# Synthesis and Antiplasmodial Activity of Novel Chloroquine Analogues with Bulky Basic Side Chains

Bruno Tasso,<sup>\*[a]</sup> Federica Novelli,<sup>[a]</sup> Michele Tonelli,<sup>[a]</sup> Anna Barteselli,<sup>[b]</sup> Nicoletta Basilico,<sup>[c]</sup> Silvia Parapini,<sup>[d]</sup> Donatella Taramelli,<sup>[d]</sup> Anna Sparatore,<sup>[b]</sup> and Fabio Sparatore<sup>[a]</sup>

Chloroquine is commonly used in the treatment and prevention of malaria, but *Plasmodium falciparum*, the main species responsible for malaria-related deaths, has developed resistance against this drug. Twenty-seven novel chloroquine (CQ) analogues characterized by a side chain terminated with a bulky basic head group, i.e., octahydro-2*H*-quinolizine and 1,2,3,4,5,6-hexahydro-1,5-methano-8*H*-pyrido[1,2-*a*][1,5]diazo-

cin-8-one, were synthesized and tested for activity against D-10 (CQ-susceptible) and W-2 (CQ-resistant) strains of *P. falcipa-rum*. Most compounds were found to be active against both strains with nanomolar or sub-micromolar  $IC_{50}$  values. Eleven

compounds were found to be 2.7- to 13.4-fold more potent than CQ against the W-2 strain; among them, four cytisine derivatives appear to be of particular interest, as they combine high potency with low cytotoxicity against two human cell lines (HMEC-1 and HepG2) along with easier synthetic accessibility. Replacement of the 4-NH group with a sulfur bridge maintained antiplasmodial activity at a lower level, but produced an improvement in the resistance factor. These compounds warrant further investigation as potential drugs for use in the fight against malaria.

## Introduction

Recent estimates suggest that more than 198 million clinical cases of malaria occur every year worldwide, with 584000 deaths in 2013. Most of the casualties are children living in sub-Saharan Africa or pregnant women.<sup>[1]</sup> *Plasmodium falciparum (Pf)*, which is the main species responsible for the observed fatalities, has developed resistance to conventional antimalarials such as chloroquine (CQ) and antifolates. Resistance even to the artemisinin derivatives has been recently reported in southeast Asia.<sup>[2]</sup> To overcome the drug resistance issue, extensive modification of existing antimalarial chemotypes and the search for novel targets (and new chemotypes interacting with them) have been undertaken and are currently being pursued. Thus, many interesting compounds have been selected for use in monotherapy or, even better, in combination chemotherapies (particularly artemisinin-based combination thera-

[a]	Dr. B. Tasso, Dr. F. Novelli, Dr. M. Tonelli, Prof. F. Sparatore Dipartimento di Farmacia, Università degli Studi di Genova Viale Benedetto XV 3, 16131 Genova (Italy) E-mail: bruno.tasso@unige.it
[b]	Dr. A. Barteselli, Prof. A. Sparatore Dipartimento di Scienze Farmaceutiche, Università degli Studi di Milano via Mangiagalli 25, 20133 Milano (Italy)
[c]	Dr. N. Basilico Dipartimento di Scienze Biomediche, Chirurgiche e Odontoiatriche Università degli Studi di Milano via C. Pascal 36, 20133 Milano (Italy)
[d]	Dr. S. Parapini, Prof. D. Taramelli Dipartimento di Scienze Farmacologiche e Biomolecolari Università degli Studi di Milano via C. Pascal, 36, 20133 Milano (Italy)
	Supporting Information for this article is available on the WWW under

pies) able to improve efficacy and delay the onset of resistance. The different approaches are illustrated and discussed in some recent reviews.<sup>[3]</sup>

Particularly, the 4-aminoquinoline derivatives continue to attract interest because resistance appears to be compound specific and not related to changes in the structure of the drug targets.<sup>[4]</sup> Indeed, even close analogues of CQ and amodiaquine retain potent activity against CQ-resistant (CQ-R) strains of *P. falciparum*, such as short side-chain compounds  $A-C^{[5]}$ and metabolically stable isoquine (**D**), *tert*-butylisoquine (**E**), and 4'-fluoro-*N-tert*-butylamodiaquine (**F**)<sup>[6]</sup> (Figure 1). Peculiar modification of the side chain of CQ is observed in ferroquine (**G**),<sup>[7]</sup> which incorporates a ferrocenyl moiety, and in piperaquine (**H**), an old bisquinoline analogue that has been resumed as an efficient partner of artemisinin derivatives.<sup>[8]</sup>

A valuable approach to obtain novel CQ analogues is represented by the assembly of hybrid molecules containing two pharmacophores joined through a linker that should not disturb the interaction of each component with the relevant target.<sup>[9]</sup> The two pharmacophores may act in different ways on the same target, as in trioxaquine derivatives I and J,<sup>[10]</sup> which are able to alkylate heme and to inhibit the formation of hemozoin, or they may act on different targets, as is the case for "reversed chloroquines" K and L; these derivatives incorporate different kinds of inhibitors of *P. falciparum* chloroquine resistance transporter (*Pf*CRTs), which inhibit the formation of hemozoin and also the efflux of the drug from the digestive vacuole of the parasite.<sup>[11]</sup>

Particularly interesting hybrid antimalarial agents have been obtained by combining the scaffold of chloroquine and primaquine (PQ) through different linkers (e.g., **M**).<sup>[12]</sup> Primaquine, an



Figure 1. Examples of chloroquine (CQ) and amodiaquine (AQ) analogues with modified side chains.

old 8-aminoquinoline derivative, acts against all exoerythrocytic forms of *Plasmodium* and displays gametocytocidal activity, which is still the only transmission-blocking antimalarial that is clinically available.<sup>[13]</sup> Moreover, PQ synergizes chloroquine activity against CQ-R *P. falciparum* by inhibiting the *Pf*CRT.<sup>[14]</sup> The synthesized CQ–PQ hybrids display good to excellent activity against both the liver and blood stages of the *Plasmodium* infection and against gametocytes, which thus inhibits the transmission cycle.

Additional examples of hybrid molecules are combination products of the 4-aminoquinoline scaffold with polyarylmethyl system N; these hybrids are able to form radical intermediates that are toxic to the parasite, and additionally, they block the formation of hemozoin.<sup>[15]</sup> Finally, CQ analogues O and P bearing a side chain ending with bulky, nonbasic moieties are noteworthy.<sup>[16,17]</sup> Some of these compounds display potent activity against different strains of P. falciparum, which suggests that a basic side chain is not very essential for antiplasmodial activity, and this opens new possibilities for the design of novel chloroquine analogues. Interestingly, the former group of compounds<sup>[16]</sup> inhibits (similar to CQ) the formation of hemozoin, whereas the latter<sup>[17]</sup> inhibits, albeit weakly, falcipain 2 (a cysteine protease critical for hemoglobin catabolism) as an alternative or additional target for antimalarial activity.[18]

Over the past several years we have been engaged in the search of novel antimicrobial agents, and recently, we studied the antimalarial activity of some 4aminoquinoline and 9-aminoacridine derivatives (e.g., Q-T; Figure 2) characterized by the presence of a bulky, strongly basic, and lipophilic bicyclic moiety, that is, a quinolizidine (octahydro-2H-quinolizine) or ring.[19-22] (hexahydro-1*H*-pyrrolizine) pyrrolizidine These moieties appear as interesting structural features that are able to overcome the resistance mechanism. Indeed, these novel CQ analogues exhibited high antimalarial activity against CQ-susceptible (CQ-S) and CQ-R strains of Pf in vitro through a CQ-like mechanism, and some of them also showed efficacy against P. berghei and P. yoelii if given orally or intraperitoneally in a murine standard four-day test.[23,24]

Pursuing our efforts to obtain novel antimalarial agents containing a bulky basic head, we deemed it interesting to investigate the effects on the activity of the following structural changes, operated either singularly or in combination: 1) Further elongation of the chain between the NH group and the quinolizidine nucleus (e.g., compounds 1–5; Figure 3), which has already been shown to improve activity and/or to alter the selectivity for CQ-R or CQ-S parasites.<sup>[5a, 19]</sup> 2) The shift in the side chain from the 1-position to the 9a-position of the quinolizidine ring to avoid stereochemical issues (e.g., compounds **6–8**; Figure 3). 3) The exchange of the quinolizidine nucleus with an

even bulkier, but less lipophilic, moiety such as that of cytisine [(1R,5S)-(1,2,3,4,5,6-hexahydro-1,5-methano-8H-pyrido[1,2-a][1,5]diazocin-8-one] (e.g., compounds **9–20**; Figure 3). The pyridone ring present in the cytisine moiety might act as a Michael



Figure 2. Previously studied 4-aminoquinoline and 9-aminoacridine derivatives with bulky basic head groups.



Figure 3. Structures of novel quinoline derivatives tested as antimalarial agents (see Table 1 for structural details).

acceptor and, as such, could affect the thiol group of the active site of cysteine protease (as falcipain).<sup>[25]</sup> Interestingly, some *N*-benzylcytisine derivatives have been recently shown to display activity against the parasite *Leishmania donovani*.<sup>[26]</sup> 4) The replacement, in the foregoing compounds, of the NH group with a sulfur bridge (e.g., compounds **22–27**; Figure 3), which in addition to decreasing the basicity of the molecule should also increase its lipophilicity. The right balance of these physicochemical characteristics is fundamental for the accumulation of the drug in the digestive vacuole and for its association with hematin to inhibit the formation of hemozoin.<sup>[4c]</sup> 7-Chloro-4-(pyrrolidin-1-yl)quinoline (**21**), incidentally obtained during the syntheses, was also considered. The structures of the 27 tested compounds are collected in Figure 3. On the basis of previous observations,<sup>[27–29]</sup> the bulky basic heads of

# CHEMMEDCHEM Full Papers

the relevant compounds are thought to be fairly resistant to metabolic attack.

### **Results and Discussion**

#### Chemistry

The preparation of the longchain  $\omega$ -(quinolizidin-1-yl)alkylamines required for the synthesis of higher homologues of com-

pounds of general structure **Q** (Figure 2) may be a difficult and time-consuming task; thus, we resorted to the use of  $\omega$ -(lupi-nylthio)alkylamines { $\omega$ -[(1*R*,9a*R*)-(octahydro-2*H*-quinolizin-1-yl)-methylthio]alkylamines} taking into account the well-known bioisosterism of a methylene unit for a sulfur atom. This approach was already used profitably for the preparation of long-chain *N*-( $\omega$ -quinolizindinyl)alkanoyl- and *N*- or *O*-( $\omega$ -quinolizidinyl)alkyl derivatives of several aromatic and heteroaromatic moieties (e.g., phenothiazine and other tricyclic systems, anilines, 9-aminoacridines, and 6-hydroxycoumarin), to obtain muscarinic M1 and M2 ligands,<sup>[30,31]</sup> antiarrhythmics,<sup>[32]</sup> antiprion toxicity agents,<sup>[34]</sup>

Thus, thiolupinine<sup>[35]</sup> (28) [(1*R*,9*aR*)-(octahydro-2*H*-quinolizin-1-yl)methanthiol] was treated with iodoacetamide,<sup>[34a]</sup> acrylonitrile,<sup>[33]</sup> chloroacetone, and methyl vinyl ketone to obtain compounds 29, 31, 33, and 36, respectively. The last two ketones were converted into corresponding oximes 34 and 37, which in addition to compounds 29 and 31 were reduced with LiAlH<sub>4</sub> to required long-chain amines 30, 32, 35, and 38. These amines were, finally, treated with 4,7-dichloroquinoline (39) or 4-chloro-7-trifluoromethylquinoline (40) to obtain compounds 1–5 (Scheme 1).

For the synthesis of compounds **6–8**, 4,7-dichloroquinoline was treated with  $\omega$ -(quinolizidin-9a-yl)alkylamines (dihydrochlorides) **41–43** in the presence of diisopropylethylamine (DIPEA). 4/6-(Octahydro-2*H*-quinolizin-9a-yl)butan/hexan-1-amines (**41** and **42**) were recently described by Barteselli et al.,<sup>[36]</sup> and the homologue 8-(octahydro-2*H*-quinolizin-9a-yl)octan-1-amine (**43**) was obtained in a similar way starting from quinolizidinium perchlorate<sup>[37]</sup> through hydroxyalkylation with the method of McIntosh<sup>[38]</sup> and final amination according to Becker et al.<sup>[39]</sup> (Scheme 2).

Compound **9** was obtained by treating *N*-aminocytisine<sup>[40]</sup> (**48**) with 4,7-dichloroquinoline, whereas compounds **10–16**, **19**, and **20** were obtained from the reactions of 4-[*N*-( $\omega$ -bromoalkyl)amino]-7-chloroquinolines **54–58** with cytisine (**59**), 9nitrocytisine (**60**),<sup>[41]</sup> and 9,11-dibromocytisine (**61**)<sup>[42]</sup> eventually in the presence of DIPEA. The reaction of 4-[*N*-(4-bromobutyl)amino]-7-chloroquinoline with cytisine or its derivatives produced, instead of the expected compounds, 7-chloro-4-(pyrrolidin-1-yl)quinoline (**21**) by intramolecular loss of hydrogen bromide. The catalytic reduction of 9-nitrocytisine derivatives **15** and **16** gave corresponding amino derivatives **17** and **18** (Scheme 3).



Scheme 1. Reagents and conditions: a) iodoacetamide, EtOH, RT, 3 days; b) acrylonitrile, Triton B, dioxane, 60 °C, 1 h; c) chloroacetone, EtOH, RT, 1 h; d) methyl vinyl ketone, dry dioxane, RT, 1 h; e) LiAlH<sub>4</sub>, THF, reflux, 72 h; f) hydroxylamine hydrochloride/6 N NaOH, EtOH, reflux, 2 h; g) 4-chloro-7-substituted quinoline, phenol, 180 °C, 4 h.



Scheme 2. Reagents and conditions: a) phenol, DIPEA, N<sub>2</sub>, 130 °C 5 h; b) ZnBr<sub>2</sub>, dry THF, 50 °C  $\rightarrow$  reflux, 20 h; c) 1 N BH<sub>3</sub> in THF, 2-methyl-2-butene, dry THF, RT, 4.5 h; d) H<sub>2</sub>O, 6 N NaOH, 35 % H<sub>2</sub>O<sub>2</sub>, RT, 1 h; e) phthalimide, Ph<sub>3</sub>P, DEAD, dry THF, RT, 20 h; f) 6 N HCl, reflux, 20 h.

