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Synthesis, antifungal activity of caffeic acid derivative esters, and their synergism with fluconazole and nystatin against *Candida* spp.

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Abstract

We tested the antifungal potential of caffeic acid and eight of its derivative esters against *C. albicans* ATCC 90028 and nine clinical isolates, and carried out a synergism assay with fluconazole and nystatin. Propyl caffeate (**C3**) showed the best antifungal activity against the tested strains. When in combination, **C3** markedly reduced the MIC of fluconazole and nystatin with synergistic effect up to 64-fold. Finally, **C3** showed a high IC₅₀ value and Selective Index (SI) against oral keratinocytes, demonstrating low toxicity against this cell type and selectivity for yeast cells. Further research should confirm its antifungal potential for development of combined therapy to treat *C. albicans* infections.

Introduction

Oral candidiasis is one of the most common opportunistic infections afflicting humans, with *Candida albicans* as the major causative agent of this disease (Garcia-Cuesta *et al.*, 2014). The complexity of interactions between *Candida* and other microorganisms in the host, mainly bacteria, suggest that several mechanisms are involved in yeast fitness to the oral cavity. Some studies have shown that *Candida* spp. can co-aggregate with bacteria in dental plaque. This feature may be an important factor for the onset of oral candidiasis as well as fungal colonization of carious cavities and periodontal pockets (Sardi *et al.*, 2012; Thurnheer *et al.*, 2015). The presence of yeasts in subgingival regions may contribute to the pathogenesis of periodontal disease or increase the chance of candidemia, especially in cases of immunosuppression (Hannula *et al.*, 2001; Reynaud *et al.*, 2001; Al Mubarak *et al.*, 2013). In addition, it has been well documented that systemic diseases such as diabetes and AIDS; physiological conditions such as pregnancy, infancy or old age; nutritional factors; treatment with broad-spectrum antibiotics; use of immunosuppressive drugs and corticosteroids; xerostomia, and use of dentures, may predispose the individual to develop candidiasis (Soll, 2002; Tekeli *et al.*, 2004; Manfredi *et al.*, 2006).

The current therapy with antifungals has serious drawbacks, in particular due to toxic effects to human cells and adverse effects (Epstein *et al.*, 2002; Gabler *et al.*, 2008). As the drugs used to treat candidiasis are not always specific and properly prescribed (targeting the causative agent of infection), there has been a significant increase in resistance of *Candida* spp. to traditional antifungal drugs. The increasing microbial resistance rates may also be a result of long-term drug exposure or selection of strains with

intrinsic resistance mechanisms (Ying *et al.*, 2013; Fernandez-Ruiz *et al.*, 2015; Liao *et al.*, 2015; Seifi *et al.*, 2015; Freitas *et al.*, 2015). Therefore, the development of novel strategies to minimize the toxic effects of current antifungals and improve their effectiveness, has been strongly encouraged.

Natural products have continued to be a rich source of new drugs with clinically significant biological targets. Over the past 34 years, 49% of FDA-approved chemotherapeutic drugs were either natural products or directly derived therefrom (Newman & Cragg, 2016). There is a great interest of the pharmaceutical industry in the discovery of new molecules of natural origin or even their combination with existing drugs, in order to improve efficacy, potency, safety, tolerability, and decrease production costs, side effects and selection of resistant strains (Svetaz *et al.*, 2016). A number of studies in the literature have established the value of combined antifungal therapy against resistant strains, in particular standard drugs with naturally-occurring agents (Pippi *et al.*, 2015; Han *et al.*, 2016).

Caffeic acid (3,4-dihydroxycinnamic acid) is an important phenolic compound commonly found in plants, foods and propolis samples, particularly in the form of caffeic acid phenethyl ester (CAPE) (Paracatu *et al.*, 2014; Rzepecka-Stojko *et al.*, 2015). It is better known for its pharmacological properties, including antimicrobial, antioxidant, anti-inflammatory and anticancer (Balachandran *et al.*, 2012; Kuo *et al.*, 2015). Nevertheless, modifications of the caffeic acid structure into esters or amides, for instance, may generate novel analog molecules with enhanced and desired biological activity (Touaibia *et al.*, 2011), particularly as antimicrobials (Fu *et al.*, 2010).

Herein, we investigated the antifungal potential of caffeic acid and eight of its derivative esters against *C. albicans* ATCC 90028 and nine oral clinical isolates. The most active molecule, propyl caffeate (**C3**), was selected for a synergism assay with fluconazole and nystatin against the *C. albicans* strains, and tested for its toxicity on oral keratinocytes (NOK cells).

Material and Methods

Synthesis of Esters. Caffeic acid (0.2 mM) solution and corresponding alcohols (20 mM) were prepared at 5°C with a solution of *N, N'*-dicyclohexylcarbodiimide (DCC, 1.0 mM) in *p*-dioxane (3.0 mL). After the solution was stirred for 48 h, the solvent was removed under reduced pressure. The residue was partitioned three-times with EtOAc and filtered. The

filtrate was serially washed with saturated aqueous citric acid solution (three times), saturated aqueous NaHCO_3 (three-times), water (two-times), dried over MgSO_4 , and evaporated under reduced pressure. The crude products were purified over a silica gel column using an isocratic system of CHCl_3 –MeOH (98:2). The modifications made in caffeic acid molecule are shown in Table 1.

