. S. and Rapoport, H. *J. Org. Chem.* **1963**, *28*, 3261 and references therein.
18. In the one-pot procedure the use of a 1.5 molar excess of NaBH₄ and a reaction time of 1 h were found necessary for the complete reduction of the aldehyde without appreciable reduction of the ester function.

(Received in the U.S.A. 15 May 1998)