Required 4-[*N*-( $\omega$ -bromoalkyl)amino]-7-chloroquinolines **54**– **58** were obtained by treating 4,7-dichloroquinoline with the suitable  $\omega$ -hydroxyalkylamines and subsequent bromination by means of 48% hydrobromic acid in the presence of concentrated sulfuric acid, as previously described for some of them.<sup>[43–46]</sup>

Compounds **22** and **23** were obtained by treating thiolupinine (**28**)<sup>[35]</sup> with the proper 4-chloro-7-substituted quinolines, whereas compounds **24–27** were obtained from 7-chloro-4-thioquinoline (**62**),<sup>[47]</sup> which was treated with *S*-(haloalkyl)thiolupinines **63** and **64**<sup>[32,48]</sup> and *N*-( $\omega$ -haloalkyl)cytisines **65** and **66**<sup>[49]</sup> (Scheme 4).

#### Antimalarial activity

Compounds **1–27** were tested in vitro against D-10 (CQ-S) and W-2 (CQ-R) strains of *P. falciparum*. The antimalarial activity was evaluated by inhibition of parasite growth, as measured by the production of parasite lactate dehydrogenase. Table 1 shows the  $IC_{50}$  values against the D-10 and W-2 strains of *P. falciparum*, as well as the means of the ratios between the  $IC_{50}$  of CQ and that of each compound against the D-10 and W-2 strains, calculated for each single experiment.

The ratios between the  $IC_{50}$  values of each compound against the two strains are also indicated. These last values are suggestive of the susceptibility of the drug to the resistance mechanism (resistance factor, RF). The results listed in Table 1 indicate that many of novel 4-aminoquinoline derivatives 1-20 display high antimalarial activity with  $IC_{50}$  values < 100 nmagainst both the CQ-S and CQ-R strains, which thus resulted in 11 compounds that were from 2.7- to 13.4-fold more active than chloroquine against the CQ-R strain W-2. The remaining compounds show an appreciable level of activity with sub-micromolar IC<sub>50</sub> values with the only exception of compounds **10** and 13. On the other hand, 4-pyrrolidinylquinoline 21 and compounds 22-27, which are six derivatives of 4-thioquinoline, display only modest or moderate antimalarial activity with IC<sub>50</sub> values in the low (1.4-12.5) µm range; nevertheless, these compounds exhibit an interesting improvement in the resistance factor.

With regard to the structure-activity relationships, a general enhancement in the activity with increasing distance between



**Scheme 3.** *Reagents and conditions*: a) phenol, N<sub>2</sub>, 180 °C 7 h; b) hydroxyalkylamine, 150 °C, 3 h; c) 48% HBr/H<sub>2</sub>SO<sub>4</sub>, 160 °C, 3.5 h; d) CH<sub>3</sub>CN, 100 °C, 36 h; e) EtOH, H<sub>2</sub>, 10% Pd/C, RT.



Scheme 4. Reagents and conditions: a) Dowtherm A, 170  $^\circ\text{C}$ , 2 h; b) KOH, CH\_3CN, N\_2, RT, 24 h.

the quinoline ring and the basic nitrogen atom of the side chain was observed, even if with some peculiarities in each subgroup of compounds. Indeed, we previously showed<sup>[19]</sup> that the increasing length of the methylene chain in compounds of structure **Q** (Figure 2) influenced negatively the antimalarial activity, especially against the CQ-R strain (for n=0,  $IC_{50}=23.3 \text{ nm}$ ; for n=3,  $IC_{50}=384.4 \text{ nm}$ ). Such a trend is now inverted in compounds **1** and **2**, which are characterized by a longer linker of four and five atoms, respectively. This behavior is similar to that observed in a set of diethylaminoalkyl analogues of CQ that first exhibit decreasing activity in the medium-length compounds but then recover the high activity if the number of methylene units is further increased.<sup>[5a]</sup>

Interestingly, although stereochemical issues are overcome by shifting the linker (between the quinoline and quinolizidine moieties) from the 1-position to the 9a-position of the quinolizidine ring, this shift produces only minor variations in the antimalarial potency (compare compounds 1 and 2 with 6 and 7). However, if the linker is elongated up to eight methylene units (as in 8), a strong increase in potency is observed ( $IC_{50} =$  9 and 25.9 nm for the CQ-S and CQ-R strains, respectively), even if the interest in this compound is restrained by its complex synthesis.

In the set of cytisinyl derivatives 9-13, elongation of the linker produces an alternating effect on the activity, which is initially decreasing and then steadily increasing. The minimal activity is observed in compound 10 (linker with two methylene units), with  $IC_{50}$  values of 941 and 1776 nm for the CQ-S and CQ-R strains, respectively. The maximal activity is found in compound 12 (linker with five methylene units) with  $IC_{50}$ values of 32.9 and 77.1 nm for the CQ-S and CQ-R strains, which are 28- and 23-fold enhancements in potency, respectively. The introduction of substituents on the pyridone ring of the cytisine moiety (i.e., compounds 14-20) produces a clear increase in antimalarial activity that is cumulative with the positive effect of elongation of the linker. Thus, compounds 16, 18, and 20 are the most potent among those presently reported, and these derivatives are 5-, 10-, and 8-fold more potent than chloroquine, respectively, and are from 1/3 to 2/3 as potent as artemisinin, against the W-2 (CQ-R) strain of P. falciparum. Moreover, the cytisinyl derivatives are characterized by low cytotoxicity (Table 2) versus two human cell lines (human microvascular endothelial cells, HMEC-1, and human Caucasian hepatocellular carcinoma, HepG2), with selectivity indices (SI) in the ranges of 400 to 880 and of 143 to 540, respectively (CQ, SI > 120 and 62, respectively). Additionally, comparably with CQ and artemisinin, they produced less than 10% hemolysis upon incubation for 72 h at a concentration of 200 μм with uninfected erythrocytes. Among the most potent compounds, only 8 displayed some hemolytic activity with a CC<sub>50</sub> value of (148.6  $\pm$  1.2)  $\mu$ M.

Therefore, the cytisinyl derivatives of 4-aminoquinoline appear worthy of further evaluation as antimalarial agents, also in consideration of the easy availability of the starting material, which is extracted from the seeds of the widespread *Laburnum anagyroides* and of other plants (*Sophora* and *Thermopsis* species). Particularly, the complex structure of the cytisine moiety that characterizes the best compounds warrants thorough investigation of their mechanism of action. The high activity of



# CHEMMEDCHEM Full Papers

Table 1. Antimalarial activity of compounds 1–27 tested in vitro against D-10 (CQ-S) and W-2 (CQ-R) P. falciparum strains.								
$\begin{array}{c} \begin{array}{c} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\$								
Compd	R	Х	R′	D-10 (СС IC <sub>50</sub> [nм] <sup>[a]</sup>	Q-S) Ratio to CQ <sup>[b]</sup>	W-2 (С IС <sub>50</sub> [nм] <sup>[а]</sup>	Q-R) Ratio to CQ <sup>[b]</sup>	Ratio IC <sub>50</sub> CQ-R/CQ-S <sup>[c]</sup>
1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 chloroqui	CI CI CI CF3 - - - - - - - - - - - - - - - - - - -	$\begin{array}{l} NH-CH_2-CH_2-S-CH_2\\ NH-CH(CH_2)_3-S-CH_2\\ NH-CH(CH_3)-CH_2-S-CH_2\\ NH-CH(CH_3)-CH_2-S-CH_2\\ NH-CH(CH_2)_3-CH_2-S-CH_2\\ NH-CH(CH_2)_4\\ NH-(CH_2)_6\\ NH-(CH_2)_6\\ NH-(CH_2)_8\\ NH\\ NH-CH_2-CH_2\\ NH-CH_2-CH_2\\ NH-CH(CH_2)_3\\ NH-CH(CH_2)_5\\ NH-CH(CH_2)_5\\ NH-CH(CH_2)_5\\ NH-CH(CH_2)_5\\ NH-CH(CH_2)_5\\ NH-CH_2-CH_2\\ NH-CH_2-CH_2\\ NH-CH_2-CH_2\\ NH-CH_2-CH_2\\ NH-CH_2-CH_2-S-CH_2\\ S-CH_2\\ S-CH_2-S-CH_2\\ S-(CH_2)_3\\ S-(CH_2)_4\\ \end{array}$	- - - - - - - H H H H H H 9-NO <sub>2</sub> 9-NO	$\begin{array}{c} 53.7\pm13.5\\ 27.1\pm6.9\\ 24.6\pm8.3\\ 23.1\pm8.7\\ 56.1\pm17.3\\ 22.5\pm9.6\\ 27.15\pm8.0\\ 9.0\pm3.7\\ 370.1\pm110\\ 941.5\pm262.1\\ 116.5\pm38.4\\ 32.9\pm2.4\\ 902.3\pm263.4\\ 115.3\pm23.2\\ 62.7\pm5.3\\ 16.5\pm5.2\\ 96.2\pm7.5\\ 11.9\pm6.0\\ 47.5\pm13.9\\ 16.8\pm5.5\\ 886.6\pm25.9\\ 7190.0\pm2620\\ >10000\\ 2412.0\pm192.1\\ 1747.0\pm63.0\\ 9841.0\pm2023\\ 3922.0\pm162.2\\ 20.0\pm5\\ \end{array}$	0.61 1.21 1.25 1.33 0.55 0.80 1.50 2.34 0.06 0.02 0.17 0.28 0.01 0.08 0.15 0.56 0.14 1.11 0.28 0.79 0.02 0.003 NA <sup>[d]</sup> 0.013 0.018 0.003 0.008 -	$\begin{array}{c} 259.4\pm 60.1\\ 155.4\pm 49.0\\ 82.6\pm 15.5\\ 119.9\pm 48.3\\ 83.5\pm 37.8\\ 475.3\pm 145.7\\ 194.3\pm 74.6\\ 25.9\pm 3.7\\ 635.8\pm 113\\ 1775.9\pm 304.4\\ 298.7\pm 35.7\\ 77.1\pm 22.8\\ 1048.6\pm 360.8\\ 874.8\pm 366.2\\ 546.6\pm 174.3\\ 42.0\pm 21.1\\ 478.4\pm 140.0\\ 24.3\pm 10.1\\ 90.7\pm 42.1\\ 31.1\pm 16.4\\ 810.9\pm 162.8\\ 4530.0\pm 2557\\ 7820.0\pm 2539\\ 181.80\pm 660.1\\ 1416.0\pm 379.8\\ 12281.0\pm 400.3\\ 5190.0\pm 2028\\ 320\pm 51\\ \end{array}$	3.15 5.25 6.20 4.27 6.13 0.68 1.67 13.45 0.50 0.16 0.94 2.96 0.22 0.26 0.42 5.43 0.52 10.32 2.76 8.06 0.31 0.07 0.04 0.28 0.36 0.04 0.10	4.83 5.72 3.36 5.19 1.49 21.12 7.16 2.88 1.72 1.89 2.56 2.34 1.16 7.59 8.72 2.55 4.975 2.04 1.91 1.85 0.915 0.63 0.62 0.75 0.81 1.25 1.32 16
artemisini	n	proceed as the maps $\pm$ SD of	at least three	$33.2 \pm 11.3$ $27.7 \pm 8.9$	- -	$16.9 \pm 7.3$ $14.5 \pm 4.2$	- - ate [b] Moon of t	0.51 0.52

[a] Results are expressed as the mean  $\pm$  SD of at least three different experiments, each performed in duplicate or triplicate. [b] Mean of ratios between the IC<sub>50</sub> of CQ and that of each compound against *P. falciparum* strains D-10 or W-2, calculated for each single experiment. [c] Resistance factor: Ratios between the IC<sub>50</sub> values of each compound against the two strains of *P. falciparum*. [d] Not available.

these compounds might be related to strong association of the molecules with hematin, similar to that observed for Solomon's<sup>[16]</sup> compounds (e.g., **O**, Figure 1) bearing a side chain with a bulky, highly lipophilic head. However, owing to the Michael acceptor characteristics of the pyridone ring embodied in the cytisine moiety, interaction with the thiol group of the active site of falcipain cannot be excluded. Actually, even compound **P** (Figure 2) inhibited, albeit weakly, falcipain 2, which should attack one of its carboamide groups.<sup>[17]</sup>

Notably, pyridone-derived falcipain inhibitors have been described,<sup>[50]</sup> for which, however, the pyridone ring served mainly as a rigid peptidomimetic backbone, and additional more potent electrophilic groups were included in the structures. In both sets of quinolizidinylalkyl and cytisinylalkyl derivatives, branching of the linker produces a slight increase in activity, as shown by comparison of the activities of unbranched compounds 1, 2, and 10 with those of corresponding methyl homologues 3, 4, and 13.

Replacement of the chlorine atom in the quinoline ring with a trifluoromethyl group decreases the activity on the CQ-S strain, but it does not affect the activity on the CQ-R strain (compare compounds **3** and **5**). Such a decrease in activity was already observed by  $us^{[19]}$  in compounds of structure **Q** (Figure 2) and by several authors in other 4-aminoquinoline derivatives. Kaschula et al.<sup>[4c]</sup> correlated the decreased activity with the stronger electron-withdrawing character of the trifluoromethyl group, which decreases the basicity of both nitrogen atoms of the side chain. However, in the case of the CQ-R strain the enhanced lipophilicity, and subsequent cell penetration, of the fluorinated compounds might compensate the negative effect of the decreased basicity on the activity.<sup>[51]</sup> **Table 2.** Cytotoxicity against human microvascular endothelial cells (HMEC-1) and human Caucasian hepatocyte carcinoma (HepG2), and selectivity index of the most potent and other representative compounds.

Compd		IC <sub>50</sub> [nм] <sup>[a]</sup>		SI <sup>[b]</sup>		
	HMEC-1	HepG2	W-2 (CQ-R)	HMEC-	HepG2	
				1		
3	$18673 \pm 5915$	$9773 \pm 1912$	$82.6 \pm 15.5$	226	118.3	
6	>26000	NT	$475.3\pm145.7$	>54.7	-	
7	$9211\pm938$	NT	$194.3\pm74.6$	47.4	-	
8	$5495 \pm 697$	$1869 \pm 628$	$25.9\pm3.7$	212.2	72.2	
12	$54800 \pm 14547$	$11165{\pm}2263$	$77.1\pm22.8$	710.8	144.8	
16	$23010\pm\!6797$	$10113\pm217$	$42.0 \pm 21.1$	547.8	240.8	
18	$21388\pm 5245$	$13142{\pm}1402$	$24.3\pm10.1$	880.5	540.8	
20	$12413{\pm}2066$	$4454\pm1211$	$31.1 \pm 16.4$	399.1	143.8	
24	$75282\pm11316$	NT	$1818.0 \pm 660.1$	41.4	-	
CQ	> 38 000	$19942 \pm 1643$	$320\pm51$	>118.8	62.3	
Arte	NT	>106260	$14.5\pm4.2$	-	>7328.3	
[a] Results are the mean $\pm$ SD of three experiments each performed in duplicate; NT: not tested. [b] Selectivity index: [IC <sub>50</sub> (HMEC-1 or HepG2)]/[IC <sub>50</sub> (W-2(CQ-R)].						