Microorganisms. *Candida albicans* ATCC 90028 strain and nine highly virulent clinical isolates of *C. albicans* obtained from the oral cavity of patients with diabetes and periodontitis (Sardi *et al.*, 2012), were used in this study. This study was approved by the research ethics committee at Piracicaba Dental School, University of Campinas, Piracicaba, SP, Brazil (protocol no. 062/2008).

Determination of Antifungal Activity. The Minimum Inhibitory Concentration (MIC) of caffeic acid and its eight derivatives against *C. albicans* ATCC 90028 was determined using 96-well microplates based on the protocol M27-A3 of the CLSI (2008), with modifications. The esters that showed the lowest MIC values against *C. albicans* ATCC 90028 (C3, C4 and C6) were then tested against nine clinical isolates of *C. albicans*. The synthetic compounds of caffeic acid were diluted in DMSO and tested in concentrations ranging from 250 $\mu\text{g/mL}$ to 0.48 $\mu\text{g/mL}$ (Scorzoni *et al.*, 2007). The inoculum was prepared (λ 530 nm, Abs 0.08–0.1) and diluted to 2.5×10^3 CFU/mL. The plates were incubated at 35°C for 24 h. The MIC₁₀₀ was determined as the lowest concentration of the compound inhibiting visible fungal growth as indicated by 0.1% resazurin (Sigma-Aldrich, St. Louis, MO, USA). Aliquots from the wells corresponding to the MIC and higher concentrations were sub-cultured on Sabouraud Dextrose Agar (Difco®, Detroit, MI, USA) for determination of the Minimum Fungicidal Concentration (MFC). The MFC was defined as the lowest concentration of the compound causing no visible growth on the agar plate.

Combinatorial Antifungal Activity (Synergism Assay). The ester which showed the best activity against *C. albicans* strains (C3) was combined with conventional antifungals commonly used for the treatment of candidiasis, fluconazole and nystatin. Their combinatorial antifungal activity was determined through the checkerboard method using 96-well microplates (Dai *et al.*, 2015). A mathematical calculation was used to generate the fractional inhibitory concentration index (FICI), as follows: $\text{FICI} = (\text{MIC compound 1 in combination} / \text{MIC compound 1 alone}) + (\text{MIC compound 2 in combination} / \text{MIC compound 2 alone})$. The combinations were classified as synergistic ($\text{FICI} \leq 0.5$), additive

($0.5 < \text{FICI} \leq 1.0$), indifferent ($1.0 < \text{FICI} < 4.0$) and antagonistic ($\text{FICI} \geq 4.0$) (Soares *et al.*, 2014).

Cytotoxic Effects on Oral Keratinocytes. The most active ester derivative (**C3**) as well as fluconazole and nystatin were tested for their cytotoxicity against keratinocytes from the oral mucosa of humans (NOK cells) provided by the Department of Medicine at Harvard Medical School (Dr. Karl Munger). NOK cells were maintained in culture medium for keratinocytes without fetal bovine serum (Gibco, Life Technologies) at 36.5°C and 5% CO_2 . Cells were seeded onto a 96-well plate at a density of 5×10^4 cells/well for 24 h. Then the cells were exposed to the treatments (**C3**, Fluconazole, Nystatin, and their combinations) for 24 h. Cell viability was determined by adding aliquots of 10 μL of MTT solution (5 mg/mL) (Sigma-Aldrich, St. Louis, MO) to each well. The plates were incubated at 37°C for 4 h to allow for visualization of precipitated formazan crystals. Aliquots of 100 μL of isopropyl alcohol were added *per* well and absorbance was read using a spectrophotometer at 560 nm (Mosmann *et al.*, 1983). Hydrogen peroxide (10%) was used as a positive control for cytotoxicity and untreated cells were considered as the negative control. Based on this cell viability assay, we next determined the half maximal inhibitory concentration (IC_{50}) for the esters, standard-drugs and their combinations. The IC_{50} was defined as the effective concentration of the compound able to inhibit 50% of the NOK cells. The establishment of this concentration was essential to calculate the selectivity index (SI) ($\text{IC}_{50}/\text{MIC}$ ratio), which was used as an indicator of potential toxicity against normal cells lines at the effective therapeutic concentration for each strain. Hence, the selectivity index (SI) indicated the relationship between drug toxicity against *C. albicans* and the host cells (Gullo *et al.*, 2012; Mora-Navarro *et al.*, 2016).

Statistical Analysis. All assays were performed in triplicate of independent experiments. The data were analyzed on Graphpad Prism 5.0 (San Diego, CA, USA) by one-way analysis of variance (ANOVA) followed by Bonferroni's multiple comparison test, with a significance level of 5%.

Results

Structure-Activity Relationship of Caffeic Acid Derivatives. A screening for antifungal activity of caffeic acid and eight derivative esters against *C. albicans* was carried out.

These esters differ by the number of constituent carbons in the molecule, as shown in Table 1. Caffeic acid and all of its synthesized derivatives showed a fungicidal activity against *C. albicans* ATCC 90028 and presented a structure-activity relationship. As seen in Table 2, the caffeic acid molecule (**C0**) without structural change and the derived **C1** showed MIC and MFC values of 125 µg/mL. The derivatives **C2**, **C5**, **C7** and **C8** showed MIC and MFC values of 31.25 µg/mL. **C3** and **C4** showed strong fungicidal activity against *C. albicans* ATCC 90028, with MIC and MFC values of 15.62 µg/mL, together with **C6** which showed MIC and MFC equal to 7.81 µg/mL. Thus, the compounds **C3**, **C4** and **C6** – which were better than their originating caffeic acid molecule – were further tested against nine oral clinical isolates of *C. albicans*. As shown in Table 3, the compound **C3** demonstrated the best activity when compared with the other caffeic acid derivatives (**C4** and **C6**), with MIC and MFC values ranging between 7.81 and 62.5 µg/mL. The MIC and MFC of **C4** and **C6** ranged between 31.25-125 µg/mL and 7.81-≥250 µg/mL, respectively.

Combinatorial Antifungal Activity with Standard Drugs. Given the strong antifungal activity of **C3** against *C. albicans* strains, we next investigated its combinatorial activity with the standard drugs fluconazole and nystatin. A synergistic activity was observed for the combination between **C3** and fluconazole against most strains, with FICI values ranging from 0.06 to 0.5. Indifferent activity was observed against two clinical isolates, with FICI values close to 2.0. When comparing the MIC values of **C3** and fluconazole alone and combined, we observed a decrease in their MIC and potentiation of fluconazole of 2 to 64-fold (Table 4). The combination of **C3** with nystatin was advantageous (synergistic) against three out of the ten strains when compared with the MIC values of the compounds tested alone, with potentiation of nystatin by 8- to 64-fold. Additive activity was observed against four clinical isolates and *C. albicans* ATCC 90028 strain, with FICI values ranging from 0.53 to 1.0. The additive activity of the combination showed potentiation of nystatin between 32- and 64-fold.

Toxic Effects on Oral Keratinocytes. A viability assay was carried out to evaluate the toxicity of **C3** and standard drugs alone and in combination, against NOK cells. The antifungal compound **C3** showed low toxicity at the tested concentrations, with a high IC₅₀ value of 420 µg/mL. Likewise, fluconazole and nystatin showed high IC₅₀ values of 320

µg/mL and 400 µg/L, respectively. When combined, both **C3** and the antifungals had their SI value increased, indicating selectivity for yeasts rather than NOK cells (Table 5).

Figure 1 shows the percentage of cell viability of NOK cells treated with the antifungal compounds at different concentrations. The data showed that treatment with **C3**, fluconazole and nystatin maintained over 80% cell viability at all tested concentrations. Hydrogen peroxide, used as a positive control, markedly affected cell viability, with a significant difference when compared to **C3** and standard drugs ($P < 0.05$).

Discussion

Opportunistic infections caused by *Candida* spp. have still been considered a recurrent health issue with high burden worldwide (Rodriguez-Tudela *et al.*, 2015; Denning & Gugnani, 2015). Thus, novel therapeutic approaches are much needed to treat *Candida* infections, including the use of naturally-occurring agents. In this study, we demonstrate the antifungal potential of caffeic acid derivatives against *C. albicans* and their successful synergism with antifungals commercially available.

Caffeic acid is commonly found in fruits (Balachandran *et al.*, 2012) and propolis samples (Freires *et al.*, 2016) used in daily life products and folk medicine. We showed that caffeic acid molecule has fungicidal activity against *C. albicans* ATCC 90028. Nevertheless, the structural modifications performed in this study rendered the caffeic acid molecule much more effective in terms of fungicidal activity, showing a structure-activity relationship. The addition of 2 to 8 carbon atoms in the caffeic acid molecule increased its anti-*Candida* activity. Other studies have also demonstrated the relationship between the number of carbons and the antifungal activity of the molecule (Nihei *et al.*, 2003; Soares *et al.*, 2014). Among the synthesized compounds, **C3** showed better activity when compared with other caffeic acid derivatives (**C4** and **C6**), showing lower minimum inhibitory and fungicidal concentrations against most strains. Balachandran *et al.* (2012) isolated methyl caffeate from *Solanum torvum* plant and showed antibacterial activity against Gram-negative and Gram-positive bacteria, in addition to antifungal activity against *C. albicans* and *Aspergillus flavus*. These findings confirm the antimicrobial potential of caffeic acid derivatives.

Of note, the clinical isolates used in this study showed lower susceptibility to nystatin and fluconazole when compared with the reference strain of *C. albicans* ATCC

90028. This could be due to acquisition of resistance mechanisms as the isolates belonged to diabetic patients. A recent study showed that this group of patients is prone to develop *Candida* infections in cases of poor glycemic control (Zomorodian *et al.*, 2016).