Finally, in compounds **22–27** the replacement of the 4-NH group by a sulfur bridge should increase the lipophilicity of the molecule but still allow resonance of the positive charge from the protonated quinoline nitrogen atom to the sulfur bridge, as suggested by the UV spectra of other 7-chloro-4-thioquinoline derivatives described by Clinton and Suter.<sup>[52]</sup> The presence of this resonance should favor interaction of the molecules with ferriprotopophirin IX (Fe<sup>III</sup>PPIX)<sup>[4a]</sup> and should therefore increase antimalarial activity, even if no activity was observed upon screening the above 4-S analogues of CQ in vivo against avian malaria.<sup>[52]</sup> Actually, the lack of in vivo activity may be related to inadequate dosage or unfavorable pharmacokinetic characteristics of the particular compounds.

Indeed, S-bridged compounds **22–27** display, in vitro, moderate antimalarial activity against both of the CQ-S and CQ-R strains of *P. falciparum*, with most IC<sub>50</sub> values in the low-micromolar range. This activity is in line with that exhibited by a set of 7-chloro-4-(*S*-diethylaminoalkyl)thioquinolines (described by Natarajan et al.<sup>[53]</sup>), despite the largely different bulkiness of the terminal heads of the respective side chains.

In both series of compounds, the activity improves somewhat with elongation of the linker, in conformity with the trend observed for the respective NH analogues. However, differently from the latter, most of these sulfur-containing compounds exhibit a resistance factor (RF = IC<sub>50</sub>CQ-R/IC<sub>50</sub>CQ-S) significantly below 1 (RF=0.62-0.81 for compounds 22-25), which suggests low susceptibility of the S-bridged compounds to resistance mechanisms. Thus, once more, within a given type of compounds the enhanced lipophilicity seems to balance the negative effect of the reduced basicity on the activity against the CQ-R strain. However, comparing the S-bridged compounds with the 4-aminoquinoline derivatives, the reduced activities of the former appear to be mainly due to the strongly decreased basicity of the quinoline nitrogen atom (mean  $pK_{a2} = 6.40 \rightarrow pK_{a2} = 3.18$ ), not balanced by the increased lipophilicity (Table 3, values calculated with ACD/Labs 7.00).

## Conclusions

Twenty-seven novel chloroquine analogues characterized by a side chain bearing a bulky basic head (i.e., octahydro-2*H*-quinolizine and 1,2,3,4,5,6-hexahydro-1,5-methano-8*H*-pyrido[1,2-*a*][1,5]diazocin-8-one) were synthesized and tested for antimalarial activity against the D-10 (CQ-susceptible) and W-2 (CQ-resistant) strains of *P. falciparum*. Most compounds were active against both strains with nanomolar or sub-micromolar IC<sub>50</sub> values; in particular, 11 compounds were 2.7- to 13.4-fold more active than chloroquine against the chloroquine-resistant strain.

The influence on activity of some structural features was analyzed, and in each subset of compounds, the positive effect of increasing the length of the spacer between the quinoline and the bulky basic head was confirmed. Moreover, exchange of the 4-NH group with a sulfur bridge maintained the antimalarial activity but at a lower level of potency; however, this structural modifi-

<b>Table 3.</b> Calculated $\log D$ and $pK_a$ values for representative compounds (quinolizidine derivatives: <b>Q-8</b> and <b>22–25</b> ; cytisine derivatives: <b>11–20</b> and <b>26</b> , <b>27</b> ).							
Compd	log D <sub>7.4</sub> [b]	log D <sub>5.2</sub> <sup>[c]</sup>	p <i>K</i> <sub>a1</sub> <sup>[d]</sup>	p <i>K</i> <sub>a2</sub> <sup>[e]</sup>	p <i>K</i> <sub>a3</sub> <sup>[f]</sup>		
NH bridged							
<b>Q</b> ( <i>n</i> = 1)	1.98	0.73	10.18	6.35	-4.48		
1	2.67	1.41	10.15	6.30	-4.69		
2	3.01	1.73	10.16	6.43	-4.13		
3	3.02	1.79	10.15	6.23	-3.99		
5	3.70	2.84	NC <sup>[h]</sup>	NC <sup>[h]</sup>	NC <sup>[h]</sup>		
8	4.13	2.92	10.59	6.51	-3.66		
11	2.40	-0.35	8.80	6.32	-4.94		
12	2.88	0.23	8.92	6.48	-3.86		
16	2.82	-0.12	8.88	6.48	-3.86		
18 <sup>[g]</sup>	2.32	-0.43	8.78	6.48	-3.84		
20	4.47	1.44	8.08	6.48	-3.83		
S bridged							
22	2.81	2.13	9.91	3.30	-		
23	2.99	2.32	9.91	2.49	-		
24	3.31	2.79	10.13	3.25	-		
25	3.65	3.15	10.15	3.37	-		
26	3.21	1.29	8.07	3.28	-		
27	3.33	1.51	8.38	3.37	-		
[a] Calculated with ACD/Labs yor 7.00 [b] Log D at plasma pH [s] Log D							

[a] Calculated with ACD/Labs ver. 7.00. [b] Log *D* at plasma pH. [c] Log *D* at *P*. falciparum digestive vacuole pH. [d]  $pK_a$  of side chain N atom. [e]  $pK_a$  of quinoline N atom. [f]  $pK_a$  of 4-NH group. [g]  $pK_a$  of 9-NH<sub>2</sub> group: 3.61. [h] Not calculated.

cation produced an interesting improvement in the resistance factor (RF = 0.6–0.8) that appears worthy of further investigation. The introduction of a cytisinyl head and a spacer containing five methylene units produced the most interesting compounds endowed with high activity, low cytotoxicity on two human cell lines (HMEC-1 and HepG2), and nonhemolytic at concentrations up to 200  $\mu$ M. Therefore, these compounds deserve further investigation to define their ADME profiles and their in vivo antimalarial activities in a murine standard fourday test.



## **Experimental Section**

#### Chemistry

General: All commercially available solvents and reagents were used without further purification unless otherwise stated. Melting points were determined with a Büchi apparatus. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded with a Varian Gemini 200 or Varian Mercury 300VX spectrometer in CDCl<sub>3</sub> or [D<sub>6</sub>]DMSO with Me<sub>4</sub>Si as an internal standard; in the assignments, Q denotes the quinolizidine ring and bisp denotes the bispidine moiety. HRMS was performed with a FT-Orbitrap mass spectrometer in positive electrospray ionization (ESI). Elemental analyses were performed with a Carlo Erba EA-1110 CHNS-O instrument in the Microanalysis Laboratory of the Department of Pharmacy of Genoa University. All final compounds and some intermediates were characterized by <sup>1</sup>H NMR spectroscopy and elemental analyses or HRMS, which indicated that their purity was always  $\geq$  95%. The remaining intermediates were characterized by either <sup>1</sup>H NMR spectra or elemental analyses. For the most interesting compounds (i.e., 12, 16, 18, and 20) the <sup>13</sup>C NMR spectroscopy data are also included.

#### General method for the preparation of 4-(*N*-{[(1*R*,9a*R*)-(octahydro-2*H*-quinolizin-1-yl)methylthio]alkyl}amino)-7-substituted

**quinolines 1–5**: Amino compound **30**, **32**, **35**, or **38** (6.5 mmol) was mixed with equimolar amounts of 4,7-dichloroquinoline (1.28 g) or 4-chloro-7-trifluoromethylquinoline (1.50 g) and phenol (4 g). The mixture was heated at 180 °C for 4 h. After cooling, the mixture was treated with 2 N NaOH solution until strongly alkaline and was extracted with Et<sub>2</sub>O (3×40 mL). The Et<sub>2</sub>O solution was washed with 2 N NaOH solution, then with H<sub>2</sub>O, and finally with 5% acetic acid solution (2×20 mL). The acid solution was alkalized with 2 N NH<sub>3</sub> solution (to pH 8) and was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3× 50 mL) to yield a light-brown solid that was washed with boiling dry Et<sub>2</sub>O (5 mL), which left the title compound as white crystals. By concentration of the Et<sub>2</sub>O solution, some more compound was recovered. In the cases of compounds **4** and **5**, the extracted raw compounds were chromatographed on neutral alumina (1:20) eluting with CH<sub>2</sub>Cl<sub>2</sub>.

#### 7-Chloro-4-(N-{2-[(1R,9aR)-(octahydro-2H-quinolizin-1-yl)me-

**thylthio]ethyl}amino)quinoline (1)**: Yield: 37%; mp: 160–165 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.03–2.14 (m, 14H, Q), 2.64–2.91 (m, 2H, 2H<sub>a</sub> near N of Q+2H, SCH<sub>2</sub>), 2.93 (t, J=6.2 Hz, 2H, CH<sub>2</sub>S), 3.42–3.61 (m, 2H, Ar-NH-*CH*<sub>2</sub>), 5.66 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.44 (d, J=5.4 Hz, 1H, arom), 7.41 (dd, J=9.0, 2.0 Hz, 1H, arom), 7.50 (d, J=9.0 Hz, 1H, arom), 7.98 (d, J=2.0 Hz, 1H, arom), 8.56 ppm (d, J=5.4 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>21</sub>H<sub>28</sub>ClN<sub>3</sub>S (389.99): C 64.68, H 7.24, N 10.77, S 8.22, found: C 64.76, H 7.36, N 10.81, S 8.27.

#### 7-Chloro-4-(N-{3-[(1R,9aR)-(octahydro-2H-quinolizin-1-yl)me-

**thylthio]prop-1-yl}amino)quinoline** (2): Yield: 38%; mp: 95–100 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$ =1.04–2.14 (m, 14H, Q+2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.69 (t, J=6.4 Hz, 2H, CH<sub>2</sub>S), 2.76–2.93 (m, 2H, 2H<sub>a</sub> near N of Q+2H, SCH<sub>2</sub>), 3.49–3.58 (m, 2H, Ar-NH-CH<sub>2</sub>), 5.60 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.44 (d, J=5.4 Hz, 1H, arom), 7.36 (dd, J=9.0, 2.0 Hz, 1H, arom), 7.61 (d, J=9.0 Hz, 1H, arom), 7.96 (d, J=2.0 Hz, 1H, arom), 8.53 ppm (d, J=5.4 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>22</sub>H<sub>30</sub>CIN<sub>3</sub>S (404.01): C 65.40, H 7.48, N 10.40, S 7.49, found: C 65.22, H 7.75, N 10.34, S 7.89%.

#### 7-Chloro-4-(N-{3-[(1R,9aR)-(octahydro-2H-quinolizin-1-yl)me-

**thylthio]prop-2-yl}amino)quinoline (3)**: Yield: 50%; mp: 110–112°C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 0.88–2.12 (m, 14 H, Q+d superimposed at 1.44, *J*=6.2 Hz, 3 H, CH-*CH*<sub>3</sub>), 2.48–2.96 (m, 2H<sub>a</sub> near

N of Q+2H, SCH<sub>2</sub>+2H, CH<sub>2</sub>S), 3.72–4.08 (m, 1H, CH-CH<sub>3</sub>), 5.33 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.45 (d, J=5.4 Hz, 1H, arom), 7.39 (dd, J=8.8, 2.0 Hz, 1H, arom), 7.72 (d, J=8.8 Hz, 1H, arom), 7.97 (d, J=2.0 Hz, 1H, arom), 8.55 ppm (d, J=5.4 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>22</sub>H<sub>30</sub>ClN<sub>3</sub>S (404.01): C 65.40, H 7.48, N 10.40, S 7.94, found: C 65.26, H 7.54, N 10.37, S 8.06.

#### 7-Chloro-4-(N-{4-[(1R,9aR)-(octahydro-2H-quinolizin-1-yl)me-

**thylthio]but-2-yl}amino)quinoline (4)**: Yield: 9%; mp: 77–82 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 0.95–2.14 (14H, Q+2H, CHCH<sub>2</sub>CH<sub>2</sub>+ d superimposed at 1.36, *J*=6.4 Hz, 3H, CH-CH<sub>3</sub>), 2.42–2.96 (m, 2H<sub>a</sub> near N of Q+2H, SCH<sub>2</sub>+2H, CH<sub>2</sub>S), 3.74–4.08 (m, 1H, CH-CH<sub>3</sub>), 5.20 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.50 (d, *J*=5.6 Hz, 1H, arom), 7.37 (dd, *J*=9.0, 1.6 Hz, 1H, arom), 7.72 (d, *J*=9.0 Hz, 1H, arom), 7.97 (d, *J*=2.0 Hz, 1H, arom), 8.53 ppm (d, *J*=5.6 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>23</sub>H<sub>32</sub>ClN<sub>3</sub>S (418.04): C 66.08, H 7.72, N 10.05, S 7.67, found: C 66.15, H 7.89, N 10.08, S 7.67.

#### 4-(N-{3-[(1R,9aR)-(octahydro-2H-quinolizin-1-yl)methylthio]prop-

**2-yl}amino)-7-trifluoromethylquinoline (5)**: Yield: 59%; mp: 91– 93 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 0.92–2.15 (14H, Q+d superimposed at 1.46, *J* = 6.6 Hz, 3 H, CH-*CH*<sub>3</sub>), 2.47–2.96 (m, 2 H<sub>a</sub> near N of Q+2 H, *SCH*<sub>2</sub>+2 H, *CH*<sub>2</sub>S), 3.78–4.12 (m, 1 H, *CH*-CH<sub>3</sub>), 5.41 (s, 1 H, NH, collapsed with D<sub>2</sub>O), 6.55 (d, *J* = 5.4 Hz, 1 H, arom), 7.62 (dd, *J* = 8.8, 1.6 Hz, 1 H, arom), 7.91 (d, *J* = 8.8 Hz, 1 H, arom), 8.29 (s, 1 H, arom), 8.65 ppm (d, *J* = 5.4 Hz, 1 H, arom); elemental analysis calcd (%) for C<sub>23</sub>H<sub>30</sub>F<sub>3</sub>N<sub>3</sub>S (425.55): C 63.13, H 6.91, N 9.60, S 7.33, found: C 63.04, H 7.03, N 9.70, S 7.15.

General method for the preparation of 7-chloro-4-{[ $\omega$ -octahydro-2*H*-quinolizin-9a-yl]alkyl]amino}quinolines 6–8: A mixture of amine 41, 42,<sup>[36]</sup> or 43 as the dihydrochloride or free base (0.88 mmol), 4,7-dichloroquinoline (192 mg, 0.97 mmol), phenol (415 mg, 4.41 mmol), and DIPEA (0.25 mL, 1.76 mmol) was heated under a N<sub>2</sub> atmosphere for 5 h at 130 °C. After cooling, the mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, and the resulting solution washed with 2 N NaOH solution (3×5 mL) and then with brine (10 mL). After drying (Na<sub>2</sub>SO<sub>4</sub>), the solvent was evaporated, and the crude solid was purified by column chromatography (silica gel, different mixtures CH<sub>2</sub>Cl<sub>2</sub>/MeOH as indicated for each compound). For the preparation of compound 8, it was more convenient to use the free base (obtained preliminary from the dihydrochloride), which avoided the use of DIPEA and heating the mixture for only 3 h.