Microbial resistance has raised concern over the last decades as the investment in antibiotic discovery is declining considerably over time when compared to high-priced drugs such as chemotherapeutics (Bax & Green, 2015). To make it worse, failure in prescribing the appropriate drug, misuse, and long hospital stay have led to the emergence of azole-resistant isolates, particularly in cases of invasive infections (Liao *et al.*, 2015; Sanguinetti, Posteraro & Lass-Flörl, 2015). This opens avenues for the development of combined antifungal therapy, in which drugs with different mechanisms of action (or not) are combined to enhance their antifungal potency and avoid selection of resistant strains. Here, we demonstrated the successful combination of propyl caffeate (**C3**) with fluconazole and nystatin against *C. albicans* strains. A study performed by Lee & Han (2005) demonstrated the synergistic effect of the combination between amphotericin B and berberine (alkaloid) against *C. albicans*. These authors also tested this combination in mice with candidemia and observed that the combined treatment prolonged the lives of mice in 22 days compared to those treated with amphotericin B alone. Other studies have reported the successful combination of essential oils with conventional drugs, including nystatin, fluzonazole and micafugin (Aprotosoiaie *et al.*, 2008; Rosato *et al.*, 2009; Rodrigues *et al.*, 2012; Stringaro *et al.*, 2014).

With the purpose of future clinical use, we also investigated the effects of **C3** against oral keratinocytes and compared them with those of the standard drugs. NOK cells were chosen for this study as they constitute the main epithelial cell type lining the oral mucosa, which would be highly exposed in case of administration of an oral suspension or a solution for oral candidiasis. Overall, low toxicity was found for **C3**, fluconazole and nystatin, with IC₅₀ values higher than their MIC/MFC. These agents also showed a high SI value, which means that if the index is greater than 10.0 the compound exhibits selectivity for killing yeasts rather than the host's cells. The combination between **C3** and fluconazole led to a considerable increase in their SIs, highlighting their selectivity for yeast cells.

Collectively, our findings indicate that the propyl ester modification of caffeic acid molecule (**C3**) has strong fungicidal activity and potentiated the effects of fluconazole and nystatin against *C. albicans* strains, with little effects against oral keratinocytes. Further

studies should focus on the effects of these combinations against *Candida* biofilms and establish C3 mechanism(s) of action.

The authors declare no conflict of interest.

References

Al Mubarak S, Robert AA, Baskaradoss JK, Al-Zoman K, Al Sohail A, Alsuwyed A, Ciancio S (2013) The prevalence of oral *Candida* infections in periodontitis patients with type 2 diabetes mellitus. *J Infect Public Health*. 6: 296-301.

Aprotosoia AC, Hăncianu M, Poiată A, Tuchiluş C, Spac A, Cioană O, Gille E, Stănescu U (2008) In vitro antimicrobial activity and chemical composition of the essential oil of *Foeniculum vulgare* Mill. *Rev Med Chir Soc Med Nat Iasi*. 112: 832-836.

Balachandran C, Duraipandiyar V, Al-Dhabi NA, Balakrishna K, Kalia NP, Rajput VS, Khan IA, Ignacimuthu S (2012) Antimicrobial and Antimycobacterial Activities of Methyl Caffeate Isolated from *Solanum torvum* Swartz. *Fruit Indian J Microbiol*. 52: 676-681.

Bax R, Green S (2015) Antibiotics: the changing regulatory and pharmaceutical industry paradigm. *J Antimicrob Chemother*. 70: 1281-1284.

Clinical and Laboratory Standards Institute (CLSI) (2008) Protocol M27-A3. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts. 3.ed. Pennsylvania: CLSI.

Cordeiro RA, Teixeira CE, Brilhante RS, Castelo-Branco DS, Alencar LP, de Oliveira JS, Monteiro AJ, Bandeira TJ, Sidrim JJ, Moreira JL, Rocha MF (2015) Exogenous tyrosol inhibits planktonic cells and biofilms of *Candida* species and enhances their susceptibility to antifungals. *FEMS Yeast Res* 15: doi: 10.1093/femsyr/fov012.

Dai L, Zang C, Tian S, Liu W, Tan S, Cai Z, Ni T, An M, Li, R, Gao Y, Zhang D, Jiang Y (2015) Design, synthesis, and evaluation of caffeic acid amides as synergists to sensitize fluconazole-resistant *Candida albicans* to fluconazole. *Bioorg Med Chem Lett* 25:34-37.

Denning DW, Gugnani HC (2015) Burden of serious fungal infections in Trinidad and Tobago. *Mycoses*. 58: 80-84.

Epstein JB, Dawson JR, Buivids IA, Wong B, Le ND (2002) The effect of a disinfectant/coolant irrigant on microbes isolated from dental unit water lines. *Spec Care Dentist* 22:137-141.

Fernández-Ruiz M, Puig-Asensio M, Guinea J, Almirante B, Padilla B, Almela M, Díaz-Martín A, Rodríguez-Baño J, Cuenca-Estrella M, Aguado JM, CANDIPOP Project; GEIH-GEMICOMED (SEIMC); REIPI. (2015) *Candida tropicalis* bloodstream infection: Incidence, risk factors and outcome in a population-based surveillance. *J Infect* 71: 385-394.

Freires IA, Queiroz VC, Furletti VF, Ikegaki M, de Alencar SM, Duarte MC, et al. (2016) Chemical composition and antifungal potential of Brazilian propolis against *Candida* spp. *J Mycol Med*. doi: 10.1016/j.mycmed.2016.01.003.

Freitas EM, Monteiro LC, Fernandes MB, Martelli Junior H, Bonan PR, Nobre SA (2015) Antifungal susceptibility in vitro determined by the Etest® for *Candida* obtained from the oral cavity of irradiated and elderly individuals. *Braz Dent J* 26: 99-104.

Fu J, Cheng K, Zhang Z, Fang R, Zhu H (2010) Synthesis, structure and structure–activity relationship analysis of caffeic acid amides as potential antimicrobials. *Eur J Med Chem* 45:2638-2643.

Gabler IG, Barbosa AC, Velela RR, Lyon S, Rosa CA (2008) Incidence and anatomic localization of oral candidiasis in patients with AIDS hospitalized in a public hospital in Belo Horizonte, MG, Brazil. *J Appl Oral Sci* 16: 247-250.

Garcia-Cuesta C, Sarrion-Pérez MG, Bagán JV (2014) Current treatment of oral candidiasis: A literature review. *J Clin Exp Dent* 6: e576-82.

Gullo FP, Sardi JC, Santos VA, Sangalli-Leite F, Pitangui NS, Rossi SA, de Paula E Silva AC, Soares LA, Silva JF, Oliveira HC, Furlan M, Silva DH, Bolzani VS, Mendes-Giannini MJ, Fusco-Almeida AM (2012) Antifungal activity of maytenin and pristimerin. *Evid Based Complement Alternat Med* 2012:340787.

Han Y, Lee JH (2005) Berberine synergy with amphotericin B against disseminated candidiasis in mice. *Biol Pharm Bull* 28: 541-544.

Han B, Chen J, Yu Y, Cao Y, Jiang Y (2016) Antifungal activity of *Rubus chingii* extract combined with fluconazole against fluconazole-resistant *Candida albicans*. *Microbiol Immunol* 60: 82-92.

Hannula J, Dogan B, Slots J, Okte E, Asikainen S (2001) Subgingival strains of *Candida albicans* in relation to geographical origin and occurrence of periodontal pathogenic bacteria. *Oral Microbiol Immunol* 16: 113-118.

Kuo YY, Jim WT, Su LC, Chung CJ, Lin CY, Huo C, Tseng JC, Huang SH, Lai CJ, Chen BC, Wang BJ, Chan TM, Lin HP, Chang WS, Chang CR, Chuu CP (2015) Caffeic Acid phenethyl ester is a potential therapeutic agent for oral cancer *Int J Mol Sci* 16: 10748-10766.

Liao X, Qiu H, Li R, Guo F, Liu W, Kang M, Kang Y, China-SCAN Team (2015) Risk factors for fluconazole-resistant invasive candidiasis in intensive care unit patients: An analysis from the China Survey of Candidiasis study. *J Crit Care* 30: 862.e1-5.

Manfredi M, McCullough MJ, Al-Karaawi ZM, Vescovi P, Porter SR (2006) In vitro evaluation of virulence attributes of *Candida* spp. isolated from patients affected by diabetes mellitus. *Oral Microbiol Immunol* 21:183-189.

Mora-Navarro C, Méndez-Vega J, Caraballo-León J, Lee M-r, Palecek S, Torres-Lugo M, *et al.* (2016) Hydrophobicity of Antifungal β -Peptides Is Associated with Their Cytotoxic Effect on *In Vitro* Human Colon Caco-2 and Liver HepG2 Cells. *PLoS ONE* 11(3): e0149271. doi:10.1371/journal.pone.0149271.

Newman DJ, Cragg GM (2016) Natural Products as Sources of New Drugs over the 30 Years from 1981 to 20140. J Nat Prod. DOI: 10.1021/acs.jnatprod.5b01055. [In press].

Nihei K, Nihei A, Kubo I (2003) Rational design of antimicrobial agents: antifungal activity of alk(en)yl dihydroxybenzoates and dihydroxyphenyl alkanoates. Bioorg Med Chem Lett 13: 3993-3996.

Paracatu LC, Bonacorsi C, de Farias CM, Nazare AC, Petronio MS, Regasini LO, Silva DH, Raddi MS, da Fonseca LM, Ximenes VF (2014) Alkyl caffeates as anti-*Helicobacter pylori* and scavenger of oxidants produced by neutrophils. Med Chem. 10: 74-80.

Pippi B, Lana AJD, Moraes RC, Güez CM, Machado M, Oliveira LFS, *et al.* (2015) *In vitro* evaluation of the acquisition of resistance, antifungal activity and synergism of Brazilian red propolis with antifungal drugs on *Candida* spp. J Appl Microbiol. 118:839-850.