# **7-Chloro-4-{[4-(octahydro-2***H***-quinolizin-9a-yl)butyl]amino}quinoline (6):** Column chromatography (CH<sub>2</sub>Cl<sub>2</sub>/MeOH 94:6); solid washed with PE. Yield: 64%; mp: 157.8–159.3 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): $\delta$ =1.19–1.84 (m, 14H, Q+4H, *CH*<sub>2</sub>*CH*<sub>2</sub>), 2.42–2.64 (m, 2H, 2H<sub>a</sub> near N of Q+2H, *CH*<sub>2</sub>C), 3.31–3.37 (m, 2H, Ar-NH-*CH*<sub>2</sub>), 4.96 (s, 1H, NH), 6.43 (d, *J*=5.5 Hz, 1H, arom), 7.36 (dd, *J*=2.2, 8.8 Hz, 1H, arom), 7.65 (d, *J*=8.8 Hz, 1H, arom), 7.95 (d, *J*=2.2 Hz, 1H, arom), 8.54 ppm (d, *J*=5.5 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>22</sub>H<sub>30</sub>ClN<sub>3</sub> (371.95): C 71.04, H 8.13, N 11.30, found: C 70.89, H 8.28, N 11.45.

#### 7-Chloro-4-{[6-(octahydro-2H-quinolizin-9a-yl)hexyl]amino}qui-

**noline (7)**: Column chromatography (CH<sub>2</sub>Cl<sub>2</sub>/MeOH 92:8); amorphous solid crystallized and washed with Et<sub>2</sub>O. Yield: 46%; mp: 144.6–146.0 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.12–1.82 (m, 14H, Q+8H, *CH*<sub>2</sub>*CH*<sub>2</sub>*CH*<sub>2</sub>*CH*<sub>2</sub>), 2.41–2.65 (m, 2H, 2H<sub>a</sub> near N of Q+2H, *CH*<sub>2</sub>C), 3.29–3.35 (m, 2H, Ar-NH-*CH*<sub>2</sub>), 4.96 (s, 1H, NH), 6.42 (d, *J* = 5.5 Hz, 1H, arom), 7.36 (dd, *J*=2.2, 8.8 Hz, 1H, arom), 7.65 (d, *J*= 8.8 Hz, 1H, arom), 7.96 (d, *J*=2.2 Hz, 1H, arom), 8.53 ppm (d, *J*= 5.2 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>24</sub>H<sub>34</sub>ClN<sub>3</sub> (400.00): C 72.06, H 8.57, N 10.51, found: C 72.23, H 8.81, N 10.19.



7-Chloro-4-{[8-(octahydro-2H-quinolizin-9a-yl)octyl]amino}quinoline (8): Column chromatography (CH<sub>2</sub>Cl<sub>2</sub>/MeOH 94:6); amorphous solid crystallized and washed with Et<sub>2</sub>O. Yield: 55%; mp: 116.8-118.3 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.12–1.77 (m, 14 H, Q + 12 H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.47–2.63 (m, 2H, 2H<sub>a</sub> near N of Q+2H,  $CH_2C$ ), 3.28–3.32 (m, 2 H, Ar-NH- $CH_2$ ), 4.95 (s, 1 H, NH), 6.42 (d, J =5.5 Hz, 1 H, arom), 7.36 (dd, J=2.2, 8.8 Hz, 1 H, arom), 7.65 (d, J= 8.8 Hz, 1 H, arom), 7.96 (d, J = 2.2 Hz, 1 H, arom), 8.54 ppm (d, J =5.2 Hz, 1 H, arom); elemental analysis calcd (%) for C<sub>26</sub>H<sub>38</sub>CIN<sub>3</sub> (428.05): C 72.95, H 8.95, N 9.82, found: C 73.14, H 9.08, N 9.56. The compound was converted into the corresponding dihydrochloride; mp: 250–252 °C (dec.); <sup>1</sup>H NMR (300 MHz,  $[D_6]DMSO$ ):  $\delta = 1.05-1.91$ (m, 26 H), 2.86-3.09 (m, 2 H), 3.35-3.40 (m, 2 H), 3.49-3.52 (m, 2 H), 6.85 (d, J=7.2 Hz, 1 H, arom), 7.76 (d, J=8.8 Hz, 1 H, arom), 8.06 (s, 1H, arom), 8.52 (d, J=7.2 Hz, 1H, arom), 8.70 (d, J=8.8 Hz, 1H, arom), 9.62 (brs, 1H, collapsed with D<sub>2</sub>O), 10.20 ppm (brs, 2H, collapsed with D<sub>2</sub>O); elemental analysis calcd (%) for C<sub>26</sub>H<sub>38</sub>ClN<sub>3</sub>·2HCl (500.98): C 62.33, H 8.05, N 8.39, found: C 62.01, H 8.32, N 8.15.

7-Chloro-4-(cytisin-3-yl)aminoquinoline (9): A mixture of N-aminocytisine<sup>[40]</sup> (332 mg, 1.62 mmol), 4,7-dichloroquinoline (333 mg, 1.62 mmol), and phenol (1.03 g, 4.41 mmol) was heated under a N<sub>2</sub> atmosphere for 7 h at 180 °C. After cooling, the mixture was taken up with  $2 \times \text{NaOH}$  to strongly alkaline pH and then with Et<sub>2</sub>O (3× 20 mL). The insoluble compound was filtered and washed with 2 N NaOH (3 mL),  $H_2O$  (2×3 mL), and thoroughly with Et<sub>2</sub>O (3×5 mL) and finally crystallized from EtOH. Yield: 94%; mp: 296-298°C; <sup>1</sup>H NMR (300 MHz, [D<sub>6</sub>]DMSO)  $\delta$  = 1.70–1.98 (m, 2H, bisp), 2.54 (s, 1H, bisp), 2.92-3.05 (m, 2H, bisp), 3.08-3.25 (m, 2H, bisp), 3.30 (s, 1H, bisp), 3.75 (dd, J=5.5, 14.6 Hz, 1H, bisp) 4.04 (d, J=14.6 Hz, 1 H, bisp), 6.02 (d, J = 4.4 Hz, 1 H,  $\alpha$ -pyr), 6.13 (d, J = 6.3 Hz, 1 H,  $\alpha$ pyr), 6.33 (d, J = 8.8 Hz, 1 H, arom), 7.30–7.53 (m, 1 H,  $\alpha$ -pyr+1 H, arom), 7.74 (s, 1 H, arom), 8.09 (d, J=8.8 Hz, 1 H, arom), 8.14 (d, J= 5 Hz, 1 H, arom), 8.47 ppm (s, 1 H, NH collapsed with D<sub>2</sub>O); HRMS (ESI): m/z: calcd for C<sub>20</sub>H<sub>20</sub>ClN<sub>4</sub>O: 367.13256 [M + H]<sup>+</sup>, found: 367.13245.

**General method for the preparation of 7-chloro-4-{***N*-[ω-(citisin-**3-yl)alkyl]amino}quinolines 10–13**: In an Aldrich pressure tube, cytisine (0.19 g, 1 mmol) was dissolved in CH<sub>3</sub>CN (3 mL) and 4-[(ωbromoalkyl)amino]-7-chloroquinoline **54–58** (0.5 mmol) was added;<sup>(43–46]</sup> the closed tube was heated at 100 °C for 36 h. The cooled mixture was filtered from cytisine hydrobromide, and the organic phase was evaporated to dryness. The residue was taken up in H<sub>2</sub>O (10 mL), alkalized (2 N NaOH until strongly alkaline), and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×40 mL). After drying (Na<sub>2</sub>SO<sub>4</sub>), the solvent was removed, and the oily residue was subjected to chromatography on alumina (1:25) eluting with CH<sub>2</sub>Cl<sub>2</sub>.

**7-Chloro-4-{***N***-[**2-(citisin-3-yl)ethyl]amino}quinoline (10): Yield: 78%; mp: 79–81 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.77–2.10 (m, 3H, bisp), 2.34–2.92 (m, 2H, NH-*CH*<sub>2</sub>+3H, bisp), 2.94–3.33 (m, 2H, Ar-NH-*CH*<sub>2</sub>+2H, bisp), 4.01 (dd, *J*=15.6, 6.4 Hz, 1H, bisp), 4.23 (d, *J*=15.6 Hz, 1H, bisp), 5.36 (s, 1H, NH, collapses with D<sub>2</sub>O), 5.95 (d, *J*=6.8 Hz, 1H, α-pyr), 6.28 (d, *J*=5.4 Hz, 1H, arom), 6.50 (d, *J*= 9.2 Hz, 1H, α-pyr), 7.04 (d, *J*=9.2 Hz, 1H, α-pyr), 7.12–7.23 (m, 1H, arom), 7.46 (dd, *J*=8.8, 1.6 Hz, 1H, arom), 7.91 (d, *J*=1.6 Hz, 1H, arom), 8.49 ppm (d, *J*=5.4 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>22</sub>H<sub>23</sub>ClN<sub>4</sub>O·0.75 H<sub>2</sub>O (408.41): C 64.69, H 6.05, N 13.79, found: C 64.85, H 6.25, N 13.56.

**7-Chloro-4-{***N***-[**3-(citisin-3-yl)propyl]amino}quinoline (11): Yield: 74%; mp: 143–145 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.65–2.10 (m, 4H, bisp+2H CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.24–2.61 (m, 2H, NH-CH<sub>2</sub>+2H, bisp), 2.86–3.23 (m, 2H, Ar-NH-CH<sub>2</sub>+2H, bisp), 3.86–3.93 (dd, *J*=15.6, 6.4 Hz, 1H, bisp), 4.13–4,18 (d, J=15.6 Hz, 1H, bisp), 4.97 (s, 1H, NH, collapses with D<sub>2</sub>O), 6.01 (d, J=6.9 Hz, 1H,  $\alpha$ -pyr), 6.20 (d, J=5.5 Hz, 1H, arom), 6.46 (d, J=9.0 Hz, 1H,  $\alpha$ -pyr), 7.27–7.42 (m, 1H,  $\alpha$ -pyr+1H, arom), 7.56 (dd, J=8.8, 1.9 Hz, 1H, arom), 7.91 (d, J=1.9 Hz, 1H, arom), 8.45 ppm (d, J=5.5 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>23</sub>H<sub>25</sub>ClN<sub>4</sub>O (408.92): C 67.55, H 6.16, N 13.70, found: C 67.65, H 6.45, N 13.38.

**7-Chloro-4-{***N***-**[**5-**(**citisin-3-y**]**)penty**]**]amino}quinoline** (12): Yield: 80%; mp: 61–64 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 0.86–1.58 (m, 2H, bisp + 6H, *CH*<sub>2</sub>*CH*<sub>2</sub>*CH*<sub>2</sub>*)*, 1.62–2.06 (m, 2H, bisp), 2.12–2.53 (m, 2H, NH-*CH*<sub>2</sub>+3H, bisp), 2.71–3.32 (m, 2H, Ar-NH-*CH*<sub>2</sub>+1H, bisp), 3.84 (dd, *J* = 15.2, 6.4 Hz, 1H, bisp), 4.19 (d, *J* = 15.2 Hz, 1H, bisp), 5.85 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.04 (d, *J* = 6.8 Hz, 1H,  $\alpha$ -pyr), 6.34 (d, *J* = 5.14 Hz, 1H, arom), 6.45 (dd, *J* = 9.2, 1.4 Hz, 1H,  $\alpha$ -pyr), 7.21–7.37 (m, 1H,  $\alpha$ -pyr+1H, arom), 7.39 (dd, *J* = 8.8, 1.8 Hz, 1H, arom), 7.96 (d, *J* = 1.8 Hz, 1H, arom); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$  = 162.80, 150.94, 150.26, 149.46, 147.58, 137.95, 133.89, 126.81, 124.12, 121.37, 116.30, 115.20, 103.92, 97.56, 59.71, 58.98, 55.70, 49.22, 42.55, 34.62, 27.06, 26.94, 24.93, 24.61, 23.92 ppm; elemental analysis calcd (%) for C<sub>25</sub>H<sub>29</sub>ClN<sub>4</sub>O (436.98): C 68.71, H 6.69, N 12.82, found: C 68.89, H 6.90, N 12.81.

#### 7-Chloro-4-{N-[2-(citisin-3-yl)-1-methylethyl]amino}quinoline

(13): Yield: 53 %; mp: 221–223 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.13 (d, *J*=6.4 Hz, 3H, CHCH<sub>3</sub>), 1.63–2.02 (m, 3H, bisp), 2.33–2.66 (m, 2H, NH-*CH*<sub>2</sub>+2H, bisp), 2.77–3.12 (m, 3H, bisp), 3.55–3.78 (m, 1H, *CH*CH<sub>3</sub>), 3.82–4.04 (m, 2H, bisp), 5.27 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.06 (d, *J*=6.8 Hz, 1H,  $\alpha$ -pyr), 6.34 (d, *J*=5.6 Hz, 1H, arom), 6.53 (dd, *J*=9.2, 1.4 Hz, 1H,  $\alpha$ -pyr), 7.18–7.44 (m, 2H, arom, +1H,  $\alpha$ -pyr), 7.94 (d, *J*=0.8 Hz, 1H, arom), 8.49 ppm (d, *J*=5.4 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>23</sub>H<sub>25</sub>ClN<sub>4</sub>O (408.92): C 67.55, H 6.16, N 13.70, found: C 67.72, H 6.56, N 13.42.

General method for the preparation of 7-chloro-4-{[ $\omega$ -(9-nitrocytisin-3-yl)alkyl]amino}quinolines 14–16 and 7-chloro-4-{[ $\omega$ -(9,11dibromocytisin-3-yl)alkyl}quinolines 19 and 20: In an Aldrich pressure tube, the substituted cytisine<sup>[41,42]</sup> (0.61 mmol) was dissolved in CH<sub>3</sub>CN (4 mL). Subsequently, equimolar amounts of DIPEA and 4-[( $\omega$ -bromoalkyl)amino]-7-chloroquinoline 54–58 were sequentially added;<sup>[43–46]</sup> the closed tube was heated at 100 °C for 24 h. After cooling, the solvent was evaporated, and the residue was taken up with H<sub>2</sub>O (10 mL), alkalized, (2 N NaOH until strongly alkaline), and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×40 mL). The organic phase was dried (Na<sub>2</sub>SO<sub>4</sub>) and evaporated to dryness; the residue was finally chromatographed on alumina (1:30) eluting with CH<sub>2</sub>Cl<sub>2</sub>. The obtained compound was, eventually, further purified by trituration with dry Et<sub>2</sub>O.

**7-Chloro-4-{[2-(9-nitrocytisin-3-yl)ethyl]amino}quinoline** (14): Yield: 66%; mp: 182–184°C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.86–2.12 (m, 2H, bisp), 2.43–2.90 (m, 2H, NH-*CH*<sub>2</sub>+2H, bisp), 2.98–3.37 (m, 2H, Ar-NH-*CH*<sub>2</sub>+3 H, bisp), 4.07 (dd, *J* = 16.2, 6.6 Hz, 1H, bisp), 4.32 (d, *J* = 16.2 Hz, 1H, bisp), 5.16 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.04 (d, *J* = 8.2 Hz, 1H, α-pyr), 6.28 (d, *J* = 5.4 Hz, 1H, arom), 7.07 (d, *J* = 9.0 Hz, 1H, arom), 7.38 (dd, *J* = 8.0, 2.0 Hz, 1H, arom), 7.92 (d, *J* = 2.0 Hz, 1H, arom), 8.17 (d, *J* = 8.0 Hz, 1H, α-pyr), 8.49 ppm (d, *J* = 5.4 Hz, 1H arom); elemental analysis calcd (%) for C<sub>22</sub>H<sub>22</sub>ClN<sub>5</sub>O<sub>3</sub> (439.90): C 60.07, H 5.04, N 15.92, found: C 60.17, H 5.33, N 15.96.

**7-Chloro-4-{[3-(9-nitrocytisin-3-yl)propyl]amino}quinoline** (15): Yield: 77%; mp: 103–105°C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.65–2.08 (m, 2H, bisp+2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.32–2.62 (m, 2H, NH-CH<sub>2</sub>+2H, bisp), 2.91–3.22 (m, 2H, Ar-NH-CH<sub>2</sub>+3H, bisp), 4.01 (dd, *J* = 16.0, 6.4 Hz, 1H, bisp), 4.24 (d, *J* = 16.0 Hz, 1H, bisp), 4.98 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.13 (d, J=8.2 Hz, 1 H,  $\alpha$ -pyr), 6.24 (d, J= 5.4 Hz, 1 H, arom), 7.36 (dd, J=9.0, 2.2 Hz, 1 H,  $\alpha$ -pyr), 7.64 (d, J= 9.0 Hz, 1 H, arom), 7.96 (d, J=2.2 Hz, 1 H, arom), 8.33 (d, J=8.2 Hz, 1 H,  $\alpha$ -pyr), 8.51 ppm (d, J=5.4 Hz, 1 H, arom); elemental analysis calcd (%) for C<sub>23</sub>H<sub>24</sub>ClN<sub>5</sub>O<sub>3</sub>•0.5 H<sub>2</sub>O (462.93): C 59.67, H 5.44, N 15.13, found: C 59.94, H 5.75, N 14.75.

**7-Chloro-4-{[5-(9-nitrocytisin-3-yl)pentyl]amino}quinoline** (16): Yield: 74%; mp: 84–86°C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$ =0.82–2.04 (m, 2H, bisp+6H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.08–2.62 (m, 2H, NH-CH<sub>2</sub>+2H, bisp), 2.82–3.35 (m, 2H, Ar-NH-CH<sub>2</sub>+3H, bisp), 3.94 (dd, J=16.0, 6.4 Hz, 1H, bisp), 4.22 (d, J=16.0 Hz, 1H, bisp), 5.44 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.11 (d, J=8.2 Hz, 1H,  $\alpha$ -pyr), 6.34 (d, J=5.4 Hz, 1H, arom), 7.36 (dd, J=9.0, 2.2 Hz, 1H,  $\alpha$ -pyr), 7.82–7.99 (m, 2H, arom), 8.32 (d, J=8.2 Hz, 1H,  $\alpha$ -pyr), 8.51 ppm (d, J=5.4 Hz, 1H, arom); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$ =159.87, 154.30, 149.90, 149.41, 147.07, 136.73, 134.12, 133.54, 126.54, 124.29, 121.02, 116.08, 101.76, 97.66, 58.77, 58.54, 55.66, 50.39, 42.24, 35.52, 28.63, 27.22, 26.57, 24.89, 24.22, 23.62 ppm; elemental analysis calcd (%) for C<sub>25</sub>H<sub>28</sub>ClN<sub>5</sub>O<sub>3</sub> (481.98): C 62.30, H 5.86, N 14.53, found: C 62.33, H 5.87, N 14.25.

7-Chloro-4-{[3-(9,11-dibromocytisin-3-yl)propyl]amino}quinoline

(19): Yield: 68%; mp: 103–105 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.47–1.98 (m, 4H, bisp+2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.24–2.53 (m, 2H, NH-CH<sub>2</sub>+2H, bisp), 2.94–3.13 (m, 2H, Ar-NH-CH<sub>2</sub>+1H, bisp), 3.96 (dd, J=16.0, 6.2 Hz, 1H, bisp), 4.17 (d, J=16.0 Hz, 1H, bisp), 4.86 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.21 (d, J=5.5 Hz, 1H, arom), 7.37 (dd, J= 8.8, 2.0 Hz, 1H, arom), 7.58 (d, J=8.8 Hz, 1H, arom), 7.82 (s, 1H, α-pyr), 7.95 (d, J=2.0 Hz, 1H, arom), 8.54 ppm (d, J=5.5 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>23</sub>H<sub>23</sub>Br<sub>2</sub>ClN<sub>4</sub>O (566.72): C 48.75, H 4.09, N 9.89, found: C 48.67, H 4.19, N 9.63.

7-Chloro-4-{[5-(9,11-dibromocytisin-3-yl)pentyl]amino}quinoline

(20): Yield: 69%; mp: 104–106 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 0.92–2.01 (m, 4H, bisp+6H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.08–2.52 (m, 2H, NH-CH<sub>2</sub>+1H, bisp), 2.74–3.52 (m, 2H, Ar-NH-CH<sub>2</sub>+3H, bisp), 3.92 (dd, J = 16.0, 6.2 Hz, 1H, bisp), 4.20 (d, J = 16.0 Hz, 1H, bisp), 5.41 (s, 1H, NH, collapsed with D<sub>2</sub>O), 7.21 (d, J = 5.5 Hz, 1H, arom), 7.39 (dd, J = 9.0, 2.2 Hz, 1H, arom), 7.86 (s, 1H,  $\alpha$ -pyr), 7.95 (d, J = 2.0 Hz, 1H, arom), 8.50 ppm (d, J = 5.5 Hz, 1H, arom); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta$  = 157.92, 149.68, 147.66, 146.92, 142.69, 134.27, 126.49, 124.39, 121.07, 116.09, 111.29, 97.70, 96.03, 58.49, 55.91, 55.64, 51.72, 42.47, 33.89, 27.25, 26.82, 24.86, 23.79 ppm; elemental analysis calcd (%) for C<sub>25</sub>H<sub>27</sub>Br<sub>2</sub>ClN<sub>4</sub>O·0.5H<sub>2</sub>O (603.78): C 49.72, H 4.67, N 9.18, found: C 49.60, H 5.04, N 8.92.

**7-Chloro-4-(pyrrolidin-1-yl)quinoline (21)**: Reaction of 4-[(4-bro-mobutyl)amino]-7-chloroquinoline with cytisine or 9-nitrocytisine, either in the presence or absence of DIPEA, only the intramolecular loss of hydrogen bromide was obtained to give. Yield: 88%; mp: 90–91 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.82–2.21 (m, 4H, CH<sub>2</sub>CH<sub>2</sub>), 3.42–3.56 (m, 4H, N-CH<sub>2</sub>), 6.44 (d, *J*=5.4 Hz, 1H, arom), 7.28 (dd, *J*=9.2, 2.2 Hz, 1H, arom), 7.95 (d, *J*=2.2 Hz, 1H, arom), 8.16 (d, *J*= 9.2 Hz, 1H, arom), 8.47 ppm (d, *J*=5.4 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>13</sub>H<sub>13</sub>ClN<sub>2</sub> (232.71): C 67.10, H 5.63, N 12.04%, found: C 67.31, H 5.88, N 12.10%.

General method for the preparation of 4-{*N*-[ $\omega$ -(9-aminocytisin-3-yl)alkyl]amino}-7-chloroquinolines 17 and 18: Nitro compound 15 or 16 (0.25–0.44 mmol) was dissolved in EtOH (40 mL), and 10% palladium on charcoal (20 mg) was added. The mixture was hydrogenated at atmospheric pressure. The catalyst was filtered, and the solvent was removed under vacuum. The residue was rinsed with dry Et<sub>2</sub>O (3 mL) to give the title compound as a white powder that rapidly became pale yellow.

#### 4-{*N*-[3-(9-Aminocytisin-3-yl)propyl]amino}-7-chloroquinoline

(17): Yield: 86%; mp: 173-175°C; <sup>1</sup>H NMR (300 MHz, [D<sub>6</sub>]DMSO):  $\delta = 0.78-1.98$  (m, 4H, bisp + 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.04–2.45 (m, 2H, NH-CH<sub>2</sub>+1H, bisp), 2.69–3.58 (m, 2H, Ar-NH-CH<sub>2</sub>+3H, bisp), 3.74 (dd, J = 16.0, 6.2 Hz, 1H, bisp), 3.95 (d, J = 16.0 Hz, 1H, bisp), 4.67 (s, 1H, NH, collapsed with D<sub>2</sub>O), 5.89 (d, J = 7.4 Hz, 1H,  $\alpha$ -pyr), 6.12 (d, J = 5.5 Hz, 1H, arom), 6.43 (d, J = 7.4 Hz, 1H,  $\alpha$ -pyr), 7.15 (s, 2H, NH<sub>2</sub>, collapsed with D<sub>2</sub>O) 7.40 (dd, J = 9.0, 1.8 Hz, 1H, arom), 7.72 (d, J = 1.8 Hz, 1H, arom), 8.21 (d, J = 9.0 Hz, 1H, arom), 8.34 ppm (d, J = 5.5 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>23</sub>H<sub>26</sub>ClN<sub>5</sub>O (423.94): C 65.16, H 6.18, N 16.52, found: C 64.72, H 6.38, N 16.29.

#### 4-{*N*-[5-(9-Aminocytisin-3-yl)pentyl]amino}-7-chloroquinoline

(18): Yield: 76%; mp: 137–139°C; <sup>1</sup>H NMR (300 MHz, [D<sub>6</sub>]DMSO):  $\delta = 0.78-2.01$  (m, 4H, bisp + 6H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.11–2.42 (m, 2H, NH-CH<sub>2</sub>+1H, bisp), 2.64–3.49 (m, 2H, Ar-NH-CH<sub>2</sub>+3H, bisp), 3.73 (dd, J = 16.0, 6.2 Hz, 1H, bisp), 3.90 (d, J = 16.0 Hz, 1H, bisp), 4.76 (s, 1H, NH, collapsed with D<sub>2</sub>O), 5.84 (d, J = 7.4 Hz, 1H,  $\alpha$ -pyr), 6.32–6.47 (m, 1H, arom+1H,  $\alpha$ -pyr), 7.19 (s, 2H, NH<sub>2</sub>, collapsed with D<sub>2</sub>O) 7.40 (dd, J = 9.0, 2.2 Hz, 1H, arom), 7.74 (d, J = 2.0 Hz, 1H, arom), 8.24 (d, J = 9.0 Hz, 1H, arom), 8.36 ppm (d, J = 5.5 Hz, 1H, arom); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>):  $\delta = 157.23, 150.27, 149.43, 147.58, 138.19, 133.89, 133.20, 126.86, 124.10, 121.17, 116.31, 112.67, 103.59, 97.59, 60.13, 59.03, 55.31, 49.52, 42.44, 33.99, 28.63, 27.16, 26.87, 25.55, 24.50, 23.61 ppm; elemental analysis calcd (%) for C<sub>25</sub>H<sub>30</sub>ClN<sub>5</sub>O (451.99): C 66.43, H 6.69, N 15.49, found: C 66.72, H 6.62, N 15.12.$ 

General method for the preparation of 4-[(1*R*,9a*R*)-(octahydro-2*H*-quinolizin-1-yl)methyl)thio]-7-substituted quinolines 22 and 23: A suspension of 4,7-dichloroquinoline (0.51 g, 2.6 mmol) or 4chloro-7-trifluoromethylquinoline (0.6 g) in Dowtherm A (4 mL) was heated at 100 °C until a clear solution was obtained, and then a solution of thiolupinine<sup>[35]</sup> (0.48 g, 2.6 mmol) in Dowtherm A (1 mL) was added. The tube was closed and the temperature was raised and maintained at 170 °C for 2 h. After cooling, the pasty material was partitioned between Et<sub>2</sub>O and 0.5 N HCl (30 mL each). The aqueous phase was extracted with Et<sub>2</sub>O (3×40 mL), alkalized, (2 N NaOH (20 mL) and extracted again with Et<sub>2</sub>O (3×40 mL). The dried Et<sub>2</sub>O solution was evaporated to leave an oil that slowly crystallized.

#### 7-Chloro-4-[(1R,9aR)-(octahydro-2H-quinolizin-1-yl)methyl)thio]-

**quinoline (22)**: Yield: 61%; mp: 114–115°C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.16–2.28 (m, 14H, Q), 2.75–3.02 (m, 2H, CH<sub>2</sub>S), 3.18–3.54 (m, 2H<sub>a</sub> near N of Q), 7.21 (d, *J*=5.0 Hz, 1H, arom), 7.47 (dd, *J*=9.0, 2.2 Hz, 1H, arom), 8.03–8.08 (m, 2H, arom), 8.67 ppm (d, *J*=5.0 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>19</sub>H<sub>23</sub>ClN<sub>2</sub>S (346.92): C 65.78, H 6.68, N 8.07, found: C 66.02, H 6.78, N 8.11. Elemental analysis calcd (%) for C<sub>19</sub>H<sub>23</sub>ClN<sub>2</sub>S (346.92): C 58.78, H 6.68, N 8.07, found: C 66.02, H 6.78, N 8.11. Elemental analysis calcd (%) for C<sub>19</sub>H<sub>23</sub>ClN<sub>2</sub>S·HCl·0.5H<sub>2</sub>O (392.39): C 58.16, H 6.42, N 7.14, found: C 58.45, H 6.36, N 7.13.