Reynaud AH, Nygaard-Østby B, Bøygard GK, Eribe ER, Olsen I, Gjermo P (2001) Yeasts in periodontal pockets. J Clin Periodontol 28: 860-864.

Rodrigues FF, Oliveira LG, Rodrigues FF, Saraiva ME, Almeida SC, Cabral ME, Campos AR, Costa JG (2012) Chemical composition, antibacterial and antifungal activities of essential oil from *Cordia verbenacea* DC leaves. Pharmacognosy Res 4: 161-165.

Rodriguez-Tudela JL, Alastruey-Izquierdo A, Gago S, Cuenca-Estrella M, León C, Miro JM, *et al.* (2015) Burden of serious fungal infections in Spain. Clin Microbiol Infect. 21: 183-189.

Rosato A, Vitali C, Piarulli M, Mazzotta M, Argentieri MP, Mallamaci R (2009) *In vitro* synergic efficacy of the combination of Nystatin with the essential oils of *Origanum vulgare* and *Pelargonium graveolens* against some *Candida* species. Phytomedicine 16: 972-975.

Rzepecka-Stojko A, Kabała-Dzik A, Moździerz A, Kubina R, Wojtyczka RD, Stojko R, Dziedzic A, Jastrzębska-Stojko Ż, Jurzak M, Buszman E, Stojko J (2015) Caffeic Acid phenethyl ester and ethanol extract of propolis induce the complementary cytotoxic effect on triple-negative breast cancer cell lines. *Molecules* 20: 9242-9262.

Sanguinetti M, Posteraro B, Lass-Flörl C (2015) Antifungal drug resistance among *Candida* species: mechanisms and clinical impact. *Mycoses* 58:2-13.

Sardi JC, Duque C, Höfling JF, Gonçalves RB (2012) Genetic and phenotypic evaluation of *Candida albicans* strains isolated from subgingival biofilm of diabetic patients with chronic periodontitis. *Med Mycol* 50: 467-475.

Scorzoni L, Benaducci T, Fusco-Almeida AM, Silva DHS, Bolzani VS, Mendes-Giannini MJS (2007) "The use of standard methodology for determination of antifungal activity of natural products against medical yeasts *Candida* sp and *Cryptococcus* sp. *Brazilian Journal Microbiology* 38: 391–397.

Seifi Z, Zarei-Mahmoudabadi A, Zarrin M (2015) Extracellular enzymes and susceptibility to fluconazole in *Candida* strains isolated from patients with vaginitis and healthy individuals. *Jundishapur J Microbiol* 8: e20162.

Soares LA, Gullo FP, Sardi Jde C, Pitangui Nde S, Costa-Orlandi CB, Sangalli-Leite F, Scorzoni L, Regasini LO, Petrônio MS, Souza PF, Silva DH, Mendes-Giannini MJ, Fusco-Almeida AM (2014) Anti-*trichophyton* activity of protocatechuates and their synergism with fluconazole. *Evid Based Complement Alternat Med*. 2014: 957860.

Soll DR (2002) *Candida* commensalism and virulence: the evolution of phenotypic plasticity. *Acta Trop* 81: 101-110.

Stringaro A, Vavala E, Colone M, Pepi F, Mignogna G, Garzoli S, Cecchetti S, Ragno R, Angiolella L (2014) Effects of *Mentha suaveolens* Essential Oil Alone or in Combination with Other Drugs in *Candida albicans*. *Evid Based Complement Alternat Med*. 2014:125904.

Svetaz LA, Postigo A, Butassi E, Zacchino SA, Sortino MA (2016) Antifungal drugs combinations: a patent review 2000-2015. *Exp Opin Therapeut Patent*. DOI: 10.1517/13543776.2016.1146693. [In press].

Tekeli A, Dolapci I, Emral R, Cesur S (2004) *Candida* carriage and *Candida dubliniensis* in oropharyngeal samples of type-1 diabetes mellitus patients. *Mycoses* 47: 315-318.

Thurnheer T, Bostanci N, Belibasakis GN (2015) Microbial dynamics during conversion from supragingival to subgingival biofilms in an in vitro model. *Mol Oral Microbiol*. doi: 10.1111/omi.12108.

Touaibia M, Jean-François J, Doiron J (2011) Caffeic Acid, A Versatile Pharmacophore: An Overview. *Mini-Rev Med Chem* 11:695-713.

Ying Y, Zhao Y, Hu X, Cai Z, Liu X, Jin G, Zhang J, Zhang J, Liu J, Huang X (2013) In vitro fluconazole susceptibility of 1,903 clinical isolates of *Candida albicans* and the identification of ERG11 mutations. *Microb Drug Resist* 19: 266-273.

Zomorodian K, Kavooosi F, Pishdad GR, Mehriar P, Ebrahimi H, Bandegani A, *et al.* (2016) Prevalence of oral *Candida* colonization in patients with diabetes mellitus. *J Mycol Med*. DOI:<http://dx.doi.org/10.1016/j.mycmed.2015.12.008>.

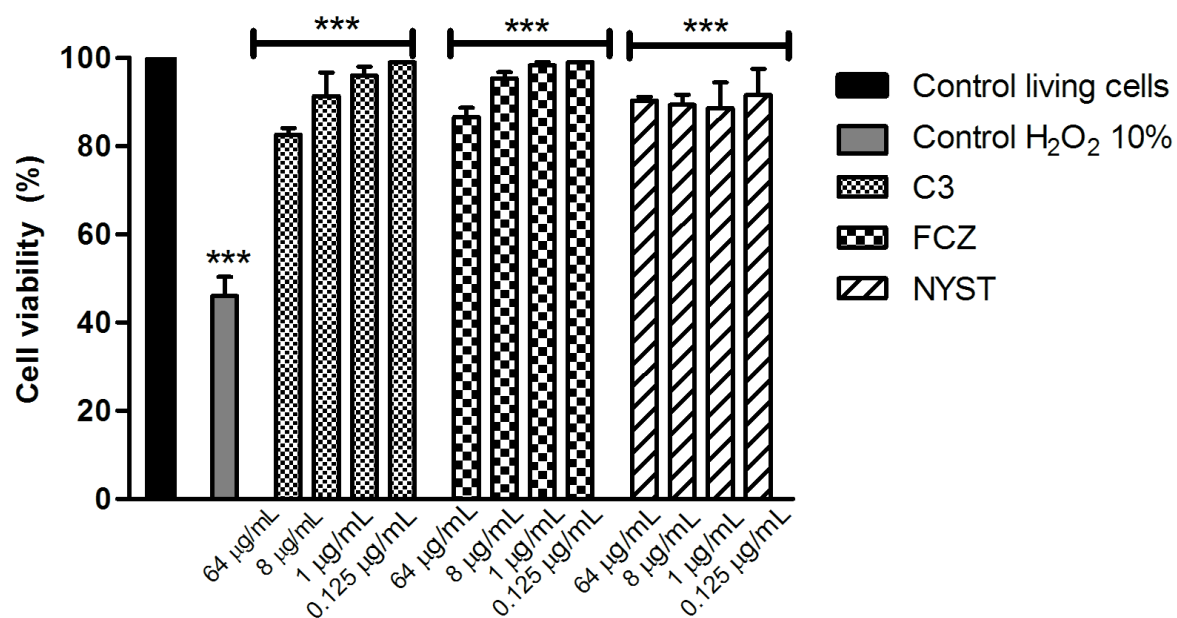
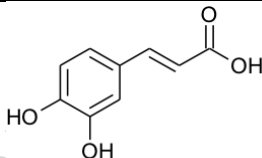
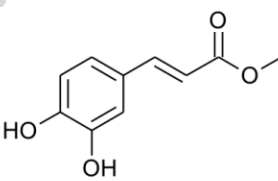
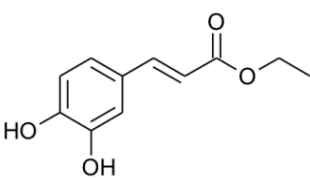
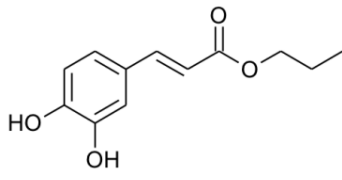
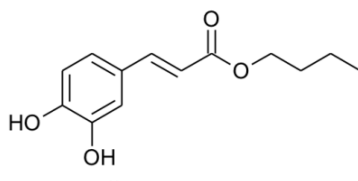
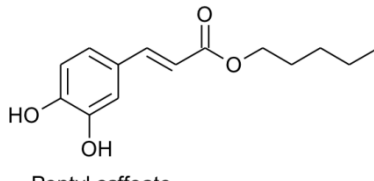
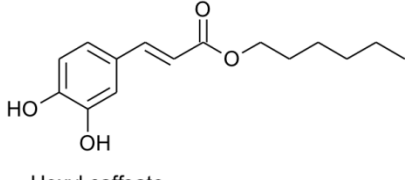
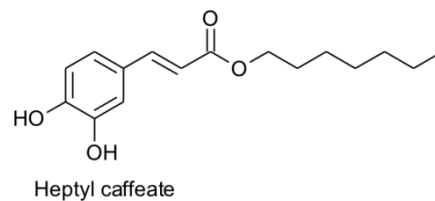


Figure 1

Table 1. Nomenclature, molecular formulas and the chemical structures of the caffeic acid derivative esters tested in this study.

Code	Nomenclature	Molecular Formula	Chemical Structure
C0	Caffeic acid	$C_9H_8O_4$	 <p>Caffeic acid</p>
C1	Methyl caffeate	$C_{10}H_8O_4$	 <p>Methyl caffeate</p>
C2	Ethyl caffeate	$C_{11}H_{12}O_4$	 <p>Ethyl caffeate</p>
C3	Propyl caffeate	$C_{12}H_{14}O_4$	 <p>Propyl caffeate</p>
C4	Butyl caffeate	$C_{13}H_{16}O_4$	 <p>Butyl caffeate</p>
C5	Pentyl caffeate	$C_{14}H_{18}O_4$	 <p>Pentyl caffeate</p>
C6	Hexyl caffeate	$C_{15}H_{20}O_4$	 <p>Hexyl caffeate</p>

C7 Heptyl caffeate $C_{16}H_{22}O_4$



C8 Octyl caffeate $C_{17}H_{24}O_4$

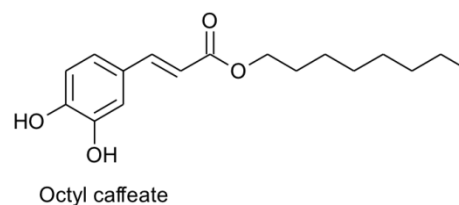


Table 2. Antifungal activity of caffeic acid derivative esters against *Candida albicans* ATCC 90028.