**4-{[(1***R***,9***aR***)-(octahydro-2***H***-quinolizin-1-yl)methyl]thio}-7-trifluoromethylquinoline (23): Yield: 69%; mp: 78–79°C (pentane); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>) \delta = 1.06–2.24 (m, 14H, Q), 2.70–2.99 (m, 2H, CH<sub>2</sub>S), 3.14–3.50 (m, 2H<sub>a</sub> near N of Q), 7.32 (d,** *J***=5.5 Hz, 1H, arom), 7.69 (dd,** *J***=8.8, 2.0 Hz, 1H, arom), 8.25 (d,** *J***=8.8 Hz, 1H, arom), 8.35 (s, 1H, arom), 8.77 ppm (d,** *J***=5.5 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>20</sub>H<sub>23</sub>F<sub>3</sub>N<sub>2</sub>S (380.47): C 63.14, H 6.09, N 7.36, found: C 63.18, H 6.10, N 7.35.** 

General method for the preparation of *S*-(7-Chloro-4-quinolinyl)- $\omega$ -{[(1*R*,9a*R*)-(octahydro-2*H*-quinolizin-1-yl)methyl]thio}alkanethiols 24 and 25: A mixture of 7-chloro-4-thioquinoline<sup>[47]</sup> (0.21 g, 1.09 mmol) and KOH (ground pellets, 61 mg, 1.09 mmol) in CH<sub>3</sub>CN (5 mL) was stirred for 30 min under an atmosphere of nitrogen. A solution of an equimolar amount of *S*-(2-chloroethyl)thiolupinine<sup>[32]</sup> (63) or S-(3-bromopropyl)thiolupinine<sup>[48]</sup> (64) in CH<sub>3</sub>CN (3 mL) was dropwise added slowly, and the solution was stirred at room temperature for 24 h. The solvent was removed, and the residue was taken up in 1 N HCl (10 mL). The acid solution was extracted with Et<sub>2</sub>O (2×10 mL), alkalized (2 N NaOH, 15 mL), and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×10 mL). The dried solution was evaporated, and the oily residue was subjected to chromatography on alumina (1:30) eluting with CH<sub>2</sub>Cl<sub>2</sub>.

**S-(7-Chloroquinolin-4-yl)-2-{[(1***R***,9a***R***)-(octahydro-2***H***-quinolizin-1-yl)methyl]thio}ethanethiol (24): Yield: 43%. Yellow oil; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): \delta = 1.04-2.12 (m, 14H, Q), 2.66–2.98 (m, 2H, CH<sub>2</sub>S+2H, ArSCH<sub>2</sub>S+2H, SCH<sub>2</sub>), 3.14–3.50 (m, 2H, 2H<sub>a</sub> near N of Q), 7.19 (dd, J = 4.7, 1.6 Hz, 1H, arom), 7.49 (dd, J = 9.0, 2.0 Hz, 1H, arom), 7.96–8.15 (m, 2H, arom), 8.72 ppm (dd, J = 4.7, 1.6 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>21</sub>H<sub>27</sub>ClN<sub>2</sub>S<sub>2</sub> (407.04): C 61.97, H 6.69, N 6.88, S 15.76, found: C 61.80, H 6.75, N 6.83, S 15.28.** 

**S-(7-Chloroquinolin-4-yl)-3-{**[(1*R*,9*aR*)-(octahydro-2*H*-quinolizin-1-yl)methyl]thio}propanethiol (25): Yield: 40%; mp: 81–83 °C (dry Et<sub>2</sub>O); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$ =0.95–2.21 (m, 14H, Q+2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.56–2.95 (m, 2H, CH<sub>2</sub>S+2H, ArSCH<sub>2</sub>S+2H, SCH<sub>2</sub>), 3.11–3.35 (m, 2H, 2H<sub>a</sub> near N of Q), 7.21 (d, J=5.0 Hz, 1H, arom), 7.49 (dd, J=8.8, 2.0 Hz, 1H, arom), 7.97–8.12 (m, 2H, arom), 8.70 ppm (d, J=5.0 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>22</sub>H<sub>29</sub>ClN<sub>2</sub>S<sub>2</sub> (421.06): C 62.75, H 6.94, N 6.65, S 15.23, found: C 62.51, H 7.19, N 6.68, S 15.38. Elemental analysis calcd (%) for C<sub>22</sub>H<sub>29</sub>ClN<sub>2</sub>S<sub>2</sub>+Hcl·1.25H<sub>2</sub>O (405.90): C 55.07, H 6.77, N 5.84, S 13.36, found: C 55.21, H 6.93, N 5.56, S 11.98.

General method for the preparation of 7-chloro-4-{5-[ $\omega$ -(citisin-3-yl)alkyl]thio}quinolines 26 and 27: A mixture of 7-chloro-4-thioquinoline<sup>[47]</sup> (0.28 g, 1.43 mmol) and KOH (ground pellets, 80 mg, 1.43 mmol) in CH<sub>3</sub>CN (5 mL) was stirred for 30 min under an atmosphere of nitrogen. A solution of an equimolar amount of *N*-(2chloroethyl)cytisine<sup>[49]</sup> (65) or *N*-(4-chloropropyl)cytisine<sup>[49]</sup> (66) in CH<sub>3</sub>CN (5 mL) was dropwise slowly added, and the solution was stirred at room temperature for 72 h. The solvent was removed, and the residue was taken up in 1  $\times$  HCl (5 mL). The acid solution was filtered, alkalized (2  $\times$  NaOH until strongly alkaline) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×20 mL). The dried solution was concentrated, and the oily residue was subjected to chromatography on alumina (1:30) eluting with CH<sub>2</sub>Cl<sub>2</sub> and CH<sub>2</sub>Cl<sub>2</sub>/MeOH (98:2). Finally, the solid was crystallized with dry Et<sub>2</sub>O.

**7-Chloro-4-{S-[3-(citisin-3-yl)propyl]thio}quinoline** (26): Yield: 54%; mp: 140–141 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.78–2.16 (m, 3H, bisp+2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.41–2.72 (2H, S-*C*H<sub>2</sub>+3H, bisp), 2.84–3.21 (m, 2H, NH-*C*H<sub>2</sub>+2H, bisp), 4.04 (dd, *J*=15.6, 6.4 Hz, 1H, bisp), 4.29 (d, *J*=15.6 Hz, 1H, bisp), 6.12 (d, *J*=6.8 Hz, 1H, α-pyr), 6.51 (d, *J*=5.4 Hz, 1H, arom), 7.11 (d, *J*=5.4 Hz, 1H, arom), 7.32 (m, 1H, arom), 7.64 (dd, *J*=8.8, 1.6 Hz, 1H, arom), 8.07–8.28 (m, 2H, arom), 8.84 ppm (d, *J*=5.0 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>23</sub>H<sub>24</sub>ClN<sub>3</sub>OS (425.97): C 64.85, H 5.68, N 9.86, S 7.53, found: C 64.66, H 5.66, N 10.05, S 7.53.

**7-Chloro-4-{S-[4-(citisin-3-yl)butyl]thio}quinoline (27)**: Yield: 43%; mp: 100–101°C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.56–2.17 (m, 2H, bisp + 4H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 2.33–2.66 (2H, S-*CH*<sub>2</sub> + 3H, bisp), 2.88– 3.19 (m, 2H, NH-*CH*<sub>2</sub> + 2H, bisp), 4.02 (dd, *J* = 15.6, 6.4 Hz, 1H, bisp), 4.22 (d, *J* = 15.6 Hz, 1H, bisp), 6.11 (d, *J* = 6.8 Hz, 1H, α-pyr), 6.56 (d, *J* = 9.0 Hz, 1H, arom), 7.22 (d, *J* = 5.0 Hz, 1H, arom), 7.26– 7.42 (m, 1H, arom), 7.65 (dd, *J* = 9.0, 2.2 Hz, 1H, arom), 8.15–8.31 (m, 2H, arom), 8.84 ppm (d, *J* = 5.0 Hz, 1H, arom); elemental analysis calcd (%) for  $C_{24}H_{26}CIN_3OS$  (440.00): C 65.51, H 5.96, N 9.55, S 7.29, found: C 65.42, H 5.86, N 9.54, S 7.43.

ω-(Octahydro-quinolizin-1/9a-yl)alkylamines, intermediates for compounds 1–8: Amines 29, 32, 41, and 42 were prepared as previously described by Tasso et al.,<sup>[32]</sup> Villa et al.,<sup>[33]</sup> and Barteselli et al.,<sup>[36]</sup> whereas amine 35, 38, and 43 were prepared as follows.

#### 1-{[(1R,9aR)-(Octahydro-2H-quinolizin-1-yl)methyl]thio}-2-oxo-

**propane (33)**: Chloroacetone (0.75 mL, 6.5 mmol) was added to a solution of thiolupinine<sup>[35]</sup> (1 g, 5.4 mmol) in EtOH (5 mL) under protection from air, and the mixture was stirred at room temperature for 1 h. The solvent was removed under reduced pressure, and the residue was partitioned between Et<sub>2</sub>O (10 mL) and 1 N HCI (20 mL). The acid solution was alkalized with 2 N NaOH (15 mL) and extracted with Et<sub>2</sub>O (4×10 mL). The Et<sub>2</sub>O solution was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to leave a pale-yellow oil (1.23 g, 95%); elemental analysis calcd (%) for C<sub>13</sub>H<sub>23</sub>NOS (241.39): C 64.68, H 9.60, N, 5.80, found: C 64.81, H 9.61, N 5.65.

2-Hydroxyimino-1-{[(1R,9aR)-(Octahydro-2H-quinolizin-1-yl)me-

**thyl]thio}propane (34):**  $6 \times \text{NaOH} (1.1 \text{ mL})$  was added to a solution of ketone **33** (1.23 g, 5.1 mmol) and hydroxylamine hydrochloride (0.43 g, 6.12 mmol) in EtOH (4 mL), and the solution was heated at reflux for 2 h and then evaporated under reduced pressure. The residue was taken up in H<sub>2</sub>O (10 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 10 mL). After removing the solvent, a pale-yellow oil (1.28 g, 97%) that crystallized on standing was recovered; mp: 79–80 °C (pentane); elemental analysis calcd (%) for C<sub>13</sub>H<sub>24</sub>N<sub>2</sub>OS (256.41): C 60.89, H 9.43, N 10.93, found: C 61.15, H 9.44, N 10.86.

**2-Amino-1-[[(1***R***,9***aR***)-(<b>Octahydro-2***H***-quinolizin-1-yl)methyl]thio}propane (35)**: A solution of oxime **34** (0.90 g, 3.5 mmol) in dry Et<sub>2</sub>O (20 mL) was added dropwise to a stirred and ice-cooled suspension of lithium aluminum hydride (0.65 g, 17 mmol) in dry Et<sub>2</sub>O (20 mL), and then the suspension was heated at reflux for 20 h. After cooling, H<sub>2</sub>O (1 mL), 15% NaOH solution (1 mL), and H<sub>2</sub>O (1.5 mL) were sequentially added. The precipitate was filtered and washed thoroughly with Et<sub>2</sub>O (3×10 mL), and the organic solution was dried with KOH pellets. After elimination of the solvent, the oily residue was distilled in vacuo (0.07 hPa) collecting a colorless oil (0.73 g, 86%) at 140°C (air bath temperature); elemental analysis calcd (%) for C<sub>13</sub>H<sub>26</sub>N<sub>2</sub>S (242.42): C 64.40, H 10.81, N 11.56, found: C 64.57, H 10.97, N 11.52.

**1-{[(1***R***,9***aR***)-(Octahydro-2***H***-quinolizin-1-yl)methyl]thio}-3-oxobutane (36): A solution of methyl vinyl ketone (1.4 mL, 17.5 mmol) in dry dioxane (1.5 mL) was added dropwise to a solution of thiolupinine<sup>[35]</sup> (3.04 g, 16.3 mmol) in dry dioxane (2.5 mL). The solution was stirred at room temperature for 1 h and then maintained at 70 °C for 30 min, always under an atmosphere of nitrogen. After cooling, the solution was diluted with Et<sub>2</sub>O (20 mL) and extracted with 0.5 N HCl (3×15 mL). The acid solution was strongly alkalized (2 N NaOH (30 mL)) and extracted with Et<sub>2</sub>O (3×20 mL). After removing the solvent, the oily residue was distilled at 0.07 hPa collecting a pale-yellow oil (3.92 g, 93%) at 140 °C (air bath temperature); elemental analysis calcd (%) for C<sub>14</sub>H<sub>25</sub>NOS (255.42): C 65.86, H 9.87, N 5.48, found: C 65.76, H 9.99, N 5.63.** 

#### 3-Hydroxyimino-1-{[(1R,9aR)-(octahydro-2H-quinolizin-1-yl)me-

**thyl]thio}butane (37)**: Ketone **35** (3.48 g, 13.6 mmol) was converted into the corresponding oxime by treatment with hydroxylamine hydrochloride (1.14 g, 16.3 mmol) and  $6 \times \text{NaOH}$  solution (2.7 mL) under the conditions used for oxime **34**. Thus, a pale-yellow oil (3.55 g, 96%) was obtained. Elemental analysis calcd (%) for

 $C_{14}H_{26}N_2OS$  (270.43): C 62.17, H 9.69, N 10.36, found: C 62.22, H 9.78, N 10.36.

**3-Amino-1-{[(1***R***,9***aR***)-(octahydro-2***H***-quinolizin-1-yl)methyl]thio}butane (38): Following the procedure described for the preparation of amino compound 35, oxime 37 (2.25 g, 8.3 mmol) was reduced with lithium aluminum hydride (1.57 g, 41 mmol). Thus, after distillation at 0.07 hPa (140 °C, air bath temperature) amine 38 (1.92 g, 90%) was obtained. Elemental analysis calcd (%) for C<sub>14</sub>H<sub>28</sub>N<sub>2</sub>S (256.45): C 65.57, H 11.00, N 10.93, found: C 65.33, H 11.34, N 10.74.** 

9a-(Oct-7-enyl)-octahydro-2H-quinolizine (45): A solution of 8bromo-1-octene (3 g, 15.7 mmol) in anhydrous THF (4.5 mL) was added dropwise to a mixture of granular magnesium (15.7 mmol) and a catalytic amount (30 mg) of zinc bromide in dry THF (9 mL). After the addition of a crystal of iodine to trigger the reaction, the mixture was stirred for 4 h at 50 °C under an atmosphere of nitrogen. Once the Grignard compound was completely formed, dry THF (30 mL) and octahydroquinolizidinium perchlorate<sup>[37]</sup> (1.89 g, 7.85 mmol) were added, and the mixture was heated at reflux for 20 h. After cooling, the mixture was diluted with saturated aqueous solution of NH<sub>4</sub>Cl (9 mL) and acidified with 6 N HCl (to pH 2). The aqueous layer was extracted with Et<sub>2</sub>O (3×10 mL), then alkalized and saturated with K<sub>2</sub>CO<sub>3</sub> and extracted several times with  $Et_2O$  (4×20 mL). The collected organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to dryness to give a yellow oil (790 mg, 54%); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.20-2.26$  (m, 22 H), 2.39–2.92 (m, 6H), 4.91-5.03 (m, 2H), 5.77-5.82 ppm (m, 1H).