Caffeic Acids Derivates	<i>Candida albicans</i> ATCC 90028	
	MIC ($\mu\text{g/mL}$)	MFC ($\mu\text{g/mL}$)
C0	125	125
C1	125	125
C2	31.25	31.25
C3	15.62	15.62
C4	15.62	15.62
C5	31.25	31.25
C6	7.81	7.81
C7	31.25	31.25
C8	31.25	31.25
Nystatin	4.0	4.0
Fluconazole	0.5	0.5

Table 3. Antifungal activity of caffeic acid derivative esters (**C3**, **C4** and **C6**) against nine oral clinical isolates of *Candida albicans*. MIC/MFC values are expressed in µg/mL.

<i>C. albicans</i> clinical isolates	C3		C4		C6		Fluconazole		Nystatin	
	MIC	MFC	MIC	MFC	MIC	MFC	MIC	MFC	MIC	MFC
Ca#22	7.81	7.81	31.25	31.25	7.81	7.81	8.0	8.0	8.0	8.0
Ca#25	31.25	31.25	62.50	62.50	125	125	8.0	8.0	8.0	8.0
Ca#45	15.62	15.62	31.25	31.25	125	125	8.0	8.0	8.0	8.0
Ca#50	31.25	31.25	125	125	>250	>250	2.0	2.0	4.0	4.0
Ca#61	31.25	31.25	62.50	62.50	>250	>250	0.5	0.5	8.0	8.0
Ca#62	7.81	7.81	31.25	31.25	7.81	7.81	8.0	8.0	8.0	8.0
Ca#63	31.25	31.25	62.50	62.50	>250	>250	8.0	8.0	8.0	8.0
Ca#105	31.25	31.25	125	125	>250	>250	8.0	8.0	8.0	8.0
Ca#124	62.50	62.50	125	125	>250	>250	4.0	4.0	8.0	8.0

Table 4. Antifungal effects of propyl caffeate (**C3**) combined with fluconazole and nystatin against *Candida albicans* ATCC 90028 and nine oral clinical isolates. MIC values are expressed in µg/mL.

<i>C. albicans</i> strains	FCZ + C3				NYST + C3			
	FCZ (MIC)	C3 (MIC)	FICI	Effect /Potentiation FCZ	NYST (MIC)	C3 (MIC)	FICI	Effect /Potentiation NYST
ATCC 90028	0.25	0.12	0.50	Synergistic /2X	0.125	7.81	0.53	Additive/32X
Ca#22	0.125	15.62	2.01	Indifferent/64X	0.125	15.62	2.01	Indifferent/64X
Ca#25	0.5	0.12	0.06	Synergistic/16X	4.0	31.25	1.50	Indifferent/2X
Ca#45	1.0	0.12	0.13	Synergistic/8X	0.5	15.62	1.06	Indifferent/16X
Ca#50	0.5	3.90	0.37	Synergistic/4X	0.5	3.90	0.24	Synergistic/8X
Ca#61	0.25	0.12	0.50	Synergistic/2X	0.125	31.25	1.01	Indifferent/64X
Ca#62	0.125	15.62	2.01	Indifferent/64X	0.125	15.62	2.01	Indifferent/64X
Ca#63	1.0	1.95	0.18	Synergistic/8X	0.125	15.62	0.51	Additive /64X
Ca#105	0.5	0.48	0.07	Synergistic/16X	0.25	31.25	1.03	Indifferent/32X
Ca#124	0.5	0.12	0.12	Synergistic/ 8X	0.25	15.62	0.28	Synergistic/32X

Table 5. IC₅₀ value and Selective Index (SI) of propyl caffeate (**C3**), fluconazole (FCZ) and nystatin (NYST) showing the toxicity of the compounds against yeasts in relationship to human oral keratinocytes (NOK cells).

Compound	IC ₅₀ (µg/mL)	SI (alone)	SI (combination)
C3	420	13.5	26.9 (FCZ) / 13.5 (NYST)
FCZ	320	40	320
NYST	400	50	100

Highlights

- Caffeic acid and eight of its derivative esters were tested for their activity against *Candida* spp.
- Propyl caffeate (**C3**) was the most active compound against reference strain and clinical isolates.
- When combined, **C3** markedly reduced the MIC of fluconazole and nystatin up to 64-fold.
- **C3** showed low toxicity against oral keratinocytes and high selectivity for yeast cells.
- Further research should focus on its combined therapy to treat *C. albicans*-related infections.