8-(Octahydro-2H-quinolizin-9a-yl)octan-1-ol (46): A solution of compound 45 (750 mg) in anhydrous THF (13.5 mL) was added dropwise over 30 min to an ice-cooled mixture of 1 M borane-THF complex solution (12.6 mL, 12.6 mmol) and 2-methyl-2-butene (1.76 g, 70.1 mmol) diluted with dry THF (2.4 mL). The mixture was stirred at room temperature for 4.5 h and then diluted with  $H_2O$ (3.7 mL), 6 N NaOH (12.5 mL), and H<sub>2</sub>O<sub>2</sub> (35% solution, 3.1 mL). The mixture was stirred at room temperature for 1 h and then saturated with  $K_2CO_3$  and extracted with  $Et_2O$  (4×20 mL). The collected organic layer was concentrated to dryness, and the residue was diluted with Et<sub>2</sub>O (50 mL) and extracted with 4 N HCl (3×30 mL). The aqueous layer was alkalized, saturated with K<sub>2</sub>CO<sub>3</sub>, and extracted with  $Et_2O$  (4×20 mL). The organic phase was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to dryness to give an oil that was purified by column chromatography (neutral Al<sub>2</sub>O<sub>3</sub>, grade IV; PE/Et<sub>2</sub>O in gradient). Oil (380 mg, 48%); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.08-1.69$  (m, 26 H), 2.38-2.42 (m, 2H), 2.55-2.64 (m, 2H), 3.62-3.66 ppm (m, 2H). One H is not visible, probably because of its interaction with the solvent.

#### 2-[8-(Octahydro-2*H*-quinolizin-9a-yl)octyl]isoindoline-1,3-dione

(47): Diethylazodicarboxylate (DEAD; 0.42 mL diluted with 1.1 mL of anhydrous THF) was added dropwise to an ice-cooled mixture of the above alcohol (340 mg, 1.27 mmol), phthalimide (374 mg, 2.54 mmol), triphenylphosphine (700 mg, 2.67 mmol), and dry THF (3.9 mL). The resulting mixture was stirred at room temperature for 20 h. The solvent was evaporated, and the crude oil was purified by column chromatography (silica gel; CH<sub>2</sub>Cl<sub>2</sub>/MeOH/conc. NH<sub>3</sub> 96:3.6:0.4). Pale-yellow oil (490 mg, 96%); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.09–1.67 (m, 24 H), 2.52–2.66 (m, 4H), 3.48–3.70 (m, 4H), 7.69–7.73 (m, 2H arom), 7.81–7.85 ppm (m, 2H, arom).

8-(Octahydro-2*H*-quinolizin-9a-yl)octane-1-amine dihydrochloride (43): The above phthalimide derivative 47 (478 mg, 1.20 mmol) was dissolved in  $6 \times$  HCl (4.7 mL) and heated at reflux for 20 h. After ice cooling, the precipitated phthalic acid was filtered and washed with ice-cold H<sub>2</sub>O (2×5 mL). The aqueous solution was concentrated under vacuum to dryness to afford a brownish, very hygroscopic powder (398 mg, 84%); <sup>1</sup>H NMR (300 MHz, [D<sub>6</sub>]DMSO):  $\delta$ =1.01–1.93 (m, 24H), 2.72–3.45 (m, 8H), 7.99 (brs, 2H, collapsed with D<sub>2</sub>O), 10.27 ppm (brs, 2H, collapsed with D<sub>2</sub>O). Free base: pale yellow oil; <sup>1</sup>H NMR (300 MHz, [D<sub>6</sub>]DMSO):  $\delta$ =0.94–1.03 (m, 4H), 1.25–1.51 (m, 20H), 2.24–2.52 (m, 8H), 3.33 ppm (brs, 2H, collapsed with D<sub>2</sub>O).

General method for the preparation of 7-chloro-4-[( $\omega$ -hydroxyal-kyl)amino]quinolines 49–53: 4,7-Dicholoroquinoline (0.6 g, 3 mmol) was mixed with the  $\omega$ -hydroxyalkylamine (9 mmol) and heated in an Aldrich pressure tube at 150 °C for 3 h. After cooling, the mixture was taken up with H<sub>2</sub>O (10 mL), and the insoluble title compound was filtered and washed with H<sub>2</sub>O (2×5 mL) and finally crystallized from MeOH. Compounds 49–51 and 53 are already known.<sup>[43–46]</sup>

**7-Chloro-4-[(2-hydroxyethyl)amino]quinoline (49)**: Yield: 86%; mp: 214–215  $^{\circ}$ C (lit.<sup>[43]</sup> 214  $^{\circ}$ C).

**7-Chloro-4-[(3-hydroxypropyl)amino]quinoline (50)**: Yield: 73%; mp: 149–150 °C (lit.<sup>[44]</sup> 148–148.5 °C).

**7-Chloro-4-[(4-hydroxybutyl)amino]quinoline (51)**: Yield: 68%; mp: 176–178 °C (lit.<sup>[45]</sup> 171–174 °C).

**7-Chloro-4-[(5-hydroxypentyl)amino]quinoline (52)**: Yield: 82%; mp: 148–149°C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.42–1.98 (m, 6H, *CH*<sub>2</sub>*CH*<sub>2</sub>*CH*<sub>2</sub>*t* + 1H, OH) 3.36 (m, 2H, NH*CH*<sub>2</sub>), 3.75 (t, *J*=6.0 Hz, 2H, *CH*<sub>2</sub>O), 5.13 (s, 1H, NH, collapsed with D<sub>2</sub>O), 6.42 (d, *J*=5.4 Hz, 1H, arom), 7.37 (dd, *J*=9.0, 2.2 Hz, 1H, arom), 7.69 (d, *J*=9.0 Hz, 1H, arom), 7.98 (d, *J*=2.0 Hz, 1H, arom), 8.54 ppm (d, *J*=5.4 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>14</sub>H<sub>17</sub>ClN<sub>2</sub>O (264.75): C 63.51, H 6.47, N 10.58, found: C 63.39, H 6.70, N 10.43.

**7-Chloro-4-[(2-hydroxy-1-methylethy)amino]quinoline (53)**: Yield: 82%; mp: 207–209 °C (lit.<sup>[46]</sup> 208–210 °C).

General method for the preparation of 4-[( $\omega$ -bromoalkyl)amino]-7-chloroquinolines 53–58: An Aldrich pressure tube was sequentially charged with 7-chloro-4-(hydroxyalkylamino)quinoline (2 mmol), 48% hydrobromic acid (1.5 mL), and concentrated sulfuric acid (0.5 mL). The closed tube was heated at 160 °C for 3.5 h. After cooling, the mixture was poured on H<sub>2</sub>O and ice and then was alkalized with 2 N NaOH solution (20 mL). The mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×30 mL); the organic phase was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was finally crystallized from toluene. Compounds **54**, **55**, and **58** are already known.<sup>[43,44,46]</sup>

**4-[(2-Bromoethyl)amino]-7-chloroquinoline (54)**: Yield: 74%; mp: 139–140 °C (lit.<sup>[43]</sup> 139–140 °C).

**4-[(3-Bromopropyl)amino]-7-chloroquinoline (55)**: Yield: 71 %; mp: 158–160 °C (lit.<sup>[44]</sup> 159–160 °C).

**4-[(4-Bromobutyl)amino]-7-chloroquinoline (56)**: Yield: 66%; mp: 220–222 °C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.42–1.98 (m, 4H, *CH*<sub>2</sub>*CH*<sub>2</sub>), 3.30–3.56 (m, 2H, NH*CH*<sub>2</sub> + 2H, *CH*<sub>2</sub>Br), 5.09 (s, 1 H, NH, collapsed with D<sub>2</sub>O), 6.44 (d, *J* = 5.4 Hz, 1H, arom), 7.38 (dd, *J* = 9.0, 2.0 Hz, 1H, arom), 7.71 (d, *J* = 9.0 Hz, 1H, arom), 8.01 (d, *J* = 2.0 Hz, 1H, arom), 8.59 ppm (d, *J* = 5.2 Hz, 1H, arom); elemental analysis calcd (%) for C<sub>13</sub>H<sub>14</sub>BrClN<sub>2</sub> (313.62): C 49.79, H 4.50, N 8.93, found: C 46.69, H 4.33, N 9.18.

**4-[(5-Bromopentyl)amino]-7-chloroquinoline (57)**: Yield: 63%; mp: 112–113°C; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>):  $\delta$  = 1.42–1.98 (m, 6H, *CH*<sub>2</sub>*CH*<sub>2</sub>*CH*<sub>2</sub>), 3.29–3.58 (m, 2H, NH*CH*<sub>2</sub>+2H, *CH*<sub>2</sub>Br), 5.02 (s, 1H, NH,



collapsed with D<sub>2</sub>O), 6.45 (d, J=5.4 Hz, 1 H, arom), 7.40 (dd, J=9.2, 1.8 Hz, 1 H, arom), 7.69 (d, J=9.2 Hz, 1 H, arom), 8.00 (d, J=1.8 Hz, 1 H, arom), 8.57 ppm (d, J=5.4 Hz, 1 H, arom); elemental analysis calcd (%) for C<sub>14</sub>H<sub>16</sub>BrClN<sub>2</sub> (327.65): C 51.32, H 4.92, N 8.55, found: C 51.30, H 5.28, N 8.20.

**4-[(2-Bromo-1-methylethyl)amino]-7-chloroquinoline (58)**: Yield: 53%; mp: 128–130 °C (lit.<sup>[46]</sup> 132–133 °C).

#### **Biological methods**

Parasite cultures and drug susceptibility assays: P. falciparum cultures were performed according to Trager and Jensen<sup>[54]</sup> with slight modifications. The CQ-susceptible strain D10 and the CQ-resistant strain W2 were maintained at 5% hematocrit (human type A-positive red blood cells) in RPMI 1640 (EuroClone, Celbio) medium with the addition of 1% AlbuMax (Invitrogen, Milan, Italy), 0.01% hypoxanthine, 20 mм HEPES, and 2 mм glutamine. All the cultures were maintained at 37  $^{\circ}$ C in a standard gas mixture consisting of 1 % O<sub>2</sub>, 5% CO<sub>2</sub>, and 94% N<sub>2</sub>. Compounds were dissolved in either water or DMSO and then diluted with medium to achieve the required concentrations (final DMSO concentration < 1 %, which is nontoxic to the parasite). Drugs were placed in 96-well flat-bottomed microplates (COSTAR) and serial dilutions made. Asynchronous cultures with parasitemia of 1-1.5% and 1% final hematocrit were aliquotted into the plates and incubated for 72 h at 37 °C. Parasite growth was determined spectrophotometrically (OD650) by measuring the activity of the parasite lactate dehydrogenase, according to a modified version of the method of Makler in control and drug-treated cultures.  $^{\scriptscriptstyle [55,56]}$  The antimalarial activity is expressed as 50% inhibitory concentrations ( $IC_{50}$ ); each  $IC_{50}$  value is the mean and standard deviation of at least three separate experiments performed in duplicate.

Cytotoxicity assays: The long-term human microvascular endothelial cell line (HMEC-1) immortalized by SV40 large T antigen was maintained in MCDB131 medium (Invitrogen, Milan, Italy) supplemented with 10% fetal calf serum (HyClone, Celbio, Milan, Italy), 10 ng mL<sup>-1</sup> of epidermal growth factor (Chemicon),  $1 \mu g m L^{-1}$  of hydrocortisone, 2 mM glutamine, 100 U mL<sup>-1</sup> of penicillin, 100  $\mu$ g mL<sup>-1</sup> of streptomycin, and 20 mM HEPES buffer (EuroClone). Human Caucasian hepatocyte carcinoma (HepG2) was maintained in DMEM medium, supplemented with 10% fetal calf serum, 2 mm glutamine, 100 U mL<sup>-1</sup> of penicillin, 100  $\mu$ g mL<sup>-1</sup> of streptomycin, and 20 mm HEPES buffer. For the cytotoxicity assays, cells were treated with serial dilutions of test compounds for 72 h, and cell proliferation was evaluated by using the MTT assay already described.<sup>[54]</sup> The results are expressed as IC<sub>50</sub>, which is the concentration of compound necessary to inhibit cell growth by 50%. Each IC<sub>50</sub> value is the mean and standard deviation of least three separate experiments performed in duplicate.

Hemolysis assays: Drugs were dissolved in DMSO or medium (chloroquine) or ethanol (artemisinin) and then diluted with medium to the starting concentration of 200  $\mu$ M. Compounds were placed in 96-well plates (EuroClone) and serial dilutions were made in a final volume of 100  $\mu$ L well<sup>-1</sup>. Fresh RBC were diluted to 1% hematocrit added to the plate and left at 37 °C for 72 h. Hemolysis was evaluated by measuring spectrophotometrically the release of hemoglobin in the supernatants (absorbance at  $\lambda$  = 405 nm, Soret band). The percentages of hemolysis were estimated by referring to a standard curve prepared with serially diluted RBC, lysed with Triton X-100 (4%). **Keywords:** antiprotozoal agents · chloroquine · cytisine derivatives · medicinal chemistry · quinolizidine derivatives

- WHO, World Malaria Report 2014, World Health Organization, Geneva, Switzerland, 2014.
- [2] a) E. A. Ashley, M. Dhorda, R. M. Fairhurst, C. Amaratunga, P. Lim, S. Suon, S. Sreng, J. M. Anderson, S. Mao, B. Sam, et al., *N. Engl. J. Med.* 2014, *371*, 411–423; b) K. M. Tun, M. Imwong, K. M. Lwin, A. A. Win, T. M. Hlaing, T. Hlaing, K. Lin, M. P. Kyaw, K. Plewes, M. A. Faiz, et al., *Lancet Infect. Dis.* 2015, *15*, 415–421.
- [3] a) R. A. Jones, S. S. Panda, C. D. Hall, *Eur. J. Med. Chem.* 2015, *97*, 335–355; b) F. Calderón, D. M. Wilson, F.-J. Gamo, *Progr. Med. Chem.* 2013, *52*, 97–151; c) M. A. Biamonte, J. Wanner, K. G. Le Roch, *Bioorg. Med. Chem. Lett.* 2013, *23*, 2829–2843; d) P. M. O'Neill, V. E. Barton, S. A. Ward, J. Chadwick in *Treatment and Prevention of Malaria* (Eds.: H. M. Staines, S. Krishna), Springer, Basel, 2012, pp. 19–44; e) S. Bawa, S. Kumar, S. Drabu, R. Kumar, *J. Pharm. BioAllied Sci.* 2010, *2*, 64–71; f) T. N. C. Wells, P. L. Alonso, W. E. Gutteridge, *Nat. Rev. Drug Discovery* 2009, *8*, 879–891; g) M. Schlitzer, *ChemMedChem* 2007, *2*, 944–986.
- [4] a) T. J. Egan, R. Hunter, C. H. Kaschula, H. M. Marques, A. Misplon, J. Walden, J. Med. Chem. 2000, 43, 283–291; b) T. J. Egan, Mini-Rev. Med. Chem. 2001, 1, 113–123; c) C. H. Kaschula, T. J. Egan, R. Hunter, N. Basilico, S. Parapini, D. Taramelli, E. Pasini, D. Monti, J. Med. Chem. 2002, 45, 3531–3539.
- [5] a) D. De, F. M. Krogstad, L. D. Byers, D. J. Krogstad, *J. Med. Chem.* 1998, 41, 4918–4926; b) F. Mzayek, H. Deng, F. J. Mather, E. C. Wasilevich, H. Liu, C. M. Hadi, D. H. Chansolme, H. A. Murphy, B. H. Melek, A. N. Tenaglia, D. M. Mushatt, A. W. Dreisbach, J. J. L. Lertora, D. J. Krogstad, *PLoS Clin. Trials* 2007, 2, e6; c) P. A. Stocks, K. J. Raynes, P. G. Bray, B. K. Park, P. M. O'Neill, S. A. Ward, *J. Med. Chem.* 2002, 45, 4975–4983; d) F. E. Saenz, T. Mutka, K. Udenze, A. M. J. Oduola, D. E. Kyle, *Antimicrob. Agents Chemother.* 2012, 56, 4685–4692.
- [6] a) P. M. O'Neill, A. Mukhtar, P. A. Stocks, L. E. Randle, S. Hindley, S. A. Ward, R. C. Storr, J. F. Bickley, J. A. O'Neill, J. L. Maggs, R. H. Hughes, P. A. Winstanley, P. G. Bray, B. K. Park, J. Med. Chem. 2003, 46, 4933–4945; b) P. M. O'Neill, B. K. Park, A. E. Shone, J. H. Maggs, P. Roberts, P. A. Stocks, G. A. Biagini, P. G. Bray, P. Gibbons, N. Berry, et al., J. Med. Chem. 2009, 52, 1408–1415; c) P. M. O'Neill, A. E. Shone, D. Stanford, G. Nixon, E. Asadollahy, B. K. Park, J. L. Maggs, P. Roberts, P. A. Stocks, G. Biagini, et al., J. Med. Chem. 2009, 52, 1828–1844.
- [7] a) C. Biot, G. Glorian, L. A. Maciejewski, J. S. Brocard, O. Domarle, G. Blampain, P. Millet, A. J. Geroges, H. Abessolo, D. Dive, J. Lebibi, *J. Med. Chem.* **1997**, *40*, 3715–3718; b) C. Biot, D. Taramelli, J. Forfar-Bares, L. A. Maciejewski, M. Boyce, G. Nowogrocki, J. S. Brochard, N. Basilico, P. Olliaro, T. J. Egan, *Mol. Pharm.* **2005**, *2*, 185–193.
- [8] a) T. M. E. Davis, T. Y. Hung, I. K. Sim, H. A. Karanajeewa, K. F. Ilett, *Drugs* 2005, 65, 75–87; b) G. M. Keating, *Drugs* 2012, 72, 937–961; c) B. Zani, M. Gathu, S. Donegan, P. L. Olliaro, D. Sinclair, *Cochrane Database Systematic Rev.* 2014, 1, CD010927.
- [9] a) B. Meunier, Acc. Chem. Res. 2008, 41, 69–77; b) F. W. Muregi, A. Ishih, Drug Dev. Res. 2010, 71, 20–32.
- [10] a) O. Dechy-Cabaret, F. Benoit-Vical, A. Robert, B. Meunier, *ChemBio-Chem* 2000, 1, 281–283; b) F. Benoit-Vical, J. Lelievre, A. Berry, C. Deymier, O. Dechy-Cabaret, J. Cazelles, C. Loup, A. Robert, J. F. Magnaval, B. Meunier, *Antimicrob. Agents Chemother.* 2007, *51*, 1463–1472; c) F. Cosledan, L. Fraisse, A. Pellet, F. Guillou, B. Mordmuller, P. G. Kremsner, A. Moreno, D. Mazier, J. P. Maffrand, B. Meunier, *Proc. Natl. Acad. Sci. USA* 2008, *105*, 17579–17584.
- [11] a) S. J. Burgess, A. Selzer, J. X. Kelly, M. J. Smilkstein, M. K. Riscoe, D. H. Peyton, J. Med. Chem. 2006, 49, 5623–5625; b) S. J. Burgess, J. X. Kelly, S. Shomloo, S. Wittlin, R. Brun, K. Liebmann, D. H. Peyton, J. Med. Chem. 2010, 53, 6477–6489; c) D. H. Peyton, Curr. Top. Med. Chem. 2012, 12, 400–407.
- [12] a) G. Bringmann, R. Brun, L. Lehmann, M. Lewis, M. Lödige, H. Moll, A.-K. Mueller, G. Pradel, U. Schurigt (Julius-Maximilians-Universität Würzburg and Ruprecht-Karls-Universität Heidelberg, Germany), Int. PCT Pub. No. WO2012048894 A1, **2012**; b) M. Lödige, L. Hiersch, *Int. J. Med. Chem.* **2015**, 458319.
- [13] N. Vale, R. Moreira, P. Gomes, Eur. J. Med. Chem. 2009, 44, 937-953.



- [14] P. G. Bray, S. Deed, E. Fox, M. Kalkanidis, M. Mungthin, L. W. Deady, L. Tilley, Biochem. Pharmacol. 2005, 70, 1158–1166.
- [15] S. Gemma, G. Campiani, S. Butini, B. P. Joshi, G. Kukreja, S. Sanna Coccone, M. Bernetti, M. Persico, V. Nacci, I. Fiorini, et al., *J. Med. Chem.* 2009, *52*, 502–513.
- [16] V. R. Solomon, W. Haq, K. Srivastava, S. K. Puri, S. B. Katti, J. Med. Chem. 2007, 50, 394–398.
- [17] C. C. Musonda, J. Gut, P. J. Rosenthal, V. Yardley, R. C. Carvalho de Souza, K. Chibale, *Bioorg. Med. Chem.* **2006**, *14*, 5605–5615.
- [18] a) R. Ettari, F. Bova, M. Zappalà, S. Grasso, N. Micale, *Med. Res. Rev.* 2010, 30, 136–167; b) M. Marco, J. M. Coteron, *Curr. Top. Med. Chem.* 2012, *12*, 408–444.
- [19] A. Sparatore, N. Basilico, S. Parapini, S. Romeo, F. Novelli, F. Sparatore, D. Taramelli, *Bioorg. Med. Chem.* 2005, 13, 5338-5345.
- [20] A. Sparatore, N. Basilico, M. Casagrande, S. Parapini, D. Taramelli, R. Brun, S. Wittlin, F. Sparatore, *Bioorg. Med. Chem. Lett.* 2008, 18, 3737–3740.
- [21] M. Casagrande, N. Basilico, S. Parapini, S. Romeo, D. Taramelli, A. Sparatore, *Bioorg. Med. Chem.* 2008, *16*, 6813–6823.
- [22] C. Rusconi, N. Vaiana, M. Casagrande, N. Basilico, S. Parapini, D. Taramelli, S. Romeo, A. Sparatore, *Bioorg. Med. Chem.* 2012, 20, 5980-5985.
- [23] a) L. Lucantoni, A. Sparatore, N. Basilico, S. Parapini, V. Yardey, L. Stewart, A. Habluetzel, L. Pasqualini, F. Esposito, D. Taramelli, 3rd COST B22 Annual Congress "Drug Discovery and Development for Parasitic Diseases", Brussels (Belgium), 1–4 October 2006, Book of Abstracts, p. 137; b) A. Rusconi, M. Casagrande, V. Tazzari, N. Vaiana, Y. Corbett, N. Basilico, L. Cortelezzi, D. Scaccabarozzi, F. Omodeo Salè, S. Romeo, D. Taramelli, A. Sparatore, The Royal College of Physicians "Antimalarial Drugs: Chemistry, Development, and Future Challanges", London (UK), 15–16 March 2011, Book of Abstracts, p. 51.
- [24] F. Omodeo-Salè, L. Cortelezzi, N. Basilico, M. Casagrande, A. Sparatore, D. Taramelli, Antimicrob. Agents Chemother. 2009, 53, 4339–4344.
- [25] J. C. Powers, J. L. Asgian, G. D. Ekici, K. E. James, Chem. Rev. 2002, 102, 4639–4750.
- [26] M. A. Turabekova, V. I. Vinogradova, K. A. Werbovetz, J. Capers, B. F. Rasulev, M. G. Levkovich, S. B. Rakhimov, N. D. Abdullaev, *Chem. Biol. Drug Des.* 2011, *78*, 183–189.
- [27] E. Novacki, S. Wezyk, Rocziniki Nauk Rolniczych 1960, 75B, 385–399; Chem. Abstr. 1961, 55, 9663c.
- [28] R. Simeonova, V. Vitcheva, M. Mitcheva, Interdiscip. Toxicol. 2010, 3, 21– 25.
- [29] C. Rusconi, M. Casagrande, N. Basilico, V. Tazzari, N. Vaiana, L. Vivas, S. Romeo, D. Taramelli, A. Sparatore, Workshop in "Neglected and Orphan Diseases", Siena (Italy), 29 May 1 June 2010, Poster 35.
- [30] B. Tasso, A. Sparatore, F. Sparatore, Farmaco 2003, 58, 669-676.
- [31] F. Novelli, A. Sparatore, B. Tasso, F. Sparatore, *Bioorg. Med. Chem. Lett.* 1999, 9, 3031–3034.
- [32] B. Tasso, R. Budriesi, I. Vazzana, P. Ioan, M. Micucci, F. Novelli, M. Tonelli, A. Sparatore, A. Chiarini, F. Sparatore, J. Med. Chem. 2010, 53, 4668– 4677.

- [33] V. Villa, M. Tonelli, S. Thellung, A. Corsaro, B. Tasso, F. Novelli, C. Canu, A. Pino, K. Chiovitti, D. Paludi, C. Russo, A. Sparatore, A. Aceto, V. Boido, F. Sparatore, T. Florio, *Neurotoxic. Res.* 2011, *19*, 556–574.
- [34] a) B. Tasso, M. Catto, O. Nicolotti, F. Novelli, M. Tonelli, I. Giangreco, L. Pisani, A. Sparatore, V. Boido, A. Carotti, F. Sparatore, *Eur. J. Med. Chem.* 2011, 46, 2170–2184; b) M. Tonelli, M. Catto, B. Tasso, F. Novelli, C. Canu, G. Iusco, L. Pisani, A. De Stradis, N. Denora, A. Sparatore, V. Boido, A. Carotti, F. Sparatore, *ChemMedChem* 2015, *10*, 1040–1053.
- [35] F. Novelli, F. Sparatore, *Farmaco* **1993**, *48*, 1021–1049.
- [36] A. Barteselli, M. Casagrande, N. Basilico, S. Parapini, M. Tonelli, V. Boido, D. Taramelli, F. Sparatore, A. Sparatore, *Bioorg. Med. Chem.* 2015, 23, 55–65.
- [37] M. Rönn, Q. McCubbin, S. Winter, S. W. Veige, N. Grimster, T. Alorati, L. Plamondon, Org. Process Res. Dev. 2007, 11, 241-245.
- [38] J. M. McIntosh, Can. J. Chem. 1980, 58, 2604-2609.
- [39] D. P. Becker, D. L. Flynn, A. E. Moormann, R. Nosal, C. I. Villamil, R. Loeffler, G. W. Gullikson, C. Moummi, D.-C. Yang, *J. Med. Chem.* **2006**, *49*, 1125–1139.
- [40] G. Luputiu, F. Moll, Arch. Pharm. 1973, 306, 414-418.
- [41] a) M. Freund, A. Friedmann, Chem. Ber. 1901, 34, 605–619; b) E. Marrière, J. Rouden, V. Tadino, M.-C. Lasne, Org. Lett. 2000, 2, 1121–1124.
- [42] a) G. Luputiu, F. Moll, Arch. Pharm. 1971, 304, 151–158; b) P. Imming, P. Klaperski, M. T. Stubbs, G. Seitz, D. Gündish, Eur. J. Med. Chem. 2001, 36, 375–388.
- [43] R. C. Elderfield, W. J. Gensler, O. Birstein, F. J. Kreysa, J. T. Maynard, J. Galbreath, J. Am. Chem. Soc. 1946, 68, 1250–1251.
- [44] J. Bolte, C. Demuynck, J. Lhomme, J. Med. Chem. 1977, 20, 106-113.
- [45] B. C. Pérez, C. Teixeira, I. S. Alburquerque, J. Gut, P. J. Rosenthal, J. R. B. Gomes, M. Prudencio, P. J. Gomes, J. Med. Chem. 2013, 56, 556–567.
- [46] T. Singh, J. F. Hoops, J. H. Biel, W. K. Hoya, R. G. Stein, D. R. Cruz, J. Med. Chem. 1971, 14, 532–535.
- [47] A. R. Surrey, J. Am. Chem. Soc. 1948, 70, 2190-2193.
- [48] B. Tasso, PhD Thesis, University of Genoa, Italy, 1999.
- [49] C. Canu Boido, F. Sparatore, Farmaco 1999, 54, 438–451.
- [50] E. Verissimo, N. Berry, P. Gibbons, M. L. S. Cristiano, P. J. Rosenthal, J. Gut, S. A. Ward, P. M. O'Neill, *Bioorg. Med. Chem. Lett.* 2008, 18, 4210–4214.
- [51] P. G. Bray, S. R. Hawley, M. Mungthin, S. A. Ward, Mol. Pharmacol. 1995, 50, 1559-1566.
- [52] R. O. Clinton, C. M. Suter, J. Am. Chem. Soc. 1948, 70, 491-494.
- [53] J. K. Natarajan, J. N. Alumasa, K. Yearick, K. A. Ekoue-Kovi, L. B. Casabianca, A. C. de Dios, C. Wolf, P. D. Roepe, *J. Med. Chem.* **2008**, *51*, 3466– 3479.
- [54] W. Trager, J. B. Jensen, Science 1976, 193, 673-675.
- [55] M. T. Makler, D. J. Hinrichs, Am. J. Trop. Med. Hyg. 1993, 48, 205-210.
- [56] D. Monti, N. Basilico, S. Parapini, E. Pasini, P. Olliaro, D. Taramelli, FEBS Lett. 2002, 522, 3-5.

Received: May 4, 2015 Revised: June 16, 2015 Published online on July 24, 2015